

# Roadside Revegetation

An Integrated Approach to Establishing  
Native Plants and Pollinator Habitat



U.S. Department of Transportation  
**Federal Highway Administration**

## Technical Report Documentation Page

|   |                             |  |                     |
|---|-----------------------------|--|---------------------|
| 1. Report No.<br>FHWA-HEP-23-022  | 2. Government Accession No. | 3. Recipient's Catalog No.   |                     |
| 4. Title and Subtitle<br><i>Roadside Revegetation: An Integrated Approach to Establishing Native Plants and Pollinator Habitat</i>  |                             | 5. Report Date<br>March 2019   |                     |
|   |                             | 6. Performing Organization Code  |                     |
| 7. Contributing Author(s)<br>Amit Armstrong, Ph.D., P.E., Federal Highway Administration<br>Robin Christians, WSP USA<br>Vicky Erickson, United States Forest Service<br>Jennifer Hopwood, Xerces Society<br>Matt Horning, Ph.D., United States Forest Service<br>Andrea Kramer, Ph.D., The Chicago Botanic Garden<br>Lynda Moore, United States Forest Service<br>Deirdre Remley, Federal Highway Administration<br>Shane Roberts, WSP USA<br>Mark Skinner, United States Forest Service<br>David Steinfeld, Native Restoration Consultant<br>Todd Teuscher, WSP USA<br>Abbey White, The Chicago Botanic Garden  |                             | 8. Performing Organization Report No.  |                     |
| 9. Performing Organization Name and Address<br>WSP<br>1600 Broadway Street, Suite 1100, Denver, CO 80202<br>USDA-Forest Service, Pacific Northwest Region<br>1220 SW 3rd Avenue, Portland, Oregon 97204<br>Xerces Society for Invertebrate Conservation<br>628 NE Broadway, Suite 200, Portland, OR 97232   |                             | 10. Work Unit No. (TRAIS)  |                     |
|   |                             | 11. Contract or Grant No.  |                     |
| 12. Sponsoring Agency Name and Address<br>Federal Highway Administration, 1200 New Jersey Avenue, SE<br>Washington, DC 20590  |                             | 13. Type of Report and Period Covered  |                     |
|   |                             | 14. Sponsoring Agency Code   |                     |
| 15. Supplementary Notes<br>Contracting Officer's Representative: Amit Armstrong, Ph.D., P.E., FHWA  |                             |  |                     |
| 16. Abstract<br>Native plants are a foundation of ecological function, affecting soil conservation, wildlife habitat, plant communities, invasive species, and water quality. Establishing locally-adapted, self-sustaining plant communities can also support transportation goals for safety and efficiency. Past obstacles to establishing native plant communities on roadsides have been technical, informational, and organizational. Effective strategies and practical techniques for revegetating the disturbed conditions with limited resources must be made available to practitioners. Multiple disciplines including engineering, soil science, ecology, botany, and wildlife science must work cooperatively and not in isolation. |                             |  |                     |
| Roadsides play an important role in the conservation of declining wild pollinators and in supporting the health of managed pollinators. Throughout the revegetation process, practitioners and designers can enhance roadsides to benefit pollinators. This report offers an integrated approach to facilitate the successful establishment of native plants and pollinator habitats along roadsides and other areas of disturbance associated with road modifications. It guides readers through a comprehensive process of: 1) initiating, 2) planning, 3) implementing, and 4) monitoring a roadside revegetation project with native plants and pollinator habitat.   |                             |  |                     |
| 17. Key Words<br>pollinators, integrated roadside vegetation management, butterflies, bees, moths, nectar plants, host plants, roadside habitat, reduced mowing, native plants, roadside landscape design, roadside maintenance   |                             | 18. Distribution Statement<br>No restriction. This document is available to the public from the sponsoring agency website. |                     |
| 19. Security Classif. (of this report)<br>Unclassified  |                             | 20. Security Classif. (of this page)<br>Unclassified   |                     |
|   |                             | 21. No. of Pages   | 22. Price<br>\$0.00 |

# Roadside Revegetation: An Integrated Approach to Establishing Native Plants and Pollinator Habitat

## CONTRIBUTING AUTHORS

Amit Armstrong, Ph.D., P.E., Federal Highway Administration

Robin Christians, WSP USA

Vicky Erickson, USDA-Forest Service

Jennifer Hopwood, Xerces Society

Matt Horning, Ph.D., USDA-Forest Service

Andrea Kramer, Ph.D., The Chicago Botanic Garden

Lynda Moore, USDA-Forest Service

Deirdre Remley, Federal Highway Administration

Shane Roberts, WSP USA

Mark Skinner, Ph.D., USDA-Forest Service

David Steinfeld, Native Restoration Consultant

Todd Teuscher, WSP USA

Abbey White, The Chicago Botanic Garden

*Cover photo courtesy of Jennifer Hopwood*



U.S. Department of Transportation  
**Federal Highway Administration**

# **Roadside Revegetation**

## **An Integrated Approach to Establishing Native Plants and Pollinator Habitat**

### **Chapter 1: Introduction**

---

- 1.1 An Integrated Approach**
- 1.2 The Ecological Context of Roads**
- 1.3 Objectives of this Report**
- 1.4 Scope**
- 1.5 Approach**
- 1.6 How to Navigate through this Report**

### **Chapter 2: Initiation**

---

- 2.1 Introduction**
- 2.2 Preliminary Tasks of Initiation**
- 2.3 The Process of Road Development**
- 2.4 Road Construction Plans**
- 2.5 Interpreting Engineering Views for Revegetation Planning**
- 2.6 Understanding Technical Concepts and Terminology**

### **Chapter 3: Planning**

---

- 3.1 Introduction**
- 3.2 Defining Revegetation Objectives**
- 3.3 Gathering Pre-field Information**
- 3.4 Defining Revegetation Units**
- 3.5 Identifying Reference Sites**
- 3.6 Gathering Field Information**
- 3.7 Defining the Desired Future Condition**
- 3.8 Identifying Limiting Factors to Plant Establishment**
- 3.9 Identify Factors That Affect Pollinators**
- 3.10 Inventory of Site Resources**
- 3.11 Developing a Vegetation Management Strategy during Project Design**
- 3.12 Select Site Improvement Treatments**
- 3.13 Selecting Plant Species for Propagation**

**3.14 Select Plant Establishment Methods**

**3.15 Develop a Revegetation Plan**

**Chapter 4: Revegation Plan Example**

**Chapter 5: Implementation**

---

**5.1 Introduction**

**5.2 Soil and Site Treatments**

**5.3 Obtaining Plant Materials**

**5.4 Installing Plant Materials**

**5.5 Post-Installation Care of Plant Materials**

**Chapter 6: Monitoring**

---

**6.1 Introduction**

**6.2 Developing a Monitoring Plan**

**6.3 Plant and Soil Monitoring Procedures**

**6.4 Pollinator Monitoring Procedures**

**6.5 Photo Point Monitoring Procedures**

**6.6 Developing a Monitoring Report**

**Chapter 7: Operations & Maintenance**

---

**7.1 Introduction**

**7.2 Decision Process for Treating Unwanted Vegetation**

**7.3 Vegetation Treatment Options**

**7.4 Prevention**

**7.5 Protection**

**Chapter 8: Case Studies**

---

**8.1 I-35 Corridor (aka "The Monarch Highway") Case Studies**

**8.2 Florida resolves to protect wildflowers on roadsides**

**8.3 Mapping and planning benefit Washington State pollinators**

**8.4 Establishing native plants in Arizona**

# 1—Introduction

## **Introduction**

- 1.1 An Integrated Approach**
- 1.2 The Ecological Context of Roads**
- 1.3 Objectives of this Report**
- 1.4 Scope**
- 1.5 Approach**
- 1.6 How to Navigate through this Report**

# INTRODUCTION

The Roadside Revegetation report was written to provide current best practices for planning, designing, and implementing a revegetation project and also includes considerations for creating habitat for pollinators. The report identifies steps and considerations in developing a revegetation project from a variety of perspectives, and presents them in a typical design project order from planning through implementation and maintenance. A diverse writing team of experienced civil engineers, transportation engineers, landscape architects, botanists, geneticists, pollinator conservation specialists, soil science specialists, restoration specialists and environmental protection specialists applied their specific experience and knowledge to this report. Technical aspects of the writing were reviewed by Department of Transportation (DOT) civil engineers and landscape architects and revegetation specialists from several state and federal agencies, who also applied their regional perspective on the planning, design, installation, and maintenance processes.

Users of this report may find it beneficial to review Table 1-1 and the Primer for a quick overview of a revegetation process and an outline of the report chapter contents in order to gain an understanding of the full scope of the report and how it is organized. The Primer also provides an introduction to the online Ecoregional Revegetation Application ([ERA](#)) tool and the [Native Revegetation Resource Library](#). The ERA is an extensive ecoregional plant database that also includes pollinator value information for each plant. The ERA provides a list of all native plants within an ecoregion as well as a list of “workhorse” plant species, the primary beneficial native plants for pollinators and roadside revegetation in a specific ecoregion, and then allows designers to download either form of information in a digital spreadsheet for evaluation and manipulation for their project(s). The [Native Revegetation Resource Library](#) is an online depository that contains copies of applicable revegetation and pollinator educational resource material, referenced from other online sources, compiled in this one location for designer convenience.

This report was written by technical experts for the technical experts involved in the revegetation planning, design, and implementation process. Many report topics include regional and optional considerations and techniques that designers can evaluate for applicability to their project conditions. The writers of this report envision the following ways to use this report:

- **Primary Revegetation Reference**—Designers, revegetation specialists, and contractors can reference the document as they work through revegetation project planning, design, implementation, and monitoring and maintenance.
- **Education Tool**—The report may be used in the classroom and field demonstration setting to educate the next generation of designers, installers, and maintenance professionals.
- **Supplemental Information**—Using the information to revise and supplement existing agency best practices.
- **Planning Resource**—The Table of Contents (TOC) can be used to identify applicable revegetation tasks for specific projects.
- **Scope of Work Development**—For creating a revegetation project scope of work for a Request for Qualifications or a Request for Proposal.
- **Scope, Schedule, and Budget Resource**—The TOC can be used as a template for creating a list of revegetation tasks for an estimate of labor hours, project budget, and project schedule.

- **Project Team Evaluation**—For evaluating the project team strengths and identifying additional sub-consultant expertise that may be needed for the revegetation project.
- **Revegetation Project Agenda Outline**—Using the TOC as a project kick-off meeting agenda to discuss revegetation topics and to start team coordination.
- **Operations and Maintenance Report**—Portions of the information can be used as an outline of the revegetation project operations and maintenance tasks.
- **Project Management Resource**—The client, managers, and team members can identify the revegetation project tasks and then track progress of each task throughout the project duration.

## 1.1 AN INTEGRATED APPROACH

Integrating societal goals for safe, efficient transportation with goals for ecological health is a crucial issue that is receiving increased attention from transportation agencies (Forman et al 2003; National Research Council 2005). Today, most road projects involve modifications to existing roads rather than new construction (National Research Council 2005). As roads are modified or updated section by section, a tremendous opportunity presents itself to remedy the oversights of the past, mitigate environmental impacts, and improve conditions for healthy ecosystems (Figure 1-1).

Native plants are a foundation of ecological function, affecting soil conservation, wildlife and pollinator habitat, plant communities, invasive species, and water quality. Although all of these ecological functions are important, recent emphasis has been placed on supporting pollinators and this facet is highlighted throughout this updated report. Establishing locally adapted, self-sustaining plant communities can also support transportation goals for safety and efficiency. Protecting existing native vegetation during construction and establishing native plants on roadsides following disturbance is key to integrating road systems into natural systems.

Past obstacles to establishing native plant communities on roadsides have been technical, informational, and organizational. Designer and project technical success can be achieved when effective strategies and practical techniques for revegetating the disturbed conditions with limited resources are made available to designers. Project success and efficiency can increase when multiple disciplines, ranging from landscape architecture to engineering, soil science, ecology, botany, genetics, entomology, and wildlife conservation are able to communicate and coordinate as a team early in the project planning, working cooperatively, not in isolation. Finally, improved interagency cooperation and planning processes that consider ecological effects at every step can enhance the success of a revegetation project.

This report offers an integrated approach to facilitate the successful establishment of native plants and pollinator habitat along roadsides and other areas of disturbance associated with road modifications. It guides readers through a comprehensive process of initiating, planning, implementing, maintaining and monitoring a roadside revegetation project with native plants and pollinator habitat.



**Figure 1-1 / Pollinator habitat on roadside**

Roadsides offer an opportunity for improving ecosystems by establishing and maintaining native plant communities.

*Photo credit: Kirk Henderson*

## 1.2 THE ECOLOGICAL CONTEXT OF ROADS

Our road system infrastructure is large, covering millions of acres, with roads under the jurisdiction of Federal land management agencies alone comprising over 17 million acres (Ament, Begley, Powell, Stoy 2014). The road system is widespread and affects all but our most protected lands (National Research Council 2005). The total road corridor (paved road plus roadside or right-of-way) covers over one percent of the nation's land surface, an area equal to the size of South Carolina (Forman and Alexander 1998). If unpaved roads are also included, the percentages increase (FHWA 2008). The ecological effects of roads extend into a zone far beyond the edge of the pavement, with effects including habitat fragmentation, wildlife mortality, noise and chemical pollution, disruption of hydrologic cycles and water quality, increased erosion, and the potential creation of transportation corridors for noxious and invasive weeds that can invade adjacent lands. With these considerations, an estimated 15 to 20 percent of the United States is ecologically affected by roads (Forman and Alexander 1998). The enormous challenge of understanding and mitigating the ecological effects of roads deserves attention and dedication on local, regional, and national scales.

### 1.2.1 PAST OVERSIGHTS

Much of the existing road network in the United States was designed and constructed prior to the 1970s, in an era before ecological health became a widespread concern among American citizens and before ecological science had evolved to address large-scale issues (Forman et al 2003). Safety and efficiency were the primary goals of transportation programs in the past, and the ecological context of roadways were largely overlooked in planning, construction, and maintenance efforts. The effects of roads on natural systems (habitat fragmentation; interruption of natural flows of water; and disturbances to animals, plants and their pollinators, soils, and other resources) were not well understood or considered. Lack of awareness about these factors led to a largely antagonistic perception of the relationships between natural systems and road systems.

For example, without effective revegetation of the road disturbance with desirable plants, undesirable vegetation can encroach on the roadway. Undesirable vegetation can disrupt safety and visibility, leading to expensive and potentially hazardous maintenance measures. Undesirable vegetation can invade and wipe out areas of desirable native plants that pollinators need for food and cover to survive. Conflicts with neighboring land uses could result if corridors for invasive weeds are established or if vegetation control measures are viewed as a health or safety concern by the community. These are all issues that arise when ecology and revegetation science are not considered during road design, construction, or modification. Eventually, poorly integrated or addressed natural processes can threaten the function and structural integrity of the road itself, leading to premature deterioration of the road's infrastructure (Berger 2005).

### 1.2.2 PRESENT AWARENESS

For over 20 years, the ecological effects of roads have been increasingly recognized by the FHWA and by state and county transportation agencies (National Research Council 2005). Today, road effects on ecological processes are major concerns among private citizens, land management agencies, and the transportation community. Consequently, an emphasis has been placed on the integration of ecological

considerations into all phases of road design and construction processes. For example, fish passages have been built to reconnect natural water flows under roads. Other projects have modified roads that were deemed particularly dangerous to endangered species. These roads are being made more permeable to wildlife, greatly reducing losses by improving habitat connectivity, ensuring better visibility for drivers and animals, and creating safer underpasses or overpasses for wildlife (Forman et al 2003). Given the recent decline of pollinator species, there is now a greater emphasis on supporting pollinators by creating habitat on roadsides. A National Pollinator strategy (79 FR 35901) has recently been released, tasking federal agencies to do more with their land holdings, development practices, and maintenance operations to support pollinators in order to reverse pollinator declines ([Section 1.5.4](#)). Efforts to limit inappropriate road expansion and to obliterate unnecessary roads remain important. Where modification and increased capacity are needed, ecological health, safety, and efficient transport are not mutually exclusive goals. Understanding roadside environments, how they interface with adjoining lands, and how to minimize environmental impacts has become a key focus of the Federal Highway Administration. Given political will and proper levels of attention, integration of environmental concerns with transportation can result in significant gains.

### 1.2.3 TRENDS IN ROAD CONSTRUCTION

For the purposes of this report, the primary focus is on Federal, State, and County road corridors, that include highways, interchanges, rural routes, farm to market routes, river roads, and most roads through our National, State, and County parks and preserves. Roads are widespread, fairly permanent fixtures on the landscape and in the culture of the United States. Given current trends, road networks are expected to persist and expand over time. Current modifications predominantly involve updating infrastructure to increase capacity and to improve safety, including widening roads, replacing bridges, and reducing or altering curves and grades to make the road safer for motorists (National Research Council 2005). The opportunity to integrate ecological goals with transportation was largely overlooked when the road networks were originally constructed. However, as the nation's roads are being updated and modified, the opportunity cannot be ignored. While attempts to integrate ecological factors are positive, much of the potential for improved integration is still largely unrealized. This has been due, in part, to a shortage of practical information and the absence of an integrated approach to the challenge. The question is, what can be done to balance societal desires for safe, efficient transport with a healthy environment? In other words, what can be done to help road systems function better with natural systems?

## 1.2.4 CHALLENGES AND OPPORTUNITIES

The fact that the nation's road networks are in varying states of updates, repairs, and maintenance presents an opportunity to improve road systems so that they integrate better with natural systems. Planners and designers strive to understand detrimental effects associated with roads and how to mitigate them by minimizing the ecological footprint of roads and maximizing potential ecological benefits. Many groundbreaking resources have emerged to support these efforts. *Road Ecology: Science and Solutions* (Forman et al 2003) places the challenges into comprehensive frameworks, illuminating goals and principles for an ecological approach to transportation issues. Multiple intervention points are identified to help road systems function better with natural systems, integrating transportation goals for safety and efficiency with approaches to protect water, soil, vegetation, wildlife, and aquatic life.

The FHWA published a landmark book called *Roadside Use of Native Plants* (Harper-Lore and Wilson 2000) that brought the issue of native plant communities on roadsides to the forefront in the transportation community, highlighting the importance of native plants and their broad utility. The National Research Council of the National Academies of Science expanded on the frameworks identified in Forman et al (2003) in its publication, *Assessing and Managing the Ecological Impacts of Paved Roads* (2005). Processes within the FHWA, state DOT, and other agencies are being improved for better integration.

In addition to these advances directly related to roads, many advances in the field of restoration ecology have distilled essential principles applicable to severely degraded sites (e.g., Munshower 1994; SER 2004; Clewell et al 2005; Claassen 2006). In addition, vegetation specialists from a variety of organizations have come to consensus about what truly defines a "native" plant and have developed seed collection, transfer, and propagation principles to ensure that locally adapted materials are used for optimum results (Withrow-Robinson and Johnson 2006, Johnson et al 2010, Basey et al 2015). In both the public and private sectors, seed and plant producers and installers have been developing innovative methods to meet unique site conditions.

Recently, the opportunity to support declining pollinator species through habitat creation on roadsides has been addressed with great success (Hopwood et al 2015, Hopwood et al 2016a, and case studies in this report). Designers have created "pollinator-friendly" roadside habitat, and agency maintenance departments have altered procedures to better maintain these habitats with a focus on the needs of pollinators.

While the publications above assess the best available conceptual and practical information, each also recognizes extensive needs for further work. Central to ecosystem function is native vegetation (SER 2004). However, much of the pertinent information related to protecting and establishing native plants on roadsides has been difficult to put into practice. This report is intended to bridge some of the informational, technical, and organizational gaps to facilitate successful roadside revegetation with native plants. An integrated approach is offered to support both designers and field-based practitioners in successfully revegetating roadsides and obliterated roads with native plant communities (Figure 1-2).



**Figure 1-2 | Recently planted trees on an obliterated section of highway in Oregon**

Most road projects today do not involve building new roads, but rather modifying or obliterating existing roads. This photograph shows an abandoned road where the soils were restored and seedlings planted.

*Photo credit: Lynda Moore, USFS*

## 1.2.5 WHY REVEGETATE ROADSIDES WITH NATIVE PLANTS?

Long-term economic and ecological advantages can be gained by establishing desirable native plant communities on roadsides (Berger 2005). Roadside vegetation can support safety goals by reducing headlight glare, reinforcing the road alignment, protecting view planes and visibility, controlling snow drifts, and reducing wind speeds (Forman et al 2003). Pollinator-friendly plants species, many of which are showy flowering plants, can improve the experience of the road user by creating natural beauty via plant form and color diversity along the roadside, in addition to improving driver performance by reducing monotony and stress. Importantly, creating pollinator habitat along roadsides can directly support imperiled pollinators such as the iconic monarch butterfly (*Danaus plexippus*), economically important managed species such as the European honey bee (*Apis mellifera*), as well as a wide variety of native pollinators including wild bees, butterflies, moths, flies, beetles, and wasps. A self-sustaining native plant community on a roadside stabilizes slopes, protecting water and soil quality. In addition, the establishment of healthy native plant communities is often the best long-term defense against invasive and noxious weeds. Maintenance costs for managing problematic vegetation are reduced, as is the pollution and controversy that sometimes results from roadside herbicide use (Berger 2005). Establishing healthy roadside vegetation can also help sequester carbon dioxide, one of the factors responsible for global climate change (Palumbo et al 2004, Ament et al 2013).

Using native vegetation supports every aspect of the goals identified as best management practices by the transportation community for road design. These include goals to:

- Produce a safe, cost effective, environmentally friendly, and practical road design that is supported by and meets the needs of the users
- Protect water quality and reduce sediment loading into water bodies

- Protect sensitive areas and reduce ecosystem impacts
- Maintain natural channels, natural stream flow, and passage for aquatic organisms
- Minimize ground and drainage channel disturbance
- Control surface water runoff and stabilize the roadbed driving surface
- Control erosion and protect soil
- Implement slope stabilization measures and reduce mass wasting
- Stormproof and extend the useful life of the road (Keller and Sherar 2003)
- Create and maintain pollinator-friendly habitats (Hopwood et al 2015)



Clearly, the goals of safe and efficient transportation and the goals of establishing and protecting native vegetation overlap; when properly integrated, native vegetation supports road objectives. At the same time, considering vegetation as part of road planning processes aids in minimizing and mitigating the ecological footprint of roads during and after

construction. Native plants can provide wildlife habitat and improved connectivity for the length of the road (Forman et al 2003). Understanding vegetation and forage preferences, and careful design that accounts for visibility and safety, can guide animals to safe passageways for travel while minimizing dangerous interactions with vehicles. The presence of birds and small animals can be enhanced when appropriate plant species are established. Processes that work for roadside revegetation are also applicable to the process of obliterating roads where roads are no longer needed.

Despite the potential benefits, many past attempts at roadside revegetation have not succeeded. Although revegetation was considered important, some efforts emphasized seeding of exotic plants; these species were perceived as inexpensive, readily available, and easy to establish on disturbed sites, however, this practice has not been effective or self-sustaining on many projects; either the exotic plants spread to become problematic, or failed to persist because they were not locally appropriate species. Once established, exotics may preclude reintroduction of desirable natives. In other cases, little consideration was given to establishing roadside vegetation during or after construction; if vegetation was considered, it was often as an afterthought. A short-term approach to revegetating roadside disturbances often predominated past efforts, while efforts toward long-term development of native plant communities did not receive adequate consideration. The ineffectiveness of revegetation efforts in the past has resulted in such problems as soil erosion and landslides that affected water quality (Figure 1-3). Visually, unvegetated road disturbances diminish the experience of the road user and economically translate into high costs associated with ongoing maintenance.

Past shortcomings may be attributed to past approaches that were often piecemeal and lacking the cooperation and coordination of disciplines necessary to fully integrate native vegetation into the road planning and construction processes. Revegetation specialists typically worked in isolation from engineers, and sometimes even the biological specialists (soil scientists, botanists, wildlife biologists) failed to

**Figure 1-3 | Unsuccessful roadside revegetation on steep slopes**

Steep slopes are often difficult to revegetate and many past attempts at roadside revegetation did not succeed.

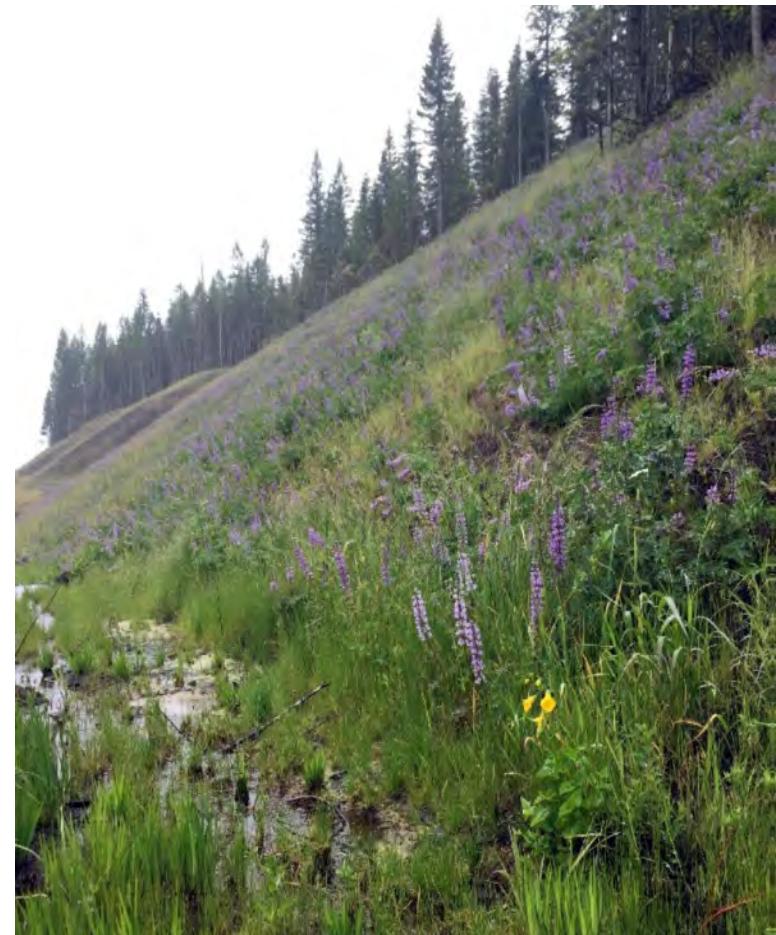
*Photo credit: Lynda Moore, USFS*

coordinate their knowledge and efforts. Success depends on both practical and technical information and a systematic, comprehensive approach.

## 1.3 OBJECTIVES OF THIS REPORT

This report brings theoretical and practical information to bear on the challenge of revegetating roadsides with native plants. Written by and for project designers and field-based practitioners, it synthesizes a comprehensive, holistic approach that can be used to effectively revegetate roadsides and other similarly disturbed areas. Given the unique ecological factors at play on each project, the report is not prescriptive, but rather provides principles and a step-by-step process for designers to use in the field to generate and implement their own locally appropriate, context-sensitive revegetation plan. Examples and proven strategies are offered to serve these goals. Topics covered include how to:

- Improve interagency cooperation in order to think ecologically about road modifications and make revegetation an integral part of road design
- Coordinate information and efforts to bring multiple disciplines such as soil science, genetics, botany, ecology, wildlife science, landscape architecture and engineering together for a holistic approach to revegetation
- Integrate goals for native vegetation establishment with transportation goals for safety, function, and efficiency
- Mitigate harsh, drastically disturbed conditions of road disturbance areas to enable native plants to establish through natural colonization and/or active replanting
- Apply a step-by-step planning, implementation, and monitoring process, including mid-course corrections, to overcome potential pitfalls, resulting in cost-effective, successful establishment of native plants
- Share knowledge, including new revegetation tools and techniques



**Figure 1-4 | Roadside native plant community**

The establishment of native plant communities is the cornerstone of ecological restoration.

*Photo credit: Lynda Moore, USFS*

## 1.4 SCOPE

The complexity of ecologically sensitive road design, implementation, and maintenance can benefit from increasing cooperation from multiple sectors of society and multiple fields of practice and expertise. This report may be of interest not only to field-level practitioners and project designers in both public and private sectors, but also to transportation and planning professionals; land managers; policy-makers; owners and operators of roads on county, state, and federal scales; and concerned citizens. Any agency or organization involved in altering, developing, operating, maintaining, or decommissioning roads will find this publication useful. This report is especially intended to serve field-based practitioners and planners of diverse backgrounds whose goal is to establish locally appropriate, low-maintenance native plant communities on roadsides.

Because integration of multiple sources of expertise is necessary for effective long-term revegetation, this report does not assume that the designer has a particular specialized background but more, a broad level understanding of such disciplines as botany, plant propagation, soil science, genetics, entomology, landscape

architecture, and engineering. The designer may be one of these specialists and may involve one or more of these other specialists during the planning process, depending on the project's complexity. The report states where specific expertise may be appropriate. Key information specifically for designers or contractors and key milestones for communication and integration between engineers and non-engineers are highlighted.

The approach in this report is applicable to any type of road-related project that involves disturbances to soil and vegetation. Revegetation of roadsides adjacent to dirt, gravel, and paved roads would involve similar processes, although there are differences in scale and intensity of efforts. This report applies to new construction or reconstruction and modifications of existing roadways. The principles and practices are also applicable in revegetating other drastically disturbed sites with similar limiting factors to roadsides, such as utility, gas, oil, or powerline rights-of-way and mine reclamation projects.

This report focuses on opportunities for integration during road construction or modification. Long-term maintenance and management of established roadsides is discussed briefly, with references to related management practices such as Integrated Roadside Vegetation Management (IRVM) (Berger 2005). Roadsides that are more permeable to natural flows of water help mitigate the ecological effects of the road. Efforts to improve habitat connectivity and road permeability, as well as storm-water drainage and created wetlands, can be supported by the revegetation practices described in this report. However, specific mitigations for these important topics are beyond the scope of this publication. Also beyond the scope are the myriad other potential ecological and social issues that affect, and are affected by, the engineering and transportation planning processes. Issues of community planning are not addressed. Larger policy-making and planning procedures are also beyond the scope of this report.

## 1.5 APPROACH

The establishment of native plant communities, in order to reinitiate natural processes of succession is a cornerstone of most ecological restoration work (Dorner 2002) (Figure 1-4). Effective revegetation on highly disturbed roadsides aims to initiate or accelerate processes of natural succession following disturbances. Three aspects are generally considered: (1) health (the functional processes of the ecosystem); (2) integrity (species composition and community structure); and (3) sustainability (resistance to disturbance and resilience) (Clewell et al 2005). While restoring plant communities to a pre-disturbance state is not typically a goal on highly disturbed roadsides, each of the above three ecosystem aspects can be improved with appropriate roadside revegetation practices. The establishment of reference sites, or natural models for the desired recovery process, is key to identifying and overcoming limiting factors and accelerating succession by establishing native plants.

Native species play an important role in ecosystem development. If native species can become established on a disturbed site, the processes of succession, including soil recovery and nutrient cycling, are initiated (Brown and Amacher 1999). In most cases, native plants are established on roadsides through seeding or planting, although sometimes passive revegetation (natural colonization) is possible where native seed banks are nearby and limiting factors are mitigated.

## 1.5.1 SUSTAINABLE REVEGETATION ON ROADSIDES

Sustainable revegetation projects attempt to integrate disturbed sites with the surrounding, non-disturbed landscape. An ideal integration is both visual and functional in nature, and the blending of the two is often considered an art.

Revegetation efforts are most likely to be sustainable if they are process based in approach, rather than form based. Process-based projects aim to create healthy and resilient ecosystems with the necessary components to develop multiple natural processes. Many desired natural processes take hundreds of years to fully develop, so it is unreasonable to think a designer can anticipate, replicate, and accelerate their development over the course of a project that might span three, five, or ten years. The objective, rather, is to install as many components of as many processes as possible, to facilitate their development. In this sense the designer is often most interested in the trajectory of the developing processes, rather than their presence in complete form.

The purpose of this section is not to discuss every possible contributor to resiliency or process building. Rather, it is to provide the designer with information that might assist in the recognition of actions they can take to potentially increase resiliency and accelerate the development of natural processes.

### Supporting Resiliency

Resiliency is a system's ability to recover quickly, and hopefully entirely, from disturbance. The use of genetically appropriate plants, collected from throughout the project and surrounding area within the provisional seed zone(s), can encourage genetic health and resilience of the restoration plant population as well as those populations surrounding the project area. Restoration plants, installed at sufficient spacing, can increase competition and facilitate the site's resistance to invasion by non-native or weedy plant species, thereby making it more resilient to plant compositional changes when they do occur. Plant species and form diversity creates redundancies in life form, functional grouping, and services; all of which increase resiliency.

### Consideration of Natural Processes

Multiple natural processes have components for which the designer can assist when re-integrating a disturbed site into the surrounding landscape. The designer may want to identify and initiate the enhancement or construction of those processes that already naturally occur, or would be expected to occur, on the surrounding landscape. There would be little point in allocating resources to try to develop a wetland in a sage steppe of a high desert upon which no wetlands occur, for example.

Soil forming processes can be considered during a number of project activities such as decommissioning roads, constructing cut and fill slopes, or when shaping the final grade of spoils areas. Knowing that wind and water erosional forces shape and sculpt land formations, attention to the contours of the surrounding landscape can inform designers and equipment operators of how best to blend the target area with its environment. By doing so the newly formed contours not only visually re-integrate the project site into the surrounding setting, but also prevents out-of-proportion, angular, or inappropriate landforms from interrupting wind and water patterns. In addition, the accumulation of plant detritus is a basic component of soil formation. It is often useful to de-compact the soil and leave the site in a roughened, irregular, and undulating condition. The resulting hummocks and low spots can seem small and inconsequential, but they provide pockets for soil to accumulate and will eventually



**Figure 1-5 | Roadsides are disturbed areas**

Disturbed roadside areas result from road construction, modification, or maintenance challenges.

*Photo credit: Lynda Moore, USFS*

support plant life. Avoiding compaction of the soils frequently provides for better water infiltration and drainage, reduced erosion, and overall stability of soil particles.

The use of annual and perennial plants in revegetation projects contributes plant litter to the system throughout a prolonged period of time compared to a site that only uses one or the other. This litter then becomes reduced and incorporated into disturbed sites, facilitating the nutrient cycling. Including various plant life forms in a revegetation plan, including shrubs and trees where appropriate, provides refuge, breeding-, feeding-, rearing-grounds, as well as domicile habitat for insects, birds, and various small and large animals. The abundance and diversity of organisms supported on the revegetation site contribute greatly to the development of a functioning nutrient cycle. In addition to facilitating nutrient cycling, the abundance and diversity of organisms supported also play critical roles in the dispersal of seeds and spores.

## Considering Maintenance Operations and Costs

Sustainability from the maintenance stand point is a component that is easy to overlook, when focused on the biology of a project. Roadside maintenance programs are designed to insure the safety of travelers and to protect the integrity of the road. If the life forms planted are inappropriate for the roadside, if setbacks or roadside zones are ignored, if the structural integrity of the selected plants is not sound, or if the restoration plantings create visibility and safety issues, then the revegetation project will not be sustainable. Maintenance operations and costs are components to be considered in all phases of a revegetation project as its success and sustainability is partially dependent on understanding the needs of roadside maintenance programs.

### 1.5.2 WHAT IS THE “ROADSIDE”?

In this report, the term “roadside” refers to any area of disturbance associated with road construction, reconstruction, waste areas, source pits, and maintenance, (Figure 1-5). The roadside includes the sides of the road corridor beyond the paved road (shoulders and verges), including impacted or maintained roadside areas within the right-of-way. The roadside area is sometimes narrow, but sometimes extends several hundred feet or more beyond the edge of the road surface, depending on the project. In some situations, revegetation efforts may encompass areas beyond the right-of-way that are affected by or affect the road. The area where the revegetation specialists will focus their efforts is usually dependent on two factors: ownership of the right-of-way and surrounding lands, and areas of disturbance (construction footprint). Most roadsides are drastically disturbed environments, where soil may be severely compacted and consist of a mixture of subsoil and parent material (Figure 1-6). Beneficial microorganisms, nutrients, and organic matter necessary to sustain plant growth may be absent or severely depleted. Often, slopes can be very steep and inaccessible, exposed to the erosive effects of wind and water. These environments represent a revegetation challenge of high intensity and magnitude.



**Figure 1-6 / Roadsides can be highly disturbed**

Roadsides are often drastically disturbed and infertile environments with no topsoil, severe compaction, and a lack of beneficial microorganisms.

*Photo credit: Matt Horning, USFS*

### 1.5.3 WHAT ARE NATIVE PLANTS?

“Native plants,” as defined in this report, are locally adapted, genetically appropriate native plant materials (Withrow-Robinson and Johnson 2006). These plants are best suited evolutionarily to the local conditions, and generally need less maintenance and persist longer than non-local species. When properly established, they form

plant communities with the potential to be self-sustaining and self-perpetuating over time, requiring little or no input from humans to persist. Native plants also support more robust communities of pollinators, birds, and other small wildlife.

#### **Inset 1-1 / Federal Lands Policies on Native Plants for Revegetation**

Many land management agencies have policies on the use of native plants. For example, the USDA Forest Service has the following policies in place applicable to road projects on Forest Service lands:

- **National Native Plant Materials Policy:** "Native plant materials are the first choice in revegetation..." (USFS 2005)
- **Burned Area Emergency Response Manual (FSM 2523):** "...when practical, use genetically local sources of native species..." (USFS 2003)
- **R6 Revegetation Policy:** "...use local native plants to the extent practicable..." (USFS 2004)
- **Vegetation Ecology (FSM 2070):** "...promote the use of native plant materials in revegetation for restoration and rehabilitation...native plant materials are the first choice in revegetation for restoration and rehabilitation efforts." (USFS 2008)
- **National Seed Strategy for Rehabilitation and Restoration:** "...ensure the availability of genetically appropriate seed..." (Federal Interagency Plant Conservation Alliance 2015)

Challenges to establishing native plants on roadsides are significant, partially due to difficulties in obtaining appropriate materials. However, the technological capacity of native plant propagation and outplanting efforts in both private and public sectors has increased significantly in the past two decades. Innovative stocktypes and application methods have made roadside revegetation more effective. Federal agencies that manage roads increasingly use native plant materials as the first choice in revegetation efforts, thereby making roadside revegetation an important and expanding frontier for native plant suppliers.

#### **1.5.4 WHY THE EMPHASIS ON POLLINATORS?**

An estimated 85 percent of the world's flowering plants depend on animals for pollination (Ollerton et al 2011). Animal pollinators visit flowering plants seeking floral resources and, in the process, incidentally transfer pollen from anthers (male reproductive structures of the flower) to stigmas (female reproductive structures), vectoring fertilization and allowing these flowering plants to reproduce. Pollinators rely on flowering plants for food, requiring nectar (a sugar-rich liquid) and sometimes pollen itself (a source of protein) as sources of energy and nutrition. Most animal pollinators are insects, such as honey bees, bumble bees, flies, and butterflies. However, vertebrates such as birds (e.g., hummingbirds), mammals (e.g., bats, rodents) and some reptiles pollinate certain plant species. The pollination services provided by pollinators are essential to the health and persistence of the plant species that depend on them, and for the wildlife and pollinators that in turn depends upon the plants for food, nesting, and shelter.

Pollinators are also important to human health and the global economy. More than 75 percent of the world's 115 principal cultivated crops are reliant on animal pollinators or benefit from animal pollination. Roughly 35 percent of global crop production is dependent on pollination by animals (Klein et al 2007). Insect-pollinated forage plants such as alfalfa and clover also provide feed for livestock. Pollinator-dependent food

crops, namely fruits, vegetables, and nuts, make up a critical component of our diet (McGregor 1976). The majority of minerals, vitamins, and nutrients needed to maintain human health (such as vitamin C, lycopene, calcium, and folic acid) come from crop plants that depend partially or fully on pollinators (Eilers et al 2011). Consequently, pollinators provide essential agricultural services with a high economic value. A recent worldwide estimate suggests pollinators in general contribute 9.5 percent (\$216 billion per year) to crop value (Gallai et al 2009). In the United States, it is estimated that pollinators (both managed and wild) contribute \$29 million in farm income annually (Calderone 2012). Importantly, native pollinators specifically have been shown to contribute approximately \$1 billion to the California crop economy (Chaplin-Kramer et al 2011).

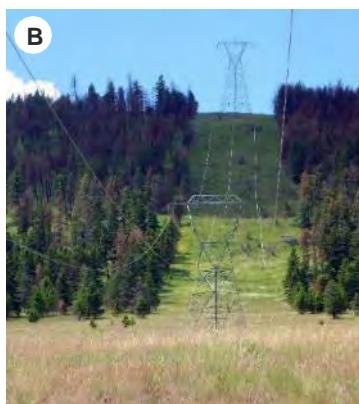
Globally, pollinators are in decline (National Research Council 2007; Potts et al. 2010). In the United States, wild pollinators such as monarch butterflies and many bumble bee species, as well as colonies of managed honey bees, are experiencing declines due to a loss of habitat, the spread of disease, overuse of pesticides, and various other factors (National Research Council 2007; Hatfield et al 2015; Jepsen et al 2015). Pollinator declines threaten the viability of agricultural productivity and the health of natural ecosystems.

With at least 17 million acres of roadsides in the United States, roadside vegetation can serve as much needed habitat for pollinators, offering food, breeding, or nesting opportunities and connectivity that can aid pollinator dispersal (Hopwood et al 2015). Roadsides can support a diversity of generalist pollinators, including bumble bees, honey bees, butterflies, and hummingbirds as well as rare or federally listed species. Roadsides sustain plants that are sources of pollen and nectar for adult pollinators as well as host plants for the caterpillars of butterflies and moths. The availability of floral resources influences the abundance and diversity of butterflies and bees found on roadsides (Saarinen et al 2005; Hopwood 2008). Pollinators on roadsides benefit in particular from native plants (Ries et al 2001; Hopwood 2008). Roadsides planted with native plants also can provide pollinators with shelter, sites for nesting or egg-laying, and overwintering habitat. Pollinators have complex life cycles, with different needs at different stages of their lives. Roadsides can provide resources for a portion of the life cycle of some species, while providing resources needed for the entire life cycle of other species.

**Figure 1-7 | Vegetated linear corridors as habitat for pollinators and other wildlife**

Linear corridors such as irrigation canals (A), utility rights-of-way (B), roadways (C), and river ways and riparian areas (D), present opportunities to reconnect fragmented pollinator habitats.

Photocredits: (A) Chris Jensen, USFS; (B) Justin Moffett, BPA; (C) Lynda Moore, USFS; (D) Lynda Moore, USFS



Evidence also suggests that the linear shape and connectivity of roadsides may help pollinators to move through landscapes in search of food or in pursuit of new habitat (Ries et al 2001; Dirid and Cryan 1991). Roadsides extend through all landscapes and can be particularly important sources of habitat for pollinators in highly altered landscapes such as intensely managed agricultural lands (Figure 1-7).

Not all roadsides are equally beneficial to pollinators. Roadsides that are intensively mown, blanket-sprayed with herbicides, or planted with introduced grasses support far fewer species of pollinators and smaller population densities than roadsides managed for native plants (Smallidge and Leopold 1997; Johst et al 2006; Reis et al 2001). Roadside vegetation management influences how pollinators use roadsides, and even influences the number of pollinators killed by vehicles. For example, butterfly vehicle mortality rates increase with more frequent mowing and decrease with high plant diversity in roadside vegetation (Skorka et al 2013) (Figure 1-8).

Roadsides play an important role in the conservation of declining wild pollinators and in supporting the health of managed pollinators. Throughout the revegetation process, practitioners and designers can enhance roadsides to benefit pollinators.

### 1.5.5 GOAL-ORIENTED, CONTEXT-SENSITIVE, AND INTEGRATED

The overall approach in every aspect of this report is goal-oriented, context-sensitive, and integrated (Clark et al 2001). The goals of establishing and protecting native plant communities are considered along with transportation goals, including safety, efficiency, and cost-effectiveness for the life of the road. This is not an idealistic approach; while recognizing that resources are limited and conditions are degraded, the approach is technically and economically feasible while still enabling the integration of roads with ecological processes.

Sensitivity and appropriateness to the local context are essential parts of successful revegetation. This report is intended to facilitate the process of developing locally appropriate, context-sensitive principles on a project-by-project basis, integrating top-down and ground-up information to meet the specific challenges at hand. For this reason, the report does not provide cookbook-type “recipes” or specific prescriptions. For example, no “one-size-fits-all” seed mix exists for roadside revegetation. The process and tools needed to arrive at context-sensitive solutions are not difficult to apply; by following the steps outlined in this report, practitioners will be able to generate the information they need.



**Figure 1-8 / Roadside mowing**

Although mowing a clear zone directly adjacent to the pavement poses little harm to pollinators, frequent roadside mowing of the entire roadside can decrease the densities of pollinators.

*Photo credit: Idaho Transportation Department*

## 1.6 HOW TO NAVIGATE THROUGH THIS REPORT

Navigating through this report can be done in several ways:

- **Search Field**—The easiest way to find a topic in this report is to use the “Search” field. If the topic is very specific, this may be the quickest method. However, if it is a broad topic, discussed throughout the report, this method may not easily narrow down the location in the report. When this is the case, referring to the table of contents and/or using search tools (for digital formats)

may be the better approach.

- **Table of Contents**—The table of content is a quick way of getting to the main sections of the report through hyperlinks.
- **Revegetation in 15 Steps**—For those who wish to see the revegetation process in a step by step approach may want to use Table 1-1 to navigate through the report. Case studies of completed revegetation projects that create habitat for pollinators are provided in [Chapter 8](#).

**Table 1-1 | Revegetation in 15 steps**

| Revegetation Process Steps              |           | Further defined here   |
|---|-----------|--|
| <b>INITIATION PHASE</b>                 | <b>1</b>  | Due diligence, plan development                                |
|   | <b>2</b>  | Objectives and desired future condition                        |
|   | <b>3</b>  | Pre-field information  |
|   | <b>4</b>  | Revegetation units and reference sites                         |
|   | <b>5</b>  | Field information  |
|   | <b>6</b>  | Limiting factors to plant establishment and pollinator habitat |
| <b>PLANNING PHASE</b>                   | <b>7</b>  | Site resources   |
|   | <b>8</b>  | Maintenance strategy   |
|   | <b>9</b>  | Site improvement treatments                                    |
|   | <b>10</b> | Plant species  |
|   | <b>11</b> | Plant establishment methods                                    |
|   | <b>12</b> | Revegetation plan  |
| <b>IMPLEMENTATION PHASE</b>             | <b>13</b> | Implementation   |
| <b>MONITORING AND MAINTENANCE PHASE</b> | <b>14</b> | Monitoring   |
|   | <b>15</b> | Operations and maintenance                                     |

# **2—Initiation**

- 2.1 Introduction**
- 2.2 Preliminary Tasks of Initiation**
- 2.3 The Process of Road Development**
- 2.4 Road Construction Plans**
- 2.5 Interpreting Engineering Views for Revegetation Planning**
- 2.6 Understanding Technical Concepts and Terminology**

## 2.1 INTRODUCTION

Incorporating ecological concepts into all aspects of road design, construction, modification and maintenance is a goal of the transportation community (National Research Council 2005; Forman et al 2003). The Federal Highway Administration (FHWA), state DOTs, and other federal, state, and county agencies that are responsible for road infrastructure all strive to achieve this goal. One successful approach for meeting this goal is to integrate issues of native plant revegetation (including protection of existing vegetation) into the larger design and construction processes of road projects. Revegetation planning is now an integral part of road planning and is an important aspect of road projects that can achieve a higher level of success and project benefits when incorporated early in project design. Experts recommend (as illustrated on the timeline Figure 2-1) that the implementation phase of revegetation begins while the overall project development process is still underway. Waiting until construction begins reduces the likelihood that locally-adapted native plant materials in the quantities needed will be able to be propagated in advance.

To increase the opportunity for successful integration of revegetation issues within the overall road project, the designer of a revegetation plan can identify the cooperators and agencies involved, discover how their processes and timelines work, and coordinate at the appropriate times and with the appropriate people. The revegetation plan designer can add greater project value if involved in planning and construction processes whenever soil and vegetation disturbances are planned. Agency schedules, milestones, and budgetary issues are commonly defined in the planning process to effectively synchronize the revegetation efforts with road development and construction.

Road projects may be administered from local, state, or federal levels, or sometimes from a combination of all three levels. In terms of timing, road projects can be complex and span many years, whereas other projects are streamlined and on a compressed timeline. It is beyond the scope of this manual to cover all the specific procedures and processes for every agency involved in road projects. This chapter, however, provides an overview to successfully navigate the various processes for a project.

Designer involvement and input is important from the inception of a project through completion. The earlier one can get involved, the more input provided. The preliminary steps for initial involvement include:

- Defining cooperators, processes, timelines, and milestones.
- Defining objectives: What is the project trying to accomplish?

This chapter provides an overview of each of these steps, followed by a discussion of typical road development processes that includes key points of involvement. This chapter also discusses the technical content, interpretation, and use of road project plans and views.

## 2.2 PRELIMINARY TASKS OF INITIATION

Roadside revegetation is a complex process, frequently involving numerous agencies and individuals. Appointing a single designer to coordinate the planning, implementing, and monitoring/adaptive management of the revegetation aspects of the road project can help streamline revegetation coordination. Typically, the designer will be the responsible professional landscape architect or civil engineer who is in charge of sealing the revegetation documents and is directly involved with the design and supervision of others who are assisting in the preparation of the design documents. Depending on the training and expertise of the designer, the project scale, level of environmental impacts of the project, and level of political and public scrutiny, the designer and owner of the project are often best served by enlisting experts from other natural resource disciplines to help with the revegetation planning so that expertise in botany, plant genetics, horticultural practices, landscape architecture, soil science, engineering, hydrology, wildlife biology (including pollinator specialists), geology, and ecology is available for the project as necessary. Project quality and efficiency can be enhanced when the designer is the coordinator of the technical and organizational aspects of the revegetation project, as well as the

### *For the Designer*

**It is beyond the scope of this manual to cover all the specific procedures and processes for every agency involved in road projects. However, this chapter provides a general overview.**

### *For the Designer*

**Incorporation of revegetation planning very early on in road project development can benefit project coordination, schedule, and budget.**

contact between revegetation efforts and the other aspects of road planning and construction.

### 2.2.1 DEFINING COOPERATORS PROCESSES, TIMELINES, AND MILESTONES

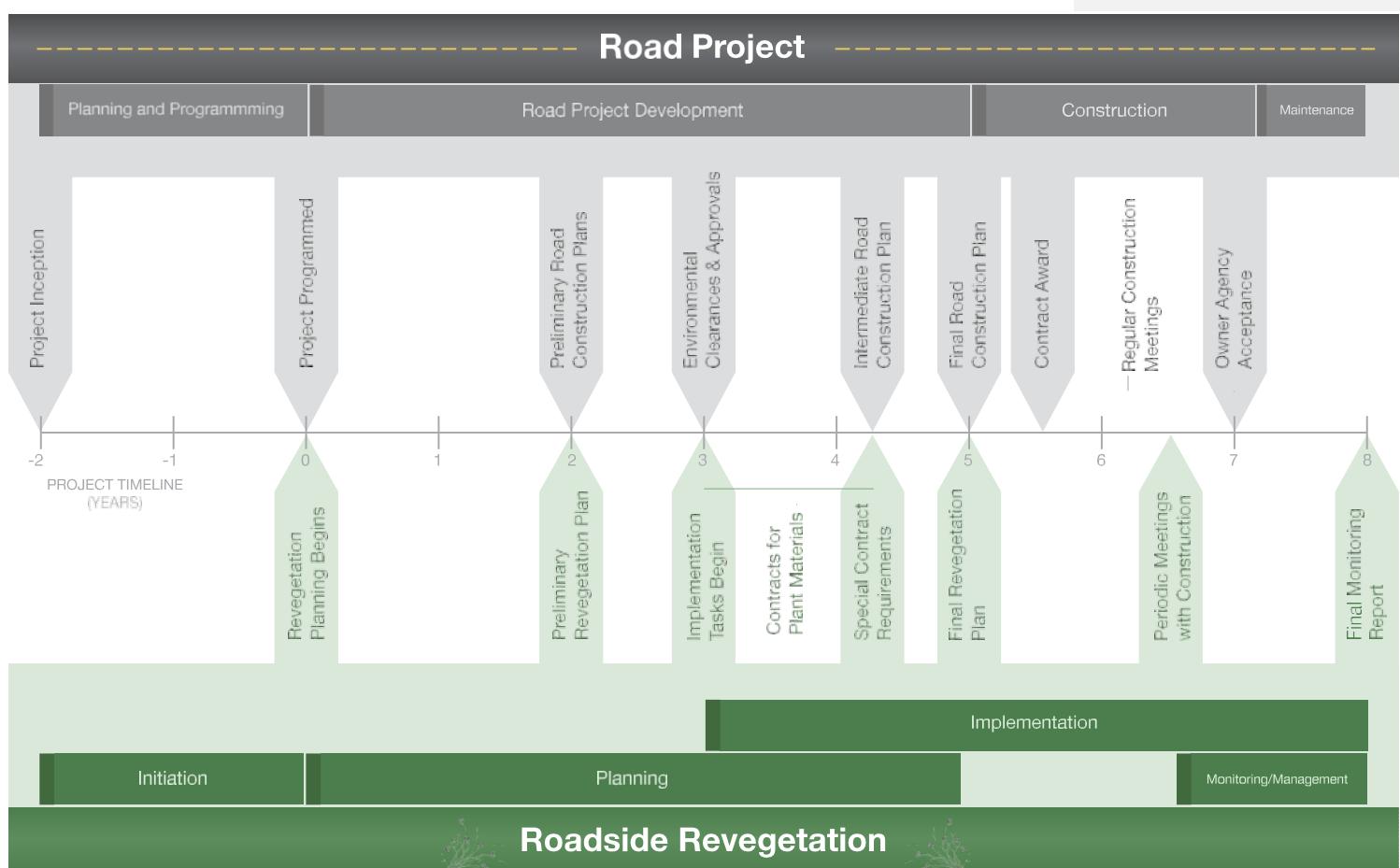
Designer due diligence early in the project planning process includes identification of the reviewing agencies and individuals involved in the road construction project, along with their respective roles and responsibilities. It is especially important to understand: (1) who the actual decision-makers are, (2) who the land owning agency is, (3) who maintains the road and roadsides, (4) who will be carrying out the road construction project, and (5) who is funding the project. Sometimes, the actual decision-makers are not the same people who attend the design meetings. It can be important for the designer and design team to confirm the agency organizational dynamics and to get key design direction approvals in writing at the appropriate times in the planning and design process.

An understanding that the timing, responsibilities, and, most importantly, the plan review and approval processes associated with each agency will vary, will allow the designer to plan, communicate, and interact more effectively with the right people at the right time. While this may seem complicated, many agencies have a procedural manual that describes how a project is carried out from conception to completion, defining the timelines, milestones, roles and responsibilities, terminology, and how funding works. Most current documents are online, however, some may only be available in hardcopy upon request. The designer may want to confirm the location of current documents with the reviewing agencies. Location and use of these documents and agency manuals to help create a project schedule can be a key due diligence item for the designer. Initial meetings with owners, maintainers, and agency plan reviewers are also beneficial for the designer, as they can create relationships that strengthen the lines of communication during the project, and are an ideal time to clarify project requirements, expectations, and various cooperator processes.

Each agency has certain approvals and procedural activities, including some that involve fulfilling environmental regulations. Early designer due diligence may include defining these activities and

**Figure 2-1 / Project coordination timeline example**

Coordinating revegetation with the larger processes of road construction is helpful. While the timelines and agencies involved will vary, this figure illustrates some of the key opportunities for communication and integration.



determining how revegetation work fits within them. The steps in the approval process are often important milestones for the agency, and they expect the designer to have that understanding and to provide input at appropriate times. Defining appropriate roles can help the designer to coordinate with the proper people, follow protocols, and avoid duplicating efforts.

Many variables affect the overall timelines from inception to construction. Timelines vary depending on the complexity of the project, the amount of controversy involved, and the availability of funds. Some projects take less than two years, while some can take over ten years. Reviewing Figure 2-1 with the assigned road project engineer and discussing milestones, timelines, procedures, budgets, and roles can be an effective approach to getting oriented to the complex process of road development.

## 2.2.2 DEFINING OBJECTIVES: WHAT IS THE PROJECT TRYING TO ACCOMPLISH?

Once the agencies and processes for each phase of the project are clarified, the designer can begin to understand how their work relates to the overall objectives of the project. Objectives can be found in the programming documents that originally identified the need for the project. These objectives often center on improving safety or updating the road infrastructure. Phase One of the planning process ([Chapter 3](#)) describes how to identify the objectives of the road project and translate them into specific goals for revegetation.

Environmental protection, pollinator habitat creation, and maximizing the ability of the roadside to regenerate native vegetation are primary revegetation goals. When a revegetation designer is involved early in a project, when disturbances to soil and vegetation are planned, the designer can be a key link to understanding the potential disturbances that might be caused and how to best minimize or mitigate them. The designer, with specialists' input, can help the roadway engineer understand what types of disturbances can be feasibly revegetated with native plants. If a disturbance to soil and vegetation will not allow for revegetation, alternatives to that type of disturbance can be considered. The input of specialists can be crucial for determining potential strategies and alternatives. The project objectives also help determine the types of native vegetation that are most appropriate for the work. Revegetation design solutions can vary widely depending if the project crosses a wildlife corridor, is a scenic drive, is in an ecologically sensitive area with more intensive recovery needed, travels through open farm land, or contains steep slopes.

Safety, efficiency, protecting and enhancing environmental health, and creating habitat for pollinators are all important priorities in road projects. While safety concerns may at times limit what is appropriate in roadside revegetation (e.g., tall trees along a roadside may be a desirable choice from an environmental and aesthetic standpoint but not from a safety or visibility standpoint), experienced design professionals recommend that these concerns not be viewed as an impediment to successfully revegetating roadsides. Some experts recommend that the designer coordinate early with the roadway engineer to gain a full understanding of safety issues, particularly regarding visibility and the ability of drivers to recover if they drive off the road and into the roadside area (see discussion of how to define roadside revegetation zones in [Chapter 3](#)).

## 2.3 THE PROCESS OF ROAD DEVELOPMENT

While each cooperating agency will divide up and define tasks differently, the process of road development generally has four stages: (1) planning and programming, (2) project development, (3) construction, and (4) maintenance. For revegetation work, the implementation phase often begins well before road construction is initiated (with the collection of plant materials for propagation).

Revegetation efforts also continue after road construction is completed. Figure 2-1 compares the revegetation process with the overall road development process, showing process steps where interface is crucial.

During road project development, a number of meetings can take place involving representatives from the agencies and interests involved with the project. Experts recommend setting revegetation and wildlife

objectives, such as developing pollinator habitat, accommodating wildlife corridor crossings, and planning stormwater features as habitat enhancement, early in the planning and programming phase, as they each have project safety, schedule, and budget implications. During the planning phase, meetings usually take place at the preliminary, intermediate, and final stages of the road plan. It is recommended that the designer attend these meetings. This ensures that good information gets into the project budget and schedule, that quality communication takes place, and that trust is built during the road planning process.

Meetings also offer the opportunity for reminders and confirmation that designers are meeting any requirements they face and that proper channels are utilized to get the job done. Regular communication between designers and quality control plan review at similar preliminary, pre-final, and final milestones can lead to developing a complete set of construction documents that can be easily interpreted and tightly bid. During the construction phase, the construction manager, design engineer, and other key players carrying out the project typically meet on a weekly basis. Attending some of these meetings can be valuable both for learning and contributing input as the project progresses, and for interacting with contractors, inspectors, and other stakeholders who may also be at the meetings or field site. Key contacts (such as the construction manager or design engineer) can help clarify the most appropriate meetings to attend, as well as the channels for communicating with other individuals who are involved with the project.

### ***For the Designer***

**Providing wildlife crossings under roadways can increase driver safety. Planning and budgeting of wildlife under-crossings is often overlooked.**

#### **2.3.1 ROAD PLANNING AND PROGRAMMING**

The process of deciding when to modify or build a section of road is often lengthy. Transportation planners identify and prioritize functional, structural, and safety issues regarding roads. If an issue is becoming problematic, alternatives to address it will be considered (FHWA 2005). The negative effects of transportation infrastructure and rights-of-way on communities and the environment are well documented. New road alignments or major road widenings are often controversial and often involve extensive study of functional, cultural, environmental, and aesthetic issues.

Context Sensitive Solutions (CSS) are design solutions, often developed through a public engagement process, that identify and address site-specific effects in an attempt to physically and visually connect transportation facilities into communities and their surrounding contexts. Depending on the project scope and context, solutions often prescribe bridge and wall structure aesthetic design, lighting and signage styles, bicycle, pedestrian and wildlife crossing accommodations, development of stormwater facilities as wildlife habitat, and providing appropriate types and amounts of vegetation. Each solution has function, safety, schedule, and cost implications for a project and are typically most successful when addressed early in the planning and programming of a road project. The revegetation design professional is often well-versed in Context Sensitive Solutions and can benefit the road project if their input is included in the early planning and programming phase.

The FHWA, several State DOTs, and several States have adopted policies to provide sustainable highway design, CSS studies, and incorporation of CSS solutions into their transportation projects. As examples, the framework to provide Sustainable and Context Sensitive Solutions is found in the State law and policy for both the Illinois DOT and the Indiana DOT. In the State of Illinois, Statute 605 ILCS 5/4-219 Context Sensitivity, states: "It is the intent of the General Assembly to ensure that Department of Transportation projects adequately meet the State's transportation needs, exist in harmony with their surroundings, and add lasting value to the communities they serve." Statute 605 ILCS 5/4-219 also states that "A hallmark [of the context sensitive design process is to include]... early and on-going collaboration with affected citizens, elected officials, interest groups, and other stakeholders to ensure that the values and needs of the affected communities are identified and carefully considered in the development of transportation projects." Further, the Illinois CSS process and design promotes "the exploration of innovative solutions, commensurate with the scope of each project that can effectively balance safety, mobility, community and environmental objectives in a manner that will enhance the relationship of the transportation facility with its setting" (State of Illinois General Assembly-a, 2013).

Similarly, the Indiana “Procedural Manual for Preparing Environmental Documents” (State of Indiana, 2008), includes section II.B.3.f, Context Sensitive Solutions (CSS), which states that “INDOT’s goal is to incorporate CSS into development, construction, and maintenance processes for improvements to the state jurisdictional transportation system.” The manual also states that “CSS promotes the following key principles:

- Use a full range of communication methods early and often, to effectively engage stakeholders and the public.
- Use interdisciplinary teams.
- Seek consensus on purpose and need.
- Document, track, and address all commitments.
- Use all resources effectively in the decision making process.
- Allow for design flexibility while considering a safe facility for all modes

While the CSS process works to identify both broad and detailed impacts of a project and proposes appropriate mitigation and enhancements, the process also aims to accomplish the prime goal of the project and be sustainable over the long term. Sustainable design in the design-build environment seeks to balance environmental, functional and financial needs and impacts. All can be accomplished through thoughtful and efficient design that seeks to do no harm, minimize its footprint, and strives to incorporate dynamic functional solutions. Ideal sustainable design solutions often accomplish their intended function, are aesthetically pleasing, endure and improve over time, and reduce future costs and impacts.

Many DOTs have committed to utilize FHWA’s Infrastructure Voluntary Evaluation Sustainability Tool (INVEST). This tool facilitates the development and tracking of sustainability measures throughout a project’s life, including overall planning, project development, and operations and maintenance. A project is measured for “triple bottom line” Social, Environmental, and Economic accomplishments with the INVEST scorecard; for example, the Rural Extended Project Development module lists 25 weighted items on which the project can score. A “Platinum” rating is achieved when 60 percent or more of the possible sustainability rating points are achieved. More information on FHWA’s INVEST tool is available at the website [www.sustainablehighways.org](http://www.sustainablehighways.org).

Design studies and tools such as CSS and INVEST attempt to create projects and budgets that address the “triple bottom line” for the benefit of the public and the environment. With or without these tools, wildlife corridor crossings and bicycle/pedestrian connections are two issue areas that are often overlooked when roadway budgets are initially set. The accommodations for wildlife corridors and bicycle/pedestrian connections through transportation routes are detailed functional, structural, and safety issue items, with significant project cost implications. Wildlife crossings can require taller bridge heights, longer spans, and may be needed at multiple bridges to accommodate safe movement of large animals across the right-of-way. Bicycle/ pedestrian connections can require wider bridges for bicycle lanes and sidewalks, protective medians or barriers, and may be needed over or under multiple roadway bridge structures. Ignoring these accommodations can have long-lasting functional, safety, and financial impact on communities, motorists, and wildlife.

Once it has been determined that a road will be built, modified, or updated, how it will be built or modified, and an alignment selected, a budget and schedule are created. At this point, the project has been “programmed” for a specific delivery year. This process usually identifies the following:

- Project purpose and need
- Roles and responsibilities of partnering agencies
- List of project alternatives established
- Primary contacts for project

#### **Inset 2-1 / Roadside vegetation and driver safety**

Greater safety for the traveling public is the primary objective of many road projects. The designer can support road safety when he/she does not propose vegetation strategies that might make the roadway less safe. Integrating revegetation goals with safety goals can mean balancing an awareness of visibility issues, wildlife interactions, and other factors. Highway roadside design and revegetation efforts are subject to clear zone requirements that follow the American Association of State Highway and Transportation Officials (AASHTO) recommendations. FHWA and State DOT agencies are interested in the concept of a “forgiving” roadside: a roadside environment that allows a driver to recover safely if they drive off the road onto the roadside. The agencies expect a clear zone of low vegetation adjacent to the road, preferably low grassy surfaces, or shrub masses instead of trees. The roadside distance required to make a roadside forgiving depends on the speed limit of the highway, the traffic volume, and surrounding conditions. The AASHTO Roadside Design Guide (RDG) recommends clear zone widths based on the road design speed, average daily traffic, the up or down slope of the roadside and horizontal curve radius. The presence of curbs does not affect the clear zone distance along high speed roadways. Highway clear zones are generally in the 20-46 foot range, but vary based on speed and noted conditions.

- Preliminary project delivery schedule with milestones
- Collection and analysis of traffic data (accident history, average daily traffic volumes, etc.)
- Preliminary construction cost estimate

Environmental concerns for the project (cultural and natural resource) and estimation of the affected environment (WFLHD 2005).

Experts recommend that a revegetation designer be involved throughout the early planning and programming of a roadway project in order to identify issues and solutions, refine the budget, and help assess the feasibility of various alternatives.

### 2.3.2 ROAD PROJECT DEVELOPMENT

The road project construction (contract) document development process begins after the project is programmed and ends with the beginning of construction. Depending on environmental concerns and right-of-way issues, the project development process may take between one and five years. Contract documents are typically defined in the Owner-Contractor Agreement within the specifications Division 00. Contract documents typically consist of drawings, specifications, addendums to 100 percent documents distributed to bidders, and supplemental drawings provided by the designer to the contractor.

The road project development phase usually has three document review and approval sub-phases. These involve developing, analyzing, and considering approaches and alternatives to various design details within the project until a strategy and specifications of how to best proceed are shared in the final documents. The process usually involves:

- **Preliminary**—review of road construction documents that are approximately 30 percent complete
- **Intermediate**—review of road construction documents that are approximately 50 to 70 percent complete
- **Final**—review of road construction documents 70 to 100 percent complete. Final reviews often have a 95 percent or 100 percent pre-final review and the designer is able to make final corrections before plans are distributed for bidding or construction. Ideally, only the approved 100 percent plans will be advertised for bids and then given to the awarded contractor for construction or in a design-build process, the 100 percent plans will be reviewed and approved by the contractor and project owner team and construction will begin.

#### Preliminary Phase

The preliminary development phase involves collecting information and initiating contacts with “stakeholders.” Stakeholders are individuals or parties who are interested in or affected by the road construction (local and adjacent landowners, resource agencies, regulatory agencies, and any other potentially affected parties). The local jurisdiction will often be able to provide a detailed contact list of stakeholders who will likely be interested in the road project. The preliminary phase is necessary to refine purpose and need, to develop a range of alternatives to address purpose and need, and to obtain the approvals and clearances to allow the project to proceed. This includes commitments for CSS safety, cultural, aesthetic, functional, and environmental mitigations. The preliminary phase takes the road construction plans to about 30 percent completion. Once approvals and clearances are obtained, funds can typically be committed to continue development of the project.

Usually the preliminary phase will include the development and identification of:

- Preliminary construction plans (usually drafts about 30 percent complete) of the proposed alternatives (plan/profile sheets, typical sections, major work items identified and located)
- Preliminary construction cost estimates for alternatives

- Resource surveys (wetlands, archeological sites, and biological assessments)
- Preliminary construction schedule
- Identification of impacts and mitigation
- Environmental approvals and selection of alternatives for implementation
- List of contacts for the project

For the revegetation designer, the preliminary phase is a crucial one. This phase often represents the best opportunity for input regarding issues associated with revegetation, including disturbances planned for existing soil and vegetation on the site. Significant features of the preliminary revegetation plan will need to be incorporated during this road planning phase. By the time of environmental approvals, the vegetative concepts and the necessary commitments of resources and funds will need to be integrated with the documents, as revegetation is an important aspect of environmental protection and mitigation. The appropriate level of detail for the revegetation plan during the preliminary phase depends on the project. State and local agencies may specify environmental goals and requirements for the project regarding issues of soil stabilization, percent native vegetative cover, and protection of water quality (Inset 2-2). The designer is often asked to assist in preparation of the Storm Water Pollution Prevention Plan (SWPPP) for the project based on these goals, and to design them into the final revegetation plan. Environmental approvals are key milestones for the project team in order to maintain the project schedule, and for some designer teams, important in regard to availability of funds to carry out site assessments and revegetation planning work including preliminary mapping and seed collections.

## Intermediate Phase

The next phase of road development moves towards 50 to 70 percent completion of the road construction documents. This phase involves refining plans and specifications, obtaining rights-of-way and permits, and creating detailed plan and profile sheets. The intermediate set of documents will include major budget items and quantities, final information pertaining to environmental concerns (such as SWPPP or erosion control plans), and major elements such as grading, drainage, and other issues defined.

At the intermediate stage, the designer is typically far along in the development of the revegetation drawings. The SWPPP is submitted to the State or local jurisdiction for review and approval. Mitigating measures have been identified for affected areas and owner or design-build contractor contracts for seed and seedling production have begun. The intermediate set of road documents will include specifications for how the road project will be carried out. These specifications and contract requirements are key tools for the designer.

### Special Contract Requirements: A Key Tool for Revegetation

In every phase of road development, there are two key components of contract documents: (1) drawings (plans) and (2) specifications (contract descriptions). The drawings are visual representations of the proposed work with dimensions, labels, and notes as described later in this chapter. Specifications describe and define materials, equipment, systems, procedures, performance, workmanship, standards, provisions, and requirements for the work that each agency provides to contractors or employees to carry out the work. A “special provision” (FHWA, State DOT) or “special contract requirement” (US Forest Service) is a type of specification. Standard specifications are uniformly carried out for most projects.

Special provisions or special contract requirements address local, project specific, context-sensitive concerns for a particular project. They are modifications of existing specifications found within the agency manual, or newly written specifications that are designed to address special concerns not adequately addressed in the standard contract specifications. For example, a standard specification may exist for chipping woody debris; however, the standard specification does not address size requirements of the chipped material. A project may require a uniform size of material be shredded and screened, rather than

*For the Designer*  
**Special Provisions and special contract requirements often need pre-approval by the reviewing agency prior to inclusion in construction documents. The approval process can take several months.**

chipped, to create optimal mulch for the project. Accordingly, the designer can create a special provision or special contract requirement that will specify the size (such as three inches or less in length) and processing needs (such as shredded and screened rather than chipped). Careful research is often needed to adequately develop and describe a special provision or special contract requirement, and ultimately to achieve the desired results in the field. In the future, generically applicable special provisions or special contract requirements may become adopted as standard specifications if they come to be utilized on most projects.

#### **Inset 2-2 / Example of an environmental regulation requirement**

"Finalstabilization" means that all soil disturbing activities at the site have been completed and that a uniform perennial vegetative cover with a density of at least 70 percent of the native background vegetative cover for the area has been established on all unpaved areas and areas not covered by permanent structures, or equivalent permanent stabilization measures.

"Final Stabilization" (adapted from EPA 2006) means that:

- All soil disturbing activities at the site have been completed and either of the two following criteria are met:
  - *a uniform (i.e., evenly distributed, without large bare areas) perennial vegetative cover with a density of 70 percent of the native background vegetative cover for the area has been established on all unpaved areas and areas not covered by permanent structures, or*
  - *equivalent permanent stabilization measures (such as the use of riprap, gabions, or geotextiles) have been employed.*
- When background native vegetation will cover less than 100 percent of the ground (e.g., arid areas, beaches), the 70 percent coverage criteria is adjusted as follows: if the native vegetation covers 50 percent of the ground, 70 percent of 50 percent (i.e., 35 percent ( $0.70 \times 0.50 = 0.35$ ) is total cover for final stabilization. On a beach with no natural vegetation, no stabilization is required.
- In arid and semi-arid areas only, all soil disturbing activities at the site have been completed and both of the following criteria have been met:
  - *Temporary erosion control measures (e.g., degradable rolled erosion control product) are selected, designed, and installed along with an appropriate seed base to provide erosion control for at least three years without active maintenance, and*
  - *The temporary erosion control measures are selected, designed, and installed to achieve 70 percent vegetative coverage within three years.*

Special provisions or special contract requirements are an important tool for designers to communicate special expectations with contractors, to clearly define contracting responsibilities to reduce duplicate efforts, and to set standards for performance. The designer can specify to contractors what the requirements are, and how they might be met, measured, and paid for. The agency with jurisdiction over the specifications will review and approve special provisions or special contract requirements and assign them specification numbers, a process that typically can take weeks or months. An effective practice can be to give attention to modifying or creating special provisions or special contract requirements to meet the revegetation needs of the project by the intermediate phase. Special provisions or special contract requirements that are approved may be included in the final contract documents for the road construction project.

## **Final Phase**

The final set of road construction documents will include the detailed design elements of the Revegetation Plan, as well as all the details for road construction. Drawings and contract specifications will be fully developed. Special provisions and special contract requirements will have been submitted

and approved. Environmental approvals and permits will have been accomplished. Final cost estimates will be provided along with a comprehensive schedule. The work of the designer in developing the revegetation plan, as well as efforts to reduce the construction footprint and protect native vegetation on the project site, will be an integral part of the road construction documents. At this point, the revegetation documents, including a budget, are finalized. Final road construction documents are submitted for distribution to bidders or, in the case of design-build, are submitted to the contractor who distributes them to his or her team of sub-contractors. Once the documents are in the contractor's hands, there is a good opportunity for the revegetation designer to connect and coordinate with the contractor, landscape sub-contractor, and agencies in order to confirm plant material sources and to schedule availability of plant materials with outplanting windows.

### 2.3.3 CONSTRUCTION

Following project development, the construction phase begins. Road construction can take one to three years. Sometimes there is a formal milestone when the project is handed off to construction personnel. If so, the construction manager or project engineer becomes an essential contact for the designer, who may attend some of the weekly meetings that will take place during road construction. The construction phase of a road is completed when there is a formal acceptance of the road by the road owning agency. For the designer, implementation and monitoring phases of revegetation may begin before road construction (e.g., with plant materials procurement) and continue following completion of construction.

### 2.3.4 MAINTENANCE

Following construction of the road, the work of the designer will usually continue for an additional one to three years until the revegetation is fully implemented. Also, the activities centered on monitoring and adaptive management of the establishing vegetation continue to take place, perhaps up to five years after the road construction is complete. The submission of a final monitoring report is the milestone marking the end of the designer's formal efforts on the project. Although the designer's efforts are contractually complete, valuable information can be obtained by periodic visits to the project site in order to see how the restoration efforts develop over time.

Coordination with the road owning agency and the individuals who carry out road maintenance is important to ensure that native vegetation continues to thrive on the site. In many instances once the state DOT hands off the project to the county, the state DOT does not provide further input. For example, the agency taking ownership, often the county, could have maintenance methods that may ultimately undo portions of the revegetation, such as blanket herbicide use as standard practice along roadsides. The designer can protect the revegetation plan by identifying the ultimate roadside vegetation management agency in the planning phase, checking what maintenance methods are currently utilized, and to continually coordinate efforts with the ultimate maintaining agency in order to ensure that future management is appropriate for the native vegetation. The transportation agencies can continue to monitor the road to ensure that the project adequately addressed the problem (infrastructure decay, safety issues, etc.) that led to it.

## 24 ROAD CONSTRUCTION PLANS

This section explains how to use roadway engineering views for revegetation planning, including determining the vegetation zones that begin where the pavement ends. A glossary with illustrations is provided in order to understand technical concepts and terminology for effective communication with others involved in road design and construction. This section also describes how to read and interpret:

- Plan views

- Profile views
- Cross-section views
- Typical views
- Summary of quantities tables

## 2.5 READING PLANS

The plan set consists of construction drawings and specifications for each section of road or project. The four most common views of plans utilized by the designer are plan views, cross-section views, profile views, and typical views. Each of these is defined in Table 2-1. Examples and descriptions for interpreting each of these views are provided below. Each engineering plan includes a legend defining abbreviations and symbols as well as a summary of plan quantities table.

### 2.4.1 PLAN VIEW

The plan view shows the existing and proposed road locations from a bird's eye view. It is important to note that plan sets, in particular road plans, historically displayed distances in meters. This practice was discontinued nationwide in the early 2000s and distances are now displayed in US Customary Units (feet). The proposed road is usually designated with solid lines (Figure 2-2A). The solid centerline (of the road to be constructed) is divided into 100 foot sections (large ticks), often with 20 foot subdivisions also designated (small ticks—not shown). Each 100-foot division is called a station, representing a discrete, surveyed, and identifiable point within the road corridor. Each station is identified with a unique number that indicates its distance from the beginning of the project. For example, the station 19+000 indicates this point is 19,000 feet from the start of the project; 19+040 indicates this point is 19,040 feet from the start. This short-hand identifier is also used to indicate the placement of road-related infrastructure, such as culverts, the beginning and end of guard-rail construction, or the placement of a sign. In the field, stations are identifiable as vertically aligned numbers written on wooden stakes and driven into the ground, facing the roadway. Detailed location of elements off of the roadway can then be identified by station along the centerline of the alignment plus the offset distance dimension from the edge of roadway pavement. This will appear as a +0.00 note on the field stake. Not only do the stations provide locations, they help to locate revegetation units. The plans also show the top of the cut slope (Figure 2-2B, dotted line), bottom of the fill slopes (Figure 2-2C, dashed lines), and the location of the original road, which will be obliterated in this example (Figure 2-2D, shaded area). Plans also include temporary construction easement lines (Figure 2-2D (outside line)) and right-of-way lines (ROW or R/W). These are, in effect, the property lines of the roadway and an important boundary for the designer.

**Table 2-1 | Definitions of views**

| View          | Definition  |
|---------------|---|
| Plan          | A drawing depicting a portion of the road project from a bird's eye view.   |
| Profile       | A drawing depicting the vertical plane section along the longitudinal centerline of the road, expressed in elevation or gradient. |
| Cross-section | A drawing depicting a horizontal section of the road viewed vertically, as if cut across the width of the road.                   |
| Detail        | A drawing depicting features of a particular design, installation, construction or methodology.                                   |

*Source: Keller and Sherar 2003*

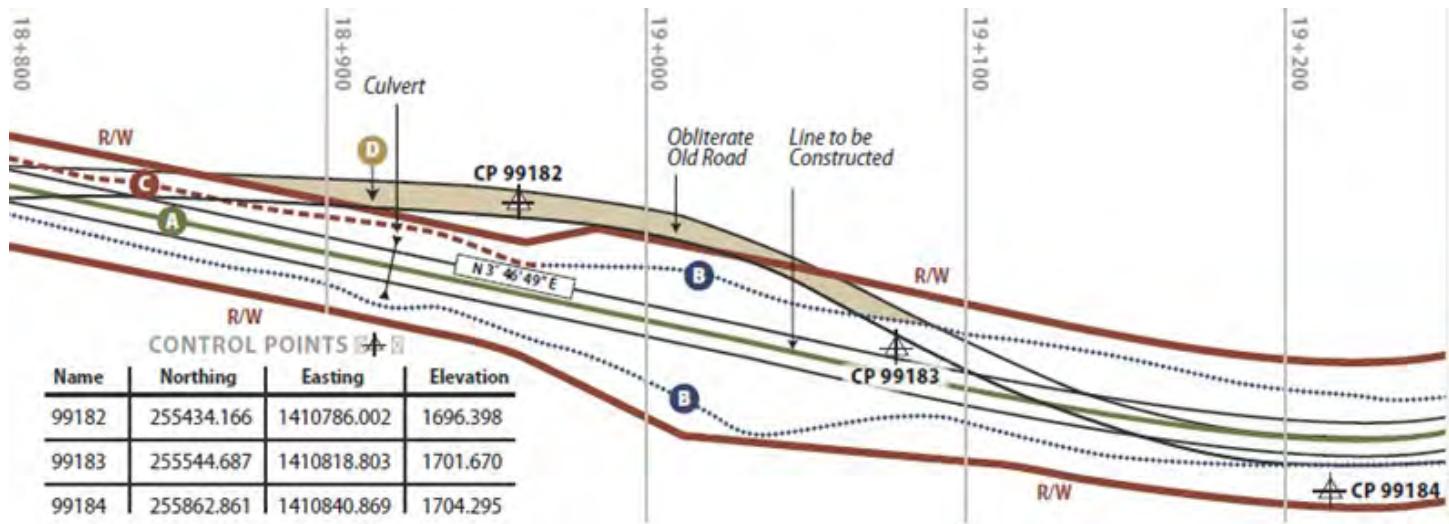


Figure 2-2 | Example plan view

## 2.4.2 PROFILE VIEW

The profile view is a trace of a vertical plane intersecting a particular surface of the proposed road construction (Figure 2-3E). It corresponds to the longitudinal centerline of the road bed in the plans.

Profile grade means either elevation or gradient of the trace, depending on the context. The trace of the existing road is shown as a dashed line (Figure 2-3F) and a dotted line (Figure 2-3G). A vertical scale provides useful information about the profile of construction grades throughout the project. This view shows where the proposed road will be lower than the existing road (Figure 2-3H) and areas where it will be higher (Figure 2-3 I). Where the planned road is lower, material will usually be removed and used in areas needing fill. Additional information is often displayed adjacent to and locatable by the station numbers, such as volumes of excavation and embankment work, guard-rail placement, or wall placements.

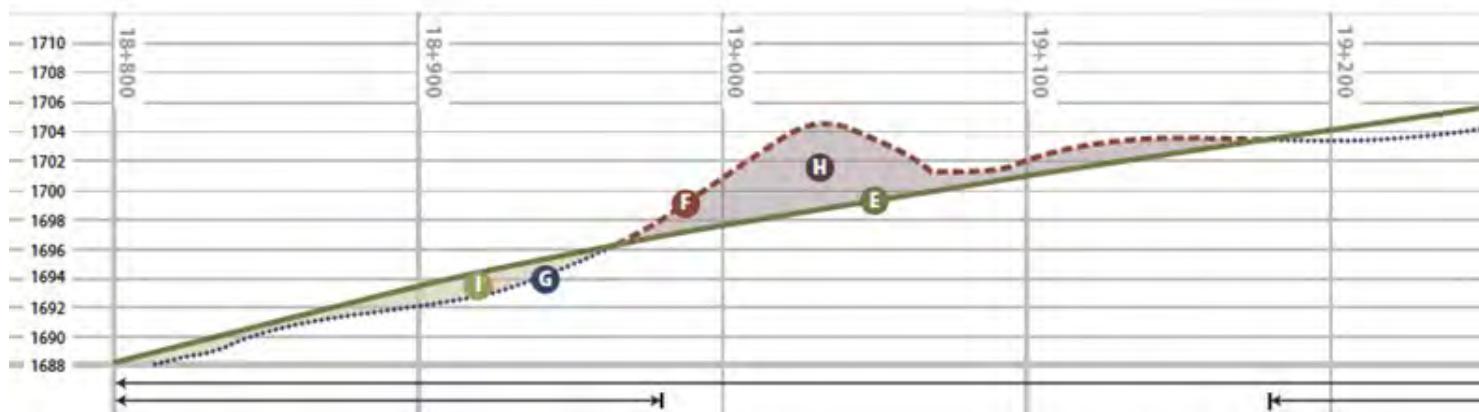
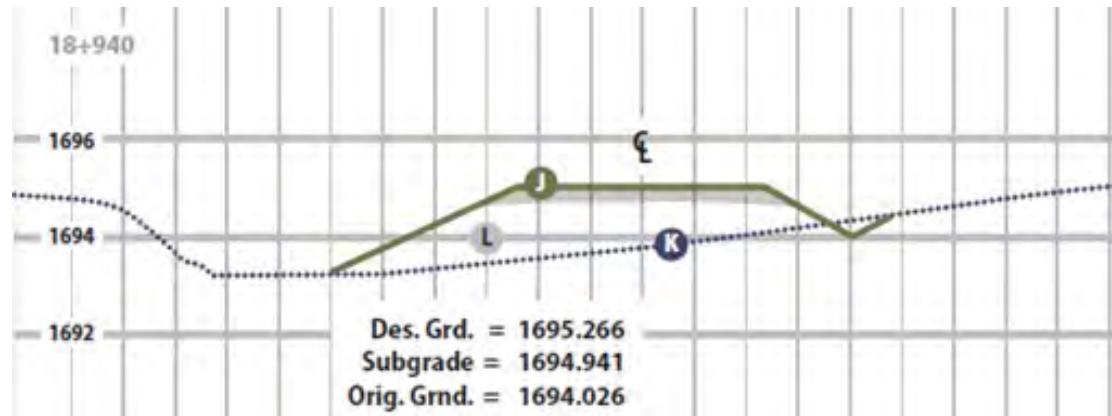


Figure 2-3 | Example profile view

### 2.4.3 CROSS-SECTION VIEW

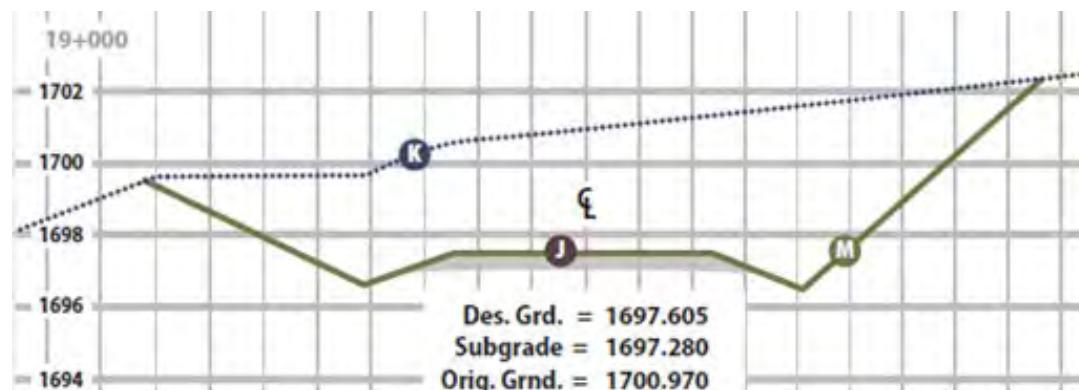
Cross-sections are views of the slopes perpendicular to the direction of the road. They display a vertical section of the ground or structure at right angles to the centerline or baseline of the roadway.

Depending on the length and topographic complexity of the road, there can be hundreds of cross-sections. Each cross-section is referenced back to a station. For example, the cross-section shown in Figure 2-4 depicts the slope at Station 18+940. It shows the proposed road (Figure 2-4 J), and the natural ground line as a dotted line (Figure 2-4 K). This section will have material brought in and placed as fill (Figure 2-4 L).



**Figure 2-4 / First example cross-section**

The cross-section in Figure 2-5 shows a through cut at 19+000. Material will be removed from the natural ground line (Figure 2-5K) to the proposed ground line – solid line (Figure 2-5 M).



**Figure 2-5 / Second example cross-section**

Cross-section and plan views are used together to view the proposed road three-dimensionally. From these views, a more detailed revegetation plan can be developed. Each cross-section can be reviewed and a set of revegetation criteria can be developed for similar cross-sections throughout the project.

Cross-sections show the proposed slope gradients for cut and fill slopes and provide the designer a means to determine slope steepness. Like stationing, the method of depicting slopes has changed over the years. Older plan sets often depicted slopes as a ratio of one unit horizontal to one unit vertical.

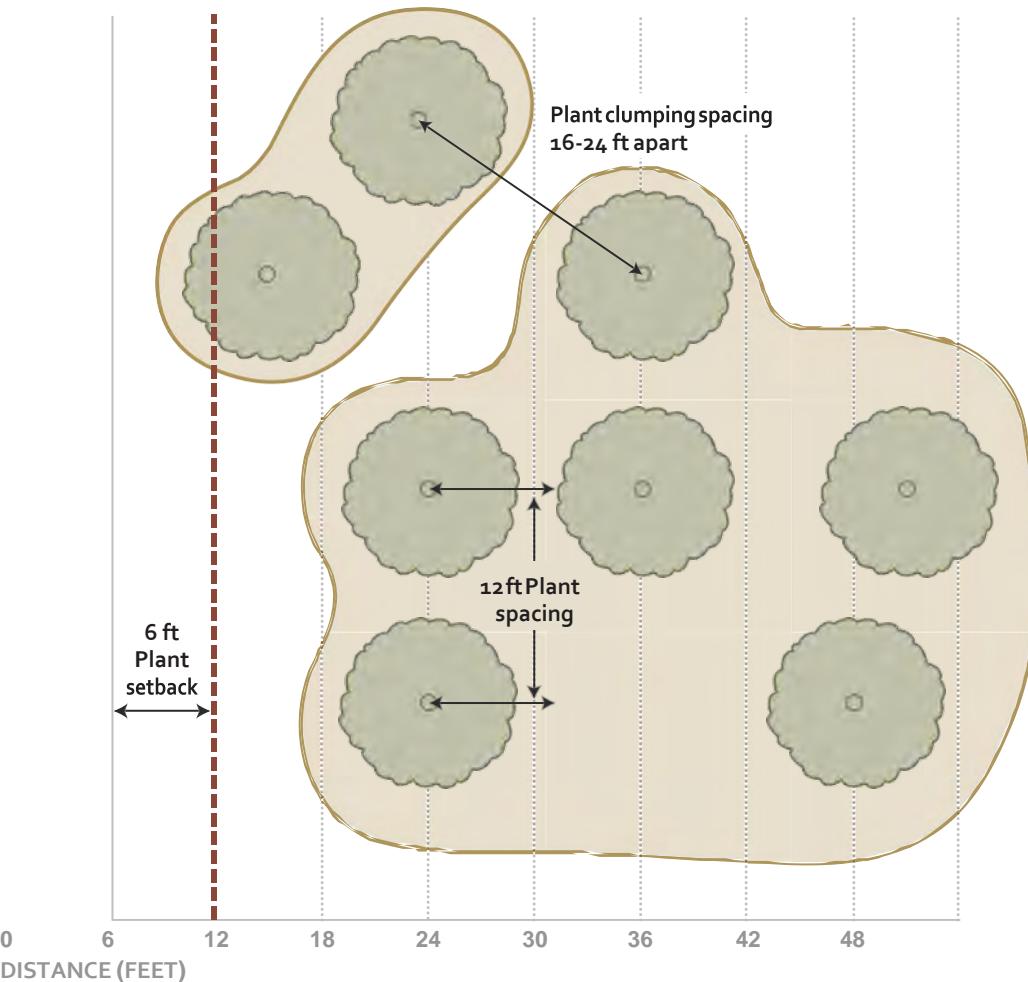
Several years ago, however, slope ratios were brought more in line with other disciplines and are now depicted as one unit vertical to one unit horizontal (vertical:horizontal). When slopes are flatter than 1:1 ( $45^\circ$  or 100 percent), the slope is expressed as the ratio of one unit vertical to the number of units horizontal. For slopes steeper than 1:1, the slope ratio is expressed as number of units vertical to one unit horizontal. To avoid confusion, it is wise to notate the ratio by indicating the vertical and horizontal, for example 1V:2H, and to think in terms of rise over run (Figure 3-61).

#### 2.4.4 TYPICAL VIEWS

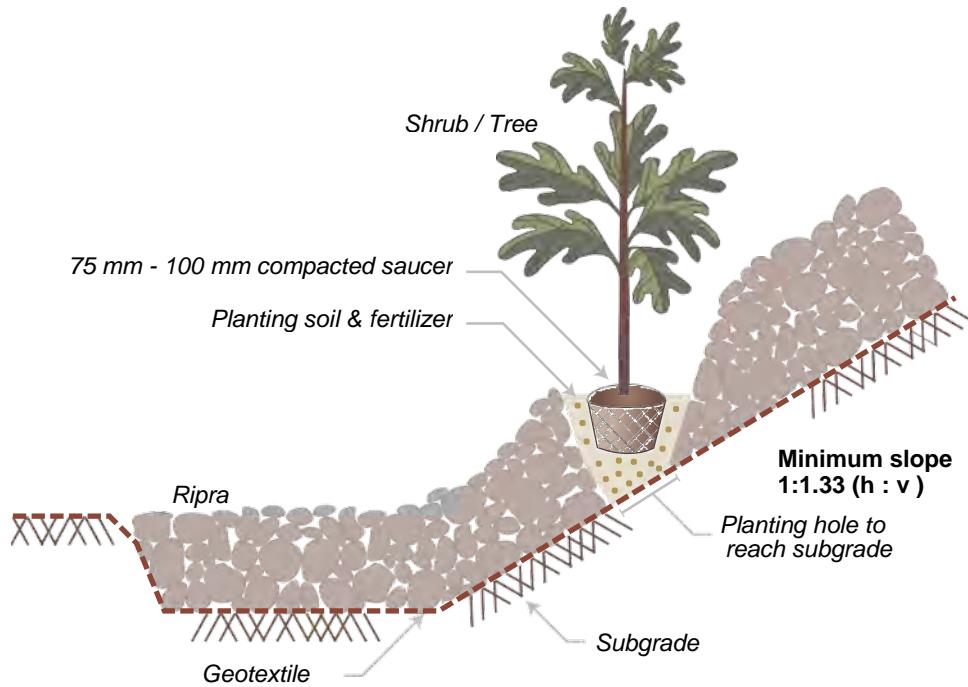
Typical views graphically illustrate the design or construction details of the structures or other components that will be encountered in the road project (Figure 2-6). They can cover such structures as retaining walls, road surfaces, guardrails, ditch lines, plant installation, etc. They may be shown in profile, cross-section, or plan views (Figure 2-7). Like special contract requirements (see above), typical views are useful in helping communicate a new or modified approach to an existing methodology or construction technique.

#### 2.4.5 SUMMARY OF QUANTITIES TABLE

Tabulation of plan quantities tables contain details on quantities, types of materials, and performance specifications. Standardized for Federal Lands Highway projects, specifications for construction of federal roads are described in the FHWA handbook: Standard Specifications for Construction of Roads and Bridges on Federal Highway Projects. They are cited as "FP-03," or "FP-14", indicating "Federal Project" specifications issued in 2003 or 2014, respectively. The State DOTs have analogous manuals. Tabulation of plan quantities references not only the particular item specification number in the FP manual, but also the station number(s) of the planned work. Information of special interest for the designer includes the number of hectares of clearing and grubbing, hectares of obliterated roads listed by station, and the number of cubic meters of wood mulch to be produced. The summary of quantities table provides a summary of all tabulation of plan quantity tables contained within the plan. It generally does not include station numbers.



**Figure 2-6 | Example typical view**

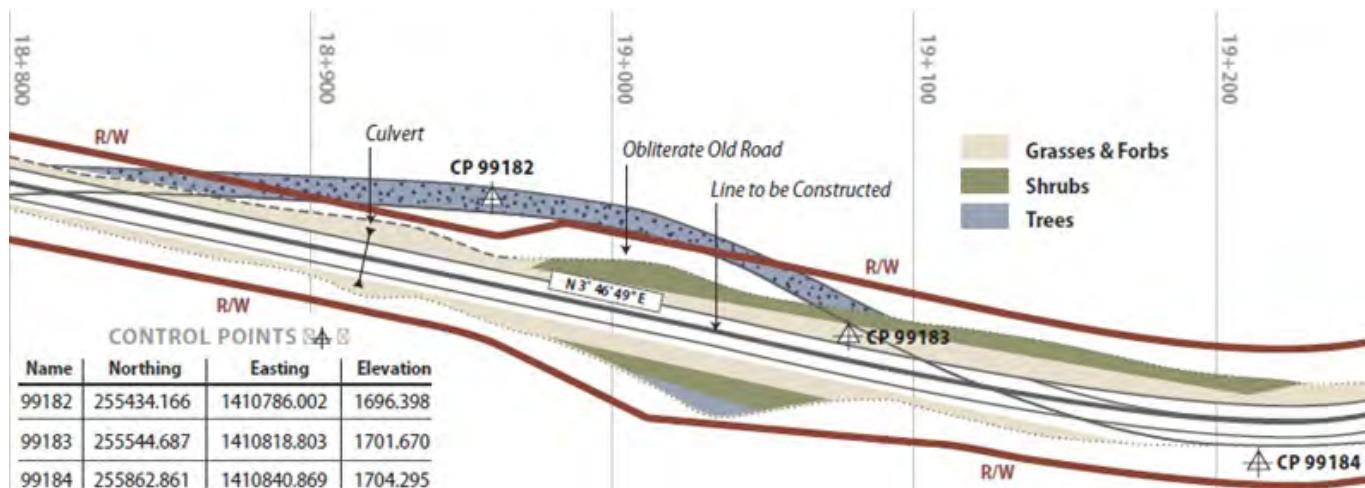


**Figure 2-7 | Example typical view of installation trail and turnout**

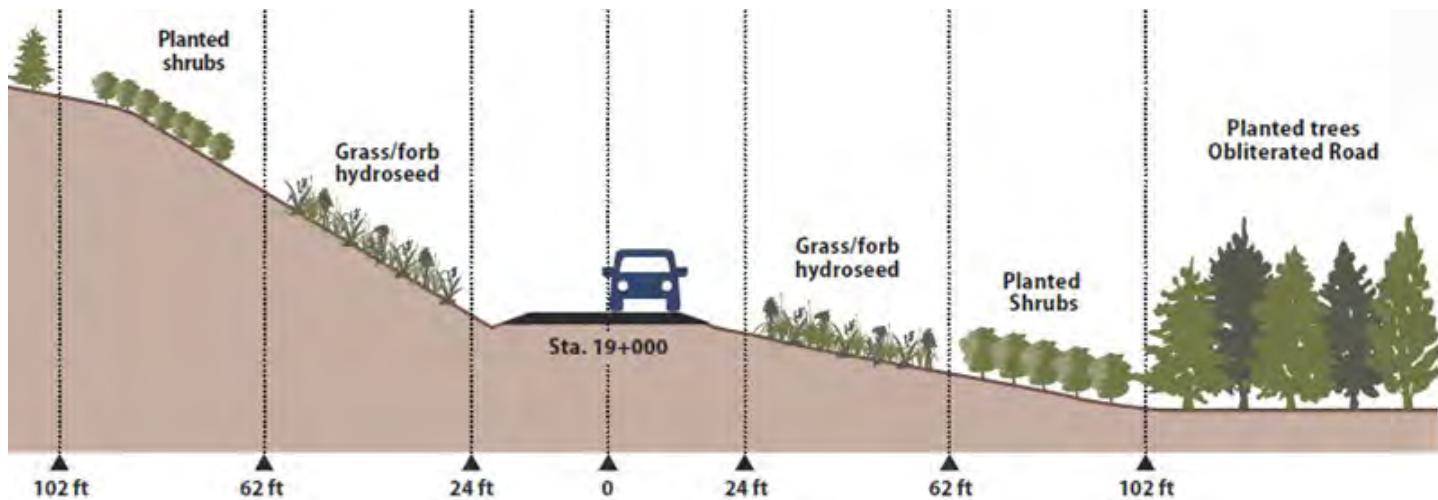
## 26 INTERPRETING ENGINEERING VIEWS FOR REVEGETATION PLANNING

Road construction, management, and safety concerns result in distinct revegetation zones along roadsides. Properly interpreting plans helps define where these zones may be located and what types of vegetation should be established. While the sizes and characteristics will vary, the zones that parallel the road can be grouped into four categories. Zone 1 begins immediately adjacent to the road surface (concrete or asphalt) and includes the road shoulder (concrete, asphalt, compacted gravel, coarse subsoil, etc.), the bottom of the drainage ditch, and portions of cut and fill slopes. This first zone is generally considered to be up to 10 feet from the pavement edge and is often barren of vegetation due in part to herbicide application, road salts, frequent ditch cleaning, and/or mowing. Local, state, and federal road managing agencies will have limitations to the types and proximity of vegetation that can be established next to the road. Zone 2 begins at roughly 10 feet from the pavement edge and continues laterally to about 30 feet. This zone may begin at the ditch bottom or at some point on a cut or fill slope, and may continue to the limit of the construction zone. Within Zone 2, grasses and forbs can thrive, but larger forbs, shrubs, and trees usually are not planted or encouraged due to safety, maintenance, and visibility issues. Beyond 30 to 50 feet from the pavement edge, Zone 3 begins in which larger forbs and shrubs can be planted. Past about 75 feet from the pavement edge, Zone 4 begins with largely unrestricted revegetation potential. Understanding these zones is necessary to coordinate revegetation with road management practices and safety considerations (Forman et al 2003).

To define the zones and begin to interpret engineering plans for revegetation work, information from plan sheets and quantity tabulations is applied to the plan map as shown in Figure 2-8. Each area can be considered a revegetation unit or subunit. An estimate of the area in each of these units can be calculated and used in determining how many seedlings or pounds of seeds will be needed. These criteria can be graphically displayed on a typical cross-section (Figure 2-9). On this cross-section, the criteria can be expressed as follows: from 20 to 60 feet grasses/forbs will be hydroseeded; from 60 to 100 feet shrubs will be planted; and on obliterated roads, trees will be planted. The slopes given in cross-sections can help define the types of revegetation methods available.



*Figure 2-8 | Interpretation of plan view showing revegetation zones*



*Figure 2-9 | Cross-section showing revegetation zones as interpreted from engineering plans*

## 2.7 UNDERSTANDING TECHNICAL CONCEPTS AND TERMINOLOGY

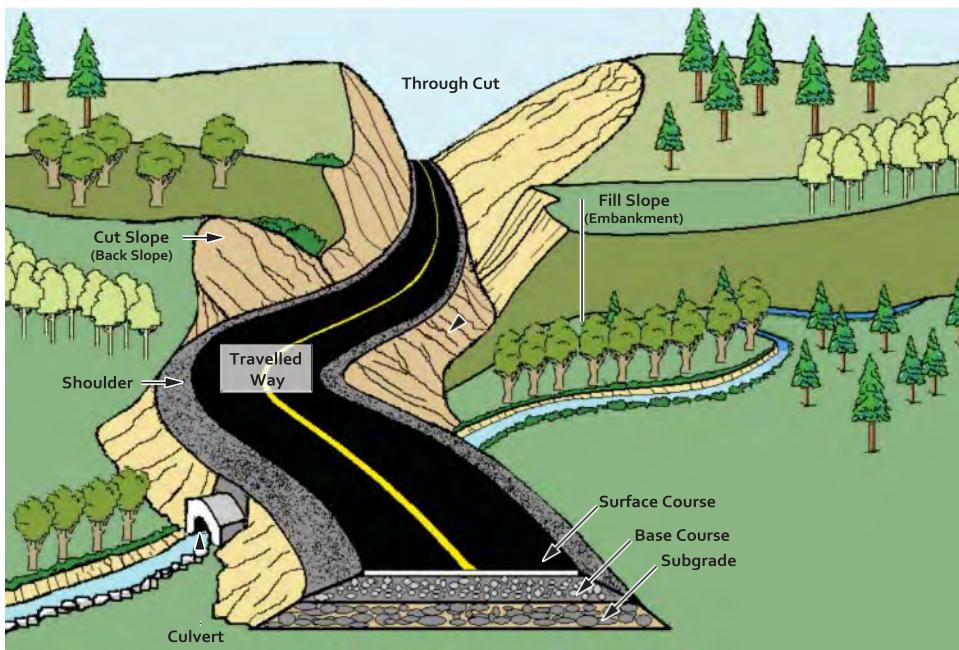
The ability to understand and utilize the technical concepts and terminology used by road engineers is essential to roadside revegetation planning. This ability enables the designer to contribute effectively to road design and construction processes, as well as to communicate revegetation needs and goals to others involved with the project. The following section introduces key technical concepts and terminology relating to road design and construction.

### Road Concepts and Terminology

The following terminology was adapted from Keller and Sherar 2003.

## 2.7.1 ROAD COMPONENTS

- **Berm**—A ridge of rock, soil, or asphalt, typically along the outside edge of the road shoulder, used to control surface water. It directs surface runoff to specific locations where water can be removed from the road surface without causing erosion.
- **Buttress**—A structure designed to resist lateral forces. It is typically constructed of large riprap rock, gabions, or well-drained soil to support the toe of a slope in an unstable area.
- **Cross-Section**—A drawing depicting a section of the road sliced across the whole width of the road. Can also apply to a stream, slope, or slide.
- **Cut Slope (Back Slope or Cut Bank)**—The artificial face or slope cut into soil or rock along the inside edge of the road
- **Cut-and-fill**—A method of road construction in which a road is built by cutting into the hillside and spreading the spoil materials in adjacent low spots and as compacted or side-cast fill slope material along the route. A “balanced cut-and-fill” utilizes all of the “cut” material to generate the “fill.” In a balanced cut-and-fill design there is no excess waste material and there is no need for hauling additional fill material. Thus, cost is minimized.
- **Ditch (Side Drain)**—A channel or shallow canal along the road intended to collect water from the road and adjacent land for transport to a suitable point of disposal. It is commonly along the inside edge of the road. It also can be along the outside edge or along both sides of the road.
- **End Haul**—The removal and transportation of excavated material off-site to a stable waste area (rather than placing the fill material near the location of excavation).

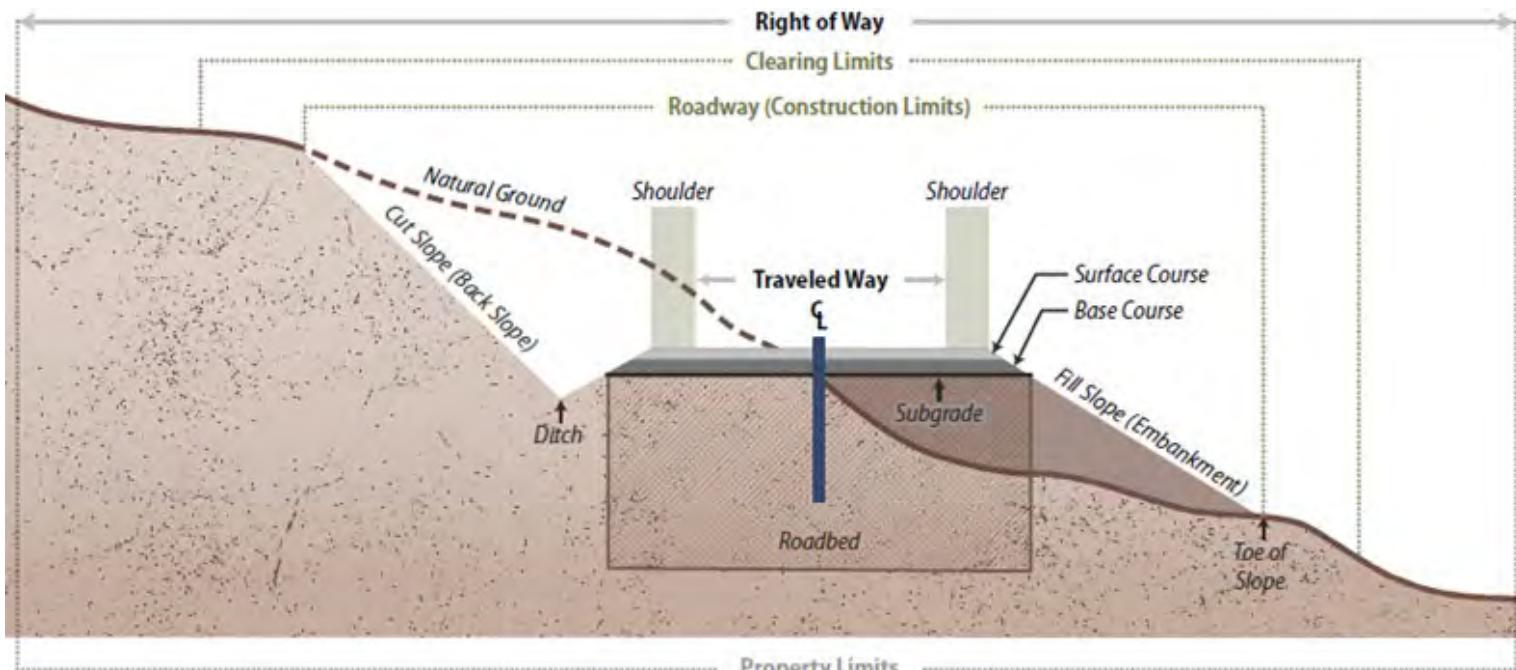


**Figure 2-10 | Terms used to define roads**

- **Fill Slope (Embankment Slope)**—The inclined slope extending from the outside edge of the road shoulder to the toe (bottom) of the fill. This is the surface formed when excavated material is placed on a prepared ground surface to construct the road subgrade and roadbed template.
- **Grade (Gradient)**—The slope of the road along its alignment. This slope is expressed as a percentage and is the ratio of elevation change compared to distance traveled. For example, a +4 percent grade indicates a gain of 4 units of measure in elevation for every 100 units of measure traveled.
- **Natural Ground (Original Ground Level)**—The natural ground surface of the terrain that existed prior to disturbance and/or road construction.
- **Plan View (Map View)**—View seen when looking from the sky towards the ground (bird's-eye

view).

- **Reinforced Fill**—A fill that has been provided with tensile reinforcement through frictional contact with the surrounding soil for the purpose of greater stability and load carrying capacity. Reinforced fills are comprised of soil or rock material placed in layers with reinforcing elements to form slopes, walls, embankments, dams, or other structures. The reinforcing elements range from simple vegetation to specialized products such as steel strips, steel grids, polymeric geogrids, and geotextiles.
- **Retaining Structure**—A structure designed to resist the lateral displacement of soil, water, or any other type of material. It is commonly used to support a roadway or gain road width on steep terrain. They are often constructed of gabions, reinforced concrete, timber cribs, or mechanically-stabilized earth.
- **Right-of-Way**—The area or footprint of land over which facilities such as roads, railroads, or power lines are built. Right-of-way is an easement that grants the right to pass over the land of another.
- **Road Center Line**—An imaginary line that runs longitudinally along the center of the road.
- **Roadbed**—Width of the road used by vehicles, including the shoulders, measured at the top of subgrade.



**Figure 2-11 | Terms used to define roads: cross-section**

- **Roadway (Construction Limits or Formation Width)**—Total horizontal width of land affected by the construction of the road, from the top of cut slope to the toe of fill or graded area.
- **Side-Cast Fill**—Excavated material pushed on a prepared or unprepared slope next to the excavation to construct the roadbed. The material is usually not compacted.
- **Shoulder**—The paved or unpaved area along the edge of the traveled way of the road. An inside shoulder is adjacent to the cut slope. An outside shoulder is adjacent to an embankment slope.
- **Traveled Way (Carriageway)**—That portion of the road constructed for use by moving vehicles including traffic lanes and turnouts (excluding shoulders).
- **Through Cut**—A road cut through a hill slope or, more commonly, a ridge, in which there is a cut slope on both sides of the road.
- **Through Fill**—Opposite of a through cut, a through fill is a segment of road that is entirely

composed of fill material, with fill slopes on both sides of the road.

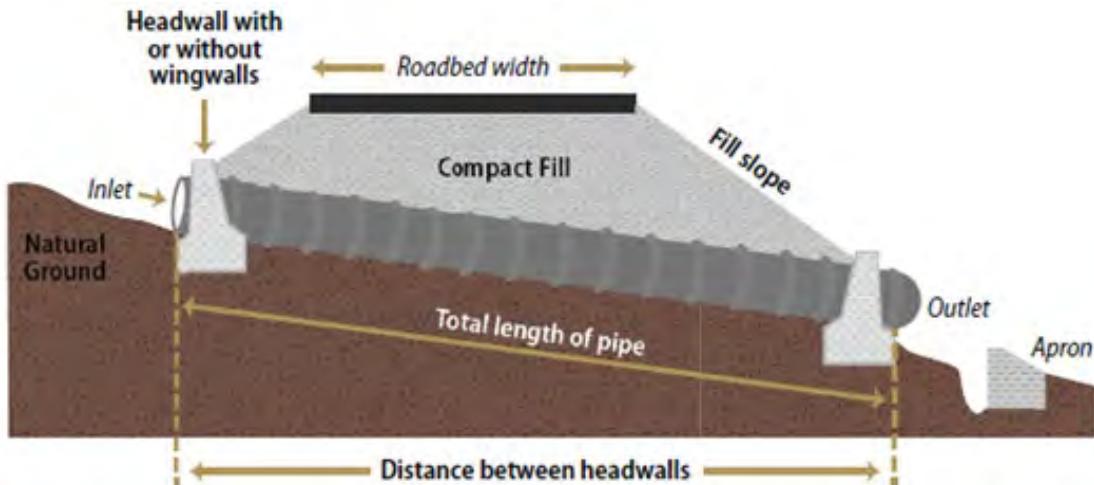
## 2.7.2 ROAD STRUCTURAL SECTION AND MATERIALS

- **Borrow Pit (Borrow Site)**—An area where excavation takes place to produce materials for earthwork, such as a fill material for embankments. It is typically a small area used to mine sand, gravel, rock, or soil without further processing.
- **Quarry**—A site where stone, riprap, aggregate, and other construction materials are extracted. The material often has to be excavated with ripping or blasting, and the material typically needs to be processed by crushing or screening to produce the desired gradation of aggregate.

## 2.7.3 SURFACE DRAINAGE

- **Armor**—Rocks or other material placed on headwalls, on soil, or in ditches to prevent water from eroding and undercutting or scouring the soil.
- **Drainage Structure**—A structure installed to control, divert, or move water off or across a road, including but not limited to culverts, bridges, ditch drains, fords, and rolling dips.
- **French Drain (Underdrain)**—A buried trench, filled with coarse aggregate, and typically placed in the ditch line along the road to drain subsurface water from a wet area and discharge it to a safe and stable location. French drains may use variable sizes of rock but do not have a drain pipe in the bottom of the trench.
- **Inslope**—The inside cross-slope of a road subgrade or surface, typically expressed as a percentage. Inslope is used to facilitate the draining of water from a road surface to an inside ditch. An insloped road has the highest point on the outside edge of the road and slopes downward to the ditch at the toe of the cut slope, along the inside edge of road.
- **Outslope**—The outside cross-slope of a road subgrade or surface, typically expressed as a percentage. Outslope is used to facilitate the draining of water from a road directly off the outside edge of the road. An outsloped road has the highest point on the uphill or inside of the road and slopes down to the outside edge of the road and the fill slope.
- **Riprap**—Well-graded, durable, large rock, ideally with fractured surfaces, sized to resist scour or movement by water and installed to prevent erosion of native soil material.

## 2.7.4 CULVERTS AND DRAINAGE CROSSINGS



**Figure 2-12 | Culvert components**

- **Catch Basin**—The excavated precast, or constructed basin at the inlet of a culvert cross-drain

pipe for storing water and directing it into the culvert pipe.

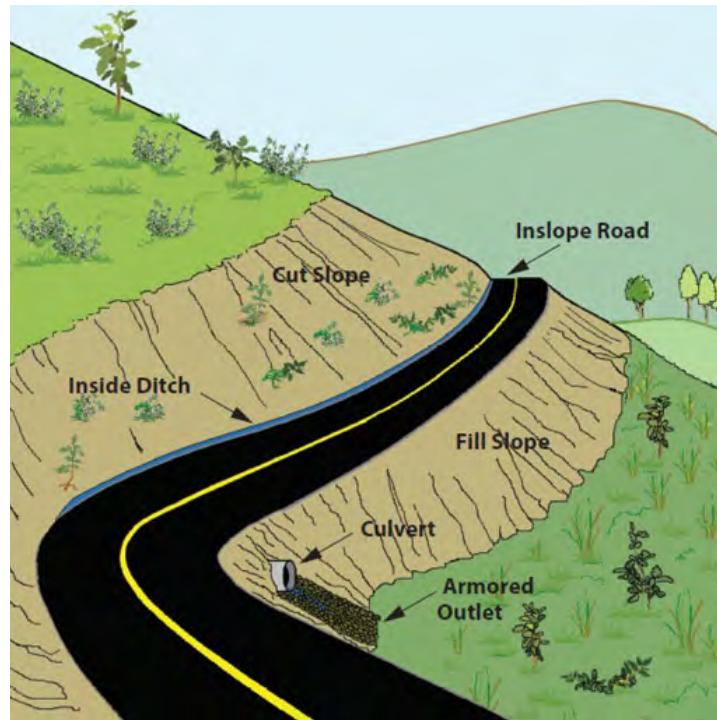
- **Culvert**—A drainage pipe, usually made of metal, concrete, or plastic, set beneath the road surface to move water from the inside of the road to the outside of the road, or under the road. Culverts are used to drain ditches, springs, and streams that cross the road. The invert is the floor or the bottom of the structure at its entrance.
- **Headwall**—A concrete, gabion, masonry, or timber wall built around the inlet or outlet of a drainage pipe or structure to increase inlet flow capacity, reduce risk of debris damage, retain the fill material, and minimize scour around the structure.
- **Inlet**—The opening in a drainage structure or pipe where the water first enters the structure.
- **Outlet**—The opening in a drainage structure or pipe where the water leaves the structure. The outlet is usually lower than the inlet to ensure that water flows through the structure.

## 2.7.5 MISCELLANEOUS TERMS

- **Angle of Repose**—The maximum slope or angle at which a granular material, such as loose rock or soil, will stand and remain stable.
- **Gabions**—Baskets (usually made of heavy-gauge wire) filled with rocks or broken pieces of concrete (~10-20 cm in size), used for building erosion control structures, weirs, bank protection, or retaining structures.
- **Road Decommissioning**—Permanently closing a road through techniques that may include blocking the entrance, scattering limbs and brush on the roadbed, replanting vegetation, adding waterbars, removing fills and culverts, or reestablishing natural drainage patterns. The basic road shape, or template, is still in place. The end result is to terminate the function of the road and mitigate the adverse environmental impacts of the road.
- **Road Obliteration**—A form of road closure that refills cut areas, removes fills and drainage structures, restores natural contours, revegetates the area, and ultimately attempts to restore the natural ground shape and condition. Most adverse environmental impacts of the road are eliminated.
- **Silt Fence**—A temporary barrier used to intercept sediment-laden runoff from slopes. It is typically made of porous geotextile material.

- **Streamside Management Zone**

The land, together with the associated vegetation, immediately in contact with the stream and sufficiently close to have a major influence on the total ecological character and function of the stream. It is a buffer area along a stream where activities are limited or prohibited.



**Figure 2-13 | Terms used to describe road slopes**

# 3—Planning

- 
- 3.1 Introduction**
  - 3.2 Defining Revegetation Objectives**
  - 3.3 Gathering Pre-field Information**
  - 3.4 Defining Revegetation Units**
  - 3.5 Identifying Reference Sites**
  - 3.6 Gathering Field Information**
  - 3.7 Defining the Desired Future Condition**
  - 3.8 Identifying Limiting Factors to Plant Establishment**
  - 3.9 Identify Factors That Affect Pollinators**
  - 3.10 Inventory of Site Resources**
  - 3.11 Developing a Vegetation Management Strategy during Project Design**
  - 3.12 Select Site Improvement Treatments**
  - 3.13 Selecting Plant Species for Propagation**
  - 3.14 Select Plant Establishment Methods**
  - 3.15 Develop a Revegetation Plan**

## 3.1 INTRODUCTION

Careful planning is essential to the success of any roadside revegetation project. There are a series of steps that are important to consider in developing a comprehensive revegetation plan. These are shown in Table 3-1 and outlined in this chapter.

**Table 3-1 | Planning Phase Steps**

| Activity  | Definition   |
|---|--|
| Defining revegetation objectives                    | Description of the general purpose and goals of the road project as determined by societal, ecological, and transportation needs, environmental regulations, and other factors. Development of revegetation objectives, including pollinator habitat enhancement, erosion control, water quality enhancement, weed control, and carbon sequestration.  |
| Gathering pre-field information                     | Prior to field surveys, the review of reports and websites that describe soils, vegetation, climate, and pollinators for the project area.   |
| Defining revegetation units                         | Classification of areas within the project site that are similar enough to be appropriate for similar strategies and treatments. Homogenous sites will have only a few units; sites with greater diversity (different soil types, microclimates, vegetation types, and management needs) will have more revegetation units.<br>Each revegetation unit is distinct in terms of ecology, management requirements, or both. |
| Identifying reference sites                         | Location of natural or revegetated areas that will serve as models for desirable recovery of native plant communities and pollinator habitat. One or more reference sites are identified for each revegetation unit in the project area.   |
| Gathering field information                         | Survey of reference sites, as well as the road project area, for vegetation, soils, climate, and pollinator habitat that will provide baseline ecological data for developing the revegetation plan.   |
| Defining the desired future condition (DFC) target  | Creation of specific, measurable goals for each revegetation unit, usually defined in terms of the percentage of vegetative cover, ground cover, species composition, plant growth, plant density, and pollinator diversity and abundance.   |
| Identifying limiting factors to plant establishment | Review of pre-field and field information to determine which site factors may be limiting for plant growth based on water input, water storage, water loss, nutrient cycling, surface stability and slope stability.   |
| Identifying factors that affect pollinators         | Review of pre-field and field information to identify the limiting factors affecting pollinators. These include nectar and pollen sources, breeding habitat, water sources, shelter, landscape connectivity, nesting habitat, and vegetation management.   |
| Inventory of site resources                         | Assessing physical resources that may be available or generated for use in the revegetation plan. These resources include topsoil, duff, litter, woody materials, logs, and plant materials  |
| Developing a vegetation management strategy         | A maintenance strategy is developed to assess how the revegetation project will affect the management and maintenance of the roadside after the road project has been completed and integrate this into the revegetation plan. Ideally, the planning team or designer meets with local maintenance personnel, to learn what problems can be expected in reestablishing roadsides with native plants.                     |
| Selecting site improvement treatments               | The treatments that will improve the site for plant growth or pollinator enhancement are selected.   |
| Selecting plant species for propagation             | Native plant species that will be used on the project are selected based on project objectives and how well they will perform on the site. Genetic diversity and local adaptation is considered in the reproductive sources that will be used to propagate the plant materials.  |
| Selecting plant establishment methods               | Optimal propagation methods are determined for each plant species. These include the plant materials that will be produced (seeds, cuttings, plants), the method of plant material installation, and when to install the plant materials (seeding and planting windows).   |
| Developing a revegetation plan                      | A written revegetation plan captures the most important information and decisions that were made on revegetating the project site. It typically outlines project objectives, revegetation units, treatments, plant species, planting methods, roles, responsibilities, timelines, and budget.  |

## 3.2 DEFINING REVEGETATION OBJECTIVES

The design objectives of a road project guide the development of the revegetation plan. As discussed in Chapter 2, road objectives usually involve goals of improving safety and efficiency, as well as environmental health. Revegetation objectives develop from road objectives and become the foundation of the revegetation and monitoring plans. It is important to develop a clear set of revegetation objectives early in the planning phase. When these objectives are understood and expectations are clear, the development and implementation of a revegetation plan are easier and more successful. Most roadside revegetation projects share the common objective of initiating and/or accelerating the process of natural succession near the roadside in order to establish self-sustaining native plant communities (Brown and Amacher 1999; Clewell et al 2005). This objective usually reflects larger project goals, stated in terms of increasing pollinator habitat, protecting soil and water resources, carbon sequestration, enhancing roadside aesthetics, limiting invasive plants, and improving road safety and function while protecting environmental health. Later in the planning process, revegetation objectives are used to develop specific goals (stated as DFC targets) for evaluating the success of the revegetation work. Table 3-2 defines some terms commonly used in defining revegetation objectives. Clarifying whether the overall goal is reclamation or restoration, for example, is an essential distinction for defining revegetation objectives.

**Table 3-2 | Terms used in defining revegetation objectives**

| TERM           | DEFINITION   |
|----------------|--|
| Revegetation   | To reestablish vegetation on a disturbed site. This is a general term that may refer to restoration, reclamation, and rehabilitation.  |
| Restoration    | This is the re-creation of the structure and function of the plant community identical to that which existed before disturbance. The goal of restoration is conservation, with the intention of maximizing biodiversity and functioning.   |
| Reclamation    | This is the re-creation of a site that is designed to be habitable for the same or similar species that existed prior to disturbance. Reclamation differs from restoration in that species diversity is lower and projects do not re-create identical structure and function to that before disturbance. However, a goal of long-term stability with minimum input is implied. |
| Rehabilitation | This process creates alternative ecosystems that have a different structure and function from the pre-disturbance community, such as a park, pasture, or silvicultural planting.   |

Adapted from Allen et al 1997

Table 3-3 illustrates some of the most common road-related revegetation objectives as they relate to the road design goals. Most revegetation projects state several objectives to address both short-term and long-term outcomes. For example, short-term, immediate revegetation objectives on most projects include erosion control and water quality protection through mulch and vegetative cover. A long-term revegetation objective would be to establish a native plant community, with a range of plant species that benefit pollinators by increasing foraging, breeding, and nesting habitats. Table 3-4 outlines roadside objectives specific to enhancing pollinator habitat. While short-term objectives might rely on quick-growing ground covers such as grasses and forbs, long-term objectives are often broadened to include such revegetation treatments as planting deep-rooted tree and shrub seedlings to stabilize roadsides, creating visual screens of road infrastructure, and/or supporting sustained native plant community development.

**Table 3-3 | Native plants are used to meet road and revegetation objectives**

| REVEGETATION OBJECTIVE         | FUNCTION OF NATIVE PLANTS   |
|--------------------------------|---|
| Pollinator habitat enhancement | An important revegetation objective is to improve pollinator habitat by selecting a mix of plant species and site improvements that encourage foraging, breeding, nesting, and overwintering of a variety of pollinator species (Table 3-4).  |
| Erosion control                | Controlling surface erosion and thereby protecting soil and water quality is a high priority on road construction projects. Native grasses, forbs, and other herbaceous plants can help meet this challenge, particularly when they are accompanied by appropriate mulching treatments. Deep-rooted native trees and shrubs can also enhance stability of cut and fill slopes.  |
| Water retention                | Runoff from road surfaces and cut slopes concentrate water into ditches during rainstorm events, increasing the amount of water that normally enters natural drainage ways. Practices that use native plants in the design, such as constructed wetlands and bioretention swales, amended ditches and fills, filter strips, can help retain much of this water on the project site, reducing the amount of sediments and road pollutants from entering stream courses. The additional water increases the productivity of the established plants. |
| Weed control                   | Roadsides can be corridors for the transport and establishment of noxious or invasive weed species. Once established, weeds are hard to eradicate and become seed sources for further encroachment. Revegetating with desirable native species minimizes opportunities for problem species to establish.  |
| Carbon sequestration           | Roadside revegetation with native plants can help improve air quality and the health of the public and environment by plants taking in and reducing the amount of carbon dioxide in the atmosphere. Plants store the carbon in the soil long-term and release beneficial oxygen. Native roadside vegetation typically requires less mowing maintenance, herbicides and pesticides, which reduces carbon in the atmosphere and reduces maintenance costs and associated emissions.   |
| Visual enhancement             | Vegetation is often used to enhance the aesthetic experience of the traveler. Wildflowers add color and beauty throughout the growing season; deciduous trees provide shade, vertical structure, and change color in fall; and evergreen species stay green all year, adding visual interest, structure, and green color all year. Vegetation can also be used to frame views, soften views or hide structures such as gabion walls or slopes covered by riprap.  |
| Wildlife enhancement           | Many roads intercept animal corridors. Designing native plantings into animal underpasses or overpasses can make roads more permeable to wildlife. The presence of birds and small animals can be enhanced when appropriate plant species are reestablished.  |
| Cost management                | Advanced planning, an integrated approach, and the use of appropriate stocktypes and equipment all facilitate successful and cost-effective revegetation.   |

**Table 3-4 | Roadside objectives for enhancing pollinator habitat**

Roadsides planted with native plants also can provide pollinators with shelter, sites for nesting or egg-laying, and overwintering habitat. Pollinators have complex life cycles, with different needs at different stages of their lives. Roadsides can provide resources for a portion of the life cycle of some species, while providing resources needed for the entire life cycle of other species.

| POLLINATORS                   | FOOD  | SHELTER  | REVEGETATION GOALS   |
|-------------------------------|---|--|--|
| Bats (nectar feeding species) | Nectar, pollen, fruit   | Caves and mines  | <ul style="list-style-type: none"> <li>▪ Include food plants</li> </ul>  |
| Bees: Bumble                  | Nectar for adults; nectar and pollen collected as provisions for larvae | Nest in small cavities, underground in abandoned rodent nests, under clumps of grass, or in hollow trees, bird nests, or walls | <ul style="list-style-type: none"> <li>▪ Increase density and diversity of native flowering plants</li> <li>▪ Provide native bunch grasses for bumble bee nesting habitat</li> </ul> |
| Bees: Ground-nesting          | Nectar for adults; nectar and pollen collected as provisions for larvae | Nest in bare or partially vegetated, well-drained soil   | <ul style="list-style-type: none"> <li>▪ Provide areas with partially vegetated well-drained soil</li> <li>▪ Provide living and dead pithy and woody vegetation</li> </ul>           |
| Bees:                         | Nectar for adults; nectar   | Nest in narrow tunnels in dead standing  |  |

|                                   |  |  |   |
|-----------------------------------|--|--|---|
| Tunnel-nesting                    | and pollen collected as provisions for larvae  | trees, or excavate nests in pith of stems and twigs. Some construct domed nests of mud, plant resins, saps, or gums on the surface of rocks or trees |   |
| Beetles                           | Pollen and nectar as adults; vegetation or prey such as aphids, slugs, insect eggs, as larvae or adults  | Larvae overwinter in loose soil or leaf litter; Adults shelter under rocks, logs, brush  | <ul style="list-style-type: none"> <li>▪ Increase density and diversity of native flowering plants</li> <li>▪ Provide refuge from burning and grazing during dormant season and early spring</li> </ul>   |
| Butterflies/moths:<br>Caterpillar | Leaves of larval host plants   | Host plants  |   |
| Butterflies/moths:<br>Adult       | Nectar; some males obtain nutrients, minerals, and salt from rotting fruit, tree sap, animal dung and urine, carrion, clay deposits, and mud puddles | Protected site such as a tree, bush, tall grass, or a pile of leaves, sticks, or rocks   | <ul style="list-style-type: none"> <li>▪ Increase density and diversity of native flowering plants</li> <li>▪ Include host plants</li> <li>▪ Provide refuge from burning and grazing during dormant season and early spring</li> </ul>          |
| Flies                             | Nectar and sometimes pollen as adults; insect prey such as aphids, scales, mites, thrips   | Larvae found on plants near prey; pupae and adults overwinter in soil or leaf litter   | <ul style="list-style-type: none"> <li>▪ Increase density and diversity of native flowering plants</li> <li>▪ Provide refuge from burning and grazing during dormant season and early spring</li> </ul>   |
| Hummingbirds                      | Nectar, insects, tree sap, spiders, caterpillars, aphids, insect eggs, and willow catkins  | Trees, shrubs, and vines; typically need red, deep-throated flowers, such as twin berry or penstemons  | <ul style="list-style-type: none"> <li>▪ Increase density and diversity of native flowering plants, particularly species with deep throats</li> </ul>   |
| Wasps                             | Nectar as adults; insect prey such as caterpillars, aphids, grasshoppers, planthoppers, and true bugs as larvae                                      | Many nest in the ground; others nest in tunnel nests in wood or cavities in mud or resin   | <ul style="list-style-type: none"> <li>▪ Increase density and diversity of native flowering plants</li> <li>▪ Provide areas with partially vegetated well-drained soil</li> <li>▪ Provide living and dead pithy and woody vegetation</li> </ul> |

Revegetation objectives are often developed by the designer and design team and are supported by, or integrated with, public documents such as Environmental Assessments or Environmental Impact Statements. The objectives sometimes originate from a state or federal agency and motivated by environmental concerns and regulations regarding water quality, erosion control, and vegetation establishment. In the early stages of planning, revegetation objectives are broad and general. As the project evolves, objectives are translated into more precise and measurable goals (DFC targets). After the installation is complete, DFC targets and revegetation objectives will be used to monitor, evaluate, and manage the project.

### 3.3 GATHERING PRE-FIELD INFORMATION

The revegetation plan is developed by obtaining an understanding of the road design and by gathering pre-field information on the soil, climate, vegetation, and pollinators of the project site. Much of this information can be obtained prior to visiting the project

site. A good pre-field review of information can make the time in the field more efficient and effective.

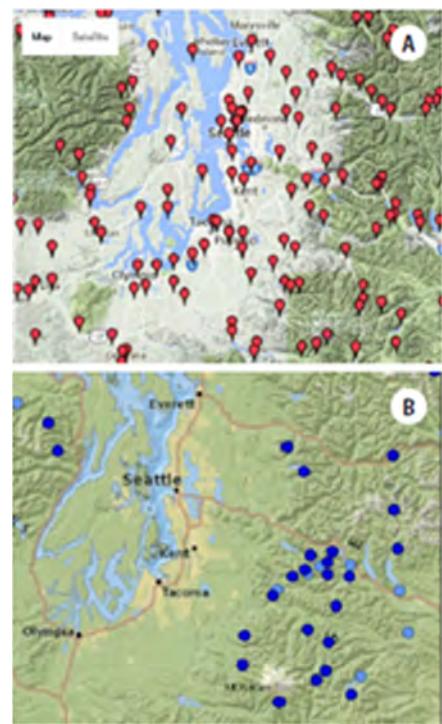
### 3.3.1 CLIMATE PRE-FIELD ASSESSMENT

Local climate plays a dominant role in the success or failure of the revegetation effort. Knowledge of local climate factors, including historic climate data and recent trends, can inform the designer and help delineate the appropriate revegetation units and develop achievable DFC targets. In later phases of the planning process, climate data will be used to determine appropriate revegetation treatments.

**Inset 3-1 | Climate change** Climate change effects, including increased frequency of extreme weather events, wildfires, invasive species, drought, increased temperatures, and altered stream flows, can affect native plants and revegetation success on both temporal and spatial scales. Measurable effects of climate change have been observed such as spring events arriving earlier, shifts in species distribution, and disruption of plant-pollinator dynamics. Parmesan and Yohe (2003) showed spring events such as budburst in plants, the arrival of migratory birds and butterflies, bird nesting, and others occurred an average of 2.3 days earlier per decade over 123 years. This same review revealed that the latitudinal and elevational range limits of several alpine plant populations had shifted northward 3.79 miles and upward approximately 20 feet per decade over the past 1,000 years. As a result of these effects of climate change plants, in particular long-lived perennials, are forced to either adapt or migrate (Parmesan 2006).

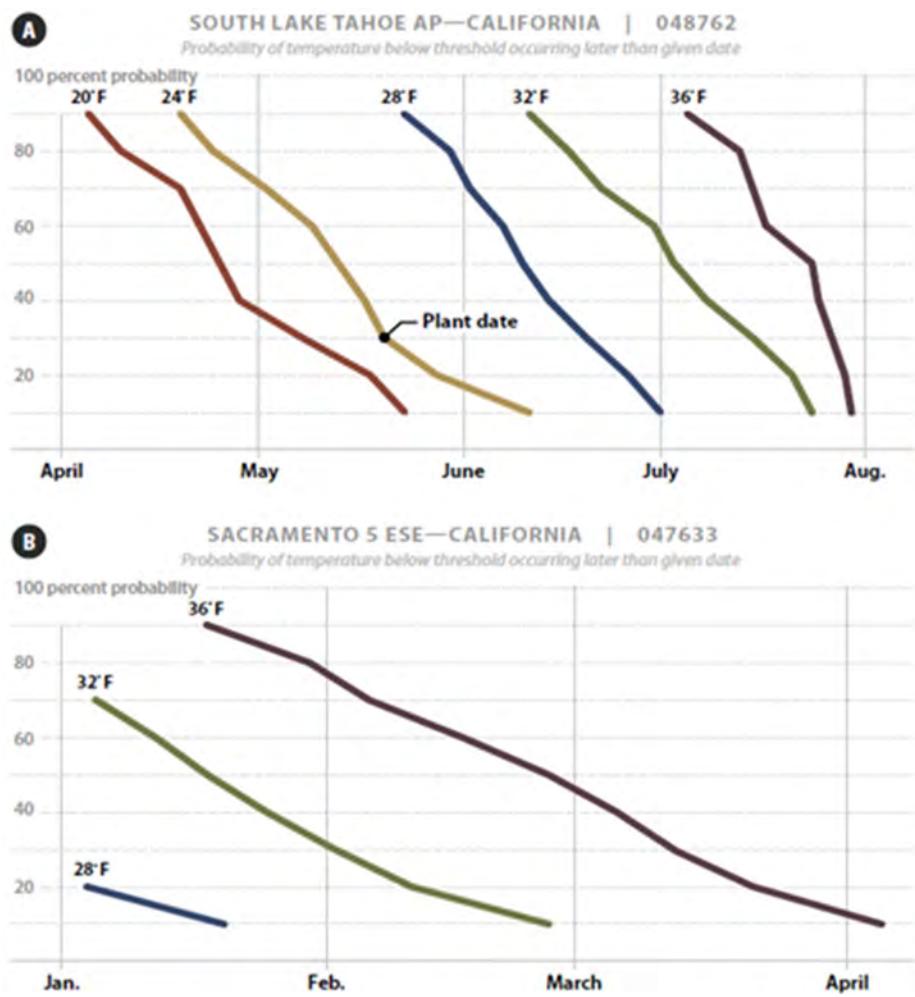
The migration of plants or changes in plant phenological events have been observed to disrupt or decouple pollinating insect interactions with their host plants. For example, host plants may senesce more quickly than caterpillars develop and other asynchronies between butterflies and their host plants (Parmesan 2007). Some species of pollinators have undergone range contractions due to climate change (Kerr and others 2015). Limitations of dispersal and establishment may mean that many species of pollinators will not be able to keep up with predicted climate change scenarios and that climate change will exacerbate other threats to pollinators, including habitat loss (Settele and others 2016).

Obtaining climate records from a variety of sources is the first step in conducting a climate assessment. There are many sources of climate records for the United States (Figure 3-1). One source is the [Western Regional Climate Center](#) website that displays the location of the National Oceanic and Atmospheric Administration (NOAA) Cooperative Stations in the United States and provides historical weather data for most stations. Each weather station has helpful graphics, such as spring and fall “freeze probabilities” (Figure 3-2) that can be used to determine the best dates for sowing seeds and planting seedlings. Another available graphic is the probability of precipitation throughout the year, which can be used to determine if supplemental irrigation is necessary (Figure 3-3).



**Figure 3-1 | NOAA and NRCS weather stations**

The United States has an extensive system of weather stations maintained by National Oceanic and Atmospheric Administration (NOAA) and Natural Resources Conservation Services (NRCS). These maps show the locations of weather stations in the Puget Sound area. Stations administered by NOAA are shown on the left (A) and those by NRCS on the right (B). Historic climate summaries and interpretative graphs for each station can be downloaded from each website.



Finding a weather station closest to the project site will be helpful in understanding the influence climate will have on the project. The PRISM website allows the user to locate the project site on a map of the United States, so instead of using data from a single weather station located miles away from the project, this website creates a weather profile specific to the project site. It does this by extrapolating data from surrounding weather stations using a digital elevation model and expert knowledge of complex climatic patterns. This website also displays weather trends and anomalies (e.g., extreme heat) that can be helpful in planning (Figure 3-4). NRCS maintains the [National Weather and Climate website](#) that reports historic and real-time weather data from automatic weather stations located in remote mountainous areas of the western United States. If a project is located near one of these stations, then it is easy to monitor current weather conditions. Historic data and more recent climate trends can be a valuable tool for the designer to consider when developing a revegetation plan. Recent studies of climate trends have noted changes that are affecting pollinators and their habitat and have offered recommendations on how designers can adapt revegetation plans to these changing conditions.

**Figure 3-2 / Spring and fall freeze probability graphs**

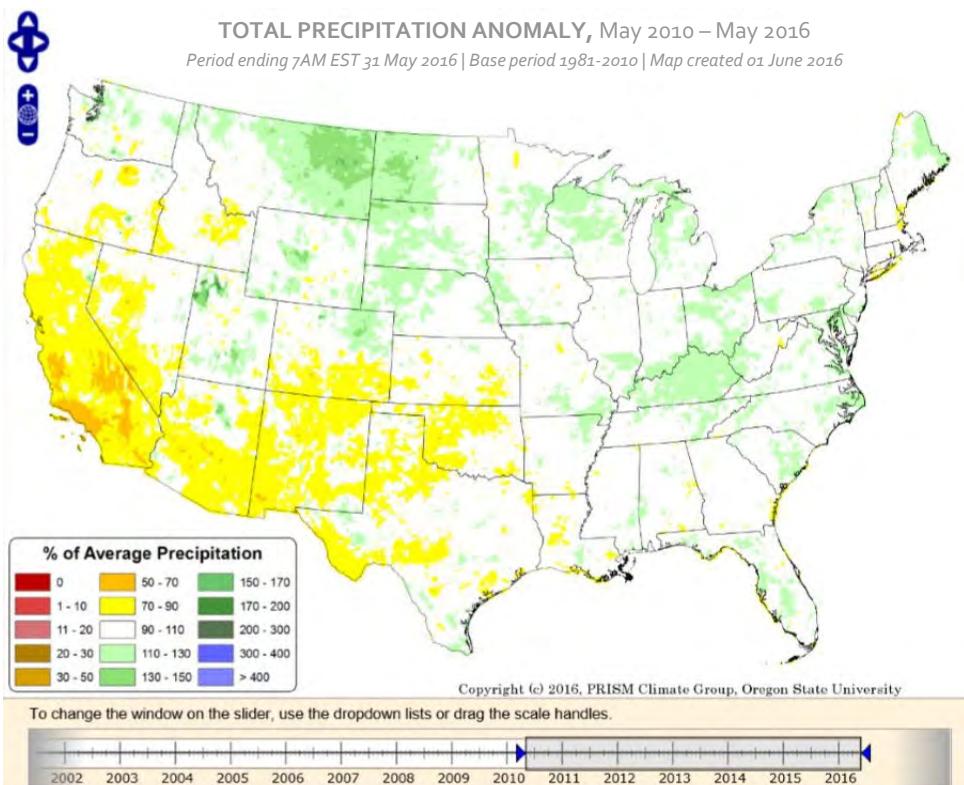
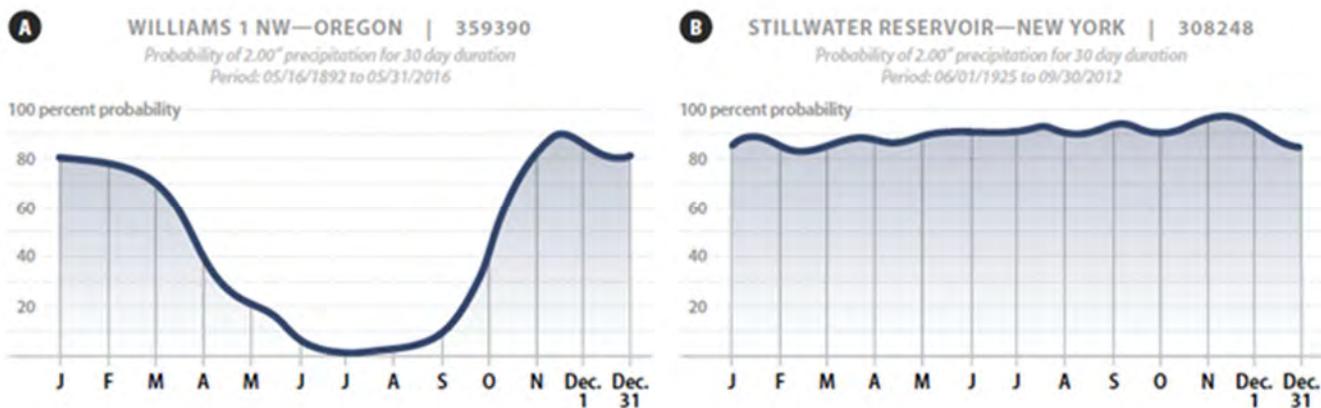
In addition to historic weather station data, the [Western Regional Climate Center](#) has many useful graphs (located on the left-hand side of the weather station data screen), such as the “Spring and Fall Freeze Probability” curves. The graph on the left shows the probability of temperatures dropping to sub-freezing temperatures during the winter through summer in South Lake Tahoe, California. At this site, planting might be planned when there is less than a 30-percent probability that temperatures will drop below 24°F to reduce the risk of seedling damage as the plants are coming out of dormancy. This would put the planting date sometime around the middle of May. In contrast, Sacramento, California, to the west and near sea level has a very different climate as shown in the lower left-hand graph. According to these curves, it is improbable that temperatures ever reach 24°F in winter and spring; therefore, plants could be installed at any time during the winter.

Source: [Western Regional Climate Center](#)

**Figure 3-3 | Rainfall probability graphs**

Another helpful graph from the [Western Regional Climate Center](#) displays the probability of receiving precipitation through the year. The probability of receiving 2 inches of rainfall in a 30-day period for a station in southwestern Oregon (A) indicates that it is highly unlikely this will ever happen during the summer, which may lead the practitioner to consider supplemental watering or some other measure to keep plants alive during the first year after the seedlings are planted. In comparison, the probability that 2 inches of precipitation would occur in 30 days in upstate New York (B) any time of the year is more than 90 percent, indicating that irrigation of newly planted seedlings may not be necessary.

Source: Western Regional Climate Center

**Figure 3-4 | Precipitation trends**

The [PRISM](#) website allows the user to obtain extrapolated weather data for any point in the United States. By identifying the project location on an interactive map of the U.S., a report is generated that summarizes site-specific climate information for that area. In addition, this website displays climate information in a variety of ways that may be helpful in revegetation planning. This map, for instance, shows precipitation trends for the past five years across the U.S., which can be useful during the development of the revegetation plan.

## Planning for Climate Change

Climate change effects, including increased frequency of extreme weather events, wildfires, invasive species, drought, increased temperatures, and altered stream flows, can affect native plants and revegetation success on both temporal and spatial scales.

Measurable effects of climate change have been observed such as spring events arriving earlier, shifts in species distribution, and disruption of plant-pollinator dynamics. Parmesan and Yohe (2003) showed spring events such as budburst in plants, the arrival of migratory birds and butterflies, bird nesting, et al occurred an average of 2.3 days earlier per decade over 123 years. This same review revealed that the latitudinal and elevational range limits of several alpine plant populations had shifted northward 3.79 miles and upward approximately 20 feet per decade over the past 1,000 years. As a result of these effects of climate change plants, in particular long-lived perennials, are forced to either adapt or migrate (Parmesan 2006).

The migration of plants or changes in plant phenological events have been observed to disrupt or decouple pollinating insect interactions with their host plants. For example, host plants may senesce more quickly than caterpillars develop and other asynchronies between butterflies and their host plants (Parmesan 2007). Some species of pollinators have undergone range contractions due to climate change (Kerr et al 2015). Limitations of dispersal and establishment may mean that many species of pollinators will not be able to keep up with predicted climate change scenarios and that climate change will exacerbate other threats to pollinators, including habitat loss (Settele et al 2016).

Revegetation project designers have an opportunity to address many of these effects of climate change during all phases of their projects. Doing so can increase the overall robustness and health of restorative plant populations, thereby increasing the success of projects. Ways in which designers might address climate change in their revegetation projects include the following.

## Diversifying Plant and Seed Sources

Revegetation efforts often occur where previously intact habitats have been disrupted or fragmented. If the area of disturbance is small, such as a localized landslide, the fragmentation might be minimal. It is often the case in roadside revegetation projects however, that the disruption to the original habitat can span miles and remain in place for decades. With these projects the original habitat has often been bisected, the hydrology disrupted, and the area might have experienced multiple disturbances or uses over time. In these instances, it's often valuable to collect plant materials along the entire length of the project area, including reference and adjacent sites, within each provisional seed zone.

Observations of, and collections from, mosaic populations of similar aged plants of the same species often provide opportunities to capture pre-disturbance genetic diversity and mimic natural gene flow patterns. Collecting and sourcing plant material in the direction of climate change conditions (i.e., up the elevation or latitudinal gradient) can potentially incorporate traits needed to compensate for predicted changes due to climate (Breed et al 2013). The [Seedlot Selection Tool](#) (SST) is a new mapping application that can assist designers in considering options for obtaining seed and matching seed sources to planting sites based on climatic information. The climates of the planting sites can be chosen to represent current climates, or future climates based on selected climate change scenarios. SST can also be used to identify planting sites that are appropriate for a particular seed source, now and into the future.

## Utilizing a Mix of Annual and Perennial Species to Meet Short- and Long-term Goals

There is often a desire to provide quick green up and stabilization to projects with annual

### **For the Designer**

The development of climate change decision support tools and their application to revegetation practices is rapidly evolving and generally beyond the scope of this manual. Designers may consult geneticists and other experts for the most current guidance and best management practices for their specific project goals and site conditions. Resiliency, diversity, and adaptability will remain important strategies for both short- and long-term revegetation success (Havens and others 2015). Monitoring will also be critically important for informing and adjusting revegetation practices in a changing climate.

plant species. Benefits of annual plants include quick germination and establishment, their seeds can be relatively inexpensive, and they are abundant in the current market. Disadvantages of annual species include the fact that they are short-lived, are often seeded in monocultures, their parental lineage and nativity may be difficult to ascertain, and they can out-compete perennial seeds. Importantly, in the context of climate change, using only annual species does not create a resilient plant community with long term persistence.

At times designers and managers can become frustrated at the slow germination and establishment of perennial seeds. Perennial plants tend to establish deeper, persistent roots and therefore provide longer lasting stabilization than do annual plants however. Native perennials are often seeded in a mixture of grasses and flowering forbs, a practice not impossible with annuals but one that seems underutilized. Due to their outcrossing, one disadvantage of long-lived perennial plants tend to be more susceptible to fitness impacts of inbreeding (Breed et al. 2013).

Developing seed mixes that contain both native annual and perennial seeds, proportional to what is appropriate for the individual project in order to avoid deleterious effects of competition and to mimic the vegetation of the surrounding environs, can exploit the best traits of each while minimizing risks.

## Developing Monitoring Plans with Climate Change in Mind

Regular assessments of plant survival and recruitment will assist designers and managers in understanding potential effects of climate change on revegetation success and outcomes. To facilitate adaptive management, keep detailed records on plant material sources, the planting scheme (e.g., seeding prescription or seedling numbers and density by species), and site preparation and seeding/planting methods. A clear summary in the monitoring report ([Section 6.6](#)) can help ensure this information is available to designers in the future.

Some projects, in particular those that involve wetland construction or enhancement, have monitoring plans ten years into the future or more. Given that plant responses to climate change trend toward upward or northward migration, stratified monitoring may be appropriate for some projects. Designers can stratify monitoring units by elevation band, latitudes, degree days, etc. in an effort to identify any changes in plant communities early. Including adaptive management strategies within the monitoring plan will help identify possible solutions to trends that are learned from monitoring.

### 3.3.2 SOILS

More than 95 percent of the counties in the United States have soil surveys either completed or in the process of completion by Natural Resources Conservation Services (NRCS).

Information from these surveys is available on the [Web Soil Survey website](#). By delineating the road project area on the Web Soil Survey map, a customized soils report specific to the project area is generated (Figure 3-5). Included in the report are profile descriptions, characteristics, and capabilities for each soil mapping unit. A typical profile description is provided and includes topsoil depth, soil textures, rock content, soil depth, available water holding capacity, permeability rates, and drainage classes. More detailed information on each soil series, such as laboratory results for nutrients, water retention curves, and other soil properties, is available at the NRCS [National Cooperative Soil Survey Soil Characterization Data website](#).



**Custom Soil Resource Report for Butler County, Kansas, and Chase County, Kansas**

**C**

**7170—Reading silt loam**

**Map Unit Setting**  
Soil area map unit number: 27psk  
Elevation: 1,000 to 1,000 feet  
Mean annual precipitation: 31 to 38 inches  
Mean annual air temperature: 54 to 57 degrees F  
Frost-free period: 175 to 230 days  
Farmland classification: All areas are prime farmland

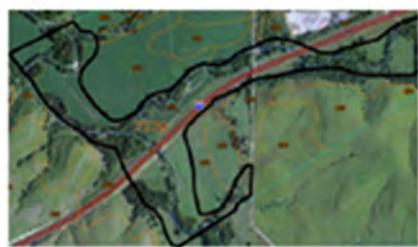
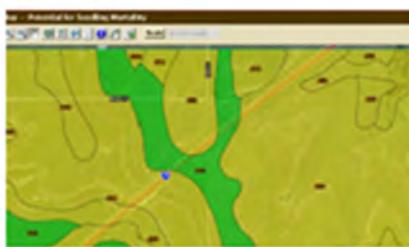
**Map Unit Composition**  
Relictic, nearly levelled, and smaller soils: 90 percent  
Minor components: 10 percent  
Estimates are based on observations, descriptions, and transects of the mapped

**Description of Reading, Rarely Flooded**

**Setting**  
Landform: Stream terrace  
Landform profile (two-dimensional): Tread  
Down-slope shape: Linear  
Across-slope shape: Linear  
Parent material: Alluvium

**Typical profile**  
Agric: 0 to 2 inches: silt loam  
A: 2 to 14 inches: silty clay loam  
Bt1: 14 to 28 inches: silty clay loam  
Bt2: 28 to 36 inches: silty clay loam  
C: 36 to 72 inches: silty clay loam

**Properties and qualities**  
Slope: 0 to 2 percent  
Depth to restrictive feature: More than 80 inches  
Natural drainage class: Well drained  
Haplotype: C  
Capacity of the moist rooting layer to transmit water (Ksat): Intermediately high (0.20 to 0.40 cm/h)  
Depth to water table: More than 100 inches  
Frequency of flooding: None  
Frequency of freezing: None  
Calcium carbonate: maximum in profile: Relatively low to very slightly saline (0.0 to 2.0 mmol/L)  
Salinity: maximum at profile: Relatively low to very slightly saline (0.0 to 2.0 mmol/L)  
Available water storage in profile: Very high (about 12.0 inches)

**B****D**

The information generated from these reports is for undisturbed soils, therefore the use of this information needs to be adapted to the type of disturbance expected to occur within the project. For example, on a project where topsoil is to be removed, it can be assumed that the soil remaining after construction will be the subsoil and not topsoil. The designer would consider the characteristics of the subsoil (B horizon) described in the Web Soil Survey report rather than the topsoil. If the topsoil is to be salvaged, the Web Soil Survey report can provide a general characterization of the topsoil that will be removed. It can also give a good description of an undisturbed reference site soil which may be helpful if restoration of the original site is the objective.

In addition, the Soil Data Explorer portion of the website creates a series of maps based on soil interpretations. Depending on the county the survey was conducted in, a wide range of maps are available, including suitability for hand planting, potential for seedling mortality, forest and range productivity, soil pH, hydrologic soil groups, depth to restrictive layers, and more.

Most lands administered by the USDA Forest Service have separate soils reports in addition to, or in lieu of, the NRCS soils report. These reports are often referred to as a Soil Resource Inventory report and can be obtained at the Forest Service District Office. The agency also maintains a national inventory and mapping ARC-GIS application that includes a soil database as well as information on geology, potential natural vegetation, and Terrestrial Ecological Units Inventory (TEUI).

**Figure 3-5 | Soils maps generated from the Web Soil Survey website**

The Web Soil Survey can be used to develop a custom soils report for most highway projects in the U.S. In this example, a 10-mile stretch of road is being proposed for reconstruction through several counties in Kansas. To understand the soils, the study area is delineated on the Web Soil Survey Interactive Map of the U.S. which generates a site-specific soil report (A). Within the report is a map of the project area with the locations of each soil mapping unit. For each mapping unit, the report describes the soil profile and soil characteristics (C). In this example, a prominent soil mapping unit in the planning area is "7170—Reading silt loam" (B). It is a deep stream terrace soil described in the report (C). The Web Soil Survey also allows the user to query the project site for areas of similar land capabilities or limitations. For this project site, one of the maps generated was areas where high seedling mortalities may be expected (D). Grouping of soil mapping units can also be used in the development of revegetation units.

### 3.3.3 VEGETATION PRE-FIELD ASSESSMENT

#### Ecoregions and Seed Zones

Ecoregions are defined areas in North America that have similar geographic, vegetative, hydrologic, and climatic characteristics. Several ecoregion systems are available; however, for the purposes of this publication, the ecosystem maps developed by the U.S. Environmental Protection Agency are used. The United States is divided into four ecoregion levels, each level representing increasing degrees of detail:

- **Level I**—12 broad ecoregions
- **Level II**—25 ecoregions
- **Level III**—105 ecoregions
- **Level IV**—967 ecoregions

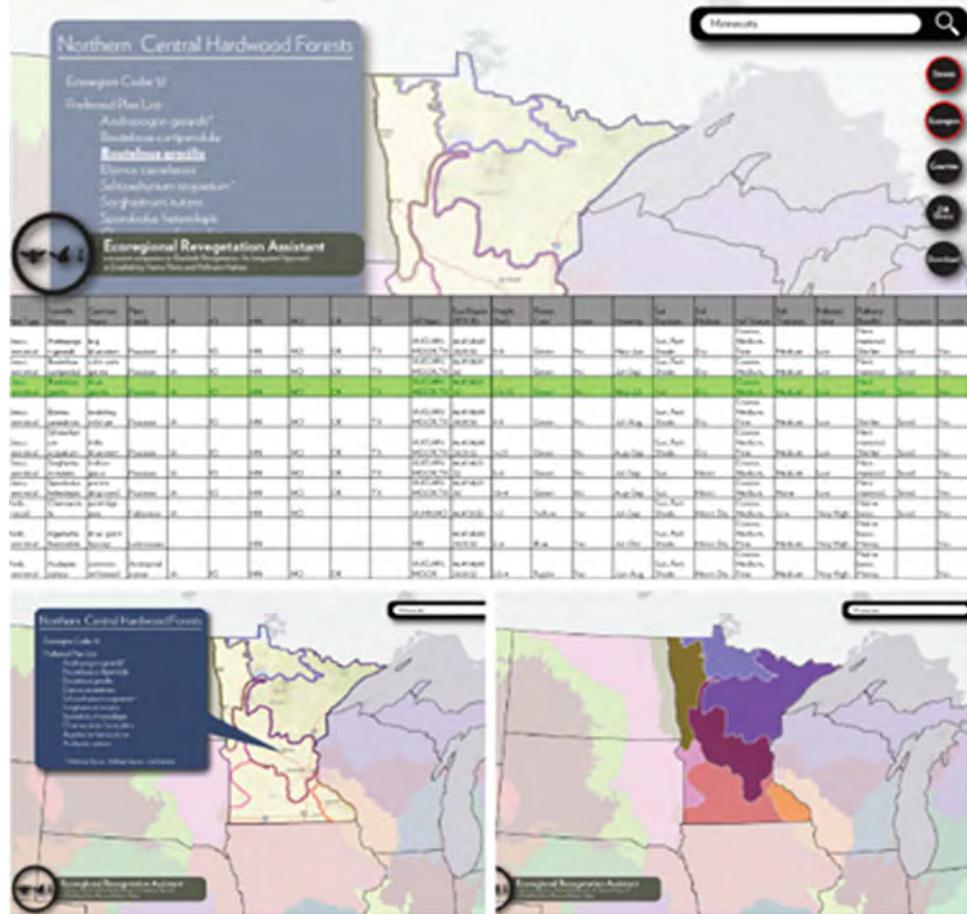
Because ecoregion maps identify areas with similar environmental characteristics, they are useful in planning, monitoring, information sharing, and management. The Level III ecoregion map has also been used to provide recommendations for seed use and movement of species and geographic areas where empirical genetic information is lacking (Bower et al 2014). Used in conjunction with climate data, such as minimum winter temperature and aridity, Level III ecoregions are a good starting point for guiding source selection of revegetation species. See [Chapter 5](#) for a more detailed discussion of seed zones and seed transfer guidelines.

#### Ecoregional Revegetation Application (ERA)

A spatially explicit online planning tool called the [Ecoregional Revegetation Application \(ERA\)](#) has been developed by the FHWA that will list the recommended workhorse and pollinator-friendly plant species for all EPA Level III ecoregions in the continental United States (Figure 3-6). The data used to create these lists were vetted by botanical experts and gathered from reliable sources such as the scientific literature, [USDA PLANTS](#) database, the USDA Agricultural Research Service [pollinating insect unit](#), and the [Xerces Society for Invertebrate Conservation](#). For each plant species, the ERA will provide attributes such as flowering season and preference for sun that will denote whether a given species is a workhorse (i.e., a reliable and available revegetation plant), or pollinator-friendly (e.g., supports larval or adult pollinators). For pollinator-friendly plant species the selector will denote which general groups of pollinators the plant species will benefit. Moreover, using data generated at the Chicago Botanic Garden (White et al 2016) the ERA will also indicate if a species of interest is commercially available. The first step in revegetation planning, specific to developing pollinator habitat, will be to use ERA to identify potential workhorse plant species for the project area. From this list, other plant species may be selected based on development of a site-specific pollinator working group ([Section 3.13.1](#)).

**Figure 3-6 | ERA**—An online planning tool to select workhorse and pollinator-friendly species

The [Ecoregional Revegetation Application \(ERA\)](#) allows the designer to obtain a list of appropriate workhorse and pollinator-friendly plant species for any location in the United States. Each workhorse species will display the plant attributes important for plant establishment and pollinator habitat enhancement. Please note that this image is a mockup of the future product and is displayed to demonstrate the concept of the online tool.



### 3.3.4 POLLINATORS PRE-FIELD ASSESSMENT

During the early stages of planning, it is important to identify "at-risk" pollinator species that may be affected by the project so that special measures can be taken to protect or enhance populations. A good source for identifying at-risk species is the [Xerces Society Red Lists](#) website. This site lists bees, butterflies, and moths that are at-risk by state. It also provides links to recovery plans for species listed under the Federal Endangered Species Act. [Nature-Serve Explorer](#) is a searchable database of plant and animal species in the U.S. that includes conservation status information. Other sources of information for at-risk pollinators are the wildlife and forestry departments of state and federal agencies.

Many good websites are devoted to pollinator species. The **BAMONA** (Butterflies and Moths of North America) (Figure 3-7) and **eButterfly** websites are citizen scientist websites that provide access to data about butterflies and moths in North America. Sightings of butterfly and moth species are shown on maps of the U.S. for many

species on these websites, including a description of each species. These maps allow the designer to determine if specific pollinator species are near the project area. The crowd-sourcing website, [BugGuide](#), is an online citizen science group that collects images of North American insects and offers an insect identification service for submitted images. Specific to the monarch butterfly is the [Monarch Joint Venture](#) website. This is a good resource for monarch butterfly biology, and the site also presents a map that displays current monarch butterfly sightings throughout the United States.

**Figure 3-7 | BAMONA website displays pollinator sightings for locations around the US**

### 3.3.5 ROAD PLANS

Understanding the design of the road project and how the site will appear after construction are important in developing a revegetation plan. Prior to a field review, consider conducting an evaluation of road plans and reports. Road plans show road cuts, road fills, drainages, ditches, disposal areas, abandoned roads, and engineered structures, which typically require different revegetation strategies. As discussed in the next section, these road components often become the basis or foundation of the revegetation unit map. Most road plans include a series of cross sections that provide slope steepness and shape, components that directly guide revegetation design. Many road projects include Storm Water Pollution Prevention Plans (SWPPP) that describe how water will be controlled, directed, and treated. These reports address the needs and expectations for soil cover and revegetation and it is helpful to understand them prior to developing a revegetation plan. Refer to Section 2.4 for how to read road plans, profiles, cross-sections, and typical views.

## 34 DEFINING REVEGETATION UNITS

Revegetation units are areas with similar revegetation treatments and environment (e.g., soils, climate, and vegetation potential). In mountainous terrain, there may be several revegetation areas in a mile of roadside due to changes in aspect, soil type, and road drainage. Roads in mid-western states, however, often have only one revegetation unit that may encompass much of a project area because of the uniformity of the landscape. The first step in developing a revegetation unit is grouping major soil types together with similar characteristics important for reestablishing native plant communities. For

example, a project site with a group of soils that are less than a foot deep would have a different set of revegetation treatments than deeper soils and for that reason would be identified as a revegetation unit based on soil depth. Grouping soils into revegetation units can also be done on websites such as the BAMONA, which can display specific locations where pollinators have been sighted. The search for a specific county in Maine, for example, brings up a list of butterfly and moth sightings.

As described in Figure 3-5, a soils map and report are created on the Web Soil Survey by delineating the project area on the interactive map of the U.S. At the same time, a map can also be produced that groups soils by similar capabilities to create distinct plant communities called “ecological sites”. The ecological sites section of the Web Soil Survey also lists the major native species for each ecological site for many parts of the U.S.

Revegetation units also designate areas that have the same revegetation objective. For example, a road project may include a constructed wetland for maintaining or improving water quality. In another area, the objective may be to enhance pollinator habitat. These areas would be designated as separate revegetation units because they would have different revegetation treatments and species, which might include a pollinator species mix for the pollinator habitat unit and an erosion species mix for the wetland unit. In addition, soil improvement treatments for the wetland would be developed to enhance wetland species and maintain the proper functioning of a constructed wetland. This may include creating manufactured soil that is specific to wetland species and water filtration.

Road components also play a large role in delineating revegetation units. In mountainous terrain, cut slopes and fill slopes are often designated as separate revegetation units because of the differences in soil depth, slope gradient, and road drainage between the two slopes. Table 3-5 shows revegetation units commonly associated with the components of a road.

The revegetation plan includes a revegetation unit map that locates revegetation units on the road project map (Figure 3-8). The revegetation plan further describes the soils, climate, and vegetation of each revegetation unit and how the revegetation objectives will be met.

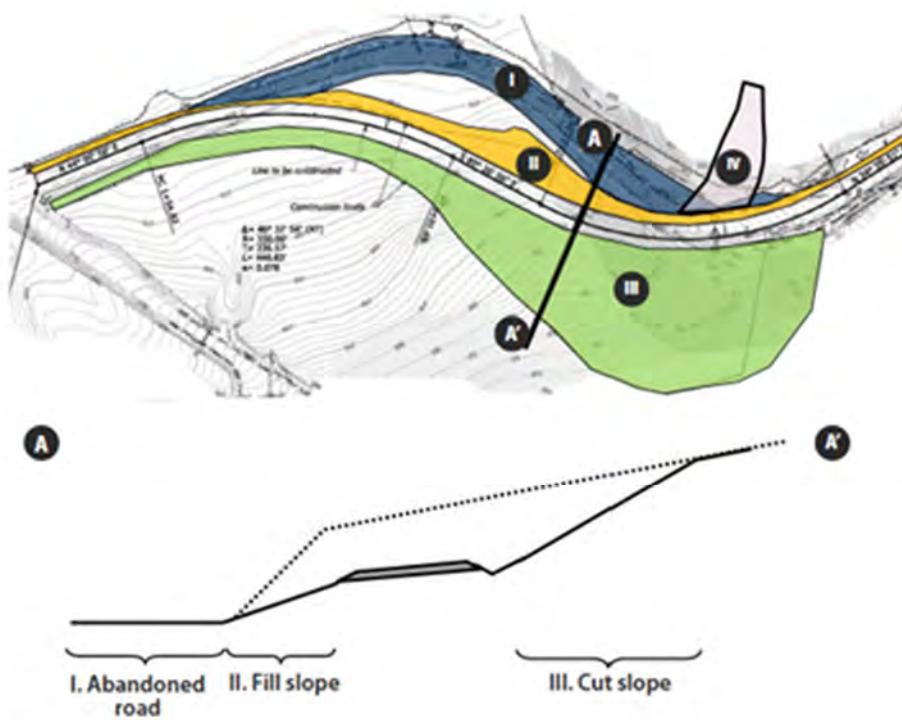
**Table 3-5 | Common revegetation units often associated with road components**

| Road Component    | Types of revegetation units   |
|-------------------|---|
| Cut slopes        | Cutslopes, living snow fence, pollinator habitat  |
| Ditches           | Amended ditches, wetlands, biorention swales, bioswales   |
| Shoulders         | Shoulders   |
| Fill slopes       | Fill slopes, filter strips, amended fill slopes, wave-attenuating bioscreens, living snow fence, pollinator habitat |
| Culvert outlets   | Vegetated culvert outlets   |
| Engineered slopes | Reinforced soil slopes, vegetated MSE walls, bioengineered slopes   |
| Bridges           | Stream restoration terraces and slopes  |

|                           |   |
|---------------------------|---|
| Disposal or staging areas | Restored areas, pollinator habitat            |
| Abandoned roads           | Restored obliterated road, pollinator habitat |

**Figure 3-8 | Example—cut and fill slopes often define revegetation units**

Major road components such as cut slopes, fill slopes and abandoned road sections often define revegetation units because of similar soils, objectives, and revegetation treatments. The road project in this example has four revegetation units that correspond to cut slopes, fill slopes, abandoned road, and bioretention swale. The objective for the road reconstruction project was to reduce existing curves for traffic safety while increasing pollinator habitat and decreasing the effects of road runoff and soil erosion on water quality. The road plans call for realigning the road, leaving an abandoned section of road, and greater area in cut slopes. During the planning phase, the revegetation units were identified based on soils and road objectives. Revegetation Unit I is the abandoned section of road to be restored by removing pavement, subsoiling the subbase, adding fill, and applying salvaged topsoil. Because the area is a pollinator habitat emphasis area, the seed mix will have more than 50 percent pollinator forb species. Flowering shrubs and trees will be planted in clumps. Salvaged logs from the road clearing operation will be randomly placed upright and on the ground for pollinator nesting habitat. Unit II includes fill slopes that will be amended with shredded wood to increase infiltration rates and capture and filtrate road surface runoff water. A low-growing native grass and forb seed mix will be applied. Unit III includes steep-cut slopes with high erosion potential that will be terraced and a seed mix primarily composed of grasses for erosion control will be applied in a bonded fiber matrix (BFM). Unit IV is a shallow draw where all road ditch water collects. It will be constructed as a bioretention swale to retain and filter sediments and road pollutants from the water before entering the stream. Wetland seedlings will be planted.



#### Revegetation Unit Descriptions

- I. **Abandoned road (pollinator habitat emphasis area)**—Deeper soils covered with 10 inches of salvaged topsoil. Large wood placed upright or down for pollinator nesting. Application of pollinator seed mix. Planting in clumps of flowing shrubs and trees.
- II. **Amended fill slopes**—Deeper soils with incorporated wood fiber for road water infiltration. Native grass/forb seed mix.
- III. **Cut slopes**—Steep, shallow soils with high runoff potential. Erosion control seed mix applied with a Bonded Fiber Matrix.
- IV. **Bioretention swale**—Engineered soils and slopes. Wtland species seed mix or plants.

## 3.5 IDENTIFYING REFERENCE SITES

Reference sites provide a natural model for possible vegetation outcomes and are important for defining DFC targets, as well as evaluating and monitoring the project following implementation (SER 2004). They can also be used to document the types and amounts of pollinator species that may be present in putatively natural environments near the project area.

Each revegetation unit can have at least one corresponding reference site that models the expected outcome or DFC target of the unit. Ideally the reference site shows how a revegetation unit might recover from disturbances at different points in time after

road construction. Reference sites can be considered a snapshot, or series of snapshots, of possible future outcomes. They demonstrate a point in time along a desirable developmental trajectory for a plant community. Using reference sites to understand the possible vegetative outcomes after disturbances will help the designer develop realistic expectations and provide a guide to the development of appropriate revegetation strategies for each revegetation unit. The most important aspect of reference sites is that they provide examples of plant communities from which designers can choose individual species for use in the revegetation project. The designer may sometimes choose to obtain baseline ecological data from several reference sites and then assemble DFC targets (SER 2004).

The two types of reference sites are disturbed and undisturbed. Disturbed reference sites are areas, typically old road cuts and road fills that have recovered, whereas undisturbed reference sites are relatively pristine sites that lack major disturbances in the recent past. For most road projects, disturbed reference sites are the most helpful because they represent sites that are ecologically similar to the revegetation unit and have recovered from disturbances similar to those planned. Undisturbed reference sites may also be used when ecological restoration is an objective or when suitable disturbed reference sites are not available. Disturbed reference sites can be categorized several ways:

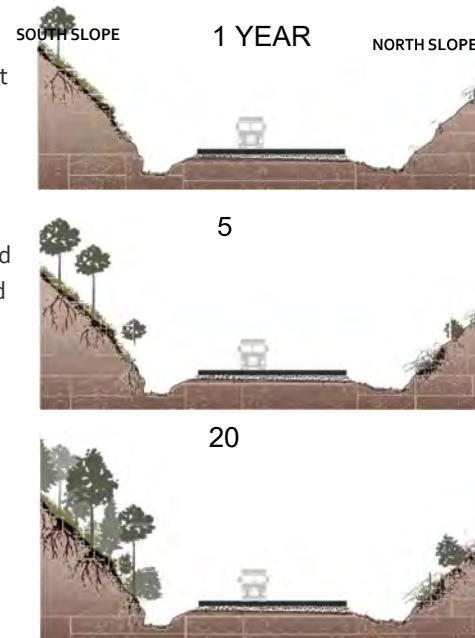
- Type of disturbance
- Length of time after the disturbance
- Desirability of the recovered vegetation

Disturbed reference sites can be old road cuts and fills, abandoned roads, ground-based logging sites, waste areas, rock source sites, ski runs, or other areas that have recovered from major soil disturbances. Disturbed reference sites often show a range of possible vegetative outcomes years after disturbance. Some sites will show good recovery and include stable soil, be visually pleasing, and populated by functioning communities of native plants. Others might show what can go wrong if revegetation is not carried out properly, including erosion, poor ground cover, weed infestation, and a lack of native vegetation. Understanding the conditions that lead to these vegetative outcomes can be a guide to avoiding them in the future.

Disturbed reference sites are the best models to demonstrate what is possible on the site in terms of vegetation, what trajectories succession might take (with or possibly without human intervention), and ways to effectively intervene in order to facilitate desired outcomes. Disturbed reference sites are invaluable in developing realistic DFC targets. Ideally, the type of disturbance on a disturbed site matches the type of road construction disturbance that will occur on the revegetation unit. For example, if the road cut after construction will be denuded of topsoil, then a disturbed reference site is to be found that lacks topsoil.

The stage of recovery is also important. It is ideal to find several disturbed reference sites that represent different successional stages of site recovery (Figure 3-9). For instance, a revegetation unit would ideally be represented by a recently disturbed site (several years after disturbance), a recently recovered site (5 to 25 years after disturbance), and a fully recovered site (over 25 years since disturbance).

While there is no such thing as a “pristine” plant community, an “undisturbed” reference site is an area that has not been heavily affected by ground-disturbing



**Figure 3-9 | Successional processes vary by microsite**

Successional processes and plant communities vary considerably based on microsite conditions. In this example, plant communities developed differently on north-facing and south-facing slopes.

activities. Undisturbed reference sites indicate the highest potential of a revegetation unit and are most often used as models when the goal is ecological restoration (re-creation of a plant community identical to that which existed before disturbance). The description of soil, climate, and vegetation in an undisturbed reference area often become the framework for the DFC target. It provides the designer with an understanding of those site characteristics or components necessary for healthy ecological

The process of selecting and describing reference sites is best accomplished in an interdisciplinary manner. The discussions that are generated between soils and vegetation specialists are generally far more thorough in knowledge and understanding of recovery processes than if surveys were conducted separately.

Disturbed reference sites can be located by driving the roads in and around the project area and finding areas that appear similar to the type of road construction being planned for a revegetation unit. Revegetated road cuts of various ages are good reference sites for cut slope revegetation units.

## 3.6 GATHERING FIELD INFORMATION

Surveying reference sites, as well as the project area in general, for soils, climate, and vegetation will provide baseline ecological data for developing the revegetation plan. If creating pollinator habitat is a revegetation objective, then reference sites and the project area are also surveyed to assess pollinator habitat quality and pollinator populations. This survey will provide insight into which pollinator species might be supported or enhanced by the revegetation project. The goal of the field survey is to obtain sufficient information from reference sites to realistically define DFC targets. During an initial survey, the appropriate survey intensity can be determined based on information needs and knowledge gaps. For example, if one of the revegetation objectives is to restore an abandoned road to a DFC target similar to a neighboring forest, then a survey of vegetation and soils of an undisturbed and disturbed neighboring forest would be conducted to describe the site characteristics and species composition.

Review the data used to define revegetation units (Section 3.3) prior to the field surveying of reference sites. Information regarding land ownership, site history, resources, and past and current management is also valuable. It is helpful to contact specialists who might have knowledge of the soils, vegetation, climate, and hydrology, as well as locals who can provide information on the site's history.

### 3.6.1 VEGETATION FIELD ASSESSMENT

The objective of assessing the vegetation of a reference site is to create a comprehensive species list that will guide in the selection of species to be used for revegetating the project area. A good method for compiling a comprehensive species list is to choose a representative cross section of each reference site that will characterize the range of plant species for that unit. Intuitively controlled surveys, such as these, maximize floristic knowledge yet are less time and effort intensive than complete floristic inventories. Usually, a few plant species are not easily identified in the field. Samples of these species can be brought back to the office for identification by specialists. If more detailed data collection is desired, such as a complete floristic inventory, surveys along transects or grids may be conducted.

Once species are identified, a comprehensive species list is developed for the project (Table 3-6). This list will be used throughout the life of the project for selecting species for plant

propagation, weed control, and plant protection. It includes some or all of the following attributes:

- **Species name (common and scientific)**—Because common names for plant species change throughout the country, it is important to list both the scientific and common names of each species. The [USDA PLANTS](#) database is a good source for obtaining the current scientific and common names. The database also includes the short species code symbol for field documentation.
- **Revegetation unit**—Identify the revegetation units where the species occurred.
- **Ecological settings**—Plants are identified by the ecological setting they are most commonly found in. A relative rating by temperature (cold, cool, warm, hot) and moisture (dry, moist, wet) gives a quick profile of the ecological setting. Some portions of the U.S. are covered by plant association maps or reports that were developed by federal agencies and are good sources for identifying the ecological setting of a species. Another way to describe the ecological setting of a species is by using the Ecological Site Assessment section of the [Web Soil website \(Section 3.3.2\)](#). This part of the website groups soil mapping units into ecological site units and dominant plant species.
- **Amplitude**—Ecological amplitude is the recurrence of a species across a wide array of ecological settings. A species found in all ecological settings would have a high ecological amplitude, while a species found in only one ecological setting would have a low ecological amplitude.
- **Abundance**—The quantity, dominance, or cover of a species found in a revegetation unit is the abundance.
- **Life form**—Group each species by life form: (1) tree, (2) shrub, (3) annual grass, (4) perennial grass, (5) annual forb, (6) perennial forb, or (7) wetland species (e.g., sedges, rushes)
- **Nativity**—Identify whether the species is native to the local area or introduced. The [USDA PLANTS](#) database identifies the nativity of all plant species in the U.S.
- **Weed status**—The [USDA PLANTS](#) database identifies the noxious weeds for each state. State-listed noxious weeds are found under the heading “Introduced, Invasive, and Noxious Plants” under the “PLANTS Topics” sidebar. Contacting the local State agency in charge of maintaining the lists, usually state departments of agriculture, is recommended.
- **Threatened and endangered species**—State and federal protected plants are found in the [USDA PLANTS](#) database under the heading “Threatened & Endangered” on the “PLANTS Topics” sidebar.
- **Succession**—Determine the seral stage a species is most commonly associated with: (1) early, (2) mid, (3) late, or (4) climax. Visiting reference sites and adjacent areas at different ages of recovery following disturbance will help provide an understanding of where each species fits into ecological succession. Figure 3-9 illustrates how plant communities develop differently over time depending on site conditions and successional processes.
- **Pollinator friendly**—Reference the [ERA](#) to determine if a species is beneficial to pollinators. Use the ERA lists of pollinators associated with each plant species to build a highly diverse pollinator community; flower color is also helpful in this regard—the more the merrier. Flowering periods for plant species can be obtained from the ERA. Use these to maximize the seasons flowers are available

to pollinators; a good minimum rule is three to five different species each of early, mid, and late bloomers. In addition to the ERA, other sources of reliable information such as species distribution maps by county from the [USDA PLANTS](#) website, or the [I-35 Corridor plants list](#) can give more detailed guidance to selection of appropriate species.

**Table 3-6 | A comprehensive species list**

Upon completion of a vegetation survey of the reference sites, a comprehensive species list is developed for the project. The spreadsheet will be used to determine the plant species mix that will be used in each revegetation unit.

| Scientific name              | Common name        | Revegetation unit | Amplitude | Abundance | Life form      | Nativity | Weed status | Threatened & endangered | Succession | Ecological setting | Pollinator friendly |
|------------------------------|--------------------|-------------------|-----------|-----------|----------------|----------|-------------|-------------------------|------------|--------------------|---------------------|
| <i>Achillea millefolium</i>  | Common yarrow      | 2,3               | High      | High      | Perennial Forb | Native   | –           | –                       | Early      | All                | Yes                 |
| <i>Abies grandis</i>         | Grand fir          | 1                 | High      | High      | Tree           | Native   | –           | –                       | Late       | All                | No                  |
| <i>Abies lasiocarpa</i>      | Subalpine fir      | 1                 | High      | Mod       | Tree           | Native   | –           | –                       | Late       | Cool               | No                  |
| <i>Agastache urticifolia</i> | Horsemint          | 2,3               | High      | High      | Perennial Forb | Native   | –           | –                       | Early      | All                | Yes                 |
| <i>Agoseris aurantiaca</i>   | Orange agoseris    | 2,3               | High      | Mod       | Perennial Forb | Native   | –           | –                       | Early      | All                | Yes                 |
| <i>Agoseris glauca</i>       | Pale agoseris      | 2,3               | High      | Mod       | Perennial Forb | Native   | –           | –                       | Early      | All                | Yes                 |
| <i>Agoseris grandiflora</i>  | Bigflower agoseris | 2,3               | High      | Mod       | Perennial Forb | Native   | –           | –                       | Early      | All                | Yes                 |
| <i>Allium acuminatum</i>     | Tapertip onion     | 4                 | Low       | Low       | Perennial Forb | Early    | –           | –                       | Early      | Wet                | ?                   |
| <i>Allium fibratum</i>       | Fringed onion      | 4                 | Low       | Low       | Perennial Forb | Native   | –           | –                       | Early      | Warm/Dry           | ?                   |
| <i>Allium macrum</i>         | Rock onion         | 4                 | Low       | Low       | Perennial Forb | Native   | –           | –                       | Early      | Wet                | ?                   |
| <i>Allium madidum</i>        | Swamp onion        | 4                 | Low       | Mod       | Perennial Forb | Native   | –           | –                       | Early      | Wet                | ?                   |

## 3.6.2 SOILS FIELD ASSESSMENT

Understanding the soil characteristics of each reference site is essential to effectively define DFC targets and develop revegetation treatments. The soils report that is generated from the [Web Soil Survey website](#) for a road project gives a close approximation of the characteristics of undisturbed soils for the project area and are to be checked in the field. It is important to remember that the soil condition after road construction will not resemble the natural soils found in the soil survey. For this reason, it is important to find disturbed reference sites that are similar to the disturbance of the revegetation unit. The following information can be collected for topsoil and subsoil:

- Soil texture
- Rock fragments
- Rooting depth
- Topsoil depth
- Nutrient levels
- Soil structure
- Litter and duff layers ([Section 5.2.3](#), see [Litter and Duff](#))
- Site organic matter
- Infiltration rates

## 3.6.3 POLLINATOR FIELD ASSESSMENT

### Habitat Assessment

During the field review, an assessment of the pollinator habitat and pollinator species populations may be conducted for the project area. The pollinator habitat assessment includes evaluating the road project plans within the context of the larger planning area for creating habitat supportive for general and at-risk pollinator species. Table 3-7 is a checklist that can be used to identify those factors important for creating pollinator-friendly habitat. Factors that improve pollinator health or habitat can be considered in design plans while factors that limit pollinator health can be mitigated or improved through management treatments or practices, presented in [Section 3.9](#).

One approach to using this checklist is for the designer to visit the project site during planning and evaluate both the current condition of the roadsides and the undisturbed reference sites ([Section 3.5](#)). Ideally these assessments can be conducted during the same visits as the vegetation assessment ([Section 3.6.1](#)) and the soil assessment ([Section 3.6.2](#)). Evaluating the quality of pollinator habitat of the existing roadsides will give some indication of what the designer can expect if standard construction practices are employed. Comparing these findings to those of undisturbed reference sites gives the designer an idea of what is possible. Comparisons of the current condition and the reference site can help the designer develop a revegetation plan for improving pollinator habitat.

The Pollinator Habitat Assessment checklist provides eight characteristics important for most pollinator habitats. The designer may want to modify the checklist based on project objectives, pollinators of interest, and the unique ecology of the roadside. Another valuable use of the checklist is that it can be used to develop target DFCs for the revegetation project ([Section 3.7](#)). For example, a DFC target from this list may state that “at least three native species will be in bloom during spring, summer, and fall”.

Field visits during the growing season would be conducted after revegetation to [Roadside Revegetation: An Integrated Approach to Establishing Native Plants and Pollinator Habitat](#)

### 3—PLANNING

determine if this target was met.

## Pollinator Monitoring

Revegetation projects, especially those specific to improving pollinator habitat, may require a pre- and post-construction assessment of pollinator species. As discussed earlier, it may be helpful to know which pollinators are present prior to project design. If imperiled pollinator species are suspected in the project area, it is important to survey for pollinators before undertaking construction. See [Section 3.3.4](#) for resources for determining imperiled pollinators in the project area and check with the state Natural Heritage Program and land managing agency, as applicable, for a list of species of conservation concern.

**Table 3-7 / Pollinator habitat assessment checklist**

This guide can be used to assess the pollinator habitat conditions at any time during the life span of a road project. The checklist gives eight characteristics important for most pollinator habitats, however, the designer may want to modify the checklist so that it addresses the climate, soils, vegetation, and pollinator species of interest specific to the project area and project objectives. It is important when using the checklist to identify the purpose of the assessment, such as whether it is describing a reference site, pre-disturbance or post-revegetation conditions.

| Components of pollinator habitat | Steps to improve pollinator habitat conditions   |
|----------------------------------|--|
| Nectar/Pollen sources            | <ul style="list-style-type: none"><li>▪ At least three blooming species in each season (spring, summer, fall)</li><li>▪ Species have overlapping and sequential bloom periods</li><li>▪ Presence of both wildflowers and woody blooming plants</li><li>▪ Aim for 45 percent plant cover of blooming plants available across seasons</li></ul>  |
| Breeding habitat                 | <ul style="list-style-type: none"><li>▪ Host plants present for target butterfly and moth species</li><li>▪ Presence of vegetation, leaf litter that can serve as egg-laying sites for other species. At least three blooming species in each season (spring, summer, fall)</li></ul>  |
| Nesting habitat                  | <ul style="list-style-type: none"><li>▪ Patches of bare ground present at site</li><li>▪ At least three species of woody plants or pithy stemmed plants that support tunnel-nesting bees</li><li>▪ Snags or downed wood present in safe location for traveling public</li><li>▪ Unmown bunch grasses present throughout growing season to support bumble bee nests</li></ul>   |
| Water source                     | <ul style="list-style-type: none"><li>▪ Water sources such as culvert outlets, ditches, draws, gullies, intermittent streams, and topographic enhancements</li></ul>   |
| Shelter and overwintering        | <ul style="list-style-type: none"><li>▪ Trees and/or shrubs present at the site</li><li>▪ Diversity of grasses to provide vegetation structure</li></ul>   |
| Vegetation management            | <ul style="list-style-type: none"><li>▪ Mowing and herbicide use is timed to reduce impact to pollinator life cycles</li><li>▪ Mowing and herbicide use is timed to support plant diversity</li><li>▪ Herbicide use in roadside beyond the safety strip is targeted to noxious and nonnative plants and other undesirable species rather than using broadcast applications</li><li>▪ Weeds are controlled before and during construction to aid in plant establishment, as well as during the establishment phase</li><li>▪ If haying (mowing and removal of biomass) by adjacent landowners is permitted on the roadside, it is conducted once at the end of the growing season</li><li>▪ Prescribed fire and prescribed grazing are timed carefully to avoid damage to life cycles of imperiled or sensitive species of pollinators</li><li>▪ Brush removal is tapered to soften transition to denser vegetation at edge of ROW, opening up the canopy to allow understory plants to bloom and leaving some stems or other sites for tunnel-nesting bees</li><li>▪ Biological and cultural control methods are integrated into vegetation management to reduce use of herbicides to control noxious and invasive weeds</li></ul> |
| Landscape connectivity           | <ul style="list-style-type: none"><li>▪ Site increases landscape connectivity by linking existing habitat parcels on nearby land</li><li>▪ Site increases roadside connectivity by linking roadside habitat</li><li>▪ Site increases diversity within the landscape and benefits agricultural activity on adjacent lands</li></ul>   |

**Road mortality**

- Site is not isolated within areas of high road density in which there are multiple barriers to pollinator movement
- Sites have reduced mowing and high plant diversity
- Clear zone width is increased within AASHTO guidelines along roadsides with high salt use and high volumes of traffic (reduces exposure of pollinators to salts, heavy metals)

It can also be helpful to monitor pollinators before construction and following revegetation in order to assess the success of the project or to perform comparisons of the effectiveness of seed mixes or revegetation techniques for different pollinators.

Monitoring techniques for these assessments are discussed in [Section 6.4](#).

## 3.7 DEFINING THE DESIRED FUTURE CONDITION

Once revegetation units and corresponding reference sites have been described, the DFC targets can be defined for each unit. The DFC target is the translation of the revegetation objectives into measurable goals for each revegetation unit. Specifically, the DFC target defines the desired or expected composition of vegetation at a particular point in time after the completion of the revegetation work.

An example DFC target would be, “one year after seeding, vegetative ground cover will be 40 percent and of this cover, 50 percent will be composed of native forb species beneficial to pollinators.” Stating expectations in this manner will (1) clarify how the site will appear after treatments, (2) narrow down the appropriate revegetation treatments to meet the DFC target, and (3) define measurable criteria, or thresholds, for monitoring the success of a project.

Commonly stated DFC criteria include the following:

- Vegetative ground cover
- Bare soil cover
- Native grass cover
- Number of species of native grasses
- Native forb cover
- Number of species of native forb species
- Seedling survival
- Seedling density (plants per area)
- Tree growth (height per year)
- Coarse pollinator diversity
- Pollinator abundance

Stating the DFC in measurable terms and with a time frame ensures that the project team, regulatory agencies, and the public have similar expectations of how the project will appear in the years following its completion. Quantifying the objectives also focuses the monitoring plan to collect only the information necessary to determine if project objectives were met. For example, if one of the objectives is erosion control, a DFC target might be, “the amount of bare soil one year after road construction will be less than 20 percent.” Monitoring procedures would focus on measuring bare soil after one year. If another objective is to increase pollinator species, then a measurable threshold for success might be, “an increase of 50 percent pollinator abundance over reference site populations three years after seeding.” Monitoring, in this case, would measure general pollinator types in reference sites and revegetation unit three years after completion of the project.

Another benefit in defining DFC targets is that it often will generate a discussion of

### 3—PLANNING

whether they are achievable without investing in soil improvement or additional plant establishment methods. Unless DFC targets are stated and discussed, individual team members will develop their own concept of what success looks like. For example, a road project was being proposed next to a river with high fisheries values. The Storm Water Pollution Prevention Plan that had been prepared for the project stated that the cut slopes would have 100 percent ground cover, which would result in very low sediment delivery to the stream after construction. Team members discussed this DFC target and concluded that it was unachievable because of the lack of topsoil and shallow soils. This left the team the choice of either modifying the DFC target or improving the soil quality.

When developing DFC targets, it is important to consider the plant community succession that is likely to occur on each revegetation unit. In some cases, planting early seral species at the outset may work. By year 3, when the early seral species begin to decline, the late seral species may be increasing. In other cases, it may be necessary to intervene immediately after seeding or planting in order to meet the revegetation objectives of the project. For example, short-term revegetation planning might call for seeding grasses and forbs to stabilize the site. One year later, the site might be revisited to remove any invasive species before they produce seeds. Two years later, the site might be revisited to interplant conifers and shrubs. These three intervention points (seeding, weeding, and planting trees) speed succession in the desired direction.

## 3.8 IDENTIFYING LIMITING FACTORS TO PLANT ESTABLISHMENT

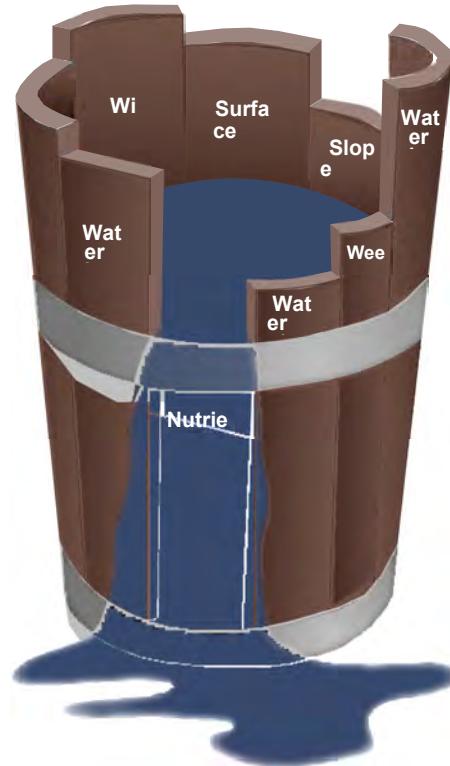
Site conditions that affect plant establishment and growth are referred to as limiting factors (Figure 3-10). Odum (1971) defines limiting factors as "any condition which approaches or exceeds the limits and tolerance (of a plant species)." He further states that "the chief value of the concept of limiting factors lies in the fact that it gives the ecologist an 'entering wedge' into the study of complex situations. Environmental relations of organisms are apt to be complex, so that it is fortunate that not all possible factors are of equal importance in a given situation or for a given organism." Not only does this simplify a complex analysis, it means the designer will need to systematically consider all site factors, focusing on those of greatest concern. For example, typical revegetation treatments conventionally call for the blanket use of fertilizers without assessing if nutrients are really limiting to plant growth. In many cases, other limiting factors to revegetation, such as low rainfall, compacted soils, low organic matter, and poor rooting depth, are more limiting. Applying fertilizer without an assessment of limiting factors, is like a physician prescribing medicine before the patient has been properly diagnosed. While soil fertility is often important on many highly-disturbed sites, it might not be the primary limiting factor to revegetation on this particular site.

This manual has grouped the site characteristics essential for plant growth into six limiting factors to revegetation typically encountered in the United States. These factors are further broken down into component parts, or parameters (Figure 3-11). In this section, each limiting factor to revegetation and corresponding parameters are discussed in terms of why they are important to plant establishment and growth, how they are assessed, and what mitigating measures or treatments can be applied to make them less limiting.

The information used in defining limiting factors for each revegetation unit can be obtained from the surveys and reports conducted during the field surveys. It is important that an assessment of every limiting factor and corresponding parameter be made for each revegetation unit based on the expected condition of the site after road

### *For the designer*

Defining the limiting factors is an essential process in developing a revegetation plan because it identifies, from a multitude of site factors, only those that are roadblocks to successful revegetation. A Limiting Factor table is available in this Planning workbook.



**Figure 3-10 | Limiting factors to revegetation**

Limiting factors to revegetation can be displayed as unequal boards of a barrel. Water can only be held to the level of most limiting factor.

### 3—PLANNING

construction. Figure 3-11 can be used as a checklist—a means of quickly assessing a site for its potential to grow plants while preventing the possibility of overlooking factors important for successful revegetation. Much like pilots or surgeons use checklists, the designer can use the limiting factors list to simplify a “complex situation” and quickly identify what is important from what is not.

From the limiting factors identified for a project, a list of mitigating measures is developed. Mitigating measures are the site treatments that will reduce or eliminate the site conditions limiting to revegetation. For example, if rainfall is limiting, a mitigating measure is to irrigate. There are usually several ways to mitigate each limiting factor. While some of the mitigating measures might seem impractical for a particular revegetation project, they are at least considered.

**How to approach this chapter**—This chapter is organized by limiting factors. It is not important to read the whole chapter, but it can be helpful to read portions, especially those that pertain to the limiting factors identified for a project. Each limiting factor section discusses how to assess or record the factor and how to mitigate for it. Many of the mitigating measures discussed in this section are presented in further detail in [Chapter 5](#). The mitigating measures described in this report are not a complete list. Consider other practices based on local or regional experience. [Section 3.12](#) discusses how to select the appropriate site improvement measures from the mitigating measures list.

#### 3.8.1 WATER INPUT

Water input refers to the moisture supplied to the soil through rainfall, snowmelt, and road drainage. This moisture recharges the soil and becomes the primary source of water for plant establishment and growth. Water input is influenced by obstacles that capture, or intercept, water before it can enter the soil, including standing live or dead vegetation and soil cover (litter, duff, and mulch). Surface infiltration rates also regulate entry of surface water. If infiltration rates are low, water that would normally enter the soil runs off the surface and is unavailable.

The primary site factors that affect water input are as follows:

- Precipitation
- Rainfall interception
- Infiltration
- Road drainage

In the western United States, water input is at its lowest levels from late spring through early fall. This is also the period when plants need the most soil moisture for survival and growth. During the summer, when water input is low, the soil profile dries out as vegetation withdraws moisture. As soil moisture is depleted, plants cease growing; if soil moisture is not recharged, plants will go into dormancy or die. It is critical that any water from precipitation arriving during the dry season enters the soil and is stored for later plant use.

#### Precipitation

In wildlands revegetation, the only source of soil water comes through precipitation in the form of rainfall or snowmelt. In the western United States, this typically occurs from late fall through mid-spring, a period when plants are dormant and least able to utilize soil moisture for growth. Water that is not stored in the soil during these events is lost from the site either to ground water or runoff. The period when plants need soil

#### Water input

- Precipitation
- Interception
- Infiltration
- Road drainage

#### Water storage and accessibility

- Soil texture
- Rock fragments
- Soil structure
- Rooting depth
- Mycorrhizal fungi

#### Water loss

- Wind
- Aspect
- Competing vegetation
- Soil cover

#### Nutrient cycling

- Topsoil
- Site organic matter
- Nitrogen and carbon
- Nutrients
- pH and salts

#### Surface stability

- Rainfall
- Wind
- Freeze/thaw
- Soil cover
- Surface strength
- Infiltration
- Slope gradient
- Surface roughness
- Slope length

#### Slope stability

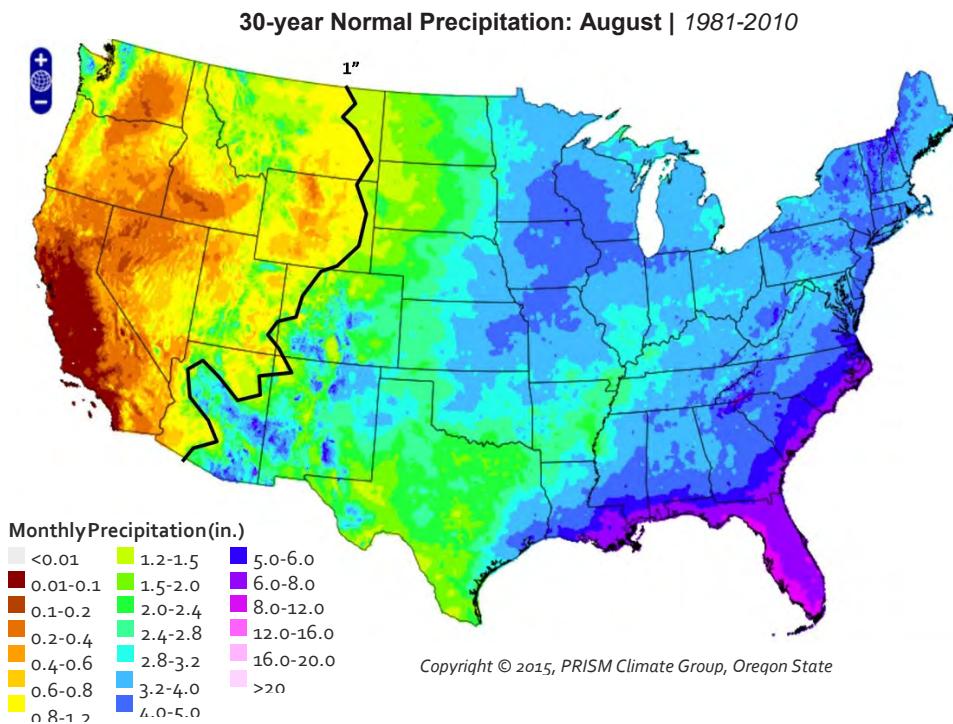
- Permeability
- Restrictive layer
- Water input
- Slope length
- Slope gradient
- Soil strength

### 3—PLANNING

moisture the most occurs during a five- to six-month period, from April through October. For most sites in the western United States, the amount of moisture that occurs in this period is less than a quarter of the total annual rainfall. Precipitation is also extremely low during the middle of the summer (Figure 3-12).

Plant survival and growth hinge on the precipitation that occurs in the years following planting or seeding. Very low precipitation in summer is common in the western United States (those areas west of the 1-inch line) but less common in the eastern United States, as shown in this map of the U.S. which depicts normal precipitation over a 30-year period for the month of August. Much of California, Oregon, Nevada, Idaho, and Washington receive less than one-half inch of rainfall in August as compared to many areas in the mid-western and eastern United States that receive more than 4 inches of rainfall in the same month. This pattern is typical of other months during the growing season (map generated from PRISM).

Vegetation native to the western United States has evolved to compensate for the limited supply of moisture during the growing season (Figure 3-13). During spring, when soils are charged with moisture from winter precipitation and soil temperatures increase, plants produce new roots, followed by new foliage. As the soil dries out and plants undergo mild moisture stresses, new root and foliage growth cease. During summer, soil continues to dry and plants respond to even greater moisture stress by shutting down their physiological functions and becoming dormant. By mid to late summer, when available soil moisture is depleted and evapotranspiration rates are high, plants will show stress symptoms (browning, loss of needles and leaves); under extreme circumstances, plants will die. By late summer and early fall, rain returns and the soil slowly moistens again, reducing plant moisture stress and signaling plants to grow new roots.



**Figure 3-12 | 30-Year Normal Precipitation: August (1981–2010)**

Plant survival and growth depend on the precipitation that occurs in the years following planting or seeding. Drought periods in the summer are common in the western United States (west of the 1-inch line) but less common in the eastern United States, as shown in this map of the U.S. which depicts normal precipitation over a 30-year period for the month of August. Much of California, Oregon, Nevada, Idaho, and Washington receive less than one-half inch of rainfall in August as compared to many areas in the mid-western and eastern United States that receive more than 4 inches of rainfall in August.

©2015 PRISM Climate Group, Oregon State University

The primary characteristic of precipitation for plant survival is the quantity of rainfall delivered in each storm event during the dry season. Storm events that deliver more than one-quarter inch of rainfall can wet the surface portion of the soil profile and reduce plant moisture stress. Precipitation events that deliver less than this amount rarely supply enough water to enter the soil, especially if interception and runoff rates

### 3—PLANNING

are high.

**How to Assess Precipitation**—Average monthly rainfall for a project area can be accessed through climate websites, as discussed in [Section 3.3.1](#). For more site-specific information, precipitation can be collected on-site using rain gauges that capture and record precipitation.

Two types of precipitation gauges are available: digital and non-digital. The advantages of digital gauges are that they record the amount and intensity of rainfall at the time it occurred; the downside is cost (although prices are falling). Many types of digital rain gauges are available, ranging in price and quality. It is important to select a digital rain gauge that is rugged, self-maintaining, and can record for long periods of time. In addition, many remote stations have the capacity to transmit data via the web so it is easy to keep current on weather events.

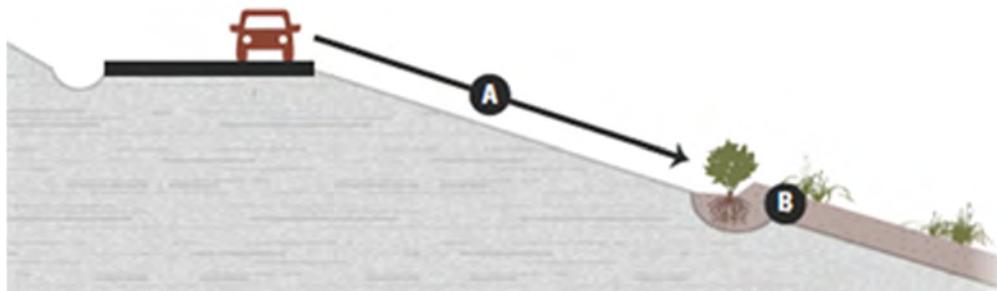
Non-digital rain gauges are basically cylinders that collect and store precipitation while preventing evaporation. The gauges are monitored by simply measuring the water in the cylinder. The disadvantage of non-digital rain gauges is that they only report the rainfall that has occurred between site visits. They do not provide the dates when rainfall occurred and do not record rainfall intensities.

**Mitigating for Low Precipitation**—For projects where rainfall is limited during the growing season, making the most of rain and snowmelt that falls throughout the year is an important art of successful revegetation planning. In most cases, supplemental watering will not be feasible. However, if very little water input occurs during the summer, temporary supplemental water could be considered during plant establishment. This can take an active form, such as irrigation, or a passive form, such as redirecting surface water to planted seedlings.

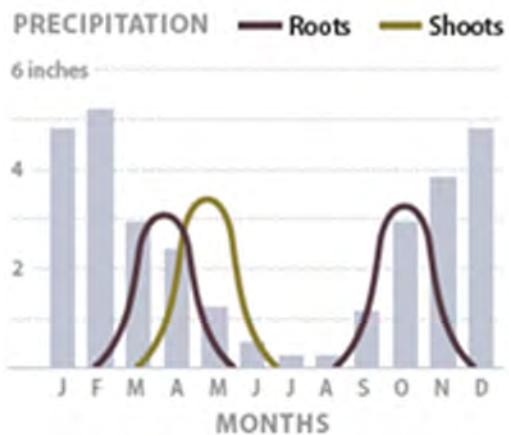
**Irrigation**—Irrigation can be expensive, and it is generally used only on projects with high visibility or when rapid establishment is necessary for slope stability. These are projects where revegetation objectives include minimizing the risk of seedling failure or enhancing vegetation growth.

Several basic types of irrigation systems are used in roadside revegetation. They are grouped into fixed systems, such as overhead sprinkler and drip irrigation, and manually applied systems. Fixed systems are discussed in [Section 5.5.5](#) (see [Drip Irrigation](#)). Manual systems involve water being delivered directly to each plant, either from a hose or water container.

If only a few applications are necessary, the entire project can be irrigated by hand.



Personnel can water each seedling or seeded area using a water truck or hydroseeding



**Figure 3-13 / Root and shoot growth**

In the western United States, root and shoot growth occur when moisture is available in the spring. Growth ceases by early summer when there is very little rainfall. Root growth takes place again from late September through November when soils are recharged by fall rainstorms.

**Figure 3-14 / Fill slope microcatchments**

Fill slope microcatchments take advantage of the low infiltration rates of compacted fill slopes. Water moves off impervious road surfaces and compacted road shoulders during rainstorms (A), and is captured in berms or flattened areas below the road shoulder (B). If this area is ripped and amended with organic matter (e.g., filter strips, amended fill slopes), it becomes a very good environment for establishing shrubs and trees. Soil and compost berms and/or flattened areas are also catchments for sediments.

### 3—PLANNING

equipment (with water only), although being mindful of not pulling hoses over establishing plants can help ensure plant survival. Creating basins around seedlings will pond the surface-applied water and keep it concentrated in the seedling root zone.

However, a better way to be certain that water will be delivered directly to the roots is to integrate the deep pot irrigation system into drip or manually applied irrigation methods (Bainbridge et al 2001). Pipes made from polyvinyl chloride (PVC) or other materials are placed at depths of 1 to 2 feet beside the seedling at the time of planting. The pipes are then filled with water when the soils dry out in summer. The advantage of deep pipe irrigation is that water is delivered directly to the root system and, because the water is placed deeper in the soil, roots are forced to extend farther into the soil for moisture. Refer to [Section 5.5.5 \(see Deep Pot Irrigation\)](#) for how to install this system.

For any irrigation method, it is important to monitor the wetting pattern of each irrigation. This will ensure that water is applied at the appropriate rates. Digging a hole where the water has been applied at least several hours after irrigation will show how far the water has moved into the soil profile. Duration of irrigations can be adjusted accordingly.

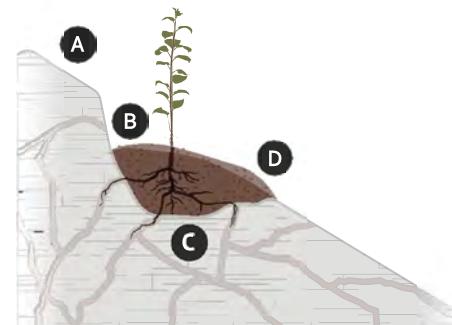
**Water Harvesting**—Water harvesting is the alteration of local topography to capture runoff water and concentrate it in areas where it can be used by plants. Water harvesting designs can be applied to roadside revegetation in several ways. They include, but are not limited to, contour bench terraces, runoff strips, and fill slope microcatchments. Fill slope microcatchments take advantage of water that drains off road surfaces and shoulders during intense rainstorms by capturing runoff in berms or depressions created at the base of the road shoulder (Figure 3-14). Shrubs and trees planted in these catchment areas will receive greater soil moisture. Contour bench terraces are structures carved out of cut and fill slopes that collect and store runoff water. When filled with topsoil or amended soil, they are referred to as planting pockets. Figure 3-15 shows how planting pockets collect water. Even very low rainfall events, which would normally be of insufficient quantity to moisten the soil surface, can recharge soil in planting pockets and fill slope microcatchments. Sediments will also be deposited on the benches and pockets during rainstorms, building the soil up over time and reducing soil erosion. Water harvesting not only supplies additional water to plants but reduces sediment and peak flow water to the stream system. Road practices that intercept water and sediments from the road surface for water quality improvement are also a source of additional water for plant growth. These include amended slopes, filter strips, amended ditches, bio-retention swales, and constructed wetlands. In addition, some of these structures create surface-water sources for pollinator species.

## Rainfall Interception

The amount of water entering the soil profile from a rainfall event can be significantly reduced by the interception of live or dead vegetation cover. Rainfall is captured through a series of layers, beginning with the tree and shrub canopy, the ground cover, litter, and duff, and is returned to the atmosphere through evaporation. During the dry season, moisture from a low rainfall event might not reach the soil.

**How to Assess Rainfall Interception**—Rain- fall interception can be determined by the soil cover and vegetation that exist on the site after construction. In most cases, there will be very little vegetation and ground cover. It is therefore important to understand the effects of various types of ground cover used in revegetation on the rainfall interception. The depth and water-holding capacity of the material will determine the effect on water input.

Water-holding capacity of a surface cover can be measured through testing labs



**Figure 3-15 | Planting pockets**

Planting pockets are designed to capture water from upslope runoff (A), which collects in a slight depression (B). Captured water wets the soil after each rainstorm and drains into the fractured bedrock (C). Soil is protected from surface erosion on the downhill side of the pocket with mulch or erosion fabric (D).

### 3—PLANNING

specializing in composts. Alternatively, it can be measured by collecting the soil layer (duff, litter, mulch) and drying it at 230° F in a drying oven (a crockpot can be used, adjusting the temperatures using a meat thermometer). When the sample is dry, it is placed in a 5-inch-long by 3-inch-round PVC pipe with a flat piece of cardboard secured to the bottom of the tube to prevent the material from falling out. The PVC pipe is weighed and placed in a bucket that is filled with water to the top of the pipe. The sample is removed and allowed to drain.

After several hours, the pipe is reweighed. The material in the pipe is removed and the pipe plus cardboard is weighed. The moisture holding capacity of the material (by percentage of dry weight) is as follows:

$$\frac{(\text{Wet weight of container and cardboard}) - (\text{Dry weight of container and cardboard})}{\text{Dry weight of container and cardboard}} \times 100$$

$$(\text{Dry weight of container and cardboard}) \times 100$$

Figure 3-16 can be used to approximate how much rainfall is intercepted based on the moisture-holding capacity of the soil cover.

**Mitigating for High Rainfall Interception**—It is important to consider the water-holding capacities of the mulches or soil covers to be used, especially on arid sites. Highly decomposed, fine-textured composts have high water-holding capacities compared to coarser-textured composts and hold more moisture after a rainstorm. Coarser materials, such as shredded wood, bark, wood chips, and wood strands, hold less water, allowing more rainwater to enter the soil. In addition, because fine-textured composts hold more water than course mulches or soil, they are good growing media for desirable native plants as well as undesirable weed species. The question to ask when selecting a soil cover is whether it is to be used as a mulch or a growing media. If a mulch, then consider using a coarse material; if growing media, consider using a fine compost.

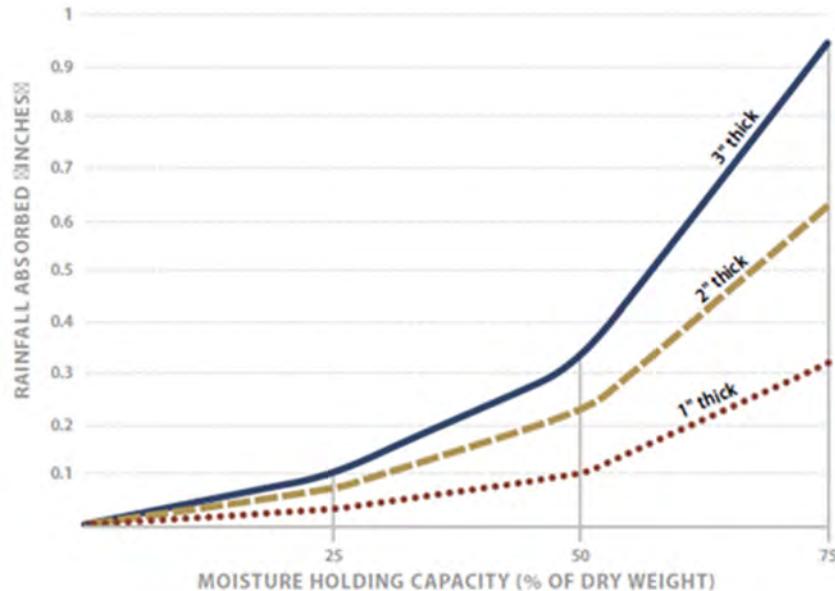
### Infiltration

Infiltration is the ability of the soil surface to absorb water from rainfall, snowmelt, irrigation, or road drainage. When infiltration rates are slower than the amount of water applied to the surface of the soil, runoff will occur and this water will not be available for plant uptake. In addition, runoff can detach and transport soil, causing soil erosion, decreased water quality, and increased peak flows. Refer to Section 3.8.5 (see **Infiltration Rates**) for a discussion of infiltration rates on surface stability.

The size, abundance, and stability of soil aggregates in the surface soil determine the infiltration rates. Large stable pores created by worms, insects, and channels left behind from decayed roots will absorb water quickly and have high infiltration rates; soils that have been compacted, had their topsoil removed, or are low in organic matter will have poor infiltration rates.

Under undisturbed conditions, infiltration rates are typically high, especially where a litter and duff cover exists. When soil cover is removed, the impact from rain splash can seal the soil surface, creating a crust that will significantly reduce infiltration rates. Infiltration rates are also reduced when the soil is compacted by heavy equipment or traffic.

**How to Assess Infiltration**—The most accurate method to measure field infiltration



**Figure 3-16 / Moisture holding capacity of mulch or litter**

The amount of rainfall intercepted by soil cover (e.g., mulch or litter) is dependent on its water-holding capacity and thickness.

### 3—PLANNING

rates the rainfall simulator (Section 3.8.5, see Infiltration Rates). This equipment is calibrated to simulate the appropriate drop size and impact velocity of many rainfall events (Grismer and Hogan 2004). The rainfall simulator is expensive to operate and is not routinely used by the designer. The most common application for this technology is in comparing different mitigating measures, such as mulches and tillage methods, on infiltration capacity.

In the absence of rainfall simulation tests, infiltration rates are inferred by measuring soil strength using a soil penetrometer, bulk density measurements (Section 3.8.6, see Soil Strength), and from site characteristics such as visual observation of compaction and the percentage of soil cover. For most construction activities that remove surface cover or disturb the topsoil, it can be assumed that infiltration rates will be reduced to levels that will create overland flow under most rainfall intensities.

## Mitigating for Low Infiltration Rates

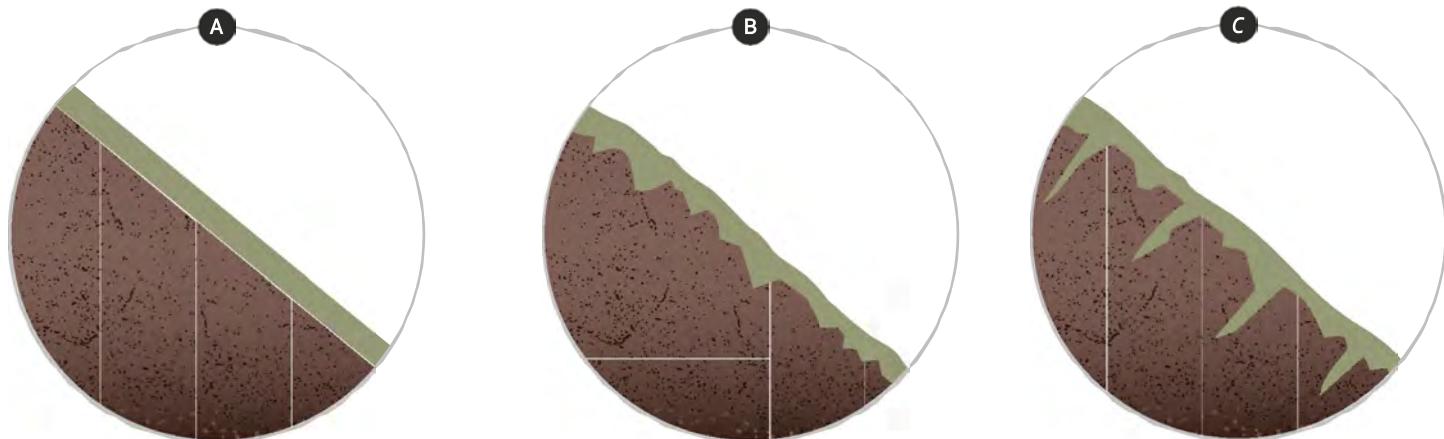
**Minimize Compaction**—Driving heavy equipment over soils causes compaction and reduces infiltration rates. After sites have been prepared for seeding or planting, avoid driving heavy equipment over soils. Practices that are often recommended for erosion control, such as trackwalking, can actually decrease infiltration rates and adversely affect the establishment and cover of native plants. These practices may not be appropriate on all soil types and can be assessed on a site-specific basis (Grismer and Hogan 2007).

**Tillage**—Infiltration rates can be increased through soil tillage, including subsoiling, ripping, and disking (Section 5.5.2). In most cases, tillage will reduce compaction and increase macro-pore space in the surface soil, as well as create surface roughness that further increases infiltration rates. Depending on the stability of the surface material and the level of organic matter, the effects of tillage on infiltration might only be effective for a short time. Under some conditions, tillage needs to be planned into the design of the road. Concentrated water from poorly designed road drainage or inadequate road maintenance has the potential to create deep gullies on tilled soils. Steep slopes in areas of high precipitation have a higher risk of slope failure if tilled slopes are not designed appropriately (Section 3.8.5, see Mitigating for Steep Slope Gradients). Deeper tillage and sculpting the subsoil are some methods to reduce these risks (Section 5.5.2).

**Organic Amendments and Tillage**—Incorporating organic amendments into the soil surface using a bucket of an excavator can create large, stable pores. However, unless the pores are interconnecting, they will not drain well (Claassen 2006). One method for creating continuous pores is to use long, slender organic material, such as shredded bark or wood, composted yard waste, straw, or hay.

### Figure 3-17 | Surface-applied compost

Surface-applied compost has greater surface area contact with the soil when it is applied to roughened surfaces (B), compared to smooth surfaces (A). Creating a rough surface prior to the application of compost creates better rooting, greater surface stability, and faster organic matter decomposition. Tilling the soil, through subsoiling and ripping, to depths of 1 to 2 feet (C) will break up compaction and create channels for compost to move into the soil, increasing soil contact and creating greater infiltration rates.



(Section 5.2.5). Compared to short organic materials such as wood chips, longer materials can increase infiltration rates. Incorporating higher quantities of organic matter in the soil will also increase porosity because of the potential of the organic material to overlap and interconnect.

**Mulch and Tillage**—Applying mulch by itself does not necessarily increase infiltration rates, although it can reduce sediment yields (Hogan and Grismer 2007). However, combined with surface tillage in the form of subsoiling or ripping prior to application of mulch, infiltration rates can be significantly increased. Mulch fills in the micro-basins left from the tillage operation (Figure 3-17).

**Establish Vegetation**—Ultimately, the best method to increase infiltration is to create conditions for a healthy vegetative cover. Good vegetative cover will produce soils with extensive root channels, aggregated soil particles, and good litter layers.

## Road Drainage

Depending on how the road is designed, surface road water from precipitation events is either dispersed or concentrated. Dispersed water is often seen on outslope or crowned roads, where water moves in sheets over the road surface during rainstorms and into the fill slopes. This water can be captured by water harvesting methods (Section 3.8.1). Concentrated water occurs where runoff from the road surface and cut slopes, as well as intercepted water from seeps and streams, is collected in ditches that flow into culverts or other road drainage structures. When designed into the road drainage system, this water can be available for plant growth. Live silt fences, bio-retention swales, and constructed wetlands are some structures that take advantage of this additional water.

**How to Assess Road Drainage**—Road drainage is assessed by identifying drainage patterns on the road plans. Often the Storm Water Pollution Prevention Plans will show the detailed direction of surface road water. Culvert outlets are the areas most likely to have concentrated water that can be considered for use for plant establishment and standing water for pollinator habitat.

## Mitigating for Road Drainage

**Species Selection**—In areas below culverts, soil moisture is typically higher than surrounding areas after rainstorms or snow melt. These areas may be suitable for more moisture-sensitive plant species that require increased soil moisture. When planted with plant species that support pollinators, these sites will increase pollinator habitat.

**Large Wood**—Obstacles, such as large wood, can be placed at the base of culverts or perpendicular to the slope to slow concentrated water and increase soil moisture in these areas. Large wood also provides nesting habitat or shelter for many pollinator species.

**Biotechnical Slope Protection**—Gullies can form below culvert outlets and, for this reason, these sites are often armored with rock. Moisture-loving vegetation, such as willows, sedges, and rushes, can be integrated into the hardened surfaces, such as live silt fences, as shown in Figure 3-18 and as discussed in Section 5.4.3.

**Water Harvesting**—Road surfaces, shoulders, and to a lesser extent, cut and fill slopes are impermeable surfaces that create runoff water during precipitation. Utilizing this water for plant growth, as shown in Figure 3-14, is a form of water harvesting.

### 3.8.2 AVAILABLE WATER STORAGE AND ACCESSIBILITY

The previous section discussed how water enters the soil surface. This section describes how water is stored in the soil and how soil water is accessed by roots. Where precipitation is low or infrequent during the growing season, the amount of water a soil can hold between rainstorms is important from a plant survival and growth standpoint.

The total available water-holding capacity (TAWHC) is the sum of all water stored in the soil profile that is available to plant roots. The amount of water that a soil can store is primarily a function of the following:

- Soil texture
- Rock fragments
- Soil structure
- Rooting depth
- Mycorrhizal fungi

The amount of water a soil stores and how easily it is accessible by roots determines the types of species and the amount of vegetative cover a site can support.

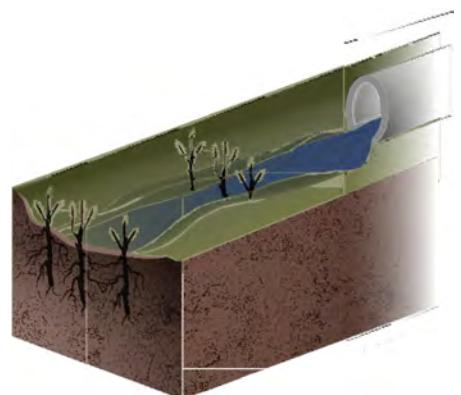
#### Soil Texture

Soils are composed of minerals of varying sizes, ranging from clays (the smallest) to sands (the largest). Each mineral particle in a soil sample can be grouped into one of three categories depending on its size:

- **Clay**—<0.00008 in (0.002 mm)
- **Silt**—0.00008–0.002 in (0.002 to .05 mm)
- **Sand**—0.002–0.08 in (0.05 to 2.0 mm)

The proportion of these size groups in a soil is called the soil texture.

Figure 3-19 shows 12 soil textural classes by their proportions of sand, silt, and clay as defined by the U.S. Department of Agriculture classification system (Soil Survey Staff 1975). Two other soil classification systems, the American Association of State Highway and Transportation Officials (AASHTO) and the Unified Soil Classification systems, are



**Figure 3-18 | Live silt fence**

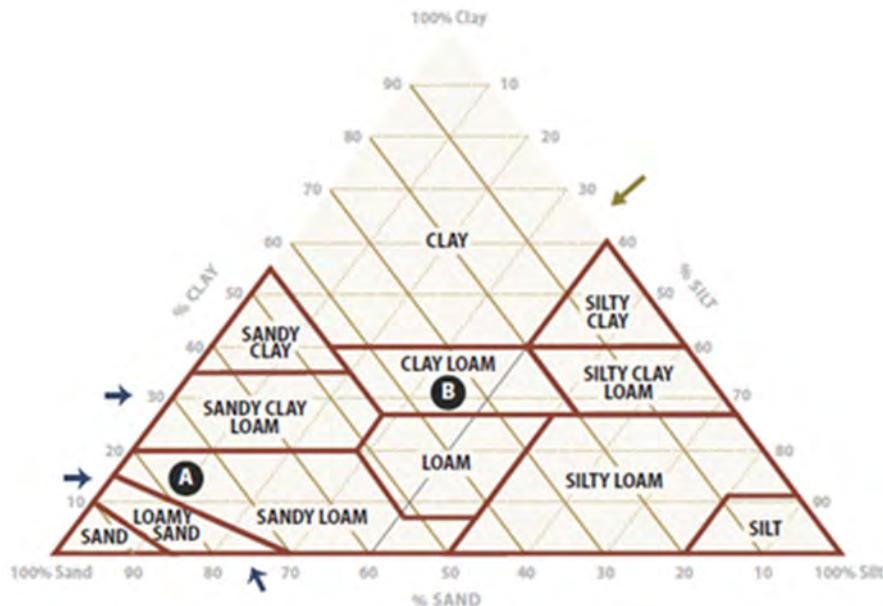
In gullies, draws, intermittent streams, or below culvert outlets, live willow stakes are placed in rows, creating what is referred to as a live silt fence, to slow water velocities and catch sediment and debris (Polster 1997). The stakes root and establish into plants over time.

used for geotechnical engineering. These two systems use different particle size ranges and include parameters such as liquid limit and plasticity in classifying soils. There is no accurate way of converting values from these systems to the USDA textural classes.

Soil texture is an important function of soil water storage because the unique arrangement of pores created in each texture class holds differing quantities of moisture. Clays are typically thin, wafer-like particles with highly charged surface areas that retain large amounts of water.

**Figure 3-19 | Soil textural triangle**

The soil textural triangle defines 12 textural classes based on the percentage of sand, silt, and clay in a soil sample. The textural classes make it easy to describe soils without having to state percentage of sand, silt, and clay. To use the textural triangle, locate the percentage of sand on the bottom side of the triangle and trace the line up to the left-hand side of the triangle. Do the same with either the silt or clay percentages on the other two sides of the triangle (follow silt diagonally down to the lower left and clay across from left to right). Where the two lines intersect is the textural class for that soil. For example, a soil with 75 percent sand and 15 percent clay would be a sandy loam (A). A soil with 30 percent clay and 35 percent silt would have a clay loam texture (B).



Clay particles are often arranged to form small void spaces, or micropores, that also store water. Sands, on the other hand, are large, rounded particles that have a very low surface area and therefore do not hold as much water. The large pores (macropores) that are created when sand particles are adjacent to each other are good for air and water flow but poor for storing water. Soils high in silts hold more water than sands because of the greater quantity of micropores. However, silt particles are not charged, therefore holding less water than clays.

**How to Assess Soil Texture**—Soil texture can be determined fairly accurately in the field using the “feel” test. This is done with the aid of a soil sieve (2 mm opening size) and a bottle of water. Obtain a fairly dry field sample and separate the fine fraction from the coarse fragments with the sieve (note the volume of gravel in the sample). Take a

#### **Inset 3-1 | Measuring available water-holding capacity**

*Modified after Wilde et al 1979*

Available water-holding capacity (AWHC) can be measured in the field by collecting soil samples from a reference site or disturbed site in mid- to late summer when soils are presumably at their driest. Collect samples in bulk density rings in the same manner as sampling for bulk density (Section 3.8.6, see Soil Strength). After removing the ring from the soil, secure a piece of card- board at each end of the ring to keep the soil from falling out, then place it an airtight plastic bag to ensure the sample stays intact during transport. When ready to take measurements, remove the top cardboard piece and weigh the sample. Place the sample in a bucket and fill with water to just the top of the ring. Allow the sample to saturate with water. Once the soil is fully saturated, remove the ring and allow it to drain. After 24 hours, remove the soil from the ring and weigh the soil (this is the wet weight), also weigh the ring and cardboard (allow the cardboard to dry first). To calculate the dry weight, subtract the weight of the dry cardboard and ring from the original dry sample weight.

**Available water-holding capacity (inches of available water per foot) =**

$$\frac{(\text{wet weight} - \text{dry weight})}{((\text{volume of cylinder}) * 12)}$$

sample of the fine fraction in the palm of the hand and moisten it with water. The soil is rubbed between the fingers and thumb and classified using the decision tree in Figure 3-20.

For a more exact determination of soil texture, a sample of soil can be sent to a soils laboratory for a particle size distribution test. This test will report the percentage of sands, silts, and clays in the sample. A Web Soil Survey report ([Section 3.3.2](#)) generated for the project area will also provide a good estimate of the soil textures.

Knowing soil texture is essential for estimating the available water-holding capacity (AWHC) of a soil. Figure 3-21 shows some typical available water-holding capacities for various soil textures. The values in this figure are generalized, but are acceptable for making recommendations on most revegetation projects. The [Web Soil Survey](#) and [Soil Characterization Data](#) websites ([Section 3.3.2](#)) may have available water-holding capacities for soils in the project area. For a more accurate assessment, samples can be sent to soils labs for moisture determination. This is a specialized test and not all labs offer this test; therefore, it is important to contact the lab prior to collecting samples. Water-holding capacity can also be measured using the methods outlined in Inset 3-1.

**Figure 3-20 | Soil texture by feel method**

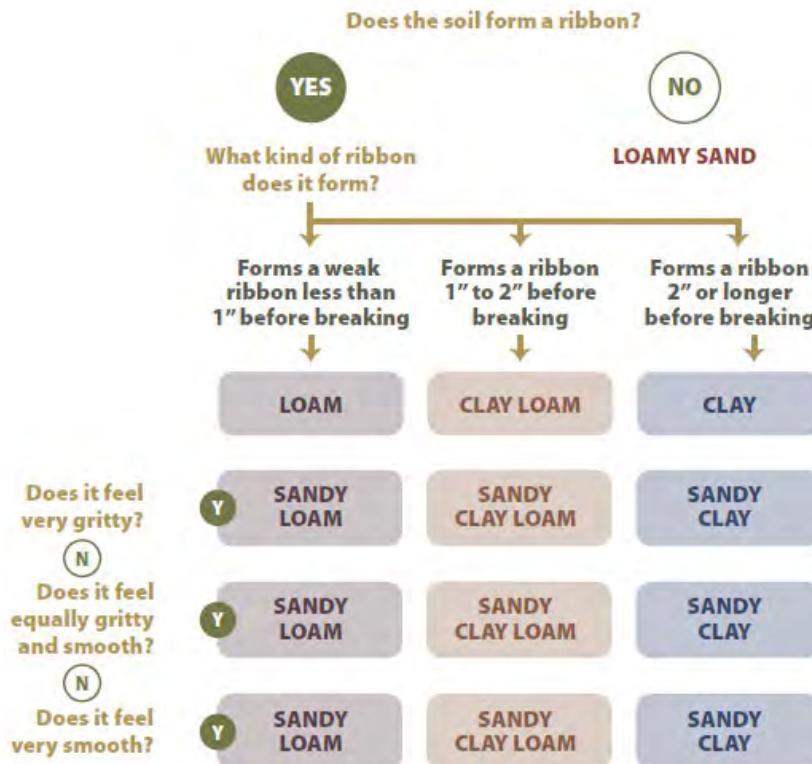
With some practice, soil textures can be determined using the following guide.

Adapted from Colorado State University Extension Publication — #7.723

*Start by placing soil in palm of hand. Add water slowly and knead the soil into a smooth and plastic consistency, like moist putty. Does the soil remain in a ball when squeezed?*



*Place ball of soil between thumb and forefinger, gently compressing the soil with the thumb, squeezing it upward into a ribbon. Form a ribbon of uniform thickness and width. Allow ribbon to emerge and extend over the forefinger, breaking from its own weight.*



## Mitigating for Textures with Low Water-Holding Capacities

**Organic Amendments.** Incorporation of organic amendments (e.g., compost, biochar, biosolids) can increase the water-holding capacity of a soil. Because the water-holding

capacity of each type of organic matter varies by composition and degree of weathering, the effect on soil water-holding capacity by any organic matter being considered is assessed prior to application (see [Section 5.2.5](#) for assessment methods). Sandy textured soils benefit most from organic matter additions, especially those with plant available water of 9 percent or less (Claassen 2006), which are typically sands, loamy sands, and sandy loam soils. Testing several different rates of incorporated organic matter on soil moisture-holding capacity prior to selection will help determine the appropriate amount of material to apply.

**Clay**—The water-holding capacity of sandy textured soils can be increased by incorporating soils with higher clay fractions into the sandy soils. These soils should have no more than 40 percent clay fraction (e.g., clay loam, sandy clay loam, and silty clay loam textures). Consider adding clays at rates that result in new soil textures similar to loams, silt loams, or sandy clay loams. Higher rates of clay addition are not recommended. It is always important to test the additions of any soil to another to understand what the effects on water-holding capacity and structure might be. Ideally this is done in the field in small plots.

**Polyacrylamides**—Polyacrylamides are hydrophilic polymers that absorb many times their weight in water. They are used to increase the water-holding capacity of greenhouse growing media. The value of using these materials in revegetation projects, however, is questionable based on the low plant response rates, high material costs, and difficulty of incorporating these materials evenly into the soil. A study, located at several semi-arid sites, showed that two rates of polyacrylamide crystals incorporated into road fills had no differences in native grass cover and survival and growth of outplanted pine seedlings from the controls (Riley et al 2013). Consider testing any full-scale use of polyacrylamides at different rates on the sites being revegetated. In addition, determining how polyacrylamide crystals would be evenly mixed into the soil is an important consideration when considering use of these materials.

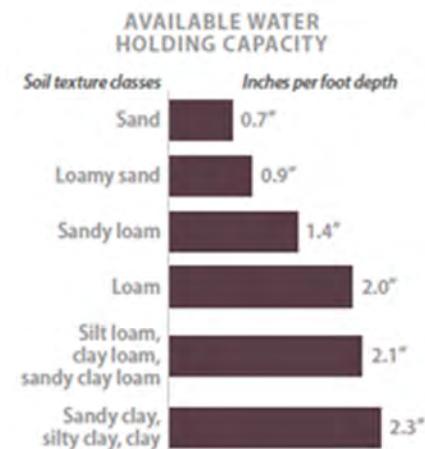
## Rock Fragments

Mountain soils and highly disturbed sites are typically high in rock fragments. The presence of rock fragments is important because rock reduces the amount of water and nutrients a soil can hold. At high volumes in the soil, rock fragments will limit the biomass and vegetative cover a site can support.

The rock classification system classifies rock fragments into the following five size ranges:

- **Fine gravels**—0.08 to 0.2 in (0.2 to 0.5 cm)
- **Medium gravels**—0.2 to 0.8 in (0.5 to 2.0 cm)
- **Coarse gravels**—0.8 to 3 in (2.0 to 7.0 cm)
- **Cobble**—3 to 10 in (7.0 to 20.0 cm)
- **Stone**—>10 in (20.0 cm)

Highly weathered rock can retain some soil moisture depending on the size of the rock fragments and degree of weathering (Flint 1983). For practical purposes, however, it is usually assumed that the presence of cobbles and stone rock fragments in the soil



**Figure 3-21 | Soil texture and available water-holding capacity**

This chart shows the general relationship between soil texture and available water-holding capacity soils (adapted after Ley et al 1994). As clays increase in a soil, so does water-holding capacity. Typically, clay loam soils hold more than twice as much water as sandy textured soils. The presence of humus in topsoil increases water-holding capacity of loams and sandy loams at a rate of 2.25 percent water to each percent rise in soil humus (Jenny 1980), which equates to an approximately 0.75 percent increase in water holding capacity for every 1 percent increase in organic matter.

will reduce the available water-holding capacity of the soil proportionally. For example, a sandy loam soil without rock fragments has a water-holding capacity of 1.4 inches per foot of soil (Figure 3-21). When 30 percent large rock fragments are added to the soil profile, the available water-holding capacity is reduced to 70 percent, or 0.98 inches of available water ( $1.4 * 0.7 = 0.98$ ). Alternatively, fine and medium gravels (0.08 to .8 inches in diameter) hold some moisture. A rule of thumb is that these fine and medium gravels reduce water-holding capacity by two-thirds of their volume. In the above example, if 30 percent of the soil were composed of medium and fine gravels, the available water in this soil would be 1.12 inches per foot ( $1.4 - (1.4 * 0.3 * 0.66)$ ).

**How to Assess Rock Fragments**—Rock fragment content is usually determined in the field. Large rock fragments, such as cobble and stones, are estimated in a variety of ways. The most common methods are surveying freshly exposed road cuts or observing soil excavation during road construction. Estimating the volume can be difficult, and often the amount of rock is over- or under-estimated. One method of estimating large rock in road cuts is to take a digital picture and lay a grid over the surface, as shown in Figure 3-22. Whenever rock is estimated from old road cuts, it is necessary to determine whether a portion of the rock is masked by soil that might have moved over the rock. A freshly exposed road cut provides the most accurate approximation of rock content.

Rock encountered while digging a soil pit will provide a more accurate estimate of larger coarse fragments. Cobbles and stones, if they can be moved, are set apart from the soil when the pit is excavated. The volume of cobbles and stones is then visually compared to the volume of soil excavated from the soil pit to estimate the percentage of rock fragments.

Gravel content is determined from the excavated soil by sieving it through several soil sieves. Sieves are available through most engineering equipment companies (Figure 3-23). The 2-mm sieve (also referred to as a #10 sieve) is the most important sieve to use because it separates the gravels from the soil fraction. Another useful sieve is the  $\frac{3}{4}$ -inch sieve because it separates the fine and medium gravels from the coarse gravels. This sieve can be used in the field to remove larger rock fragments from the soil sample to reduce the weight of the sample. When soils are dry, they are easier to sieve in the field; however, when they are moist, soils are air dried first before they can be sieved. The gravel and soil volumes are visually estimated.

It is important to include the volume of cobbles and stones estimated in the field with the gravels estimated through sieving to calculate the total rock fragments in a soil:

$$\% \text{ rock fragments in profile} = (100 - \% \text{ cobbles and stone}) * \% \text{ gravels in sample}$$

For example, a soil is estimated to have 25 percent cobbles and stones from observing road cuts and from several soil pits. Sieving shows that 50 percent of the sieved soil is composed of gravels. The soil would be composed of 25 percent cobbles and stones, 37.5 percent gravels ( $(100 - 25) * 0.50$ ), and 37 percent soil.



**Figure 3-22 | Estimating rock fragment content from roadcuts**

The amount of rock in a section of soil can be roughly estimated from road cuts. Large rock can be determined by laying a grid of 20 circles over a photograph of a road cut and recording the number of circles intercepting rock (in the center of the circle). This value is divided by the total number of circles in the grid to obtain the percentage of subsoil in rock fragments. In the picture below, subsoil contains approximately 25 percent large rock (5 intercepted rocks divided by 20 points).

*Photo credit: David Steinfeld*

**A****B****C**

**Figure 3-23 / Soil sieves for estimating rock content**

The number 10 sieve (2 mm opening) on the right separates soil particles (C) from rock particles (B and A). The 3/4-inch sieve on the left separates the fine and medium gravels (B) from the coarse gravels (A).

*Photo credit: David Steinfeld*

## Mitigating for High Rock Content

**Rock Removal**—Screening rock fragments from the soil will increase the available water-holding capacity of a soil. The greatest benefit from screening is with soils that are very high in cobble and stone, where the reduction in volume of rock in the soil would be significant. One type of screen is the “grizzly feeder” which acts as a giant sieve to sort out rock of any size depending on the screen opening widths. Screened soils have the greatest benefit where soils are shallow or a good ground cover is necessary (e.g., grasses and forbs).

**Incorporate Compost**—Compost incorporated in the soil at high rates will increase the water-holding capacity of a rocky soil (Section 5.2.5). Depending on the size of the coarse fragments, incorporation can be difficult.

**Surface Apply Compost**—A more practical method to mitigate for rocky soils is to apply composts to the soil surface without mixing. When surface applied, composts can be good growing media for seeds of grasses and forbs (Section 5.2.3). At rates greater than 3 inches applied to the surface, seeds germinate well and establish into seedlings that can access moisture and nutrients not only from the compost, but also some moisture from the rocky soil below the compost. Be aware that on steep slopes, if the site is not prepared correctly, the layer between the compost and soil can become a slip plane on slopes when compost becomes saturated with water (Section 3.8.6, see Mitigating for Low Permeability).

**Apply Topsoil**—If topsoil is available, it can also be applied over a rocky soil (Section 5.2.4). Topsoil will have to be placed deep enough to compensate for the quantity of rock in the soil being covered. On steep slopes, preparation of the site prior to application of topsoil is important to avoid slope stability problems (Section 3.8.5, see Mitigating for Steep Slope Gradients).

**Planting Islands**—On very rocky sites, rocky soils can be mitigated by focusing mitigating measures into planting islands (Section 5.2.8). Where topsoil, compost, or screened soil is limited, this material can be concentrated in mounds, pockets, or benches strategically located throughout a revegetation.

## Soil Structure

Just as soils are composed of many-sized mineral particles, they are also composed of different-sized voids (also referred to as pores) whose influence is responsible for water movement, water storage, air flow, and root penetration. Small pores (micropores) strongly influence the moisture-holding capacity of soils, while large pores (macropores) are responsible for water movement, air flow, and root penetration. Soil structure is the arrangement of soil particles into aggregated units that gives rise to the macro-porosity in the soil. It is qualitatively observed as cracks, channels, aggregates, crumbs, and clods in the soil, and described by alternative terms such as friability and tilth. Water flow and root penetration depend on good soil structure. If soil structure is poor or compacted, roots are less able to penetrate the soil to access water. Soil structure is important for other soil functions such as air flow, drainage, permeability, infiltration, and essential habitat for most soil organisms.

Soil structure is significantly reduced by operating heavy equipment over soils. The pressure applied by heavy equipment compacts the macropores, reducing soil volume and increasing soil density. This impact is called soil compaction (Figure 3-24). The effects of soil compaction on tree growth are well documented (Poff 1996). Trees growing on highly compacted soils have far less root, stem, and leaf production than those growing on non-compacted sites. Studies have shown a linear relationship between the increase in surface soil bulk density and decrease in height growth of young Douglas-fir and ponderosa pine trees (Froehlich and McNabb 1984).

It is a safe assumption that soils will be highly compacted after construction due to the use of heavy equipment. In addition to reducing the potential of a construction site to grow vegetation, compaction also increases runoff and sediment during rainstorm events, which can impact water quality. On sites where summer rainfall is limiting, there will also be less water entering the soil, reducing the amount of water available for plant growth.



**Figure 3-24 | Compacted soil**

Compacted soils are created by heavy equipment operating over soil. The large pore spaces are compressed and the impacted soils often form a platy structure as shown in this photograph.

*Photo credit: Tom Landis*



**Figure 3-25 | Poor draining soils due to soil compaction**

Compacted soils drain very slowly, as the puddles on the surface of the obliterated road in this photograph indicate. During rainfall or snowmelt, soils can stay saturated for days and even weeks. Establishing seedlings during this period can be very difficult because roots of most species cannot survive when soils are poorly drained. Seedlings shown in this photograph were dead within three months.

*Photo credit: David Steinfeld*

Compaction can occur several feet below the soil surface, depending on soil texture, moisture, and the type and weight of equipment being operated. Very compacted soil layers can significantly reduce or eliminate root penetration. Where compacted layers occur, downward water movement is restricted and water may saturate the soil layers above the compacted layer. The resulting saturated soil conditions can be very restrictive to root growth because of the lack of oxygen and the propensity for higher incidence of disease (Steinfeld and Landis 1990) and seedling mortality (Figure 3-25). Compacted layers will naturally recover to their original porosity through root penetration, animal activity, and freeze-thaw events, but recovery can take 20 to 70 years (Wert and Thomas 1981; Froehlich et al 1983).

**How to Assess Soil Structure**—It is easy to qualitatively differentiate good soil structure from compacted soil, but measuring it quantitatively can be difficult. Indirect field tests to quantify soil structure include bulk density and penetrometer tests.

The bulk density test measures the dry weight of a standard volume of soil. If the soil has a high porosity, the bulk density values will be low; if the soil is compacted, the bulk density will be high. In this method, a cylindrical tube is driven into the soil with a portable bulk density sampler and a soil core is removed (Figure 3-26). The soil is shaved evenly on both ends so that the soil is exactly the shape and volume of the cylinder. The soil is then removed from the cylinder, oven-dried, and weighed.

$$\text{Bulk Density} = \frac{\text{Weight of dry soil (g)}}{\text{Cylinder volume (cc)}}$$

Bulk density values of a disturbed site are related back to the bulk density of an adjacent reference site to make the values meaningful. Remaining within a 15 percent increase in bulk density over reference site values is ideal. Unfortunately, the bulk density method is time consuming and cannot be conducted on soils with high rock fragments.

A less quantitative, but more practical, method of measuring soil porosity is with a soil penetrometer. This equipment measures soil strength instead of density. Compacted soils have greater strength, and greater resistance to penetration by a penetrometer, than non-compacted soils. Several types of penetrometers can be purchased for field work— penetrometers that measure the resistance as a continuous pressure is applied to the probe and penetrometers (impact penetrometers) that measure the number of blows of a hammer to drive the penetrometer into a specified depth. A monitoring procedure for assessing compaction using an impact penetrometer has been developed by the NRCS (Herrick et al 2005b). The most practical and economical field method for assessing compaction, however, is simply using a long shovel, as shown in Figure 3-27. In this method, a site is traversed and, at predetermined intervals, a shovel is pushed into the ground to determine how loose the soil is. By applying the entire body weight to the shovel and observing the distance the shovel penetrates the soil, a qualitative measurement of soil compaction can be made. A rule of thumb is that a shovel penetrating over 12 inches deep indicates a soil with a very high porosity; penetration below 3 inches deep indicates a very low porosity.

Whether a shovel is used or a soil penetrometer, the readings are affected by rock content and soil moisture. When soils are dry, they have greater strength and higher resistance to penetration. This is why any comparative sampling (e.g., comparing reference site to construction site soils) using a penetrometer is done at the same moisture levels. Encountering large rocks can be confused with hitting a compacted layer. When this occurs, sampling should be done at several adjacent points until the penetrometer can be pushed into the soil without hitting rock. On very rocky soils, the penetrometer is not a practical tool.



**Figure 3-26 / A soil core is used to assess soil compaction**

Soil compaction can be assessed by measuring bulk density of a soil. The most common method is to drive a cylindrical tube into the soil, as shown in the photograph, and weighing the soil after it has been dried.

*Photo credit: David Steinfeld*

## Mitigating for Poor Soil Structure

**Tillage**—Breaking up compacted layers can be done effectively when deep tillage equipment is operated correctly ([Section 5.2.2](#)).

**Incorporate Organic Matter**—The effectiveness of deep tillage can be enhanced if organic matter is incorporated into the soil prior to tillage ([Section 5.2.5](#)). Organic matter can keep the soil from settling back to higher, pre-tillage densities. Application rates at which organic matter showed positive effects on soil structure was observed at a ratio of 25 percent incorporated organic matter to 75 percent soil by volume (Claassen 2006). Longer shreds of organic matter are preferred over smaller, chip sizes because the longer strands create interconnecting pathways for water, air, and roots while increasing soil strength (Claassen 2006). The additions of non-composted organic matter, however, will tie-up nitrogen for a period of time. While this might be problematic in the short term, the importance of developing soil structure for long-term site recovery often overrides concerns about the lack of immediately available nitrogen.

**Operate Heavy Equipment with Care**—Soil compaction is greatest when soils are moist. To limit the amount of soil compaction, schedule equipment operation during times when soils are dry. Compaction can also be minimized by using smaller equipment (Amaranthus and Steinfeld 1997) or leaving slash or deep mulch on the soil surface (which provides cushion).

**Avoid Last Minute Compaction**—Soil compaction is unavoidable during construction, but compacting soils after mitigating treatments have been implemented, such as tillage, is to be avoided. In many cases the benefits of mitigating treatments have been nullified by the lack of attention to heavy equipment operations after topsoil additions or tillage treatments have been made. For example, topsoil salvage and placement, as discussed repeatedly in this manual, benefit the site in many ways. But this expensive mitigating measure loses much of its value if the soils are compacted during or after soil placement. Once topsoil is deep-tilled, every effort should be made to avoid any further equipment operation on the site.

**Encourage Mycorrhizae Establishment**—Mycorrhizal fungi build soil structure through hyphae and water stable organic glues (e.g., glomalin). [Section 3.8.2](#) (see [Mitigating for Lack of Mycorrhizal Fungi](#)) covers methods beneficial for establishing mycorrhizae.

## Rooting Depth

Rooting depth is the distance from the surface of the soil to the lowest point that roots can penetrate. It encompasses any strata (e.g., topsoil, subsoil, and parent material) that can be accessed by plant roots. The deeper the rooting depth of a disturbed site, the greater the total available water storage and the higher the productivity of the site.

Rooting depth is affected by restrictive layers that block root penetration to lower strata ([Section 3.8.5](#), see [Mitigating for Steep Slope Gradients](#)). For example, the rooting depth of a post-construction site is estimated at 6 feet deep. However, further investigation finds that there is a highly-compacted layer at 12 inches, which would limit most, if not all, root penetration below that point. The rooting depth under these conditions has been reduced to only 1 foot of soil instead of 6 feet. Restrictive layers also include soils with very high or very low pH, toxic materials, or a high-water table.

Rooting depths vary by plant species and age of the vegetation. Most mature tree species have deep root systems that access subsoil and parent material; roots of grasses and forbs are predominantly limited to surface soils. Annual grasses and forbs



**Figure 3-27 |** A shovel can be used to determine depth to compaction

A simple, qualitative method of determining compacted layers is to mark the face of a long shovel with a ruler. Pushing the shovel in the ground with the entire body weight and observing the distance the shovel penetrates can indicate the depth to a compacted layer.

*Photo credit: David Steinfeld*

need less rooting depth than perennial grasses and forbs, with the roots of these species growing in the upper surfaces of the soil. The age of the vegetation also determines the abundance and location of roots. Newly established seedlings have shallow roots but, as the plants mature, root systems expand to access moisture deeper in the soil.

Rooting patterns and root morphology play a role in how plants access soil water. Some species have finer-textured root systems that access tightly held soil moisture; other species have aggressive root systems that can penetrate deeply into cracks between rock fragments. Grasses, for instance, have shallower root systems than trees and shrubs, but their small size and high density in the surface soil gives them an advantage in shallow soils.

**How to Assess Rooting Depth**—Rooting depth should be estimated from reference sites during planning and post-construction, but it is not always an easy parameter to measure. Observing road cuts is often the best means to determine rooting depth. Rock type (e.g., granite, sandstone, and schist), fracturing patterns, rock weathering, and the degree of rock fracturing will provide an indication of rooting depth. Observing the amount and type of roots in the fractures of existing road cuts will give a good idea of rooting depth.

Fracturing and weathering of rock can also be determined from geotechnical analysis. If the bedrock has been drilled, the drill log report can provide an indication of degree and depth of rock fracturing and weathering. One way that rock quality is assessed is through a classification called the Rock Quality Designation Index (RQD). This system rates the bedrock by how much fracturing is observed in the cores. It is calculated by measuring the pieces of rock in the core sample that are longer than 10 cm, summing the length of these pieces, and dividing by the total length of the core (Deere and Deere 1988). A small RQD means that the bedrock is highly fractured whereas a high RQD means the bedrock is massive. A RQD may be poor from an engineering standpoint because of the high fractures, but favorable from a revegetation standpoint because cracks will hold some moisture and allow root penetration. A RQD rated as “very poor,” “poor,” and even “fair” should be somewhat favorable to root penetration.

| Soil strata | Data     | Soil characteristics               | Results | Equations                       |
|-------------|----------|------------------------------------|---------|---------------------------------|
| 0 to 1'     | A<br>2   | AWHC of texture<br>(inches / foot) | 2       | From Figure 3-21 or lab results |
|             | B<br>20  | Small Rock (%)                     | 0.264   | = A * (B / 100) * 0.66          |
|             | C<br>5   | Large Rock (%)                     | 0.1     | = A * (C / 100)                 |
|             | D<br>1   | Thickness (ft)                     | 1       | Thickness of soil section       |
|             | E        | Available water by<br>section      | 1.6     | = (A - B - C) * D               |
| 1 to 2'     | F<br>2.2 | AWHC of texture<br>(inches / foot) | 2.2     | From Figure 3-21 or lab results |
|             | G<br>35  | Small Rock (%)                     | 0.5082  | = F * (G / 100) * 0.66          |
|             | H<br>25  | Large Rock (%)                     | 0.55    | = F * (H / 100)                 |
|             | I<br>1   | Thickness (ft)                     | 1       | Thickness of soil section       |
|             | J        | Available water by<br>section      | 1.1     | = (F - G - H) * I               |

**Table 3-8 | Calculating total available water-holding capacity**

The total available water-holding capacity (TAWHC) is the total amount of moisture that a soil can store for plant growth when fully charged with water. TAWHC values for each revegetation unit are helpful for determining which species will perform well and in developing the mitigating measures necessary to increase water-holding capacity or rooting depth. TAWHC is calculated by determining the texture, rock fragment content, and depth of each soil layer, and calculating how much water each layer will optimally store. The water-holding capacity of each soil layer is added together to obtain the TAWHC for the soil profile.

|         |  |     |                                    |        |                                 |
|---------|--|-----|------------------------------------|--------|---------------------------------|
|         | K  | 2.2 | AWHC of texture<br>(inches / foot) | 2.2    | From Figure 3-21 or lab results |
| 2 to 5' | L  | 35  | Small Rock (%)                     | 0.5082 | = K * (L / 100) * 0.66          |
|         | M  | 70  | Large Rock (%)                     | 1.54   | = K * (M / 100)                 |
|         | N  | 3   | Thickness (ft)                     | 3      | Thickness of soil section       |
|         | O  |     | Available water by<br>section      | 0.5    | = (K - L - M) * N               |
|         | <b>Total available water holding capacity<br/>(inches)</b> |     | 3.2                                |        | = E + J + O                     |

Rooting depth is also affected by the presence of a restrictive layer caused either naturally or by compaction. How to determine the presence of these layers is addressed in [Section 3.8.2 \(see Rooting Depth\)](#) and [Section 3.8.5 \(see Mitigating for Steep Slope Gradients\)](#).

The literature contains many references to defining the depth of soil needed to support different plant communities. For example, 18 inches of soil has been shown to support simple grassland ecosystems, but more diverse native grassland communities are reported to require up to 4 feet or more (Munshower 1994). These figures can be misleading if they are not put in the context of site climate and soil type. In many respects, it is more useful to state the TAWHC of a site rather than the rooting depth. The TAWHC is the total amount of moisture that a soil can store for plant growth when fully charged with water.

Table 3-8 shows how the TAWHC is calculated. Using the same format and equations, a similar spreadsheet can be created by copying the equations into the "Results" column. In this example, there is 1 foot of topsoil and 2 feet of subsoil over a highly-fractured basalt. The topsoil has a loam texture and available water-holding capacity of 2.0 inches (estimated from Figure 3-21 or obtained from lab results) but, because of the rock fragments, it is reduced by approximately 0.4 inch. The subsoil has a high water-holding capacity because of high clay content, but the available water-holding capacity is reduced by half due to rock. Highly fractured basalt is encountered at a depth of 2 feet, and it is estimated from the road cut that approximately 30 percent is actually fractured. Within these weathered fractures is a gravelly clay loam textured material storing approximately 0.5 inch of water. The TAWHC for this site would be the sum of all sections of soil (approximately 3.2 inches).

TAWHC is useful for comparing water relationships between revegetation units and reference sites. For example, the TAWHC for a post-construction soil is 3.6 inches compared to an adjacent reference site, which is 6 inches. If the desired future condition of the post-construction soil is to be similar to the adjacent reference site, then the TAWHC of 3.6 inches has to be increased upward toward 6 inches for the site to be capable of supporting the vegetative community occurring on the reference site. If this is not possible, then the DFC target needs to be changed to reflect the plant community that the soil can support.

## Mitigating for Poor Rooting Depth

**Increase Available Water-Holding Capacity**—Improving the water-holding capacity of the existing soil will increase TAWHC. Mitigating measures discussed in [Section 3.8.6](#) can be used to increase soil moisture.

**Tillage**—If restrictive layers due to compaction are encountered, deep tillage should be considered. [Section 5.2.2](#) provides guidelines for deep tillage.

**Apply Topsoil**—Increasing rooting depth and TAWHC can be accomplished by applying topsoil (Section 5.2.4).

**Planting Islands**—Mitigating measures, such as applying topsoil, organic matter incorporation, deep tillage, and other measures that increase water-holding capacity, can be focused in strategic locations, such as planting islands. This will conserve materials and reduce costs (Section 5.2.8).

**Blasting**—Strategic blasting to shatter the parent material has been suggested as a possible means of increasing rooting depth (Claassen 2006).

## Mycorrhizal Fungi

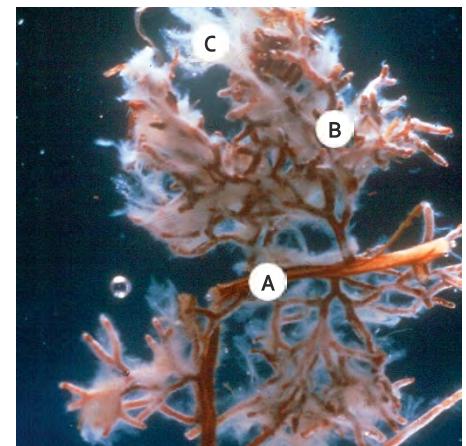
The discussion to this point has addressed the primary factors responsible for soil water storage (soil texture, rock, and rooting depth) and ease of a plant through its roots to reach this water (soil structure). In this section, the discussion turns to how plants increase the efficiency of accessing water through mycorrhizae. While mycorrhizae provide many other benefits to the site in addition to water enhancement, they are covered in depth in this section because of the importance of water to establishing vegetation on highly disturbed sites in the western United States.

Mycorrhiza is the unique symbiotic relationship between fungi (called mycorrhizal fungi) and host plants. To the naked eye, many mycorrhizal fungi appear as a fine web or netting that seems to connect the root system to the surrounding soil (Figure 3-28), and in essence, this is exactly what is occurring. The extremely small hyphae of the mycorrhizal fungi are actually taking on the form and function of an extended root. Because mycorrhizal hyphae are up to five times smaller, they are able to access spaces in the soil not easily accessible by larger plant roots. Mycorrhizal hyphae not only provide the plant with greater access to soil moisture and nutrients, they also surround and protect roots from soil pathogens. In return, the host plant supplies carbohydrates to keep the mycorrhizal fungi alive.

Mycorrhizae play a critical role in site restoration by building soil structure. Hyphae and water stable organic “glues,” such glomalin, are excreted by the mycorrhizal fungi and bind soil particles together into aggregates. These aggregates stabilize the soil (Section 3.8.6, see Soil Strength) and are important for good air exchange and water permeability. This basic soil building process, or repair, facilitates the creation of nutrient reserves and nutrient cycling essential for restoring ecosystems (Miller and Jastrow 1992). Mycorrhizal fungi can also improve survival of tree and grass seedlings (Steinfeld et al 2003; Amaranthus and Steinfeld 2005). A healthy population of mycorrhizal fungi has also been shown to increase plant biomass and cover (Wilson et al 1991; Brejda et al 1993; Sobek et al 2000), and increase the diversity of native species (Smith et al 1998; Charvat et al 2000).

Ninety percent of all terrestrial plants form symbiotic relationships with mycorrhizal fungi. Of the thousands of known species, most generally fall into two categories—ectomycorrhizal fungi and arbuscular mycorrhiza fungi.

Arbuscular mycorrhizal fungi (AMF), formerly called endomycorrhizae, are the most commonly occurring mycorrhizal fungi, forming on 75 to 85 percent of plant species. These include legumes, composites, grasses, bulbs, most shrubs, and ferns. In addition, AMF occur on many tree species, including redwoods and some cedars, and many types of tropical trees. AMF grow inside the roots of the host plant and extend hyphae out into the soil. These fungi are more general in their association with plant species, meaning that one mycorrhizal species can form an association with a broad spectrum of plant species. AMF reproduce in two ways: by forming single spores outside of the root and from fungal structures (vesicles and hyphae) present inside a colonized root system. Arbuscular mycorrhizal fungi do not disperse their spores in the wind, but



**Figure 3-28 | Mycorrhizal fungi extend root systems**

Mycorrhizal fungi can greatly increase the surface area of the root system. The ectomycorrhizal fungi attached to the pine root system (A) comprise most of the absorptive surface shown in this photograph. The mycorrhizae include brown branched structures (B) and white hyphae or filaments (C).

Photo credit: Mike Amaranthus, Mycorrhizal Applications Inc

### 3—PLANNING

instead are dispersed from root to root or by animals. For this reason, recolonization of drastically disturbed sites by arbuscular mycorrhizal fungi can be slow, especially if there are limited sources of healthy, undisturbed soils nearby to repopulate the site.

Unlike arbuscular mycorrhiza fungi, ectomycorrhizal fungi, as the name implies, coat the outside of the roots with hyphae that extend out into the soil. Ectomycorrhizal fungi form on 5 to 10 percent of plant species, the majority of which are forest trees in the western United States. Species include Douglas-fir, western larch, true firs, spruce, hemlock, oak, manzanita, willows, and cottonwood. These fungi form a netting of fine hyphae around the root system that is often observable on nursery produced seedlings inoculated with mycorrhizal spores. Unlike AMF, the relationship between ectomycorrhizal fungi and host species is very specific. Many ectomycorrhizal fungi species have evolved to associate with only one plant species. Ectomycorrhizal fungi produce fruiting bodies, such as mushrooms, puffballs, and truffles, which yield reproductive spores for wind or animal dispersal.

AMF and ectomycorrhizae do not associate with all plant species. For instance, in the western United States, arbutoid mycorrhizae forms on manzanita and madrone, while huckleberry form ericoid mycorrhizae. Still another 10 to 15 percent of the plant species in the United States do not form mycorrhizae at all. Many of these plant species have evolved root systems that function similarly to mycorrhizae and therefore give them an advantage over many mycorrhizal plant species, especially during early plant establishment when mycorrhizae inoculum may be limiting. This advantage is why many plant species, considered highly aggressive weeds, are non-mycorrhizal species.

**How to Assess Mycorrhizal Fungi**—Where soils have been drastically disturbed, it can be assumed that the mycorrhizal fungal propagules are drastically reduced or absent from the site. The size and severity of the disturbance determine the diversity and quantity of mycorrhizal fungi. As the level of disturbance increases, the density of viable fungi propagules typically decreases. Small disturbances surrounded by native forests or rangelands often reestablish quickly; in larger disturbances, where topsoil has been removed, recolonization by mycorrhizal fungi may take years.

Some laboratories offer testing for mycorrhiza fungi, but these are expensive tests. Because it is unlikely that mycorrhizal fungi will be found in recently disturbed sites lacking topsoil, conducting these tests for most projects is unnecessary.

## Mitigating for Lack of Mycorrhizal Fungi

For most construction projects, the management of mycorrhizae should be considered in the early stages of project planning. Several strategies are available to enhance mycorrhizal colonization.

**Minimize Soil Disturbance**—Operations that maintain topsoil will often preserve mycorrhizal inoculum and maintain soil nutrition. Partially disturbed topsoil is often adequate for reestablishing mycorrhizal plant species. Partial disturbances include clearing and grubbing of road right-of-way vegetation, ground-based logging, and light to moderate intensity burns. Colonized root systems left behind in these operations are sources of inoculum for endomycorrhizae.

**Leave Undisturbed Areas**—The movement of AMF into highly disturbed sites is slow. Spores are transported by soil erosion and animal movement but not by air. Leaving small areas of native vegetation and undisturbed soils within the larger disturbance reduces the travel distance of AMF, facilitating a quicker repopulation of the disturbed site. This practice is especially important where the size of the disturbance is large.

**Salvage Topsoil**—Salvaging topsoil and reapplying it to drastically disturbed sites is commonly done when quality native topsoil is available (Section 5.2.4). Topsoil obtained from non-forested sites, such as meadows, rangelands, and unforested clear-cuts, is typically high in AMF which is important for grass and forb establishment. Salvaged topsoil forested sites will have mycorrhizae suitable for tree species as well as grass and forbs.

**Apply Topsoil to Planting Holes**—If topsoil is very limiting, placing healthy topsoil into holes prior to planting seedlings is an effective method of introducing an inoculum to a disturbed site. Collecting soils as inoculum from young, actively growing forests has been shown to be suitable inoculum for young tree seedlings (Amaranthus and Perry 1987).

**Apply Commercially Available Mycorrhizal Fungi Inoculums**—Applying commercially available mycorrhizal fungi inoculum is another method used to repopulate highly disturbed sites (Section 5.2.7). Commercially available sources of mycorrhizal inoculums are available for ectomycorrhizal and AMF plant species. These inoculums can be applied in hydroseeding slurries, as seed coats and root dips, through irrigation systems, or incorporated into the soil by broadcasting or banding. Fine grades of mycorrhizal inoculum are applied to the surface of the soil and will move into the soil surface with rainfall. To make them effective, incorporate coarser-textured commercial inoculums in the soil. When purchasing live plants from a nursery, rooting media can be inoculated with mycorrhizal fungi during nursery culture (Figure 3-29).

**Reduce Fertilizer Use**—While the application of fertilizer can increase plant biomass in the short term, it can also suppress mycorrhizal infection (Jaspers et al 1979; Claassen and Zasoski 1994). However, at lower rates, fertilizers have been shown to help plant establishment and improve mycorrhizal colonization (Claassen and Zasoski 1994).



**Figure 3-29 | Non-mycorrhizae inoculated seedlings versus inoculated seedlings**

The response of adding mycorrhizae spores to non-inoculated seedlings (right) can sometimes be dramatic. Both sets of sticky currant (*Ribes viscosissimum*) seedlings shown in this photograph were stunted for months after seed germination. AMF spores were applied to the surface of each seedling container on the left with immediate growth response, while those on the right without mycorrhizae remained stressed (photo taken 50 days after inoculation).

*Photo credit: David Steinfeld*

### 3.8.3 WATER LOSS

Water loss is the depletion of soil moisture through transpiration (loss of water through leaves/ needles) and evaporation of soil moisture from the soil surface. The rate at which evaporation and transpiration draw moisture from the soil profile is the evapotranspiration (ET) rate. ET rates change daily (rates rise through the day with increasing temperatures and winds speeds and decrease at night), weekly (as weather systems move through), and seasonally (rising in spring and summer and decreasing in fall and winter) (Figure 3-30).

The plant is a conduit for water transport between the atmosphere, which demands water from the plant, and the soil, which acts as a bank of water. When ET rates are low, plants can easily pull water from the soil through the leaves to the atmosphere. As soil moisture is depleted through time, there is less moisture to draw from and plants come under greater stress. With rising temperatures, lower humidity, and lack of rainfall, plants will be under very high moisture stress levels (Figure 3-31). The amount

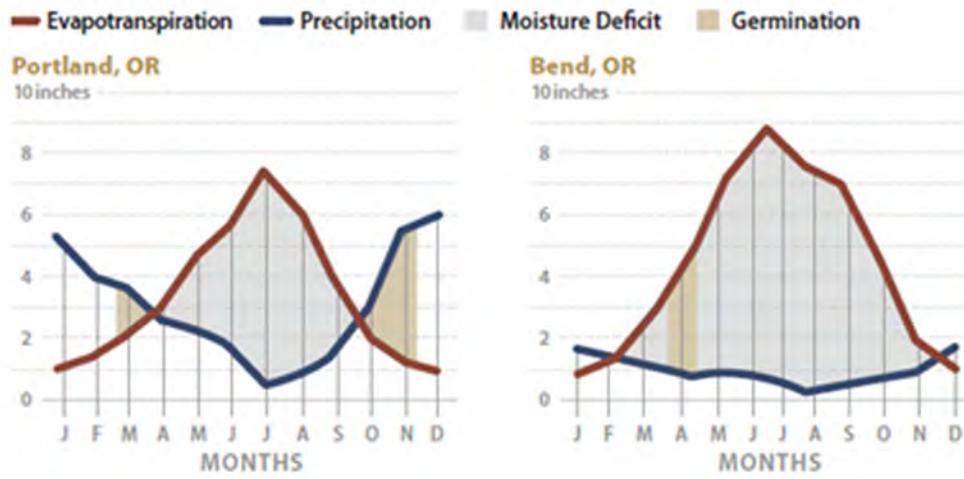
### 3—PLANNING

of stress that a plant is under is referred to as plant moisture stress (PMS) (Figure 3-32).

PMS is at its highest from middle through late summer in the western United States, when the ET rates are at their highest and soil moisture levels at their lowest.

Water loss due to ET can be influenced by a number of abiotic and biotic factors, primarily:

- Wind
- Site aspect
- Competing vegetation
- Humidity
- Soil cover



## Wind

Wind is often overlooked as a factor in the success or failure of establishing native vegetation, but it can play a major role, especially on sites where summers are hot and dry and soil moisture levels are low. Until seedlings become established, wind can place extremely high demands for moisture on newly planted seedlings, severely limit growth, and ultimately lead to death. Wind can also be an important factor for surface stability, as discussed in [Section 3.8.5](#).

**How to Assess Wind**—Wind speed equipment is available but is most likely too costly for most designers. Site visits during different times of the year, especially in the summer, can give some indication whether wind is a problem. Visiting the site during the afternoon is important because this is the time of day when hot, dry winds negatively affect plants the greatest. Other site characteristics, such as position on the slope (e.g., ridgelines are more prone than valley floor), or proximity to forested environments—as forests often reduce wind speeds—can be used to infer wind strengths and directions.

## Mitigating for High Wind

**Road Design**—Designing islands of undisturbed vegetation to help break up wind patterns can aid vegetation establishment. The taller the plants left undisturbed, the greater the wind protection. Established trees, particularly those with low-growing branches, provide the greatest protection from wind.

**Wind Barriers**—Obstacles that block wind at the soil surface can be effective for early seedling survival. These obstacles can include trees and tall shrubs, shelterbelts, filter fabric, stabilized logs, large rocks, berms, and stumps. In using these structures,

## Figure 3-30 / Evapotranspiration rates

Evapotranspiration (ET) rates can be found for many [NOAA weather stations](#).

Plotting monthly evapotranspiration rates with precipitation rates (also found on the website) gives a good indication of the climate during plant establishment and growth phases. The graphs on the left show that the climate in Portland, OR, has a very favorable environment (low evapotranspiration and high precipitation) for seed germination in March; therefore, plant establishment is likely adequate without the need for extra mitigating measures. The weather in the fall is also conducive to germination and plant establishment. The climate in Bend, OR, during April and May when seeds in that area germinate has very high ET rates and very low rainfall; mitigating measures such as applying mulch over the seeds at sowing might be critical for success.

seedlings should be planted on the windward side.

**Tree Shelters**—Tree shelters completely surround seedlings and block them from the wind (Section 5.5.4). They are an effective means of reducing ET rates created by high winds during early establishment. Once the vegetation has emerged from the top of the tube, however, tree shelters no longer protect the emerging foliage from the wind.

**Shade Cards**—Shade cards are sometimes used to block wind, but they are less effective than the fully enclosed tree shelter (Section 5.5.3). When used to block wind, shade cards are placed on the windward side of the seedling, which is not necessarily the same location that cards would be placed if protection from sun is the objective. Often two shade cards are placed around the seedling for added protection against the wind. Placement of the shade cards at the height of the foliage affords greater protection to the seedling.

**Appropriate Species Selection**—The drying and damaging effects of wind are important considerations in appropriate species selection. A simple assessment of soil type and rainfall may not account for the effects of wind. Choosing hardier, wind-tolerant, and more drought-tolerant species may be necessary to establish vegetation on windswept sites. Find reference sites in windy locations to indicate which species are adapted to wind.

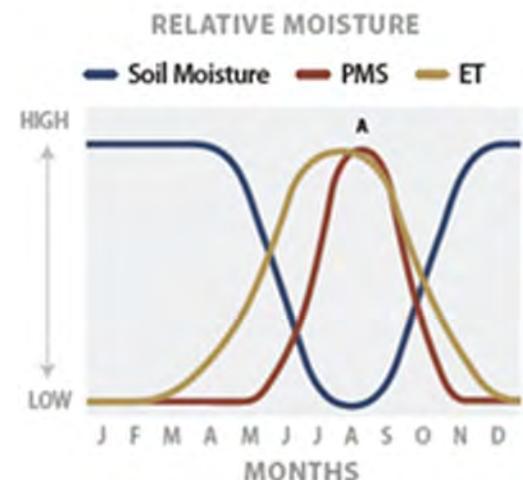
**Leave Surface Roughened**—A roughened soil surface can create a micro-basin or relief that protects young germinants from the drying effects of wind during the establishment phase (Section 5.2.2).

## Aspect

Aspect is the direction a slope is facing and is one of the predominant site characteristics affecting evapotranspiration. South and west aspects receive more solar radiation during the day and are warmer and drier, with higher ETs than north and east aspects. Soils on these south and west aspects dry out faster than north and east slopes. In spring, during seed germination, south and west aspects can dry very quickly between rainstorms, reducing the rates of germinating seeds. As seedlings emerge and grow through spring and early summer, temperatures on the south and west slopes continue to rise to very high levels, creating very unfavorable conditions for seedling establishment. Even with planted seedlings, high surface temperatures can damage stems near the ground line, severely affecting seedling survival and establishment (Helgerson et al 1992).

In climates where moisture is not the limiting factor, south and west slopes are often very productive and have greater cover. Warmer soil and air temperatures create a longer growing season, offsetting the effects of moisture stress on plant growth.

On high elevation sites, north and east aspects are cool, with much shorter growing seasons, compared to the south and west aspects, resulting in very different compositions of species. At high elevations, soils on south and west slopes stay warmer longer in the fall, providing an opportunity to plant in the late summer in time for seedlings to become established before winter arrives. During spring, at all elevations, south and west slopes warm up much sooner than north slopes, resulting in earlier seed germination and plant growth. The difference in soil temperatures between north and south aspects is a consideration for determining when to plant and what species to use.



**Figure 3-31 | Relationships among evapotranspiration, soil moisture, and plant moisture stress**

Conceptual relationship between evapotranspiration (ET), soil moisture, and plant moisture stress (PMS). In the western United States, PMS lags behind ET in late spring because soil moisture is still moderate to high from the winter rains. By midsummer (A), plant moisture stress has increased to its greatest level in the year because soil moisture is at its lowest. Newly planted seedlings undergo extreme stress during this period. Unless the seedlings are dormant or their root systems have grown deeper into the soil, where there is greater access to soil moisture, seedlings will die. In late summer and early fall, cooler weather returns and rains wet the soil, driving ET and PMS rates down again.

**How to Assess Aspect**—Aspect is

measured in the field by facing the fall line of the slope (the imaginary line a ball would roll) and taking a compass bearing downslope. A “northeast slope,” “northeast aspect,” “northeast exposure,” or “northeast-facing slope” all refer to an aspect with a compass bearing facing northeast.

Aspect can also be measured from topographic maps by drawing an arrow perpendicular to the contour lines and pointing the tip of the arrow downslope. Aspect is often a factor for delineating one revegetation unit from another due to the strong influence it has on the growth and survival of seeds, seedlings, and cuttings.

Soil and air temperatures differ greatly between aspects, and taking temperature measurements can be important for assessing the effects of aspect on revegetation. There are many types of recording devices available on the market, but only equipment that can download data to spreadsheets for analysis and graphing should be considered. Some equipment has become so inexpensive that more than one unit can be purchased (Figure 3-33).

## Mitigating for South and West Aspects

For most sites, any treatment that will shade vegetation on the south and west slopes from intense solar radiation should increase survival and growth of establishing plants.

**Overstory Vegetation**—Keeping overstory trees at a minimum density of one tree per tenth acre is a rule of thumb for reducing soil temperatures below lethal levels on south aspects (Helgerson et al 1992).

**Shade Cards**—Shade cards can significantly increase seedling survival on south aspects (Hobbs 1982; Flint and Childs 1984) (Section 5.5.3). Placing them close to planted seedlings so that the stem and lower portion of the seedlings are shaded from the afternoon sun maximizes their effectiveness (Helgerson et al 1992).

**Obstacles**—Large obstacles that cast significant amounts of shade on young seedlings will create a more favorable environment for seedling establishment and increase seedling survival (Minore 1971). These include stabilized logs, large rocks, berms, and stumps. Seedlings should be planted on the north and east side of these features to be shaded from the afternoon sun.

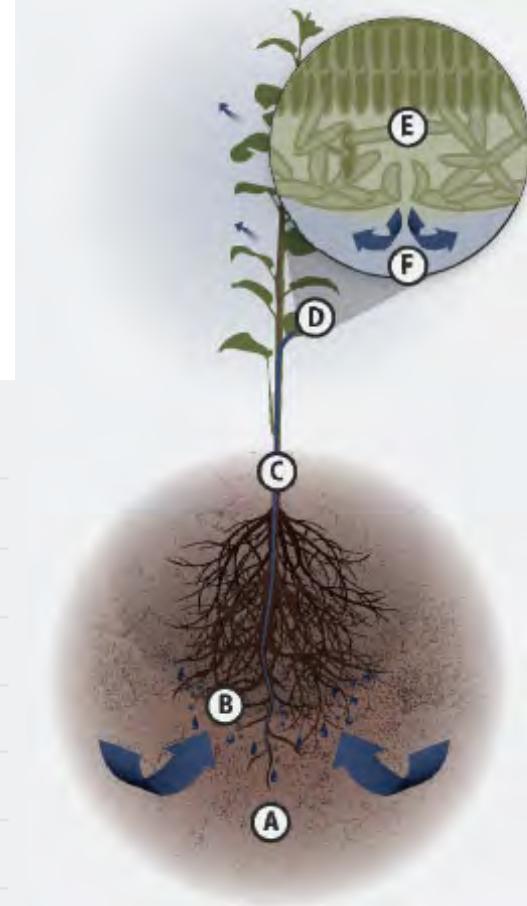
**Mulch**—On south exposures, the use of mulches as a moisture barrier should be considered for seedlings, seeds, and cuttings (Section 5.2.3). Avoid placing mulch in direct contact with the stem of the seedling.

**Species Mix**—The composition of species will probably be different for north and south aspects. Species adapted to hotter and drier environments are used for revegetating south exposures; those adapted to cool, moist environments are used on north aspects. Elevation can offset the effects of aspect. For example, species that

**Figure 3-32 | Plant moisture stress**

Plant moisture stress (PMS) is a measure of the tension or pull of moisture through a vascular plant. Much like a straw, when the demand for moisture at the surface of leaves is high, moisture is drawn from the stomata. This creates a pull of water through the leaves, stem, and down to the roots, which draws water from the soil.

| Location        | Water Potential (MPa) |
|-----------------|-----------------------|
| A Soil          | -0.1                  |
| B Plant roots   | -0.3                  |
| C Plant stem    | -0.6                  |
| D Plant leaf    | -0.9                  |
| E Plant stomata | -25.0                 |
| F Atmosphere    | -125.0                |



**Figure 3-33 | Temperature recording device**

Temperature recording technology has become smaller and very inexpensive. The iButton® shown next to the nickel can record more than a year of temperature data.

*Photo credit: David Steinfeld*

### 3—PLANNING

grow on low elevation, north aspects often occur several thousand feet higher on south aspects because of the difference in temperatures. Reference site vegetation surveys will guide in the selection of appropriate species for each exposure.

**Plant Material Rates**—South aspects often require a higher density of seedlings, cuttings, and seeds than north aspects to offset the expected higher mortality rates. Adjusting for increased mortality rates is made when calculating plant materials rate for seeds ([Section 5.4.1](#)), cuttings ([Section 5.3.5](#)), and plants ([Section 5.3.6](#)).

**Planting and Sowing Windows**—Take advantage of warmer spring and fall soil temperatures on the south exposures by sowing and planting earlier on these sites ([Section 3.14.2](#)).

## Competing Vegetation

Controlling competing vegetation around planted seedlings, whether native or non-native, reduces the rate at which water is withdrawn from the root zone and increases the potential for survival and growth. The rate at which water is depleted is a function of the type and amount of competing species. Grass species, for example, have a very fibrous root system in the upper soil horizon that allows them to withdraw moisture very quickly and efficiently during dry weather. Unless grasses are controlled, especially in the western United States, it is very difficult to achieve good survival or growth of planted seedlings in areas with high densities of grass. Perennial forbs are generally less competitive than grasses because their root systems are deeper and less concentrated in the surface where the seedlings are withdrawing moisture.

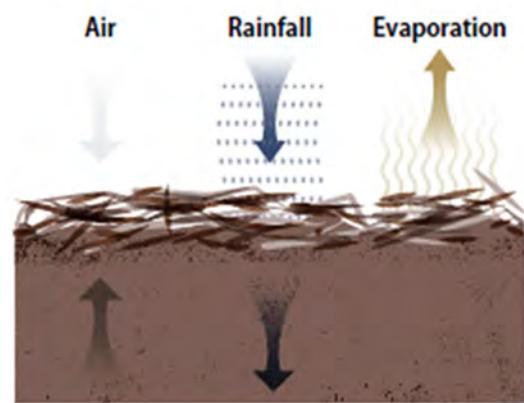
Revegetating with a seed mix is also affected by the type and quantity of competing vegetation. Those species that germinate earlier than the seeded species in the spring or fall will deplete soil moisture before the seeded species can establish. Cheatgrass is an example of an annual species that establishes quickly when soil temperatures are cool during early spring, depleting the surface soil moisture just as perennial species are beginning to germinate. How to assess and mitigate for competing vegetation is discussed in detail in [Section 3.11](#).

## Soil Cover

The thickness and composition of material that covers the soil surface influence many important soil properties covered in this manual, such as infiltration rates, interception losses, soil temperatures, surface erosion, runoff, and soil moisture loss. The following discussion focuses on soil cover as it affects soil moisture loss through evaporation.

Under undisturbed conditions, soil cover is predominantly composed of duff, litter, and stems that block the escape of soil moisture through evaporation ([Figure 3-34](#)). Mulches are also unfavorable seedbeds for weed seeds because duff and litter dry out quickly. Disturbed soils, on the other hand, are mostly composed of bare soil. Evaporation from the surface of bare soil can be high, extending to at least 6 inches below the soil surface, affecting seed germination and seedling establishment rates. Until roots of planted or seeded seedlings have extended farther into the soil profile, surface drying will negatively affect seedling establishment, especially on sites where water input and storage are already limiting.

**How to Assess Soil Cover**—Soil cover can be measured on undisturbed and disturbed reference sites or post-construction sites through the ground cover monitoring procedures outlined in [Section 6.3.1](#). In this procedure, the percentage of area in litter, duff, rock, vegetation, and bare soil is recorded and periodic measurements of litter and duff thickness are made.



**Figure 3-32 | Plant moisture stress**

Plant moisture stress (PMS) is a measure of the tension or pull of moisture through a vascular plant. Much like a straw, when the demand for moisture at the surface of leaves is high, moisture is drawn from the stomata. This creates a pull of water through the leaves, stem, and down to the roots, which draws water from the soil.

## Mitigating for Low Soil Cover

**Mulches for Seedlings and Cuttings**—Mulches create a more favorable environment for establishing seedlings and cuttings not only by reducing surface evaporation, but also by decreasing the amount of competing vegetation. There are three types of mulches for seedlings and cuttings—organic aggregate, sheet mulches, and rock mulches. The organic aggregates are thickly applied ground wood or bark, while the sheet mulches are made from non-permeable or slightly permeable plastic, newspaper, or geotextile. Rock mulches are composed of gravels, cobbles and stone. Mulches are placed around the base of the seedling and cover at least a radius of 1.5 feet from the base of the seedling ([Section 5.2.3](#) for how to install).

**Mulches for Sown Seeds**—Selecting and applying mulches over sown seeds differ from those selected for planted seedlings and cuttings. Mulch application for seedlings and cuttings is typically too thick for seeds to germinate and grow through. An ideal seedbed mulch is one that is applied at the highest rates without affecting seedling emergence. Long-fibered mulches, such as straw, hay, shredded wood, or wood strands, create the greatest loft or thickness. At the optimum thickness, these mulches allow some light to penetrate and space for seedlings to emerge. Short-fibered mulches, such as wood fiber and paper found in hydromulch products, are more compact and create less loft. While these products reduce erosion rates, they are not necessarily good as seed covers ([Section 5.4.2](#)).

### 3.8.4 NUTRIENT CYCLING

Nutrient cycling is the process by which sites store and release essential nutrients for plant survival and growth. There are 13 elements, or mineral nutrients, and each fills a specific role or function in plant development and each possesses individual characteristics of movement and storage in the soil. This manual will not attempt to explain the role and function of each mineral nutrient (there are many good textbooks on this subject). It will instead focus on how nutrients cycle through vegetation and soils; how they are captured, stored, and released; and what site components are essential to support these processes. In contrast to an agricultural system of managing optimum growth in crops through fertilization, the goal in wildlands revegetation is to create an environment that will support a self-sustaining native plant community that can develop through successional processes. This includes facilitating the establishment of nutrient cycles in a way that conserves, cycles, and builds nutrients in the system.

Mineral nutrients are stored in: soil, live or dead vegetation, and rock. They are slowly released over time at varying rates. The rates at which nutrients are released from each source will determine their availability for plant uptake. Rock and fallen trees, for example, both hold essential nutrients but release them to the soil at significantly differing rates. The fallen tree can take up to 100 years to decompose and release its nutrients; the weathering of rock might take over 100,000 years. Soil, on the other hand, can release nutrients in the order of weeks and months. Once released, these nutrients are taken up by plants, lost through leaching or erosion, or reabsorbed in the soil.

On undisturbed sites, there is a dynamic exchange of nutrients throughout the year. Plants absorb nutrients from the soil through the root system and assimilate them into vegetative biomass. As plants, or portions of plants, die, they drop leaves, branches, and stems to the ground where they eventually decompose and return nutrients to the soil. The nutrients are stored in the soil and once again become available for plant uptake. This natural process of nutrients moving from soil to vegetation and back again is referred to as nutrient cycling. The factors important in nutrient cycling are as

follows:

- Topsoil
- Site organic matter
- Soil nitrogen and carbon
- Nutrients
- pH
- Salts

In a healthy plant community, nutrients are constantly recycled with a minimum amount of nutrients lost from the site. On drastically disturbed sites, however, nutrient cycling functions poorly, if at all. The topsoil, which holds the greatest concentrations of available nutrients and supports the primary microbial activity on the site, is missing or mixed with the subsoil. Organic matter, which is a primary source of long-term nutrient supply and an energy source, has also been removed. Soil nitrogen, the most critical nutrient for plant growth and site revegetation, is lacking. Soil nitrogen governs how quickly vegetation will return to a disturbed site and how much biomass it will ultimately support (Bloomfield et al 1982). Its availability is closely regulated by the amount of carbon present in the soil, which is also in flux on highly disturbed sites.

For many sites that lack topsoil, the subsoil in its place may have pH values that are higher or lower than the pH of the original topsoil. pH values at extreme ranges affect nutrient cycling by making many nutrients insoluble and unavailable for plant uptake. In addition, increased soil salinity, which can be caused by soil disturbance and amendments, can disrupt nutrient availability and provide unfavorable conditions for plant establishment.

## Topsoil

The topsoil is the horizon directly below the litter layer that is characterized by high organic matter, abundant roots, healthy microbial activity, good infiltration rates, high porosity, high nutrient content, and high water-holding capacity (Jackson et al 1988; Claassen and Zasoski 1998). Most nutrient cycling takes place in the topsoil where the greatest biological activity occurs. Decomposing microorganisms flourish, feasting on dead vegetation and roots, releasing stored nutrients back to the soil. Most life forms occur in forest, prairie, and range top-soils, including mammals, reptiles, amphibians, snails, earthworms, insects,



**Figure 3-35 / Topsoil**

Topsoil is the upper soil horizon and is generally darker, more friable, and has more roots than subsoil.

*Photo credit: Thomas D. Landis*

nematodes, algae, fungi, viruses, bacteria, actinomycetes, and protozoa (Trappe and Bollen 1981). The top 6 inches of an acre of forest topsoil can contain as much as a ton of fungi and a half of a ton of bacteria and actinomycetes apiece (Bollen 1974). Topsoil depth is highly correlated with the nutritional status of the soil and, in forests of the western United States, has been found to be highly correlated to site productivity (Steinbrenner 1981). Topsoils possess humus, which is what gives topsoils their dark color (Figure 3-35). Humus is a stable end-product of decomposition, important for nutrient storage, soil structure, and water-holding capacity.

Sites lacking topsoil have significantly reduced productivity, and obtaining even minimal revegetation can be very difficult. Planted seedlings often fail or growth is significantly reduced, resulting in inadequate plant cover to protect the soils from erosion. Growth of planted trees can be reduced by one-third to one-half when planted in subsoil instead of topsoil (Youngberg 1981). Restoring these sites to functioning plant communities is unlikely without mitigating measures.

**How to Assess Topsoil**—On undisturbed sites, topsoil is visually differentiated from the underlying subsoil by having darker colors, less clay, better soil structure, and higher abundance of fine roots. In forest soils, topsoil depths can be more difficult to differentiate from subsoils because the color changes are not always distinct. Other attributes, such as the abundance of roots, lack of clays, soil structure, and lower bulk density, can be used instead. On construction sites, topsoil has either been removed and stockpiled or mixed in with the subsoil.

## Mitigating for Lack of Topsoil

**Minimize Soil Disturbance on “Fragile Soils”**—Where soils are especially fragile and reconstructing topsoil conditions is difficult, it is particularly important to keep the “disturbance footprint” to a minimum (Claassen et al 1995). Sites with fragile soils include decomposed granitic soils, serpentine soils, high elevation soils, and very acidic or basic soils.

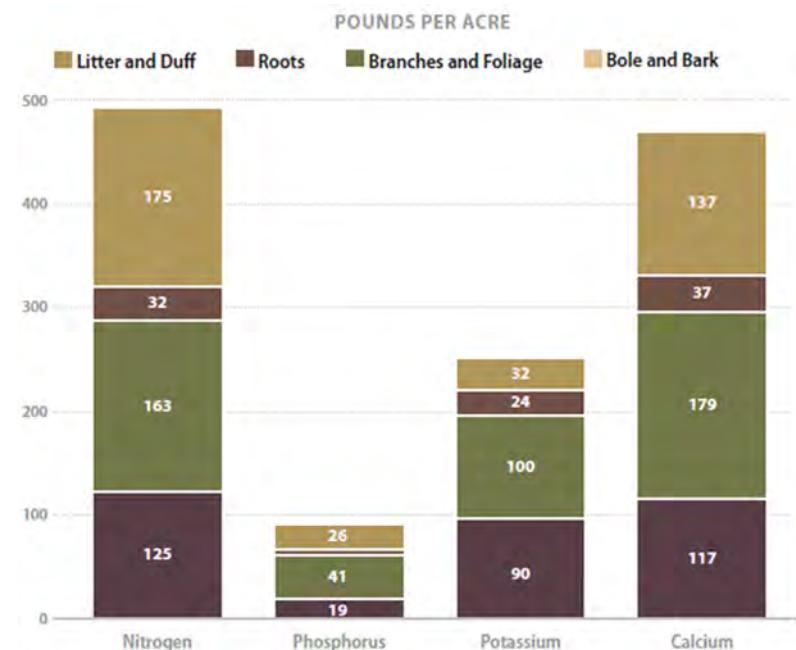
**Salvage and Reapply Topsoil**—An effective practice in revegetating highly disturbed sites is salvaging and reapplying topsoils (Section 5.2.4). This practice has been found to greatly increase plant growth and ground cover (Claassen and Zasoski 1994). Topsoil salvage and application require good planning, implementation oversight, and topsoil surveys. In the planning phase, a survey of the planned road corridor identifies the location of topsoil, the depth, and nutrient status through laboratory testing (Section 3.8.4, see Topsoil). After topsoil is removed and appropriately stored, it is reapplied to the disturbed site, ideally at depths similar to pre-disturbance reference sites.

**Create Manufactured Topsoil**—When topsoil is not available, “manufactured topsoil” can be created in situ or produced offsite and imported (Section 5.2.4, see Manufactured Topsoil). Manufactured topsoil will lack the native seed bank and some of the biological components of topsoil, but it can re-create a rooting zone high in nutrients and organic matter, with good water-holding capacity, porosity, and infiltration.

**Create Planting Islands**—If sources of manufactured or natural topsoil are scarce or too costly for broad scale applications, placing available topsoil in strategic locations, such as planting islands, can create a mosaic of productive growing sites.

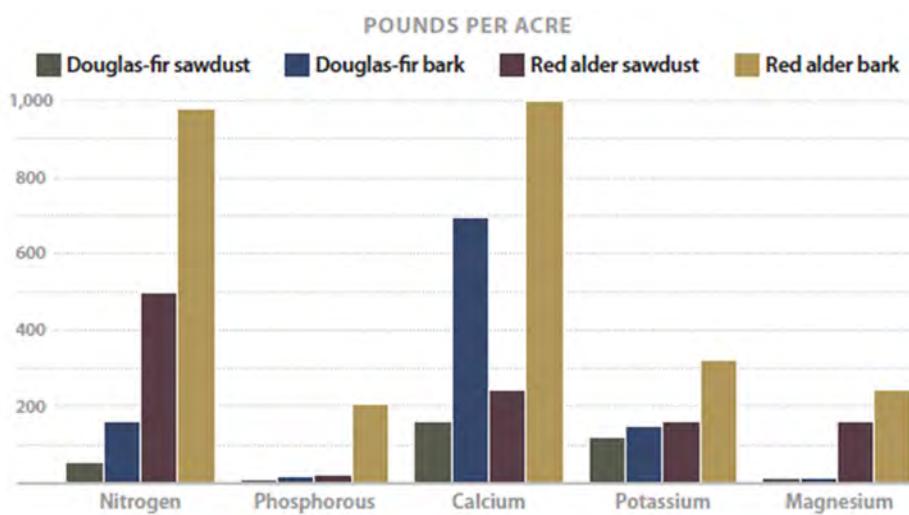
## Site Organic Matter

Site organic matter (OM) consists of plant materials in all stages of decomposition, including wood, bark, roots, branches, needles, leaves, duff, litter, and soil organisms. From a nutrient cycling standpoint, site organic matter assimilates nutrients drawn from the



**Figure 3-36 | Nutrients in forests stands**

Most of the nutrients found in a young Douglas-fir stand reside in the litter, duff, and branches (adapted from Cole and Johnson 1981).



**Figure 3-37 | Nutrients in Douglas fir and alder**

Comparison of pounds per acre of nutrients resulting from bark or wood from Douglas-fir and alder trees, based on 2 inches of applied organic matter (Rose et al 1995).

soil into live vegetation. Depending on the type of vegetation and the productivity of the site, the amount of nutrients held in organic matter can be significant. Figure 3-36 shows the quantities of four major nutrients held in the organic matter of a young Douglas-fir stand, and Figure 3-37 shows the quantity of major nutrients found in the application of 2 inches of material derived from Douglas-fir and alder. These two examples show the possible nutrient reserves that organic matter can contribute to a disturbed site if they are kept on the site or processed into mulch or soil amendments and reapplied.

While plants are essential in nutrient cycling, equally important are the decomposing organisms that release nutrients to the soil. Decomposers consist of

thousands of specialized species of animals, insects, fungi, bacteria, and actinomycetes that survive on organic matter. Decomposing organisms not only release nutrients but are essential to the development of soil structure (Section 3.8.2, see Soil Structure).

On an undisturbed site, organic matter is in all stages of decomposition, from recently dead trees to soil humus, the end product of hundreds of years of decomposition. This understanding is important for restoring a site to a functioning plant community because it is a reminder that nutrients are released throughout the life cycle of a plant community, not just at the beginning of a revegetation project. A site that has a range of organic matter in all stages of decomposition not only conserves nutrients but slowly meters them out over time. In addition, organic matter is an essential nesting habitat for many pollinator species. For example, tunnel-nesting bees nest in dead standing trees or piths of stems and twigs of shrubs, grasses, and forbs.

The rate at which organic matter decomposes and releases nutrients is a function of: soil to OM contact, OM particle size, the ratio of carbon to nitrogen (C:N ratio), temperature, and moisture. Decomposition rates of organic matter are highest in soil because the greatest microbial activity occurs when soil is in direct contact with OM (Slick and Curtis 1985; Rose et al 1995). Organic matter placed on the surface of the soil as mulch will decompose at a much slower rate than organic matter incorporated into the soil because there is less contact with the soil.

Organic matter particle size plays an important role in the rate of decomposition. Within the soil profile, the smaller-sized OM fractions decompose faster than the larger fractions due to greater surface area in contact with soil. Roots, leaves, needles, and very finely ground sawdust or bark often decompose much faster than larger materials, such as buried logs or large diameter branches. Materials with C:N ratios, such as wood or bark, decompose much slower than materials with low C:N ratios, such as green leaves and grass cuttings. As Figure 3-38 illustrates, high C:N organic matter will take much longer to decompose than low C:N material, but both will decompose faster when they are reduced in size.

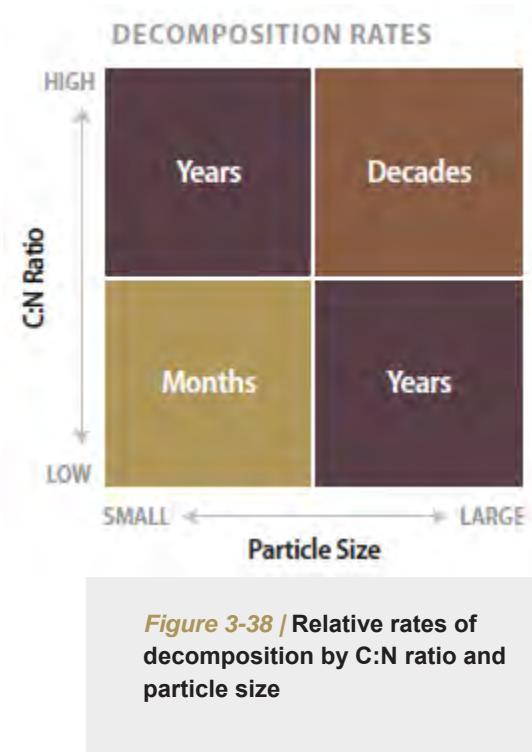
Moisture and temperature also control decomposition rates; cold and dry environments have very slow rates compared to warm, moist sites.

**How to Assess Organic Matter**—Duff and litter on reference sites can be measured through transects or plots (Section 5.2.3, see Litter and Duff). Estimating forest biomass of down, woody materials in different size classes can be done using photo series guides (Maxwell and Ward 1976a, 1976b, 1979). Fire specialists are experienced in estimating the amount of biomass in forested environments.

## Mitigating for Lack of Site Organic Matter

**Salvage and Reapply Litter and Duff**—Duff and litter can store a significant amount of nutrients, especially on sites where layers are deep. It can be salvaged separately (Section 5.2.3, see Litter and Duff) or mixed together when topsoils are salvaged (Section 5.2.4).

**Process and Apply Organic Matter**—Road projects constructed through forested sites can generate high amounts of biomass. These materials can be good sources for slow-release nutrients and carbon. Methods of processing this material and reapplying it to constructed slopes are discussed in Section 5.2.3 (see Litter and Duff). Processed organic matter can be applied directly to the soil surface as a mulch or mixed into the soil as a soil amendment. High C:N organic materials, such as sawdust and bark, can be placed on the soil surface to prevent long-term nitrogen tie-up in the soil or composted for several years to lower the C:N ratio before adding as a soil amendment. Lower C:N materials, such as leaves, needles, and branches, can be incorporated in the



soil with some addition of slow-release nitrogen to reduce the effects of nitrogen tie-up.

**Apply Composts to Soil Surface**—When compost is applied to the surface of the soil (compost blanket), it functions as a soil cover to protect the soil from surface erosion while slowly decomposing and adding nutrients as the plant community develops. Decomposition rates of surface-applied compost are slower than if it were incorporated into the soil because there is very little soil contact at the soil surface. Nutrients are released at a slower rate and are available to the site longer. Leaving the soil surface very rough prior to the application of compost creates more soil-to-compost contact, which can increase the rate of decomposition of the compost. Tackifiers are added to compost to reduce the potential for the material to move off the site. Compost blankets function very differently than mulches; compost blankets are great media for seed germination and plant establishment whereas mulches are not (Section 5.2.3).

**Salvage and Place Large Wood**—Large wood can be salvaged and placed in areas such as abandoned roads for long-term site productivity, microsite planting, pollinator nesting habitat, and soil erosion control structures (Figure 3-39). When placed in contact with the soil, large wood helps stabilize the surface of the soil from sheet and wind erosion, and can be used as buttresses to stabilize slopes. Seedlings planted on the north side of the logs can be protected from wind and sun during establishment. Large and small wood can be placed at the outlets of culverts as obstacles to capture and store sediments, reducing the amount of sediment reaching live streams (Burroughs and King 1989; Ketcheson and Megahan 1996). When large wood is placed upright, it can be used as barriers to off-road vehicle traffic, nesting habitat for many pollinator species, and feeding and nesting habitat for a range of wildlife species.

## Soil Nitrogen and Carbon



**Figure 3-39 | Large wood creates pollinator habitat and reduces soil erosion**

Placement of large wood adds long-term organic matter while creating microsites for planting seedlings and habitat for pollinators and wildlife. Large wood can also slow runoff and detain sediments from surface soil erosion.

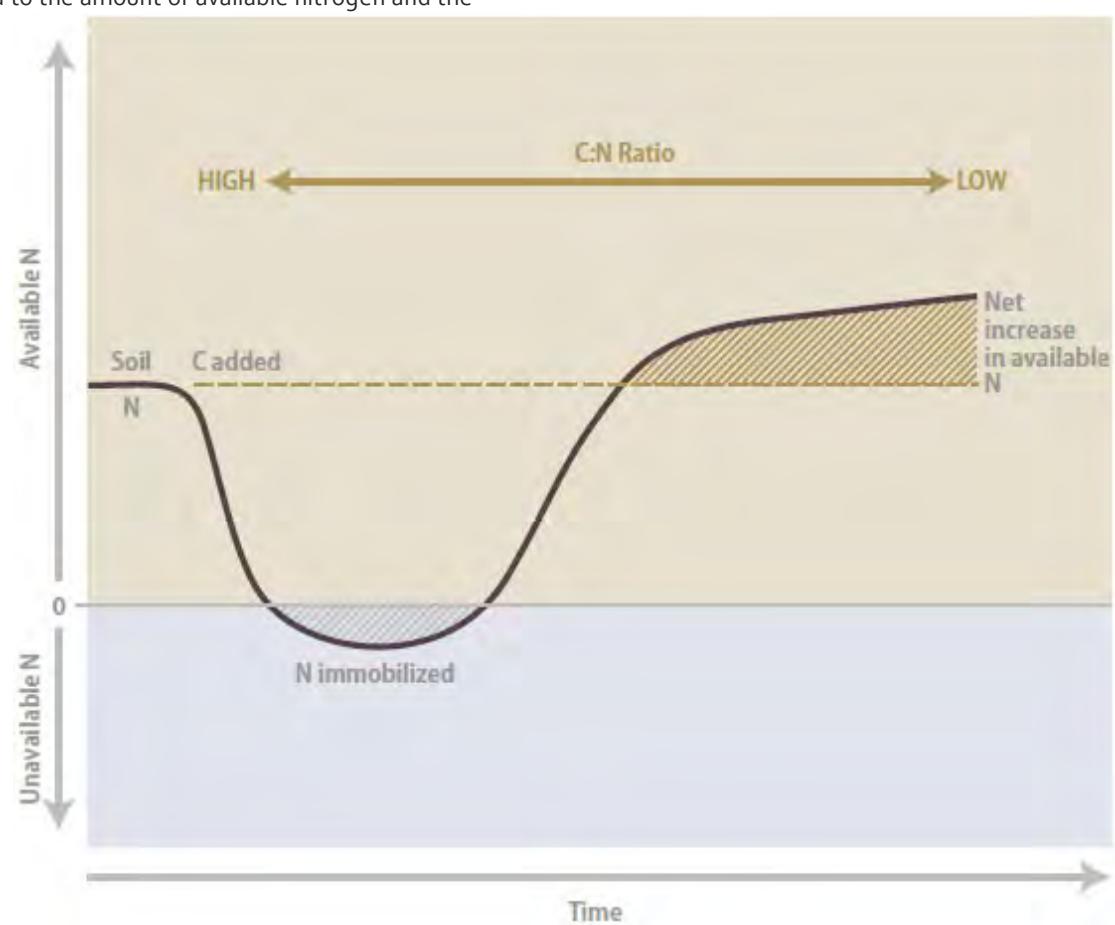
*Photo credit: David Steinfeld*

Nitrogen (N) is discussed separately from the other mineral nutrients because of its critical importance to plant growth and long-term development of plant communities. Carbon (C) is included in this discussion because of its unique relationship to nitrogen availability. Carbon governs the amount of available nitrogen in the soil while nitrogen regulates the rate at which carbon is broken down. Both factors play a critical role in microbiological activity and the development of soil properties.

**Carbon to Nitrogen Ratio (C:N Ratios)**—Carbon is the energy source for soil microorganisms, and practically all site nitrogen eventually passes through these microorganisms (Woodmansee et al 1978). The rate at which carbon (or organic matter) decomposes is directly related to the amount of available nitrogen and the type of dominant microbes present in the soil. The greater the nitrogen, the greater the decomposition rates. If decomposing organisms do not find sufficient nitrogen in the organic matter, they will withdraw it from the soil, leaving little or no nitrogen for plant growth. This is usually a temporary condition but could last several years. The tie-up of nitrogen can greatly affect the establishment of vegetation if organic amendments, such as wood chips, are incorporated into the soil without supplemental nitrogen.

With time, nitrogen is eventually released from the organic matter by microbial activity. This nitrogen, plus nitrogen released from dead and decomposing microorganisms, becomes available for plant growth. As organic matter breaks down further, there becomes a net increase in available nitrogen. In the last stage of decomposition, microorganisms move to a steady rate of decomposition, releasing a constant nitrogen supply (Figure 3-40). This process can take several years or more depending on site factors and organic matter levels. The C:N ratio is an indicator of whether nitrogen will be limiting or in surplus. A C:N ratio of 30:1 or greater indicates that decomposing organisms have consumed the available nitrogen in the soil, leaving little if any available nitrogen for plant growth. Plants respond by turning yellow and stunted. A C:N ratio below 18:1 is an indication that the decomposing organisms are releasing available nitrogen from the breakdown of organic matter at rates that exceed their need, thereby increasing nitrogen for plant uptake (Claassen 2006). For example, undisturbed topsoils typically have ratios of 10:1 to 12:1, which indicates that nitrogen is sufficiently available for plant growth (Tisdale and Nelson 1975). Dry hay, on the other hand, has a C:N ratio around 40:1, indicating nitrogen will probably be limiting for some period of time if the hay is incorporated into the soil.

The use of high C:N materials is often discouraged because of concerns about tying up nitrogen. However, there are strategies where the use of high C:N materials can aid in achieving project goals. One use of high C:N materials is to apply it to the soil surface or incorporate it into the top several inches of soil to intentionally tie up nitrogen. The lower availability of N at the surface creates a less than optimum growing environment for the establishment of annual weedy species, which thrive on high nitrogen environments.



**Figure 3-40 / Release of available nitrogen through decomposition**

Available nitrogen (N) levels change as organic matter carbon (C) is added to the soil. High ratios move to low ratios during decomposition. Nitrogen is tied up in microorganisms during the immobilization phase (blue shaded area) and unavailable to plants. With time, nitrogen becomes available again and, at some point, exceeds the original level (green shaded area). Nitrogen is then released at a constant rate (modified after Havlin et al 1999).

## Soil Nitrogen Capital

Soil nitrogen capital can be categorized into three nitrogen pools, or reserves, based

Roadside Revegetation: An Integrated Approach to Establishing Native Plants and Pollinator Habitat

on its availability in the soil:

- Available nitrogen (referred to as “extractable nitrogen”)
- Slowly available nitrogen (referred to as “mineralizable nitrogen”)
- Unavailable nitrogen (“humified organic” or “fresh” forms)

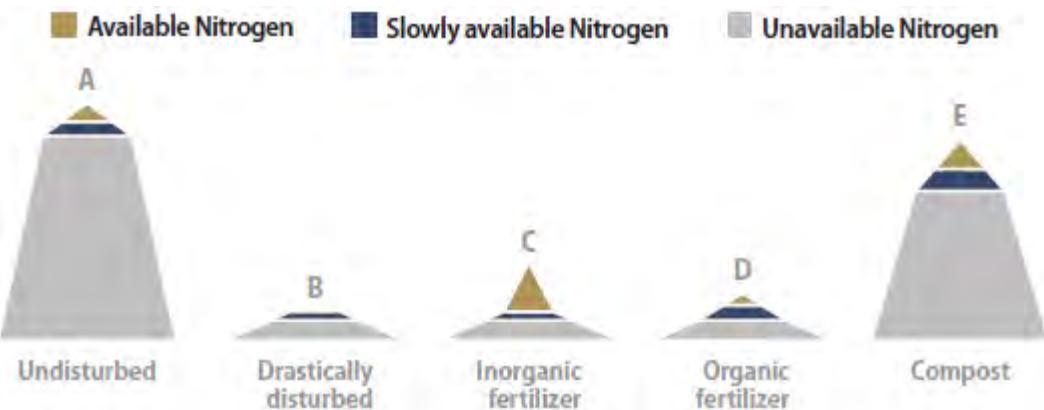
Nitrogen capital can be viewed much like our banking system. Cash received from the bank teller is comparable to “available nitrogen.” When money runs low, the teller replenishes it with money from the bank vault (similar to “slowly available nitrogen”). Banks are backed up by money held in an extremely large banking reserve system (“unavailable nitrogen”). While this money is not accessible, it is very important for the long-term stability of the banking system. Unavailable nitrogen is like the banking reserve system in that it backs up the nitrogen system and ultimately releases nitrogen to the plant community.

As with the banking reserve system, having high reserves of both slowly available and unavailable nitrogen ensures that available nitrogen levels will be released at constant rates over time which is necessary for the development of a sustainable plant community. Figure 3-41 shows the relationship of different revegetation treatments on nitrogen capital.

On highly disturbed sites, all nitrogen reserves are low. The course of action in typical revegetation projects is to apply inorganic fertilizers during the seeding operation. While this immediately makes nitrogen available, it does little for increasing long-term nitrogen reserves. Within a year of application, most soils will need more available nitrogen to sustain plant growth. Alternatively, organic fertilizers provide a combination of available and slowly available nitrogen. These fertilizers release nitrogen over several years but are typically applied at rates not great enough to bring the nitrogen reserves up to levels for long-term plant community establishment (Claassen and Hogan 1998). Applying topsoil, composts, or low C:N organic matter into the soil are mitigation treatments that create the reserves of unavailable and slowly available nitrogen important for a constant supply of available nitrogen over time.

## Minimum Soil Nitrogen Levels

Total soil nitrogen is the sum of available, slowly available, and unavailable nitrogen reserves. The level of total soil nitrogen varies by plant community and ecoregion. It can range from 20,000 lb/ac in deep forest soils of the Washington and Oregon coast (Heilman 1981) to as low as 800 lb/ac in desert grasslands of southern New Mexico (Reeder and Sabey 1987). Shortgrass prairies in northeastern Colorado and shrub-steppe prairies of the Great Basin have a range of total nitrogen from 4,000 to 5,000 lb/ac (Reeder and Sabey 1987). Sites that are drastically disturbed often have nitrogen rates below 700 lb/ac. These sites cannot fully support vegetative cover. A minimum, or threshold, level of total soil nitrogen required for a self-sustaining ecosystem has been suggested at 625 lb/ac (Bradshaw et al 1982) to 670 lb/ac (Dancer et al 1977) for drastically disturbed sites. But Claassen and Hogan (1998) suggest much higher rates might be necessary. In their research on granitic soils in the Lake Tahoe area, they found a good relationship between total soil nitrogen and the percentage of plant cover. Sites with greater than 40 percent ground cover contained at least 1,100 lb/ac total soil



**Figure 3-41 | Managing nitrogen capital**

Undisturbed sites (A) have very high total nitrogen levels, with over 95 percent tied up in organic matter and not available (gray). Slowly available nitrogen (blue) makes up 1 percent to 3 percent of the total nitrogen; available nitrogen for plant uptake (tan) is less than 2 percent of the total nitrogen. Nitrogen capital is essentially removed on drastically disturbed sites (B). The addition of inorganic fertilizer (C) dramatically increases available nitrogen but does little to build nitrogen capital. The application of organic fertilizers (D) raises the available and the slowly available nitrogen but does not add to the long-term reserves. Adding compost to the soil can increase available, slowly available, and total nitrogen reserves (E) to levels comparable to undisturbed soils (A).

nitrogen in the surface foot of soil. This implies that to maintain a minimum of 40 percent plant cover, sites like these have to contain at least 1,100 lb/ac of nitrogen, with higher nitrogen levels necessary for higher plant cover (Figure 3-42).

Target levels for available nitrogen released annually from nitrogen sources for plant growth range from below 27 to 50 lb/ac (Munshower 1994). These are nitrogen levels that should be considered when calculating fertilizer rates (Section 5.2.1, see Determine Fertilizer Application Rates).

**How to Assess Soil Nitrogen and Carbon**—Soil testing for nitrogen can be conducted for: topsoils that will be salvaged, reference sites, and post-construction soil materials. Procedures for collecting soil samples are presented in Inset 3-2. The following nitrogen tests are available:

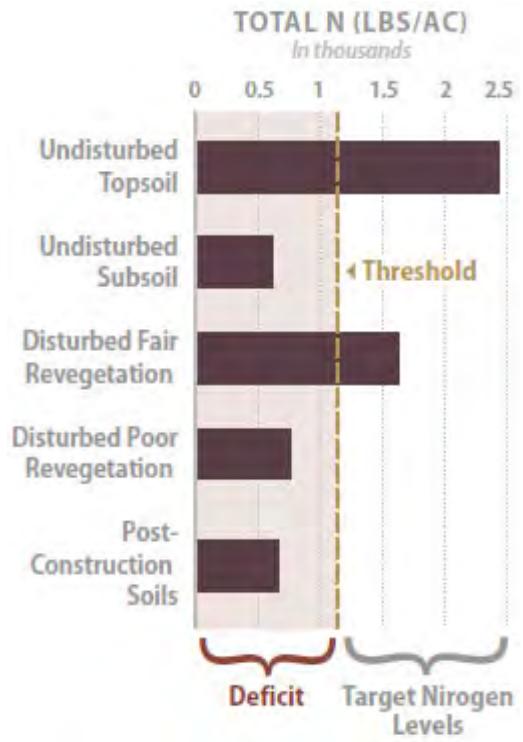
- **Total nitrogen**—Total nitrogen is an important test to request; the results will be used to determine nitrogen thresholds and nitrogen amendment needs. The common total nitrogen tests are Leco and Kjeldahl. Total nitrogen has been found to correlate well with plant cover (Claassen and Hogan 1998).
- **Mineralizable nitrogen**—This test requires the soil samples to be incubated for a period of time and then tested for available nitrogen. The results indicate the amount of slowly available nitrogen present in the sample. While this test is not widely used, it nevertheless is a very good test to perform because the results correlate well with expected plant cover (Claassen and Hogan 1998). There are several types of incubation tests, so it is good to confer with the soil laboratory as to which tests would be most appropriate.
- **Extractable nitrogen**—This test is less meaningful because it only indicates available nitrogen, not what is in reserve. This test is often included in a soil testing package. The extractable N pool has the lowest correlation to the amount of plant cover growing on a site (Claassen and Hogan 2002). The most common test for extractable nitrogen is zN KCl extract.

Nitrogen testing for composts and organic matter should be done by laboratories specializing in these tests. These laboratories should follow the testing procedures outlined in the Test Methods of the Examination of Compost and Composting (TMECC) explained in Section 5.2.3 and Section 5.2.5.

**Nitrogen Analysis**—Soils laboratories report nitrogen in a variety of units, such as gr/l, ppm, mg/kg, ug/g, and percent. Unless these values are converted to pounds per acre, it is difficult to determine rates of fertilizer, compost, or topsoil necessary to restore site nitrogen. Use Table 3-9 (Line E) to convert lab values to total pounds per acre of nitrogen. These calculations account for soil bulk density, soil thickness, and coarse fragment content, which affect the total nitrogen levels of a site.

**Nitrogen Thresholds and Deficits**—Each plant community has a total nitrogen requirement it must meet in order to develop into a functioning and self-sustaining system. For successful revegetation efforts, a practical goal is to meet the minimum target, or threshold level, for total nitrogen. Threshold values, however, are not found in textbooks and are developed from soil tests of disturbed and undisturbed reference sites. Conducting nitrogen tests on disturbed reference sites where revegetation efforts have succeeded, as well as reference sites where revegetation efforts have failed, can help determine a threshold value (Figure 3-42). Alternatively, conducting soil tests on undisturbed reference sites will define the optimum nitrogen levels and also bracket target nitrogen levels. Converting soil test results into total nitrogen per acre is shown in Table 3-9, line E.

Post-construction soils are typically deficient in nitrogen. In order to develop a strategy for bringing soil nitrogen above threshold levels, it is important to determine



**Figure 3-42 | Determining total N threshold values from reference sites**

In this example, the total N threshold was estimated at 1,100 lb/ac (average of disturbed reference sites with “poor” and “fair” revegetation). Total N in post-construction soils was 650 lb/ac, making these soils deficient by 450 lb/ac. The undisturbed topsoils of reference sites showed a total N of 2,430 lb/ac, which set the target levels of nitrogen between 1,100 and 2,430 lb/ac.

the approximate nitrogen deficit. Table 3-9 shows how this is calculated by subtracting the total nitrogen value of post-construction soils (Line E) from the threshold nitrogen value (Line F).

### **Inset 3-2 / Soil testing**

Soil testing is a means of describing those soil characteristics that cannot be observed or accurately measured in the field. The tests include analysis of chemical properties, including pH, soluble salts, macronutrients, micronutrients, and organic matter, as well as physical properties such as density, water-holding capacity, and texture. Soil testing is costly and if not sampled, analyzed, and interpreted properly can lead to unneeded and expensive soil amendments and application practices. In many respects, it is better not to test soils than to test them or interpret the results incorrectly. Some laboratory test results found for many soil series in the U.S. is available at the National Cooperative Soil Survey Soil Characterization Data website.

Soil testing is performed with topsoil recovery surveys and reference site surveys (discussed in other sections) to identify soil physical and chemical factors that will limit plant growth, develop site-specific soil quality targets, and develop a set of revegetation treatments that will increase short- and long-term soil productivity targets. The three components of soil testing are soil sampling, lab analysis, and interpretation of lab results. Adhering to an established procedure for each component of soil testing is critical for developing appropriate revegetation treatments.

**Sampling soil**—Soil sampling is the field collection of soils in a manner that best represents the soils of an area. The number of soil samples taken within a project area is usually kept to a minimum because of the expense of collecting and handling the samples and the cost of laboratory analysis. Taking too few samples to describe a project site, however, can be misleading, especially if the soils are extremely variable. This leaves the designer with the challenge of determining the best approach to collecting soil samples in a way that most accurately represents the sites being described.

The following guidelines are useful in developing a sampling strategy for soil testing:

- **Determine the area to be sampled**—The areas to be sampled are called sampling areas and they typically encompass a vegetation unit, an individual topsoil stockpile, similar topsoil salvage areas, or a reference site. For most projects, only one soil sample is collected from a sampling area. For this reason, it is important to select a collection site (an area where soil samples are collected) that best represents the sampling area. Only for small sampling areas, such as topsoil stockpiles or reference sites, will the collection site be the same as the sampling area. For larger sampling areas, such as revegetation units or topsoil salvage areas, the collection site will be a smaller, representative area within the sampling area.
- **Collect multiple subsamples**—Once a collection site has been identified, a set of subsamples are collected. Collecting soil from one point is never enough. The number of subsamples to collect within a collection site should be based on the site's variability. Small collection sites generally need fewer subsamples than larger areas because these sites are usually less variable. Undisturbed sites are typically more homogenous than disturbed sites and therefore need fewer subsamples. Guidelines for the numbers of subsamples to collect range from 6 for very homogenous sites to 35 for large, heterogeneous sites.
- ✓ **Randomly collect subsamples**—Subsamples should be collected randomly within the collection site. For small areas like reference sites or stockpiled soil, the samples can be collected on a grid system. For very large areas, samples can be collected in a zigzag or "W" pattern at predetermined intervals.
- ✓ **Determine sampling depth**—The objectives for soil sampling are reviewed and the sampling depth is determined. If the objective is to characterize the soil for topsoil recovery, then the soil samples are collected only in the topsoil horizon, in which case the depth of the topsoil will have to be determined prior to sampling. If it is known that the surface soil including duff and litter will be removed to 15 inches during topsoil salvage, then collection depth would be a sample 15 inches deep that included the duff and litter layers. If soil sampling objectives are to determine the nutrient levels of a topsoil pile, the entire pile becomes the collection site and the subsamples are collected from various depths within the piles, as well as around the pile to obtain a representative sample. If the designer felt that the interior of the pile was significantly different in nutrient status or mycorrhizae, then the pile could be stratified into two collection sites—the exterior of the pile and the interior—and sampled separately. It may also be important to sample the subsoil because this will be the condition of the soil after construction and before mitigation.

- ✓ **Collect a representative slice of soil**—It is important to evenly sample the predetermined depth of soil. For example, if the sampling depth is 0 to 15 inches deep, then the entire section of soil is equally sampled for each subsample.
- ✓ **Mix subsamples**—Subsamples for a sampling area are placed in a clean bucket and mixed thoroughly. From the composite subsamples, the required amount of soil is removed to send to the lab for analysis.
- ✓ **Determine coarse fragment content**—If the soils are high in coarse fragments, the samples can be sieved in the field. If the soils are dry, a 2mm sieve can be used. This will reduce the amount of soil to haul out of the survey area and also give an estimate of the coarse fragment content. Soils can be sieved back at the office prior to sending to the lab. If the samples are wet or moist, they will need to be air dried prior to sieving. The percent coarse fragment content is recorded, which includes large and small coarse fragments. This information will be used later to modify the lab results.
- ✓ **Selecting a lab**—The criteria for selecting a soil lab is typically based on costs, turnaround time, analytical tests, and consulting services. Most labs offer pH, nutrients, nitrogen, and organic matter tests for under \$80 per sample (2016 prices) and deliver the results within two weeks of receiving the soil samples. For an added fee, laboratories will interpret the results of the analysis. While these are important reasons for selecting a lab, the primary criteria for selecting a lab should be based on the quality of the testing facilities.

A common assumption is that all labs are of similar quality in their analytical testing, and that if a group of labs were sent the same soil sample they would report similar results for most tests. This is not typically the case, as several university reviews of laboratories have shown (Neufeld and Davison 2000; Rose 2004). In one comparison, eight reputable laboratories reported widely differing results for all soil nutrients when sent identical soil samples (Rose 2004). One reason for the variation in results is that usually several testing procedures can be used to quantify a soil parameter. Some methods have greater accuracy and precision than others. The soil testing industry at this time has not settled on an agreed upon set of analytical methods to use. Even when the same tests are performed, labs often report different levels of accuracy (Rose 2004).

Soil laboratories can voluntarily participate in the North American Proficiency Testing (NAPT) program that will assess the quality of their analytical procedures. In this program, NAPT periodically sends all participating labs identical soil samples. Each lab analyzes the samples for mineral nutrients using established analytical procedures, then sends the results back to NAPT. The results from all labs are compiled and analyzed statistically and each lab is sent a report on how their results compared to the other participating labs. NAPT suggest that the accuracy be less than 10 percent of industry values and precision no greater than 15 percent of industry values (Neufeld and Davison 2000). These reports are not available to the public, but laboratories might share them if asked. NAPT is not a certification program but is often a basis for a soil lab quality control program.

The following is a checklist for selecting a high-quality lab (modified from Neufeld and Davison 2000):

- ✓ Does the lab have a quality control program? If they do, ask them to explain it.
- ✓ Does it participate in a proficiency testing program (such as NAPT)?
- ✓ Will they share the results of proficiency testing program?
- ✓ Does the lab use established analytical methods (the most appropriate for soils in the geographic area being tested)?

If a “no” is given for the answers to any of these questions, another soil testing facility should be considered. If the selection is between a couple of labs, consider sending duplicate soil samples with known properties (“checks”) to each lab and compare the results using the NAPT suggested standards for accuracy and precision. Soil “checks” can be purchased through a proficiency testing program. Once a lab is selected, continue to ask for quality control reports. If the budget allows, periodically send duplicate “check” soil samples with regular soil samples to assess accuracy and precision.

**Table 3-9 | Calculating the nitrogen deficit of a site—an example**

| Parameter                                    | Source       |   |
|--|--------------|---|
| A Total soil nitrogen (N)                    | 0.025%       | From soil test of post construction soils (if rates are expressed as gr/l, ppm, mg/kg, ug/g, divide by 10,000 to convert to percentage) |
| B Thickness of soil layer                    | 0.5 feet     | The thickness of soil represented in (A).   |
| C Soil bulk density                          | 1.4 gr/cc    | Unless known, use 1.5 for compacted subsoils, 1.3 for undisturbed soils, 0.9 for light soils such as pumice                             |
| D Fine soil fraction                         | 70%          | 100% minus the rock fragment content (from estimates made from sieved soil prior to sending to lab)                                     |
| E N in soil layer<br>$A * B * C * D * 270 =$ | 331 lbs/ac   | Calculated amount of total nitrogen in soil layer. To convert to kg/ha: $E * 1.12$  |
| F Minimum or threshold N levels              | 1,100 lbs/ac | Determined from reference sites or literature   |
| G N deficit: $F - E =$                       | 769 bs/ac    | Minimum amount of N to apply to bring up to threshold   |

**Carbon Analysis**—Carbon is determined directly using the combustion method (Leco instrument) or indirectly with the Walkley-Black and/or loss-on-ignition methods. Depending on the testing methods, carbon will either be reported as percent of organic matter or percent of carbon. To convert percent of organic matter to percent of carbon, multiply the value by 0.5 to 0.58 (Tisdale and Nelson 1975).

When soils laboratories receive soils samples, they sieve out any materials greater than 2 mm. For this reason, it is important to sieve rock fragments, but not larger organic matter, from the soil samples prior to sending them to the lab. Request that the lab not sieve the larger organic matter from the sample so that the results report out in total carbon and nitrogen.

**C:N Ratio**—The C:N ratio is calculated by dividing the percent of carbon by percent of nitrogen from the laboratory results obtained for nitrogen and carbon tests.

## Mitigating for Low Soil Nitrogen

**Develop a Strategy**—It is important to develop a strategy for increasing nitrogen over time, especially on sites that are deficient in nitrogen. The strategy takes into account the accumulation of nitrogen by all available sources—topsoil, mulch, compost, fertilizers, and nitro-gen-fixing plants. Figure 3-43 shows an example of a strategy for increasing total soil nitrogen to a threshold level.

**Topsoil**—Salvaging and reapplying topsoil is an excellent way to increase total soil nitrogen on drastically disturbed sites. The depth to apply topsoil should be similar to the soil depth found in undisturbed reference sites or pre-construction soils. If topsoil material is limited, then using the calculations shown in Table 3-9 can help determine the minimum depths to apply topsoil. [Section 5.2.4](#) discusses methods to salvage and apply topsoil. To determine if there is a tie-up or surplus of nitrogen in the salvaged topsoil, soil tests can be conducted for C:N ratios. Topsoils with C:N ratios greater than 25:1 could benefit from the addition of nitrogen, while ratios less than 8:1 will have the necessary nitrogen for plant growth.

**Composts**—Applied on the soil surface and incorporated, composts can supply sufficient soil nitrogen for long-term site needs. Application rates for composts can be calculated using the methods shown in Table 3-9. Testing and application methods for compost are discussed in [Section 5.2.3](#) and [Section 5.2.5](#).

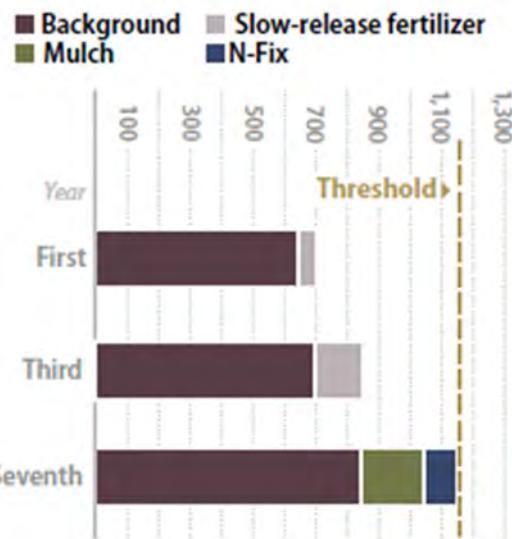
**Nitrogen-Fixing Plants**—Significant quantities of nitrogen can be supplied by nitrogen-fixing plants ([Section 5.2.7](#)). Establishing nitrogen-fixing plants is a means of meeting short-term goals by reducing the need to apply fertilizers and long-term goals by increasing the total nitrogen on the site.

**Fertilizers**—Applying nitrogen-based fertilizers to drastically disturbed soils is another means of increasing nitrogen levels, but an understanding of fertilizers (composition and release), how the soils will capture and store nutrients, and how plants will respond to increased levels of available nitrogen is necessary. As the calculations in Table 3-9 demonstrate, nitrogen-based fertilizers cannot deliver enough nitrogen in one application for long-term site recovery of drastically disturbed sites. However, applied judiciously within an overall nitrogen strategy using topsoil, composts, and nitrogen-fixing plants, nitrogen-based fertilizers can be an effective tool in site recovery.

Not all sites or conditions require fertilizers. It might not be necessary to fertilize soils that have high total N levels and low C:N ratios. In fact, applying fast-release fertilizers may be a disadvantage on some sites by favoring weedy annuals over perennial species. A discussion on selecting fertilizers, calculating application rates, and determining methods of application is provided in [Section 5.2.1](#).

**Biosolids**—Biosolids are rich in slow and fast releasing nitrogen and, if sources are available nearby, are a good means of raising soil nitrogen.

## TOTAL N (LBS/AC)



**Figure 3-43 / Raising nitrogen levels**

Raising nitrogen levels on nitrogen-deficient sites to threshold levels requires developing a long-term strategy. In this example, the site began with a background N of 650 lb/ac. After application of a slow-release fertilizer at 1,000 lb/ac during the first year, the site accumulated 50 lb N (assumes N was captured by plants or soil microorganisms and not leached from the soil). In the third year, an additional 3,000 lb slow-release fertilizer was applied, which increased total N to 850 lb/ac. By the seventh year, woody mulch that was applied during sowing had mostly decomposed, releasing approximately 200 lb N. Nitrogen-fixing plants were well established by then and had contributed approximately 100 lb N.

## Nutrients

This section broadly discusses the remaining mineral nutrients essential for plant growth (Figure 3-44). There are many references devoted to the role of nutrients in plant nutrition and the designer is directed to these sources for a more detailed discussion of each nutrient (Tisdale and Nelson 1975; Thorup 1984; Munshower 1994; Havlin and others 1999; Claassen 2006) models. For example, from an agricultural perspective, a serpentine soil has an imbalance of calcium and magnesium. Unless fertilizers containing a “correct” ratio of calcium and magnesium are applied to adjust this imbalance, the soils will be unsuitable for crop species. In wildland restoration, the approach is guided by the nutrient needs of the species endemic to the site, not to a generic agricultural crop. Because serpentine plant species have evolved on soils with these nutrient ratios, their nutrient requirements are vastly different than those of agricultural crops, or even native vegetation growing on adjacent, non-serpentine soils. In this example, the calcium-to-magnesium ratio would not be an imbalance for native serpentine plant establishment, but perhaps as a “requirement” for certain endemic species to recolonize the site. This means the designer will need to compare post-construction mineral nutrient status to that of undisturbed or recovered reference sites to determine if there are deficiencies. Amendments can then be applied to bring nutrients and other soil factors to pre-disturbance levels or to levels that meet project revegetation objectives.

**How to Assess Nutrients**—The objective of nutrient analysis is to compare nutrient levels of post-construction, disturbed soils with those of pre-disturbance, or reference site, soils. Where there are large discrepancies, a strategy can be developed to bring low post-construction levels up to minimum nutrient levels. Because this is a comparative analysis, it is essential that the sampling, collection, and testing methods are identical.

Nutrient tests are often performed on salvaged topsoil, reference sites, post-construction slopes, and areas where there have been failures in revegetation. A guide to sampling soils for nutrient analysis is presented in Inset 3-2. Nutrient testing can be used to evaluate total soil.

Success in wildland restoration is determined by its species richness, not biomass production or whether it is a self-sustaining and resilient system, not a system that requires constant energy inputs. By these standards, applying the basic agricultural model to wildland revegetation is limited.

|                                 |                      |  |
|---------------------------------|----------------------|--|
|                                 | <b>N Nitrogen</b>    | Major fertilizer element most commonly limiting in disturbed soils. Easily leached from soils. Legumes and other plants can fix atmospheric nitrogen in natural plant communities  |
| <b>Macronutrients</b>           | <b>P Phosphorus</b>  | Major fertilizer element frequently unavailable in disturbed soils. Mycorrhizal fungi can improve uptake in natural plant communities  |
|                                 | <b>K Potassium</b>   | Major fertilizer component that can be leached from soils  |
|                                 | <b>Ca Calcium</b>    | Besides being plant nutrients, calcium and magnesium can be applied as dolomitic limestone to raise soil pH  |
| <b>Secondary Macronutrients</b> | <b>MG Magnesium</b>  |  |
|                                 | <b>S Sulfur</b>      | Besides being a plant nutrient, sulfur can be applied to lower soil pH   |
|                                 | <b>B Boron</b>       |  |
|                                 | <b>Mn Manganese</b>  |  |
| <b>Micronutrients</b>           | <b>Fe Iron</b>       | More important for subsequent plant growth than for establishment. Micronutrients are frequently unavailable in disturbed soils, but can be amended with organic fertilizers or specially formulated micronutrient fertilizers |
|                                 | <b>Zn Zinc</b>       |  |
|                                 | <b>Mo Molybdenum</b> |  |
|                                 | <b>Cu Copper</b>     |  |
|                                 | <b>Cl Chloride</b>   |  |

**Figure 3-44 | The 13 essential mineral nutrients**

Success in wildland restoration is determined by its species richness, not biomass production or whether it is a self-sustaining and resilient system, not a system that requires constant energy inputs. By these standards, applying the basic agricultural model to wildland revegetation is limited.

With soil laboratory results from reference sites and post-construction sites, determine which nutrients, if any, are deficient using the process outlined in [Section 5.2.1](#) (see [Develop Nutrient Thresholds and Determine Deficits](#)). If a nutrient is found deficient, fertilizers, composts, topsoil, or other organic amendments can be applied to the soil to bring the nutrient above threshold levels. A process for determining fertilizer type, application rates, and application methods is presented in [Section 5.2.1](#).

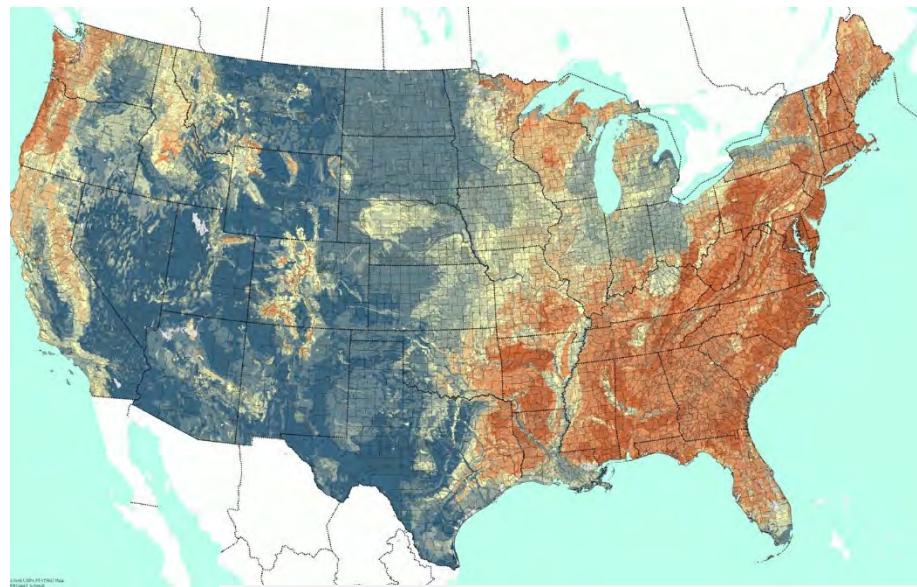
## Mitigating for Low Nutrients

**Topsoil**—Salvaging and reapplying topsoil are important for restoring nutrients to pre-construction levels, especially on sensitive soils (e.g., serpentine and granitic soils). The depth to apply topsoil should be at levels found in undisturbed reference sites or pre-construction soils, or can be calculated by methods described in Figure 5-27 in [Section 5.2.4](#).

**Compost**—Incorporating composts is a good method for increasing nutrients to pre-disturbance levels. Determining which type of compost to select and how much to apply is discussed in [Section 5.2.5](#).

**Fertilizers**—As discussed in [Section 3.8.4](#), fertilizers should be used within an overall nutrient strategy. See [Section 5.2.1](#) for a discussion on application methods, fertilizer types, timing, and other important aspects of fertilization.

**Biosolids**—Biosolids are rich in nutrients. If sources are available and transportation economical, this is a good way to add nutrients to disturbed sites.



**Figure 3-45 | Soil pH levels across the US**

The soils of the United States have a range of soil pH values, from less than to greater than 8.0. Basic (high pH) soils shown in blue are widespread in areas of the United States that receive low rainfall. Acidic soils (low pH) shown in red occur in areas of very old soils common in the eastern United States or in areas of high rainfall common to the Pacific Northwest.

Source: [Bonap.org](#)

## pH

pH (potential of hydrogen) is the measurement of soil acidity or alkalinity based on a logarithmic scale of 0 to 14. Soils with pH values below 7 are acidic, and those above 7 are basic. Basic soils have high amounts of bases (positively charged ions), such as calcium, magnesium, potassium, sodium, and phosphates. Basic soils have developed under arid and semi-arid climates and are found throughout the Basin and Range, Colorado Plateau, and portions of the Great Plains. Acidic soils have formed in wetter climates, where the continued movement of water through the soil profile leaches bases from the soil. Acidic soils are common in the eastern United States, the coast range and mountains of the Pacific Northwest, and the Gulf states (Figure 3-45). Topsoils are typically more neutral when compared to underlying subsoil, whether the soils are acidic or basic. In some cases, the topsoil buffers the plant root systems from the underlying, inhospitable subsoil conditions. When topsoils are removed during construction, subsoils become the growing environment and, unless mitigating measures are taken, plant establishment and productivity of the site is greatly reduced.

Soil acidity and alkalinity affects mineral nutrient availability, mineral toxicity (Palmer 1990), and nitro- gen fixation (Thorup 1984). In acid soils, the ability of plants to utilize many nutrients decreases, especially for calcium and magnesium. As soil pH becomes

### 3—PLANNING

more acid (less than 4.5), aluminum becomes more soluble and more toxic to plant growth. Low pH soils also hinder the establishment of nitrogen-fixing plants, such as legumes (Bloomfield et al 1982). Significant loss of rhizobia viability has been documented at pH levels less than 6 (Brown et al 1982).

**Table 3-10 | Soil testing methods**

Common soil testing methods for the western United States (Horneck et al 1989; Munshower 1994; Teidemann and Lopez 2004).

Note: Composts use a different set of tests due to high organic matter (Section 5.2.4 and Section 5.2.5).

| Tests                           | Type               | Test method                                       | Notes   |
|---------------------------------|--------------------|---|---|
| Boron                           | Available          | Hot-Water   |   |
| Boron                           | Available          | Aqueous extract of a soil paste                   |   |
| Calcium, Magnesium              | Available          | Ammonium Acetate                                  |   |
| Calcium, Magnesium              | Available          | Aqueous extract of a soil paste                   | In semi-arid to arid soils                                  |
| Molybdenum                      | Available          | Ammonium oxalate-oxi acid extraction              |   |
| Nitrate                         | Available          | Aqueous extract of a soil paste (Saturated paste) | Accepted extrant for western soils                          |
| Nitrate                         | Available          | CaO extract & Cd reduction                        |   |
| Nitrogen (ammonium and nitrate) | Available          | KCL Extraction                                    |   |
| Nitrogen (mineralizable)        | Slowly - Available | Anaerobic Incubation                              |   |
| Nitrogen (total)                | Total              | Kjeldahl N  |   |
| Nitrogen (total)                | Total              | Combustion (Leco Instrument)                      |   |
| Organic matter                  | Total              | Loss - Ignition                                   | Best used for soil high in organic matter                   |
| Organic matter                  | Total              | Walkley-Black Method                              |   |
| Organic matter                  | Total              | Combustion (Leco Instrument)                      | Reports out in Total C                                      |
| pH                              |                    | Aqueous extract of a soil paste (saturated paste) |   |
| Phosphorus                      | Available          | Olsen Sodium Bicarbonate                          | For arid and semi-arid soils                                |
| Phosphorus                      | Available          | Dilute Acid-Flouride (Bray-P1)                    | For mesic sites   |
| Phosphorus                      | Available          | AB-DIPA   | Reports out at half the rates of Olsen method               |
| Potassium                       | Available          | Sodium Acetate                                    |   |
| Potassium                       | Available          | Olsen Sodium Bicarbonate                          | For arid and semi-arid soils                                |
| Potassium                       | Available          | Ammonium Acetate                                  |   |
| Sodium                          | Available          | Ammonium Acetate Displacement                     |   |
| Sodium                          | Available          | CTPA  |   |
| Sulfate Sulfur                  | Available          | Aqueous extract of a soil paste (saturated paste) |   |
| Sulfate Sulfur                  | Available          | CaHPO & ICP                                       |   |
| Zinc, Copper, Manganese, Iron   | Available          | CTPA  | Iron is not performed in Oregon because not found deficient |

Soil with pH values of 8.0 or greater indicate the presence of calcium carbonate (Thomas 1967). Calcium and magnesium are at such high levels that they interfere with the uptake of other nutrients, notably phosphorus, iron, boron, copper, and zinc (Campbell et al 1980). High pH soils typically have high salt levels, which can also restrict the growth of many plants. For example, as soil pH approaches 9.0, sodium concentrations become toxic to plants (Tisdale and Nelson 1975).

**How to Assess pH**—The pH test is a standard analytical measurement that is typically run on soil samples sent for nutrient analysis. The pH test is also conducted on soil organic matter amendments considered for mulch or incorporation into the soil.

### 3—PLANNING

The pH test is accurate, with values differing between laboratories by 0.1 to 0.2 points (Thomas 1967). pH can also be tested by the practitioner on site or back at the office using reasonably priced equipment. Most portable pH meters can measure soluble salts and this dual capacity is important in areas with high salts (Section 3.8.4, see pH). It is important when selecting a pH meter that it has a tip that can be submerged easily in a soil slurry.

The most accurate method of assessing pH is through lab analysis. However, quick, reliable estimates can be made with a hand-held pH/electrical conductivity meter using the Saturated Media Extract (SME) method for preparing samples (Figure 3-46). With this method, a small amount of soil (50 cc) is placed in a jar. Just enough distilled water is stirred into the soil to make the surface “glisten” but not readily flow. After the mixture rests for approximately 15 minutes the pH probe is inserted into the soil so that the sensors are completely covered and the pH reading is made.

## Mitigating for Low pH Soils

**Apply Liming Materials**—Raising the pH through the application of liming materials is a common agricultural practice that can be applied to revegetating road sites (Section 5.2.6).

**Apply Appropriate Fertilizers**—Some commercial fertilizers, especially ammonium-based fertilizers such as ammonium nitrate, ammonium sulfate, and ammonium phosphate, will reduce pH (Havlin et al 1999) and should be limited on acidic soils. Fertilizers that have calcium, magnesium, or potassium in the formula are more appropriate for low pH soils. Examples of these fertilizers are calcium nitrate, potassium nitrate, and magnesium sulfate.

**Apply Lime with Organic Matter**—Incorporation of organic matter will lower pH. On acid soils, application of lime with organic matter will raise the pH of the soil (Section 5.2.6).

**Apply Topsoil**—Where topsoils have been removed leaving very basic or very acidic subsoils, reapplying topsoil or manufactured topsoil can moderate pH levels.

## Mitigating for High pH Soils

**Apply Organic Matter**—Incorporated composts or other types of organic matter can lower soil pH as the organic matter decomposes (Havlin et al 1999). For arid sites, however, the pH and conductivity of the organic matter needs to be tested prior to purchase to avoid the possibility of introducing organic matter high in salts.

**Add Nutrients**—To compensate for the tie-up of certain nutrients, the addition of nutrients through fertilization may be considered; however, some of the benefits of using fertilizers on arid soil may be offset by the possibility of creating fertilizer salt problems.

**Apply Sulfur**—Agricultural soils can be treated with sulfur to lower pH, it is necessary to apply high quantities of sulfur and irrigation to lower the pH just slightly (Havlin et al 1999). The use of sulfur in roadside revegetation therefore is not a widely practiced method.



**Figure 3-46 / pH meter**

Most portable pH meters will also measure salts (electrical conductivity). Many probes can be directly inserted into the saturated media.

*Photo credit: David Steinfeld*

## Irrigation

Applying irrigation water is another method of reducing soil pH by leaching out bases. However, the amount of water needed to lower pH levels can be very high; in most cases, using irrigation is not a viable mitigating measure on roadsides. It is also difficult to find irrigation water in arid environments that is low in bases and salts. Applying irrigation water that is high in bases will raise pH and salt levels in the soil, compounding the problem.

## Salts

Soil salinity is the measure of the total amount of soluble salts in a soil. The term soluble salts refers to the inorganic soil constituents, or ions, that are dissolved in the soil water. The principal soluble salts in most soils contain the cations—sodium, calcium, and magnesium, and the anions chloride, sulfate, and bicarbonate (Landis and Steinfeld 1990).

Almost all plants are susceptible to salt injury under certain conditions, with germinants and young seedlings being particularly susceptible to high salt levels (Figure 3-47). Soluble salts can injure plants in several ways:

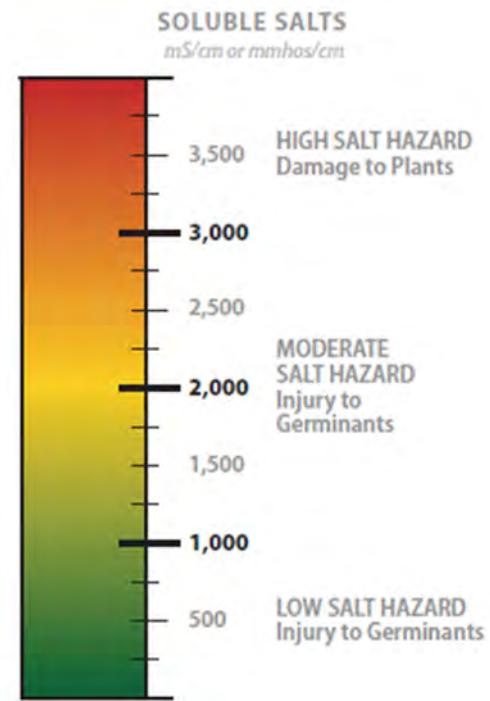
- **Reduced soil moisture**—Salts can lower the free energy of water molecules, causing an osmotic effect and thereby reducing the moisture availability to plants.
- **Reduced soil permeability**—High salt concentrations (specifically sodium salts) can change the soil structure by reducing the attraction of soil particles, causing them to disperse. Pore space is lost and air and water movement within the soil profile are restricted.
- **Direct toxicity**—High levels of certain ions, including sodium, chloride, and boron, can injure plant tissue directly.
- **Altering nutrient availability**—Certain nutrients as salts can change the availability and utilization of other plant nutrients (Landis 1981; Landis and Steinfeld 1990).

High salt levels are common in arid regions of the United States where there is inadequate precipitation to leach salts out of the plant root zone (Figure 3-48). As a result, salts move out of the topsoil and accumulate in the subsoil. At high enough concentrations, a layer of calcium carbonates form, creating an impermeable horizon called caliche. This layer restricts root growth and soil drainage. When topsoil is removed, the resulting surface soils may be very high in salts or where a caliche horizon is present, may expose a hardened calcium carbonate surface.

High salt concentrations can also be created by poor soil drainage resulting from compaction; when excessive amounts of fertilizer, manure, or compost are applied; or when de-icing chemicals applied to roads run off and enter the soil (Parent and Koenig 2003).

Deicing salts can pose a problem to plant establishment depending on the annual quantity of salt applied, distance from the road, type of salt applied, annual precipitation, and soil type (Section 3.11.9, see Deicing for Winter Safety). The sensitivity of a plant species is also important and it may be necessary to revegetate with plant species that are less sensitive to salts. These can be selected using the ERA tool.

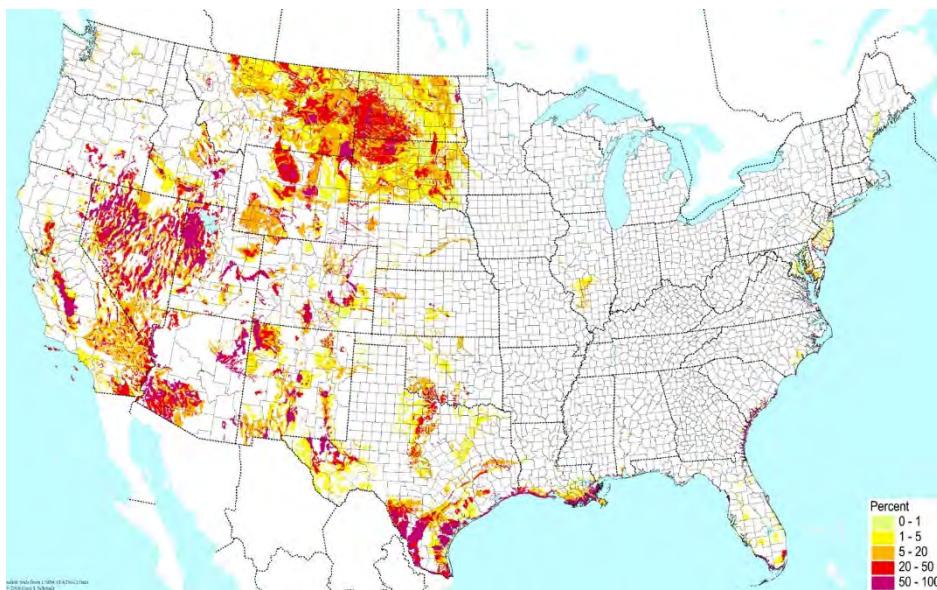
**How to Assess Salts**—There are two methods of measuring salts: Electrical Conductivity (EC) and Total Dissolved Salts (TDS).



**Figure 3-47 | Soluble salt effects on plants**

Soluble salts will injure germinants and, at higher concentrations, damage established plants. Values are based on the saturated media extract method of conductivity measurement reported in mS/cm (microSiemens per centimeter).

Modified from Fisher and Argo 2005



**Figure 3-47 | Soluble salt effects on plants**

Soluble salts will injure germinants and, at higher concentrations, damage established plants. Values are based on the saturated media extract method of conductivity measurement reported in mS/cm (microSiemens per centimeter).

Modified from Fisher and Argo 2005

EC is a relatively easy test to run and is the most commonly used test in nurseries, forestry, and agriculture. Most pH meters are equipped to measure EC. Electrical conductivity measures how strongly electrical current flows between two metal plates. The more dissolved salts there are in solution, the greater the current and higher the electrical conductivity. EC is reported as the conductance over the distance between plates. The standard unit of measure is microSiemens per centimeter ( $\mu\text{S}/\text{cm}$ ), though there are many ways that it can be expressed which can be confusing. To convert these units to  $\mu\text{S}/\text{cm}$ :

- $\mu\text{S}/\text{cm} = 0.001 \text{ dS}/\text{m}$
- $\mu\text{S}/\text{cm} = 0.001 \text{ mS}/\text{cm}$
- $\mu\text{S}/\text{cm} = 0.1 \text{ mS}/\text{m}$
- $\mu\text{S}/\text{cm} = 0.001 \text{ mmho}/\text{cm}$
- $\mu\text{S}/\text{cm} = 1 \text{ } \mu\text{hos}/\text{cm}$

The most accurate method of assessing salinity is through lab analysis, however, quick estimates can be made with a hand-held pH/electrical conductivity meter using the Saturated Media Extract (SME) method for preparing samples as described in [Section 3.8.4 \(see pH\)](#). Landis and Dumroese (2006) provide a more detailed discussion on EC and measuring methods.

The second method of measuring salinity is the TDS. In this method, a known sample (water or soil solution) is evaporated and the remaining salt is weighed. This test is more difficult to run but it is often used in reporting salinity levels in road deicing studies. Test values are reported in milligrams per liter of water (mg/l) which is equivalent to parts per million (ppm). To convert TDS values to EC:

$$EC (\mu\text{S}/\text{cm}) = TDS (\text{mg/L}) / 0.6$$

### Mitigating for Salts

Because high levels of soluble salts are often caused by poor soil management, the key to mitigating high salinity is to avoid creating the conditions that could cause those levels. In soils where internal drainage is poor, prevention may be the only feasible approach for reducing salt problems. Reducing the quantity of road deicing salts can also lower the amount of salts that enter the soil.

**Avoid Mulch or Soil Amendments with High Salinity**—Testing all materials to be applied to the site will aid in the prevention of increased salt levels in the soil. Amendment materials with electrical conductivity readings more than  $1 \text{ } \mu\text{S}/\text{cm}$  should

be avoided.

**Reduce Commercial Fertilizers**—Some commercial fertilizers, such as control-release fertilizers (CRF), can significantly increase the soluble salts found in the soil. This can be a major problem when using CRF in arid conditions. The fertilizer will begin to release following wet, warm spring conditions, but will not be leached through the soil without significant rainfall through the summer. Salts can build up to damaging levels, both on the surface and in the plant root zone.

**Apply Gypsum with Irrigation**—Incorporation of gypsum (calcium sulfate) followed by leaching can be effective in situations where sodium is the cause of high soluble salts (e.g., de-icing materials have been applied to roads). The calcium in gypsum will displace sodium, and the sodium will then leach out of the soil profile with irrigation or rainfall (UMES 2004).

**Irrigation**—Application of irrigation water is often used to leach salts from the soil. The amount of water depends on the soil type. In arid soils, application of 6 inches of water can reduce salinity levels by 50 percent; 12 inches can reduce salinity levels by 80 percent; and 24 inches can reduce salinity levels by 90 percent (UMES 2004). However, for most sites roadside project, this is not practical due to high costs.

**Select salt resistant plant species**—If salts are present or are expected to be present through deicing practices, selecting species from the ERA that are less sensitive to the effects of salts may improve revegetation.

### 3.8.5 SURFACE STABILITY

Surface stability is the tendency of the soil to remain in place under the erosive forces of rain, wind, and gravity. Good surface stability is essential for establishing plants, reducing erosion, and maintaining high water and air quality. When seeds are applied to unstable surfaces, they often move off the site through water or wind erosion leaving the site barren of vegetation. Soil is also removed in this process, which reduces the productivity of the site. Excessive erosion also affects the survival of planted seedlings by removing soil and exposing roots.

Site factors that influence surface stability and soil erosion are as follows:

- Rainfall
- Wind
- Freeze-thaw
- Soil cover
- Surface strength
- Infiltration rates
- Slope gradients
- Surface roughness
- Slope lengths

All surface erosional processes start first with the detachment of soil through the forces of rainfall, wind, or frost heave. These forces loosen seeds and soil, making them more susceptible to movement off the site. Surface runoff, during rainstorm events, is the primary factor in moving seeds and soil into stream channels, resulting in lost seeds and water quality problems. This occurs when infiltration rates (the rate water moves through the soil surface) are lower than rainfall rates. If slope gradients are steep, slope lengths are long, or surface roughness is low, surface water picks up energy and transports greater amounts of soil and seeds downslope. As this energy increases,

water becomes concentrated with enough force to cut through the surface of the soil, creating rills and gullies. The degree to which soils detach is directly related to the percentage of soil cover protecting the soil (more cover, less erosion) and to the soil strength, or the capacity for individual soil particles to hold together under erosional forces. The result of soil erosion can often be detected on road cuts by noting how much sediment is in the ditch line. If the ditch is full of recently deposited sediment (recently deposited sediment usually lacks vegetation), there is a good chance that sediment came from the cut slope. An inspection of the surface of the cut slope will indicate if the sediment originated there. Gravels, cobbles, and even small plants will show the results of soil movement (see Figure 3-52, Figure 3-54, and Figure 3-55).

## Rainfall

Each project site has a unique rainfall pattern that will affect the stability of the soil surface. Periods of high rainfall intensities can move seeds and soil particles off-site through erosional processes which begins with the raindrop. Raindrops have been likened to small bombs. In heavy rainstorms, they fall with such speed (up to 20 miles per hour) that, when they hit the soil surface, they create an impact that can blast the soil or seeds several feet away and leave behind small craters. After such events, the soils surface is compacted and sealed with fine soil particles that can significantly reduce surface infiltration rates.

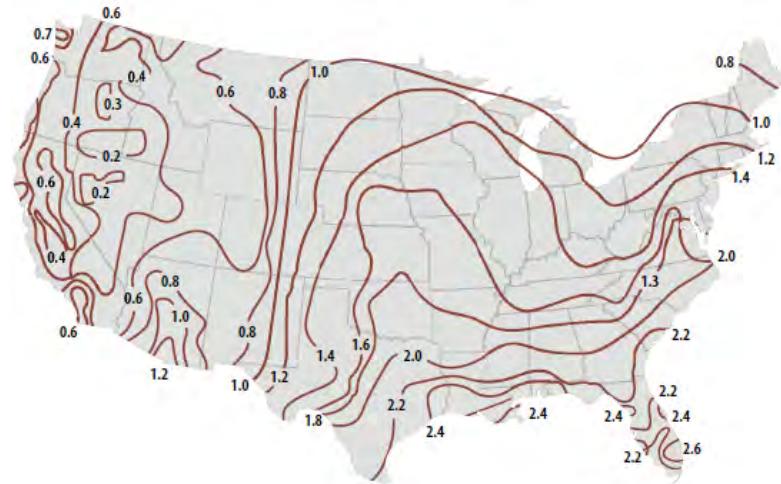
The intensity of a rainfall event determines how much soil is detached. A high intensity rainfall will detach more soil particles than a low intensity rainfall. But detachment is only one aspect of erosion; it takes surface runoff to move soil and seeds downslope. If an intense rainstorm lasts only a short period, there may be insufficient water to exceed infiltration rates and water will absorb into the soil. If the duration is long, some water will not enter the soil, and run over the surface as overland flow, carrying soil and seeds downslope. The most critical weather events are those that bring high intensity rainstorms of long duration. High intensity rainfall is common in the central and eastern United States, whereas in the western U.S., high intensity storm events are less frequent and typically occur during summer thunderstorms or major winter storm systems (Figure 3-49).

Precipitation in the form of snow is not typically a problem for surface erosion because snow cover protects the soil surface from rainfall splash and water from snowmelt usually occurs at such slow rates that even soils with low infiltration can absorb it, reducing the likelihood of runoff.

**How to Assess Rainfall**—It is unlikely that climate reports or weather records will give the duration and intensities of rainfall events for site level planning. Digital rainfall gauges are available that record the amount of rainfall and the time it occurred. This information is used to determine duration and intensity. While the cost of this equipment is high, it is becoming more affordable. Many types of digital rain gauges are available, ranging in price and quality. It is important to select a digital rain gauge that is rugged, self-maintaining, and can record for long periods of time. Some systems upload weather data to the internet where it can be accessed remotely through smart phones and computers for data summary and analysis.

## Mitigating for High Rainfall

**Minimize Disturbance**—In areas of high rainfall or sites where water quality values are high (near streams or rivers), the best engineering design is to keep the footprint of the



**Figure 3-49 | Rainfall intensities across the US**

Rainfall intensities increase moving from western to eastern United States. The differences in intensities can be dramatic, ranging from 0.2 inches/hour in Nevada to 2.6 inches/hour in Florida for a 2-year, 1-hour storm event.

*Source: FHWA*

construction project disturbance to a minimum. Not only does this reduce the risk of delivering sediment to the aquatic system, it can reduce project costs by reducing the amount of area needing revegetation.

**Integrate Erosion Practices**—On disturbed sites, especially those near streams, the integration of erosion practices with plant establishment techniques offers the best approach to stabilizing the soil surface. These include practices such as increasing soil cover, shortening slopes, reducing slope gradients, leaving roughened surfaces, reducing compaction, increasing infiltration rates, and quickly establishing native vegetative cover.

**Use Appropriate Mulching Practices**—Applying a surface mulch is one of the best practices for controlling surface erosion because it protects the soil surface from rainfall impact and reduces overland flow. The types of mulches are described in [Section 5.2.3](#).

## Wind

Wind erosion can be a major limiting factor in establishing native plants (Figure 3-50), especially in the Great Plains and portions of the Eastern Temperate Forests ecoregions (Figure 3-51). Wind erosion begins with a process called saltation in which a soil particle is lifted by the wind and bounced along the ground surface, dislodging other soil particles in its path. The resulting dislodged particles either become airborne or continue to roll or hop along the surface of the soil, dislodging more particles. Saltation occurs with very fine to medium sands, ranging from 0.003 to .04 inches (.07 to 1 mm). Particle sizes smaller than that (silts and clays) become airborne and can be carried long distances (Fifield 2004).

Wind erosion affects revegetation in several ways. Newly sown seeds and seed cover can be removed in high winds, resulting in poor germination and plant establishment. Plants that do establish, can be damaged through saltation, where dislodged soil particles continually abrade the plant stems. In extreme conditions, topsoil is removed, reducing the soil productivity and plant growth. This is more severe where topsoils are thin. Where plants are established, loss of topsoil can also expose roots, causing reduced growth and in some cases, mortality.

**How to Assess Wind**—Permanent wind speed equipment is available but is most likely beyond the reach of most project budgets. Site visits during different times of the year provide some indication if wind is a problem. Site characteristics, such as position on slope (ridgelines are more prone to high winds than a valley floor) or proximity to forested environments (forests often reduce wind speeds). Observing soil surfaces for signs of erosion can be helpful and include appearance of bare soils, exposed roots, and fine soils deposited behind stable structures. Local residents may also provide some information on local weather events.

Assessing risk of wind erosion can be determined by:

- **Wind speed**—The rate of soil movement is proportional to wind velocity. Wind speeds are considered erosive when they exceed 13 miles per hour measured 1

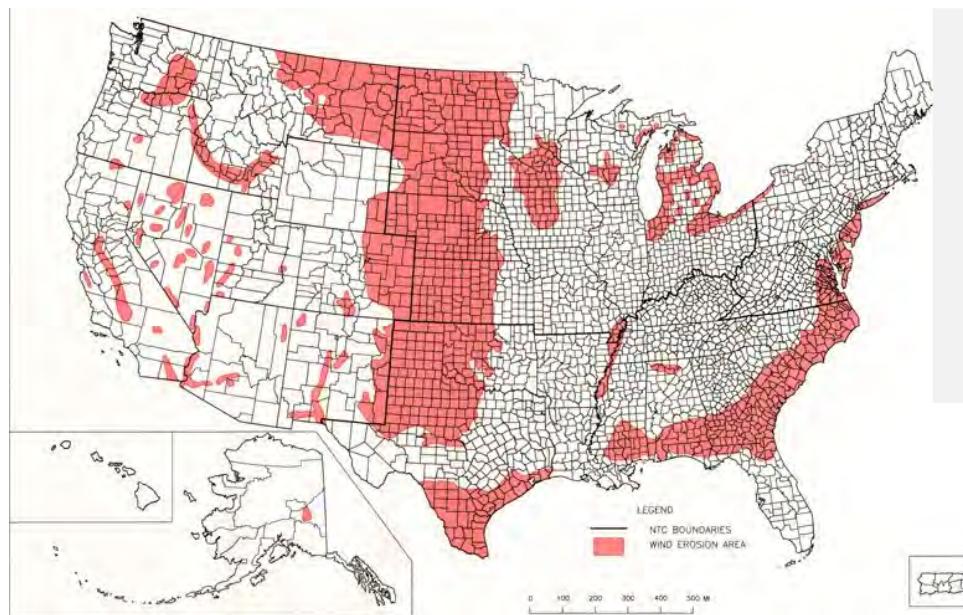


**Figure 3-50 | Wind erosion removes topsoil and exposes roots**

Wind erosion not only blows seeds, soil, and mulches off the soil surface, it will also expose roots of established plants, as shown in this photograph. Non-cohesive soils, like sands and silts, are most prone to wind erosion.

*Photo credit: David Steinfield*

foot above the soil surface for loose sands (Lyles and Krauss 1971).



**Figure 3-51 / Areas in United States that have high winds**

High winds are the primary cause of soil erosion in many areas of the United States. This map shows areas where wind erosion could strongly influence the establishment of native plants (areas in red) (FHWA 1992). The effects of wind on revegetation are very site specific, the potential of which can be determined during field surveys.

- **Soil Texture**—Silt contents have the most potential to be dislodged and become airborne. Clays can be susceptible when soils have been continually disturbed and clods have been destroyed. Sandy texture soils are less susceptible to becoming airborne but will move through surface creep and saltation.
- **Rock fraction**—The amount of rock fragments in a soil affects the severity of wind erosion; the more rock, the greater the protection of the soil surface. In addition, surface rocks may also collect windblown silts and sands over time, increasing soil depth.
- **Organic matter**—Soils that are higher in soil organic matter, especially organic cements that produce stable aggregates, have greater stability.
- **Water content**—water holds soil grains together through cohesion so when surface soils have a high moisture content, they have greater strength.
- **Surface roughness**—a roughened soil surface composed of a microtopography of ridges and valleys will trap and suspend soil particles, especially if the ridges are predominantly perpendicular to the direction of the wind.
- **Vegetative cover**—The height and density of vegetation affects the air flow on the surface of the soil; the greater the height and density the lower the wind velocity. Windbreaks, such as shelterbelts, can significantly reduce wind velocities (see Inset 3-3).
- **Soil cover**—Soils that are covered with a stable material are more resistant to wind erosion.

## Mitigating for High Wind

**Maintain vegetative cover**—Keeping the construction footprint to a minimum is one of the best practices for controlling wind erosion. Minimizing the width of the disturbances perpendicular to the direction of the prevailing winds will also reduce the effects of strong winds.

**Install shelterbelts**—A shelterbelt is a line of trees that reduce the wind velocity and reduce the potential for wind erosion (Inset 3-3). The ERA tool can be consulted for appropriate trees and shrubs suitable for shelterbelts, depending on the ecoregion.

**Create microtopography of ridges and furrows**—When the soil surface is left in a roughened condition of ridges and furrows, wind erosion can be reduced significantly, however the effectiveness depends on the stability of the soil (Fryrear and Skidmore 1985). Ridges formed in non-cohesive soils or soils that do not form clods when tilled, will have a short life span.

Tillage, using a land imprinter ([Section 5.2.2, see Roughen Soil Surfaces](#)), may be a good alternative to loosening soils with traditional tillage equipment (e.g. disks, harrows) because imprinting compresses the soil while it forms depressions, thereby creating a roughened surface and increasing soil strength that will increase resistance to wind erosion (Dixon and Simanton 1977).

**Mulch**—Applying a mulch is one of the best practices for controlling surface erosion ([Section 5.2.3](#)), however, mulches be removed with strong winds. Some materials, such as wood strand mulches ([Section 5.2.3, see Wood Strands and Wood Wool](#)), have been tested under high wind conditions and, because of higher weights and interlocking particles, may be more resistant to high winds.

Lighter mulch materials, such as straw and hay, are more susceptible to wind erosion and have to be crimped into the soil to keep them in place. Hydromulch with tackifier or just tackifier applied with hydroseeding equipment can be effective in stabilizing hay mulches in areas with lower wind speeds.

**Place Large Woody Material**—Downed woody material, such as trees and large branches, can be used to block the soil surface from wind and rainfall.

### Inset 3-3 / Wind breaks—shelterbelts and living snow fences

**Shelterbelts.** A shelterbelt is a vegetative barrier that reduces wind speeds, resulting in lower soil erosion. Shelterbelts also provide excellent pollinator and wildlife habitat. Shelterbelts work by creating wind resistance in the face of the prevailing wind. The reduction in wind velocity can be significant depending on the height of the shelterbelt. Depending on the wind speed, the effect on the leeward side of the shelterbelt (the side away from the wind), can be up to 10 times the height of the shelterbelt and on the windward side (the side toward the wind), as much as 5 times the height (see illustration below).

For greatest effectiveness, shelterbelts are designed perpendicular to the direction of the prevailing winds. Shelterbelts with multiple rows of vegetation have a greater reduction in wind speeds than single row shelterbelts because there is more vegetation to block wind flow. A two-row shelterbelt composed of shrubs and trees not only provides more wind resistance, it provides better wildlife and pollinator habitat. The effectiveness of a windbreak depends on the selection of tree and shrub species and planting density. It is important to understand wind speeds, shelterbelt effectiveness, and appropriate species specific for the project area to develop an appropriate shelterbelt design.

Wind speeds can be reduced on the leeward side of a shelterbelt by 25 to 60 percent within a distance that is 10 times the height of the shelterbelt. It also can reduce wind speeds on the windward side by 25 to 80 percent within a distance 5 times the height on the windward side. In the example shown below, a 10-foot-high shelterbelt reduces the wind velocity significantly 50 feet before the shelterbelt and 100 feet beyond the shelterbelt (modified from Casement and Timmermans 2007).

**Living Snow Fence.** When vegetation is installed for snow management, it is referred to as living snow fences. Densely planted trees and shrubs along highways can reduce the amount of snow drift on roadways and subsequently, lower winter maintenance costs. These living snow fences create a barrier which reduces wind speeds and drops snow in front of and downwind of the break. It is necessary to understand the wind speed, wind direction, and annual snowfall when designing a living snow fence. Some of this information can be obtained from historic climate data from local weather stations ([Section 3.3.1](#)) but in most cases, it can be obtained from local maintenance personnel. Visiting existing snow fences during the winter can help in designing living snow fences. The most effective living snow fence is planted perpendicular to the prevailing wind direction at a minimum of 175 feet from the road centerline (South Dakota Department of Agriculture (2006). Other States and municipalities may have guidance on the design of natural or man-made snow fencing for their area. Shrub and tree species that grow at least 6 to 8 feet tall are good for living snow fences. Optimum snow storage capacity is achieved when individual rows have a density of 50 to 60 percent cover as viewed through the winter vegetation. Density is determined by how closely plants are spaced and whether the vegetation is deciduous or evergreen. When vegetation densities are low, more snow moves through the living snow fence while snow fences with high densities run the risk of being damaged by deep snow drifts (Brandle and Nickerson accessed 2017). The most effective living snow fences are those with at least two rows of shrubs or trees.



### Freeze-Thaw

Freeze-thaw is the process of ice formation and ice melting that occurs in a 24-hour cycle within the surface of the soil. At night, temperatures drop at the soil surface and water begins to freeze within the soil pores, creating ice crystals. As ice crystals continue to form, water is drawn from the soil below through capillary action to replace the water that created the ice crystals. During freezing, ice crystals expand in the soil and push soil aggregates apart (Ferrick and Gatto 2004), weakening the internal structure of the soil. When soils thaw the following day, soil strength is greatly reduced (Gatto et al 2004), leaving the soil surface significantly less resistant to erosional forces. Freeze-thaw is considered one of the least understood aspects of soil erosion (Ferrick and Gatto 2004) and yet accounts for significant annual soil losses

### 3—PLANNING

(Froese et al 1999).



The formation of ice crystals will destabilize the seed germination environment.

Freeze-thaw cycles affect germinating seeds by creating ice crystals that physically push the new seedlings above the soil surface, exposing the emerging roots to extremely harsh conditions for seedling establishment, including low humidity, high temperatures, and sunlight. On steeper slopes, soil particles and germinating seeds move downslope after each freeze-thaw cycle, further destabilizing the seed germination environment. Freeze-thaw processes can also affect seedling establishment. Long periods of freeze-thaw cycles can push seedlings out of the ground, exposing roots and, in many cases, killing seedlings (Figure 3-52).

Soils most susceptible to freeze-thaw effects are those with a high silt content or soils that are compacted. Soils are most susceptible when they are cold and wet. Silt-sized particles have pore sizes that are small enough to pull moisture to the layers that are freezing through capillary action, yet large enough to form ice crystals (Ballard 1981). Sandy soils do not draw moisture to the freezing layer because the pores are too big. Clays, on the other hand, have good capillary characteristics, yet do not have large enough pores for ice crystals to form (Ferrick and Gatto 2004). Sands are susceptible to freeze-thaw when they are compacted because the size of the pores is reduced, encouraging capillary rise (Gatto et al 2004). Soil cover, which includes litter, duff, and organic mulches, does not typically have good capillary rise characteristics, and therefore are less frost-susceptible. In addition, soil cover offers good thermal protection, which moderates the degree of freezing and thawing at the soil surface. The effects of freeze-thaw on surface stability increases as slope gradients steepen; the steeper the slopes, the greater the movement of seeds and soil downslope each day. For example, a seed or soil particle on a steep slope rises 2 inches on top of an ice crystal (Figure 3-53) but when the crystal melts, the seed or soil particle drops to a different point farther downslope. After many freeze-thaw cycles, the distance traveled by the seed or soil particle can be significant.

**How to Assess Freeze-Thaw**—Freeze-thaw processes typically occur in the spring and fall, when soil moisture levels are high and soil temperatures are cold. Soil surfaces that have undergone freeze-thaw cycles will have a very loose crust that will collapse when touched or walked upon. Gravels are often perched on pedestals, but give way under light pressure.

Project areas with bare surface soils high in silts or compacted sands should be considered prone to freeze-thaw processes, while soils with deep mulch or litter layers are less susceptible.

**Figure 3-52 | Freeze-thaw effects on planted seedlings**

Continual freeze-thaw conditions can push root systems of planted seedlings out of the ground, reducing growth and potentially killing seedlings.

*Photo credit: David Steinfeld*



**Figure 3-53 | Freeze-thaw ice crystals**

Ice crystals that form under freeze-thaw conditions can lift soil particles over 2 inches above the original surface. Later in the day the crystals will melt and the particles will drop.

*Photo credit: David Steinfeld*

## Mitigating for Freeze-Thaw

**Apply Mulch**—Available research on the mitigation of freeze-thaw effects is slim, but it can be assumed that practices, such as applying an organic mulch, will insulate the soil surface and minimize the effects of freeze-thaw. The deeper the mulch layer, the less propensity for freeze-thaw at the surface. Hydromulch applications at typical rates of 1,000 to 2,000 lb/ac are too thin to moderate surface temperatures or strong enough to resist the destabilizing effects of ice crystal formation on surface strength. Deep application of most mulches however, will bury seeds, resulting in poor seedling emergence. An alternative to is to apply needles or wood strands because these materials can be applied at greater depths and still allow light through for seedling emergence. For planted seedlings, the application of very deep mulches will reduce the effects of freeze-thaw. Gravels, cobbles, and stone can provide a stable soil cover. These materials are often left over after rock has been screen from the soil and designed into the slope can add surface stability and reduce evaporation.



**Figure 3-54 | Soil erosion affects seed germination**

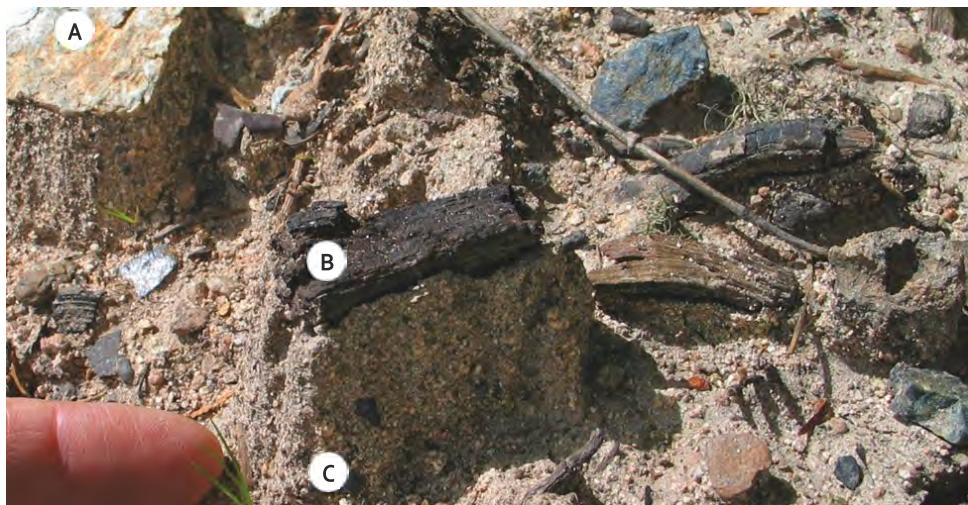
Soil cover protects the surface from rainfall impact. Not only is soil removed during rainstorms, affecting germination and seedling establishment, but seedlings that do emerge can be covered with soil that splashes from rainfall impact. Seedlings will not grow through an encasement of soil.

*Photo credit: Thomas D. Landis*

**Till Compacted Soils**—Sandy soils are very susceptible to freeze-thaw if they are compacted. Loosening soils through tillage is perhaps the best method of mitigating the effects of freeze-thaw on these soil textures (Section 5.2.2).

**Avoid Wet Soils**—Consider avoiding planting or sowing in soils with high water tables and poorly draining soils. The extra moisture in these soils will continue to supply water for ice crystal formation. Planted seedlings can be pushed out of the soil in these environments (Figure 3-54).

**Maintain Some Overstory Canopy**—Trees and shrubs will moderate surrounding temperatures, reducing the potential for freeze-thaw.



**Figure 3-55 | Soils are protected by a soil cover**

Larger materials, such as gravels (A) and wood chips (B), protect the soil and sown seeds by absorbing the energy of rainfall impact. While unprotected soil and seeds are removed through splash and sheet erosion, protected soil remains in pedestals, sometimes several inches above the surface of the soil. Seeds that do remain on or near the surface have a difficult time germinating through the surface crust created by rainfall impact (C). Plants that do establish will have roots exposed by successive rainfall events.

*Photo credit: David Steinfeld*

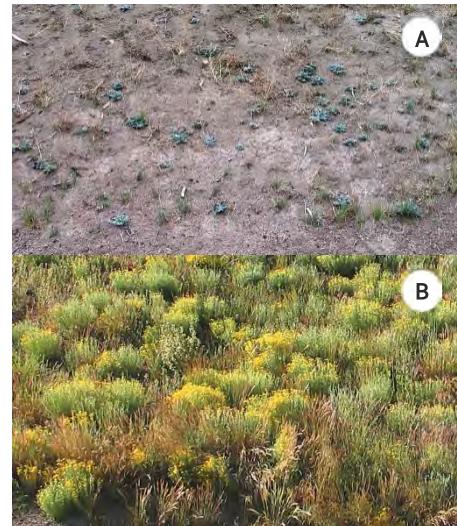
## Soil Cover

Soil cover is the layer directly above the surface of the soil. In an undisturbed environment, soil cover is composed of a combination of duff, litter, live plants, and rock. Soil cover is very important for surface stability because it dissipates energy from rain drop impact, protecting the soil from high intensity rainfall events. Furthermore, soil cover will slow the movement of runoff and capture sediments and seeds, preventing them from moving downslope. [Section 3.8.3](#) discusses how soil cover is important for reducing evaporation; this section will discuss its role in stabilizing the soil surface and reducing erosion.

When the soil surface lacks cover, it is subject to the direct forces of raindrop impact, overland flow, wind, and gravity. These forces not only move soil offsite, affecting water and air quality, but they also displace seeds or remove soil from around newly developing seedlings (Figure 3-54). A lack of soil cover will impact revegetation objectives by reducing the quantity of seeds that will germinate. Seedlings that do emerge will be negatively affected by soil splash and sheet erosion that remove soil from around the seedling roots. The severity of soil erosion and seed movement is directly related to the percentage of bare soil.

After construction, most organic soil cover is removed. What remains is bare soil and coarse fragments (gravel, cobble, and stone). Left unprotected, bare soil will erode during rainstorms, leaving a pavement of coarse fragments (Figure 3-55). If the amount of coarse fragments in the soil is high, then the percentage of the soil surface covered by coarse fragments will also be high. By the third year, erosion rates on unprotected bare soils typically fall to a tenth of the rate of the first-year rate because of the formation of a coarse fragment surface (Megahan 1974; Ketcheson et al 1999). While this process produces less sedimentation to stream systems, the high coarse fragments at the soil surface are a poor environment for seed germination. For this reason, it is important to quickly stabilize the soil surface after soil exposure and why most road contracts call for temporary road stabilization during the construction period.

**How to Assess Soil Cover**—Soil cover can be measured by establishing transects and recording the percentage of rock, vegetation, litter, duff, and bare soil. A monitoring procedure for measuring soil cover is presented in [Section 6.3.1](#).



**Figure 3-56 | Many sites take more than one year to fully revegetate**

Semi-arid, arid, and cold sites often take more than one year to fully revegetate. Photo A shows the vegetative establishment one year after hydroseeding on a semi-arid site; bare soil exceeds 60 percent. Photo B shows the same site almost two years after sowing; vegetation has fully established. Soil cover methods in these cases need to last several years for soil protection and plant establishment.

## Mitigating for Low Soil Cover

The primary objective of most revegetation projects is to stabilize the soil surface and create an optimum environment for seeds to germinate and plants to establish. To work, initial surface stabilization has to remain effective until plants become established and can protect the surface from erosion through vegetative cover.

Therefore, the selection of surface stabilization methods can be based on: how effectively does the material stabilize the soil surface, does the material allow good seed germination and plant establishment, and how long does it remain effective (longevity).

**Apply an Organic Mulch**—A variety of mulches with varying qualities and longevities should be considered based on the erosional potential and revegetation needs of each site (Section 5.2.3). Short-fiber materials, such as wood fiber and paper products found in hydro-mulches, applied with a tackifier are a very effective short-term surface cover for protecting soil from rainfall impact. However, after a few months, these products are usually no longer effective. Higher rates of hydromulch and tackifier, which make up BFM products, have a higher degree of soil protection and greater longevity (up to a year). On many sites where the environment is optimum for establishing native plants, protection for less than a year is adequate. However, on sites that are cold, arid, or semi-arid, the establishment of vegetative cover can take longer than one year (Figure 3-56) and soil surface protection will likely require longer-lasting mulches such as straw, pine needles, hay, shredded wood, wood strands, or erosion fabrics.

**Apply a Rock Mulch**—Gravels, cobbles, and stone can provide a stable soil cover. These materials are often left over after rock has been screen from the soil and designed into the slope can add surface stability and reduce evaporation.

## Surface Strength

When soil cover is removed, the surface of the soil is exposed to the erosive forces of raindrop impact, overland flow, freeze-thaw, and wind. How strongly soil particles bind together will determine the degree by which soil particles are detached and moved through soil erosion. Topsoils with good aggregation and high organic matter will be more stable than subsoils or soils low in organic matter. Clay soils have greater strength than soils dominated by sands and silts which are non-cohesive. Seeds have no cohesive properties and, when sown on the surface of the soil without mulches or tackifiers, are very susceptible to erosive forces.

**How to Assess Surface Strength**—Determining the soil texture of the surface soil is a simple way to determine soil strength (Section 3.8.6, see Soil Strength). Soils low in clays (<15 percent) and high in sands will have low surface strength (Figure 3-57). In

most cases, soils lacking topsoil will have reduced surface strength due to the lack of roots and organic matter that hold the soil particles together. The rainfall simulator is an indirect indicator of soil strength because it measures the amount of sediment that



**Figure 3-57 | Sandy soils have low surface strength**

Soils low in clays and high in sands have very low surface strength. Not only are they prone to surface erosion, but even walking on them can leave the surface in a highly disrupted condition.

*Photo credit: David Steinfeld*

is detached from surface soils under various rainfall intensities (Section 3.8.5, see Rainfall). The USDA Natural Resources Conservation Service has developed a field test for determining surface stability for water erosion (Inset 3-4). This method rates how well surface soil samples maintain their stability after being agitated in water.

#### **Inset 3-4 / Bottlecaps test for surface stability**

From Herrick et al 2005a

Place a soil fragment in a bottle cap filled with water. Watch it for 30 seconds. Gently swirl the water for 5 seconds. Assign one of three ratings:

**M** = Melts in first 30 seconds (without swirling)—Not stable

**D** = Disintegrates when swirled (but does not melt)—Moderately stable

**S** = Stable (even after swirling)—Stable

## Mitigating for Low Surface Strength

**Apply Tackifier and Hydromulch**—Surface strength can be increased for up to a year by applying tackifiers by themselves, with hydromulches, or to bond straw onto the site (Section 5.4.2). These products strengthen the bonds between surface soil particles and between the soil particles and the products. Seeds applied with a tackifier are held tightly to the soil surface, reducing the likelihood that seeds will be detached and moved.

**Apply Long-fiber Mulch**—Applying a long-fiber mulch to the soil surface can increase the overall surface strength because of the direct contact of the material with the soil surface and the interlocking nature of the fibers (Section 5.2.3). The application of erosion mats can increase surface strength by an order of magnitude when in contact with the soil surface.

## Infiltration Rates

Infiltration is the ability of the soil surface to absorb water from rainfall, snowmelt, irrigation, or road drainage. A high infiltration rate indicates that the soil surface can transmit high rates of water; a low rate indicates that the surface has low capability of absorbing water. When infiltration rates are lower than the rate of water input, runoff or overland flow will occur. Under these conditions, runoff can detach and transport soil particles, resulting in soil erosion and, in many cases, off-site water quality problems (Figure 3-58). Overland flow can also remove sown seeds.

The size, abundance, and stability of soil surface pores determine the infiltration rates of a soil. Large stable pores created by worms, insects, and channels created by decaying root systems will absorb water quickly and have high infiltration rates, while surfaces that have been compacted, have had their topsoil removed, or are low in organic matter will have poor infiltration rates.

Under undisturbed conditions, infiltration rates are usually high, especially where a litter and duff cover exists. When soil cover is removed, the impact from raindrops can seal the soil surface, creating a crust that will significantly reduce infiltration rates. Infiltration rates are also reduced when soil is compacted by heavy equipment traffic.

**How to Assess Infiltration Rates**—The most accurate equipment for measuring infiltration rates is the rainfall simulator. This equipment simulates rainstorms of different intensities under controlled conditions and measures how the soil surface responds. Infiltration rates are determined at the point when runoff occurs. The amount of runoff water is measured at the bottom of the plot to calculate runoff rates



**Figure 3-58 / Infiltration rates**

When precipitation exceeds infiltration rates, water collects on the surface of the soil and begins to move downslope, causing erosion. On this site, litter and duff layers that typically protect the surface from rainfall impact have been removed, causing low infiltration rates.

Photo credit: David Steinfeld

### 3—PLANNING

and sediment yields (Figure 3-59). While most rainfall simulators were developed for agricultural operations, several have been developed specifically for wildland conditions. These simulators were built for transportability and conservation of water because construction sites are often in remote locations and far from water sources. The “drop-forming” rainfall simulator, developed for wildland use, delivers rainfall at the drop size or impact velocity determined for the climate of the project site (Grismer and Hogan 2004).



**Figure 3-59 / Portable Rainfall Simulator**

Soils low in clays and high in sands have very low surface strength. Not only are they prone to surface erosion, but even walking on them can leave the surface in a highly disrupted condition.

*Photo credit: David Steinfeld*

Using the rainfall simulator in revegetation planning is expensive, yet it is an important tool and not to be discounted because of the cost. Specifically, rainfall simulation used to compare the effects of different mitigating measures, such as mulches or tillage, on runoff and sediment production takes the guess work out of whether such measures are effective. This quantitative evaluation of erosion control methods might be essential in areas where water quality objectives are critical.

In most cases, infiltration rates are estimated under routine field investigation by inference from site conditions. Infiltration rates can be considered high when the soil surface has not been disturbed and has a high percentage of cover. With compaction and loss of surface cover, infiltration rates are proportionally reduced. It is often assumed that construction activities that remove surface cover or disturb the topsoil will have very low infiltration rates that will create overland flow under most rainfall intensities.

## Mitigating for Low Infiltration Rates

**Avoid Compaction**—Where possible, avoid operating heavy equipment on undisturbed soils or soils that have been tilled. Coordinate with project engineers to verify that site preparation does not compromise the compacted roadbed prism which is engineered as a structural entity.

**Tillage**—Infiltration rates can be increased through soil tillage (Section 5.2.2) which reduces compaction, increases macropore space, and creates surface roughness. Depending on the erosional characteristics of the site, the positive effects of tillage on infiltration rates might only remain effective through a series of rainfall events.

**Incorporating Organic Matter**—Incorporating organic matter into the soil surface can increase the longevity of infiltration rates created through tillage by forming stable macropores (Section 5.2.5). Unless macropores are interconnecting or continuous, however, they will not drain well (Claassen 2006). One method for creating continuous pores is to incorporate long, slender organic material, such as shredded wood or hay that overlaps and interconnects within the soil. Higher application rates of shredded wood or straw will result in greater porosity and infiltration rates.

**Surface Mulch**—Applying surface mulch will reduce the effects of rainfall impact on surface sealing and reduce soil erosion rates (Section 5.2.3). It does not, however, necessarily increase infiltration rates. Studies have shown that, while sediment yield can be less with the application of mulches, runoff rates are not necessarily reduced (Grismer and Hogan 2007).

**Plant Cover**—The best long-term way to increase infiltration is to create conditions for a healthy vegetative cover. Good vegetative cover will produce soils with extensive root channels, aggregated soil particles, and good litter layers.

## Slope Gradient

Slope angle or gradient is important in surface stability because it directly affects how soil particles will respond to erosional forces; the steeper the slope, the greater the erosional forces will act on the surface of the soil. Slope gradient is a dominant factor in water erosion and dry ravel erosional processes. In water erosion, the rate of soil loss and runoff from disturbed soil surfaces increases incrementally as slope gradient steepens. In dry ravel erosion, the surface remains stable until a specific slope gradient is reached, and then soil particles move downslope under the direct effects of gravity. This critical angle is called the angle of repose and it can be likened to the angle that accumulated sands make in an hourglass. Only non-cohesive soils, composed of sands, silts, or gravels, have an angle of repose; soils with significant amounts of clays typically do not.

Dry ravel is the downslope movement of individual particles by rolling, sliding, and bouncing (Rice 1982). It occurs more frequently as soils dry out and is triggered by vibrations from vehicles, animals, planes, and wind turbulence. Dry ravel creates a constant supply of material to the ditches at the bottom of the slopes making these slopes very difficult to revegetate unless the surface is stabilized. Seeds that germinate on steep, raveling slopes typically will not have enough time to put roots down deep enough to stabilize the surface before the emerging seedling moves or is buried by soil particles rolling down from above. Figure 3-60 shows dry ravel on a cut slope composed of a sandy soil.

**How to Assess Slope Gradient**—Slope gradient is quantified in several ways, as shown in Figure 3-61. For road construction projects, slope is usually expressed as the rise (vertical distance) over run (horizontal distance). A 1:3 road cut, for example, defines a slope that rises 1 foot over every 3 feet of distance. Biologists, including range, forest, and soil scientists, use percent slope as a measure of slope angle. This is calculated by measuring the number of feet rise over a 100-foot length. Slope gradient as expressed in degrees is not commonly used.

Slope gradient can be measured in the field using a handheld instrument called a clinometer. This equipment reads slope angle in percent slope and in degrees. Readings from a clinometer can be converted to rise over run notation using the chart in Figure 3-61. Road construction plans display the slope gradients for every cross-section corresponding to road station numbers (refer to Section 2.4 and Figure 2-4). Using these cross-sections, slope gradients can be identified on the plan map by color coding the run:rise for cut and fill slopes. For instance, 1:1 cut and fill slopes might be highlighted red for areas of concern, while those areas with gradients 1:3 or less might be light green, favorable areas for mitigation work. This exercise can quickly identify areas with the highest risk for soil erosion and difficulty in establishing vegetation.



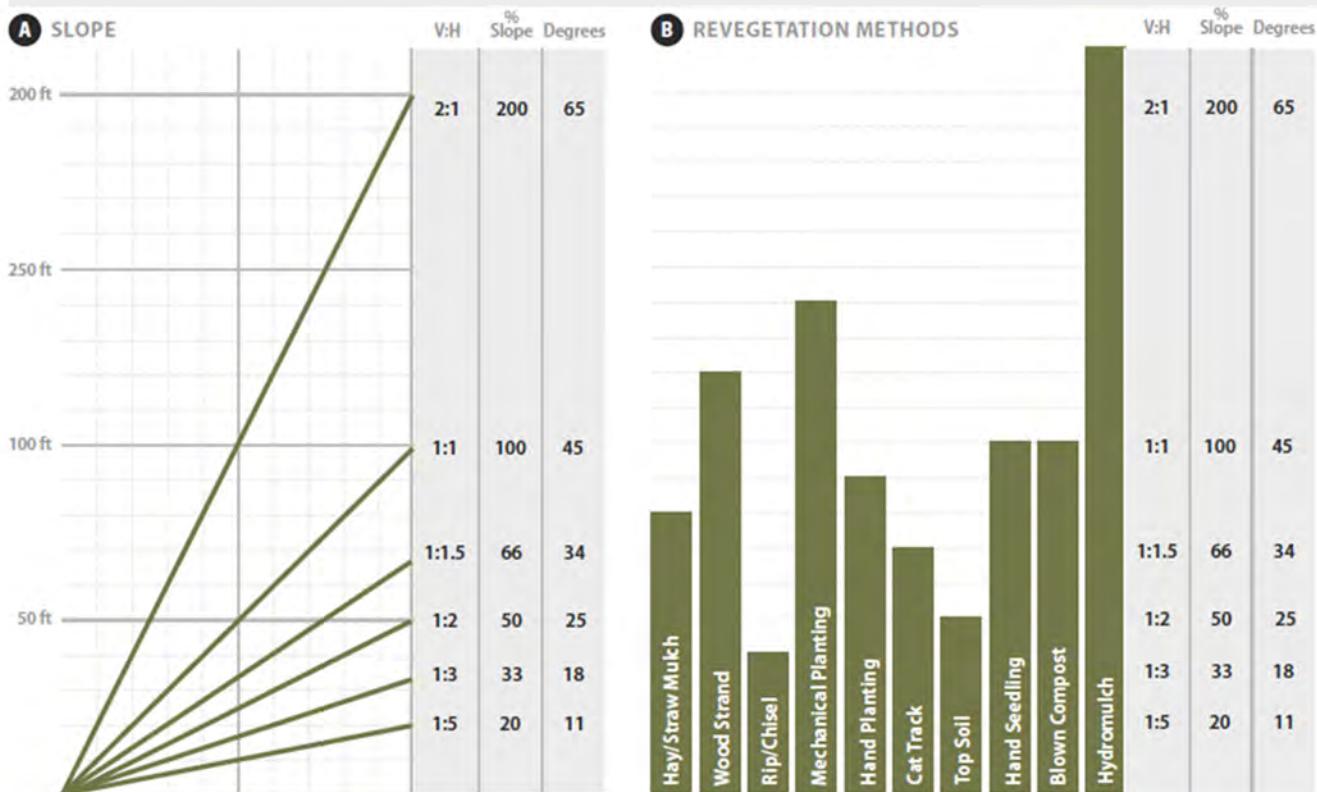
**Figure 3-60 / Dry ravel**

Steep, non-cohesive soils move downslope at a continuous rate. The soil surface is constantly moving and never stable long enough for seeds to germinate and plants to become established. These slopes can remain barren for years. Soils that are high in sands and gravels are the most susceptible to dry ravel. In this picture, trees became established below rock outcrops, where the surface was protected from dry ravel.

*Photo credit: David Steinfeld*

### Figure 3-61 | Revegetation methods and slope gradients

For engineering work, slope is generally expressed as the rise (V) over run (H). For slopes flatter than 1:1, ( $45^\circ$  or 100%), slope gradient is expressed as the ratio of one unit vertical to a number of units horizontal. For example a 1:2 slope gradient indicates that there is one unit rise to 2 units horizontal distance. For slopes steeper than 1:1, it is expressed as the number of units vertical to one unit horizontal (e.g., 2:1 indicates that there is a 2 unit rise to 1 unit horizontal distance). In general to avoid confusion, it is wise to notate the ratio by indicating the vertical (V) and horizontal (H), when defining gradient (e.g., 2V:1H). Range and forest sciences use % slope gradient to describe slope angle. Slope gradient refers to the number of feet elevation rise over 100 feet. A 66 percent slope gradient indicates that for every 100 feet, there is a 66 foot vertical distance rise. Slope gradient controls what type of revegetation treatments can be used (B). The steeper the slope gradient, the fewer tools are available.

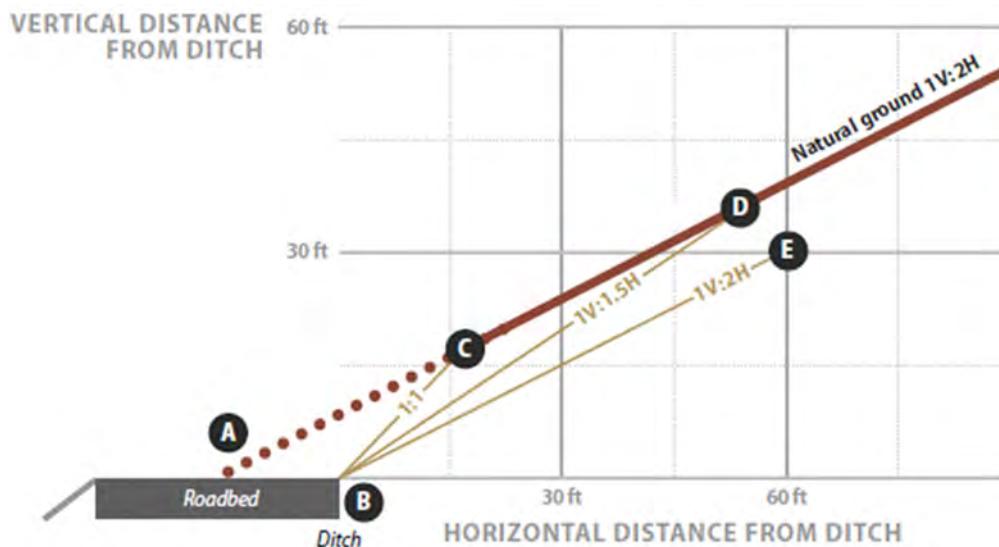


Slope angle plays a key role in revegetation planning because of its potential limitation on the types of mitigation measures that can be implemented. Figure 3-61 shows which practices are generally limited to gentle slopes and which can be implemented on steep slopes.

## Mitigating for Steep Slope Gradients

**Lengthen Cut or Fill Slopes**—Slope gradient can be reduced by increasing the length of the disturbance. This will make the revegetation effort easier and more successful, but will increase the amount of area, or construction footprint, of the project.

Road planners consider many factors when they design the steepness of cuts and fills. A common strategy is to create cut and fill slope gradients as steep as possible. This practice disturbs far less land, resulting in less area to revegetate, and because there is less exposed soil, the potential for soil erosion is considerably reduced. Furthermore, there is a substantial cost savings with less excavation and revegetation work. Figure 3-62 shows an example of how increasing the steepness of a cut slope on a typical 1V:2H natural ground slope substantially decreases the length of disturbed soils.



**Figure 3-62 | Effects of designing steep and gentle gradient slopes on size of disturbance**

The trade-off between designing steep cuts that are difficult to revegetate or creating gentle slopes that disturb more area but are easier to revegetate is demonstrated in this example. The centerline (A) of a new road intersects the 1V:2H natural ground (A to D). In the example, 15 horizontal feet of material have to be excavated from the center of the road (A) to the ditch (B) to create the road bed. The resulting road cut will have varying lengths depending on how steep it is designed. A 1:1 cut slope will expose a 25 ft cut from the ditch (B) to the top of the cut slope (C). A 1V:1.5H slope (B to D) will lengthen this exposure threefold to approximately 80 ft. A 1V:2H slope (B to E) is not achievable because it remains parallel to the natural ground slope.

The main drawback of steepening slopes from a designer's standpoint, however, is that steeper slopes are much harder to revegetate and the selection of revegetation practices available are reduced as the steepness of the slope increases. Vegetation on steeper slopes are also harder to maintain using practices such as mowing. On sites with 1V:5H slopes or less, all revegetation practices are possible; on slopes approaching 1:1, the threshold of most revegetation practices has been reached, as well as the limit of what can be successfully revegetated. It is important to work with the design engineers early in the planning stages to consider the effects of slope gradient on meeting revegetation objectives.

**Create Steep, Hardened Structures**—Creating hardened structures or walls, such as retaining walls, crib walls, gabion walls, mechanically stabilized earth walls, or log terrace structures at the base of a steep slope, will allow gentler slope gradients to be constructed above the structure. With adequate planning, these structures can be revegetated.

## Surface Roughness

Slopes that have roughened surfaces trap water during the initial stages of runoff (Darboux and Huang 2005). Roughened surfaces consist of microbasins that capture and store soil particles and seeds that detach in the erosional processes. Seeds that have been transported short distances into these depressions are often buried by sediments. Moderate seed covering from transported soil can enhance germination as long as the seeds are not buried too deeply. Microbasins can also be relatively stable during the period for seed germination and seedling establishment.

Surface roughness also reduces the effects of wind by reducing wind speed at the soil surface. Seeds may be blown into the depressions and become covered by transported soil. For both wind and water erosion, however, as the microbasins fill up with sediments, they become less effective in capturing sediments and seeds (Figure 3-63).

**How to Assess Surface Roughness**—The simplest method for assessing surface roughness is to take several measurements of the distance between the top and bottom of the microbasin and average these values. Also count the number of microbasins in a 5-foot distance perpendicular to the slope direction. Shallow, closely spaced microbasins may create more area for seed germination but will also fill up faster with sediments, while microbasins that are deeper and farther apart will take longer to fill up and be more effective for erosion control but may have less surface area for optimum plant establishment from seeds.

## Mitigation for Smooth Soil Surfaces

**Leave Surface Roughened**—Many project engineers have a tendency to “beautify” construction sites at the end of a road project by smoothing soil surfaces. While basic landscape shaping is essential, it is important to keep the soil surface as rough as possible. In many instances, leaving cut slopes “unfinished” or in the “clearing and grubbing” stage provides excellent seed bed diversity and growing environment. The diversity of micro-habitats provides greater climatic and soil environments for seed germination. An exception to this rule is where erosion control matting is used for slope stabilization. For the matting to function best, it needs complete contact with the soil surface. On roughened surfaces matting will “tent” over depressions. On smooth surfaces the correctly installed matting will contact the soil surface, hold seeds in place and stabilize the slope soils.

**Surface Imprinting**—Imprinting the soil surface to create micro-relief has been shown to be effective in reducing runoff and soil erosion and increasing plant establishment (Section 5.2.2, see *Roughen Soil Surfaces*).

**Tillage**—Tilling the surface soil layers will leave the site in a roughened condition (Section 5.2.2). This practice can have other beneficial effects on the soil, such as loosening compacted soils.

## Slope Length

Another factor influencing soil erosion and seed transport is the length of slope. As the slope lengthens, so does the potential for transport of sediments and seeds. On very long slopes, the erosive force begins as sheet erosion at the top of the disturbance and often turns into rills and, in extreme cases, gullies at the bottom of the slope (Figure 3-64).

**How to Assess Slope Length**—Slope length can be measured in the field or from road plans. From road plans, slopes can be grouped into different lengths using the road cross sections (see *Chapter 2*). In the field, the effects of different slope lengths can be assessed by observing erosional features on existing disturbed areas. New or old disturbances can be used as references for critical slope lengths. Figure 3-64 demonstrates that at some point downslope rills begin to form. The distance that this occurs is the maximum length of slope for this slope angle and soil type before severe erosion takes place. This distance will vary by other factors addressed in this section (e.g., soil cover, climate, slope gradient, surface strength, surface roughness, and infiltration rates).



**Figure 3-63 | Surface roughness creates favorable environment for germination**

Surface roughness consists of microbasins that are favorable to seed germination and early plant establishment.

*Photo credit: David Steinfeld*



**Figure 3-64 | Surface erosion increase with distance downslope**

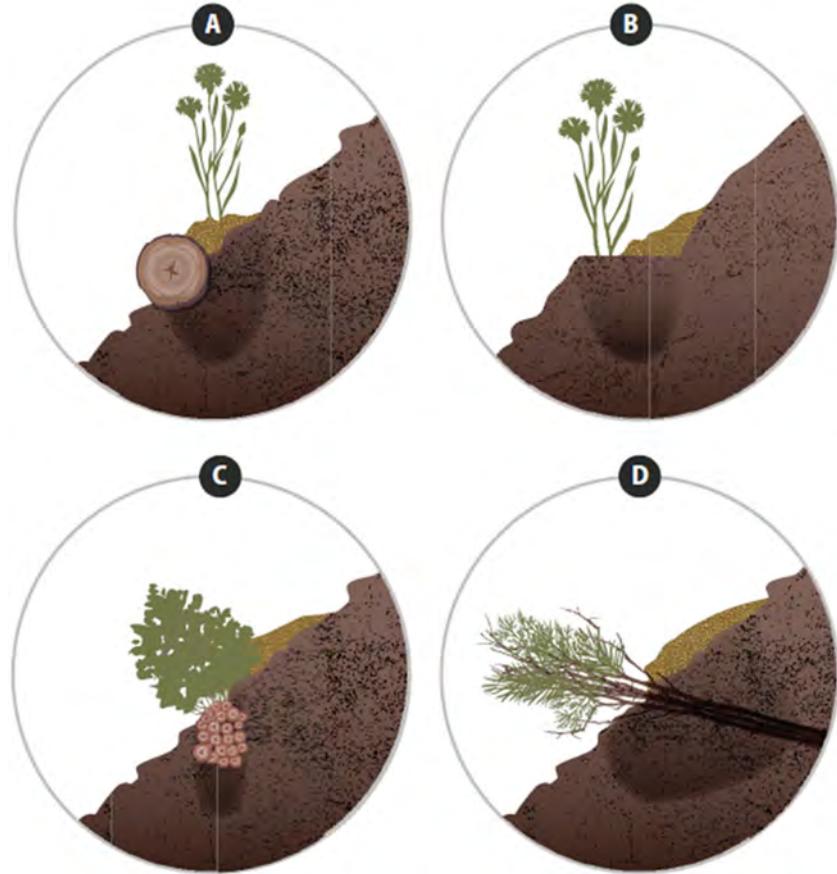
On this site, sheet erosion occurs at the top of the slope (A) and turns to rill erosion (B) as runoff collects. Mitigation that shortens slope lengths to less than the distance between A to B will reduce rill erosion.

*Photo credit: David Steinfeld*

## Mitigating Measures for Long Slopes

**Install Barriers**—Fiber rolls, straw wattles, downed wood, silt fences, or compost berms or compost socks can be laid on the surface as obstructions or barriers to reduce slope length. To be effective, they have to be placed in contact with the soil and perpendicular to the slope gradient, while straw wattles, hay bales, and silt fences have to be trenched into the soil.

**Create Benches**—The creation of slope breaks can reduce slope length. These breaks include benches, steps, or trenches cut into the slope. The reduced gradient at these breaks slows the velocity of overland flow and collects sediments (Figure 3-65).



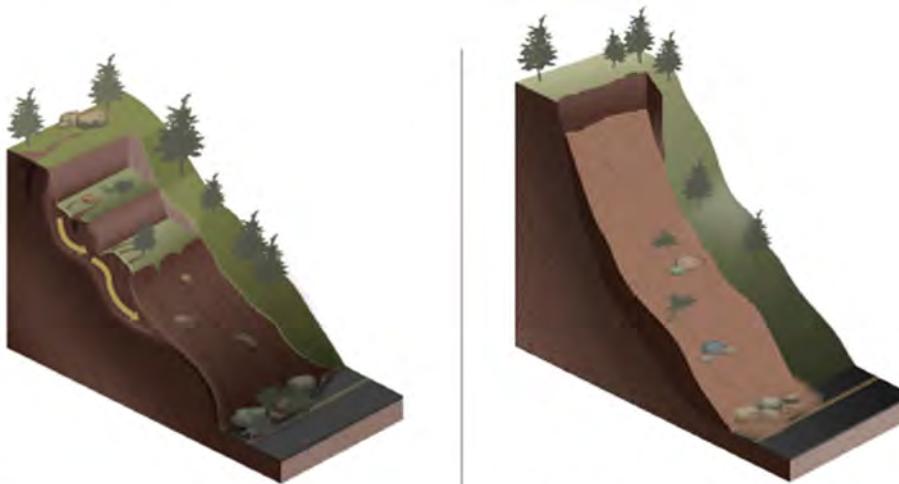
If either practice is applied in along continuous line, they are constructed on contour to the slope. If they are not level, water can collect and move along the structure, much like a channel, and eventually spill onto the slope below, creating rills and gullies. (The exception to this is the construction of live pole drains to redirect water off-site for slope stability [Section 5.4.3, see Live Fascines]). When placed properly along the contour, slope distances are shortened and the structures collect sediments and create areas for plant growth. Plants that are grown above barriers or near the bottom edge of benches can take advantage of water and sediments that collect during rainstorms. Native vegetation can be incorporated into many of these designs to take advantage of increased soil moisture and sediment accumulation. A good practice is to place these structures apart from one another less than the critical distance where rills are expected.

**Figure 3-65 | Structures that shorten slope length**

Structures that shorten the slope length can slow surface runoff, collect sediments and increase soil moisture. Typical treatments include: a) placement of fiber rolls, logs, straw wattles, and compost berms; b) benches, steps, and trenches; c) willow wattles; and d) willow brush layers. Strategic placement of plants can take advantage of increased soil moisture by planting where roots can access the additional moisture. Most species do not respond well to being buried by sediment and are to be planted above or below depositional areas (A and B). However, some species, such as willow, root where the stems are buried, and these species can be planted where sediments are expected to be deposited (C and D).

## 3.8.6 SLOPE STABILITY

This discussion is directed to non-engineers to simplify and make accessible basic slope stability concepts to be understood in developing revegetation strategies. It is by no means a substitute for professional engineering expertise. Technical references for slope stability are many (including Carson and Kirby 1972; Spangler and Handy 1973; Brunsden and Prior 1984; Denning et al 1994), to which the reader is referred for a comprehensive review of this subject. For a detailed evaluation of the role of vegetation in slope stability, the reader is referred to Gray and Leiser (1982).



### Slumps

- Moderate slope gradients
- Circular failure movement
- Cohesive soils
- Deep soils
- Slow moving

### Debris slides

- Steep slope gradients
- Planar failure movement
- Noncohesive soils
- Shallow
- Fast-moving

Creating stable slopes is essential for establishing healthy plant communities, but the reverse is equally true—establishing native vegetation is critical for stabilizing slopes. The following Cohesive soils discussion takes the latter perspective of how to create the most favorable environment for establishing vegetation on potentially unstable slopes and how this approach can be integrated into the overall strategy of slope stabilization. A vegetated slope adds stability to slopes by holding the soil together through a network of root systems and by removing water from the soil, which is the primary driving force behind most landslides.

Slope stability is the resistance of natural or artificial slopes to fail through gravitational forces. The landforms resulting from slope failures are called landslides and they are described by their morphology, movement rates, patterns, and scale. This section will focus on two general types of landslides that are common to road cuts and fills—slumps and debris slides (Figure 3-66). Slumps typically occur on deep soils that are cohesive (e.g., rich in clays). They tend to be deep seated and slow moving. Viewed in profile, the failure occurs in a circular motion, resulting in a series of tilted blocks and circular cracks. Debris slides are shallow, fast-moving landslides that form on non-cohesive soils (e.g., sandy, gravelly). These landslides occur on steep slopes.

Water is the driving force behind most slope stability problems encountered in road projects. It comes as rainstorms, snowmelt, and often as diverted surface water from road drainage. As water increases in a potentially unstable slope, the added weight of the water in the soil plus the increased pressure of water in the pores (pore water pressure) eventually exceeds the strength of the soil and the slope fails (Figure 3-67).

Increased water to a slope is especially a problem where a restrictive layer (e.g., a layer

**Figure 3-66 | Slumps and slides**

Common landslides typically associated with road construction.

Modified after Varnes (1978) and Bedrossian (1983)



**Figure 3-67 | Water pressure and slope stability**

Water pressure is greatest when soils are saturated. Slopes release this pressure through channels created by decomposing roots, animal burrows, and worm holes. When pressures become greater than the strength of the soil, slopes fail. This picture of a decomposed root releasing water pressure was taken 30 minutes before the road cut failed.

Photo credit: David Steinfeld

### 3—PLANNING

of soil that limits water movement) is close to the soil surface. As water moves through the surface horizon and encounters a restrictive layer, it builds up in the pores of the horizon above it, increasing water pressure. The saturated horizon becomes heavier and eventually the slope fails under the added weight of water and increased pore water pressure.

The faster water moves through soil, the less susceptible a slope is to failure. The measured rate at which water is transmitted through a soil mass is its permeability. When permeability is high, water quickly moves out of the soil pores, reducing the potential for increased pore water pressure and slope weight. When permeability is low, water slowly moves through soil and builds up in the soil pores. On gentle slopes, this is typically not a problem, but as slope gradients increase, gravitational forces acting on the slope raise the potential for slope failure. Slope length is another factor important to water movement because the longer the slope, the greater the buildup of water near the base of the slope. This phenomenon explains why many slope failures occur in the mid to lower portions of fill slopes.

Whether a slope fails or not is ultimately due to the strength of the soil. Soil strength is affected by the amount of clay present in the soil, the level of compaction (the greater soil compaction, the more stable the soil), and the presence of roots (more roots are better). Compaction of soils to increase soil strength usually takes priority in road design over creating optimum soil conditions for root growth. With this practice, not only are there fewer root systems to add slope stability, there is also reduced vegetative cover, which is important for intercepting and removing water from the soil. A road design that integrates vegetation into the stability of the slope can meet both road and revegetation objectives, but it will take a collaborative approach.

This section will discuss each of the following parameters as they relate to increasing slope stability through revegetation treatments:

- Permeability
- Restrictive layers
- Water input
- Slope length
- Slope gradient
- Soil strength

## Permeability

The rate at which a volume of water moves through soil material is its permeability (technically referred to as hydraulic conductivity). Soils that have moderate to high permeability rates tend to be more stable than those with low permeability rates. Where permeability is low, water fills the soil pores but does not drain quickly, adding additional weight to the slope and increasing the pore pressure. Both factors reduce the overall strength of slopes and increase the likelihood that slope movement will occur when other conditions are right.

Soils with large interconnecting pores have a higher permeability than soils with smaller pores that are less interconnecting. Soil textures that are well-graded (soils that have only one particle size) typically have a higher permeability than poorly graded soils (soils having a range of particle sizes from clays to small gravels). For example, poorly graded granitic soils have low permeability rates because the different-sized particles are neatly packed together, restricting the pathways for water flow. Well-graded soils, such as pure sands and gravels, have high permeability because pores are large and interconnecting. Alternatively, compacted soils often have low permeability rates because of the reduced or destroyed interconnecting macropores.

**How to Assess Permeability**—Simple field tests, such as percolation tests, historically utilized in assessing septic leach fields, can be used for determining permeability rates. A small hole is excavated and water is poured to a specified depth. The time to drain the water from the pit is measured in inches per hour.

These tests are run at different soil depths to determine if permeability rates change. Results from percolation tests are subject to significant variability. Nevertheless, they can indicate the relative permeability of different soil types or different soil disturbances.

Engineering laboratory tests for determining permeability include the constant head permeameter for coarse-grained soils. In this test, a soil sample is placed in a cylinder at the same density as the soils in the field. Water is introduced and allowed to saturate the sample. A constant water elevation, or head, is maintained as the water flows through the soil. The volume of water passing through the sample is collected and this provides a direct measurement of the flow rate per unit of time. The test can be repeated at various densities to determine the corresponding permeability. The test can also be repeated for various additions of organic amendments and compaction levels to determine the effects of these treatments on permeability. Consider consulting a soil lab for how to collect and submit samples for these tests.

Where testing is not feasible, engineering and soil texts can give ranges for expected permeability based upon the soil gradation and classification. A field assessment can also give some indication of permeability rates. Subsoils or soils lacking organic matter that have a range of soil particle sizes, from clays through small gravels, have a propensity for low permeability rates, especially when they are compacted ([Section 3.8.2, see Soil Structure](#)).

## Mitigating for Low Permeability

**Tillage**—Loosening compacted soil through tillage practices ([Section 5.2.2](#)) increases permeability by creating large fractures or pathways for water to flow. However, tilled soils often return to near-original permeability as the soils settle over time.

**Organic Amendments**—Long-fibered organic matter, such as shredded wood, tilled deeply into the soil will increase the infiltration and permeability of the soil because larger, interconnecting pore spaces are created ([Section 5.2.5](#)). Several studies evaluating the incorporation of unscreened yard waste suggests that an optimum rate of organic matter additions for increasing infiltration and improving soil structure is approximately 25 percent compost to soil volume (Claassen 2006). In addition, incorporating organic matter can increase slope stability because amended soils are lighter in weight than non-amended soils (mineral soils can weigh 10 to 20 times more than soils amended with 25 percent organic matter). The reduced soil weight lowers the driving forces that create unstable slopes.

## Restrictive Layer

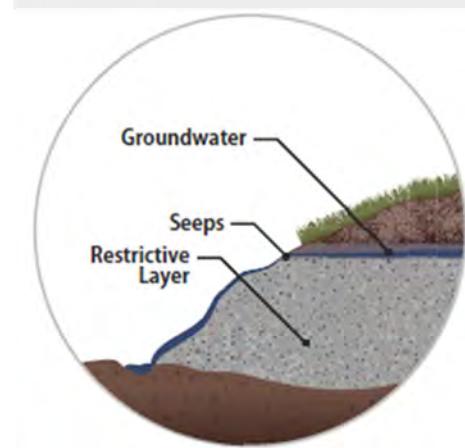
A restrictive layer is any soil horizon or stratum (including unfractured bedrock) that has very low permeability. As water flows through a surface horizon with good permeability and encounters a restrictive layer, the rate of downward water movement slows and water builds up in the pores above. The boundary between the permeable surface layer and the harder subsurface material is often a zone of weakness, sometimes referred to as a slip plane. Slope failure occurs when the pores in the soils above the restrictive layer become saturated with water to a point where the pore water pressure and soil weight exceed the soil strength. The depth of slope failure depends on the thickness of the soils above the restrictive layer. On slopes where the restrictive layer is near the surface, slope failure, if it occurs, will be shallow ([Figure 3-](#)



**Figure 3-68 | Restrictive layers can decrease slope stability**

In this photograph, a restrictive layer is very close to the soil surface. During a rain storm, water moved through the shallow soil layer and encountered a restrictive layer. Water then moved downslope, through the soil, building up pressure, until the increased pore pressure and soil weight exceeded the strength of the soil. At this point, the mid to lower slopes failed.

*Photo credit: David Steinfeld*



**Figure 3-69 | Restrictive layers and ground water**

Groundwater moves downslope above restrictive soil layers. Seeps are seen in road cuts where the restrictive layer is exposed. Increased soil water that occurs above restrictive layers can decrease slope stability.

68). Where the contact is deeper, the soil movement will be more extensive. The types of landslides that occur with restrictive layers are debris slides.

**How to Assess Restrictive Layers**—Restrictive layers in natural settings can be inferred by the presence of seeps and springs. These features occur most often at the point where a restrictive layer is intercepted or exposed to the surface. These can be intermittent features that are observed in the winter or spring but are dry in the fall or summer. The vegetation around permanent and temporary seeps and springs is typically composed of water-loving species, such as sedges (*Carex* spp.) and rushes (*Juncus* spp.). Figure 3-69 shows how a seep is created when a cut slope intersects a restrictive layer.

On construction sites, restrictive layers can be created by placing a loose soil or compost over highly compacted subsoil. These layers can be identified in the field by determining soil strength using a soil penetrometer or shovel to find compacted or dense soils (Section 3.8.6, see Soil Strength). On roadbed fill slopes or road cut slopes the compaction of the soils can be assumed. This layer is then assessed for the approximate amounts of silt and clay using the field texturing method. Dense or compacted soil layers that are high in silts and clays are likely to be very restrictive to water movement. Field permeability tests can be used to determine the rates of water flow (Section 3.8.6, see Permeability). Often restrictive layers are not observed until after construction when they manifest as intermittent seeps or wet areas in cut and fill slopes. Geotechnical investigation will determine if these features are due to shallow water movement associated with restrictive layers or interception of deeper subsurface water.

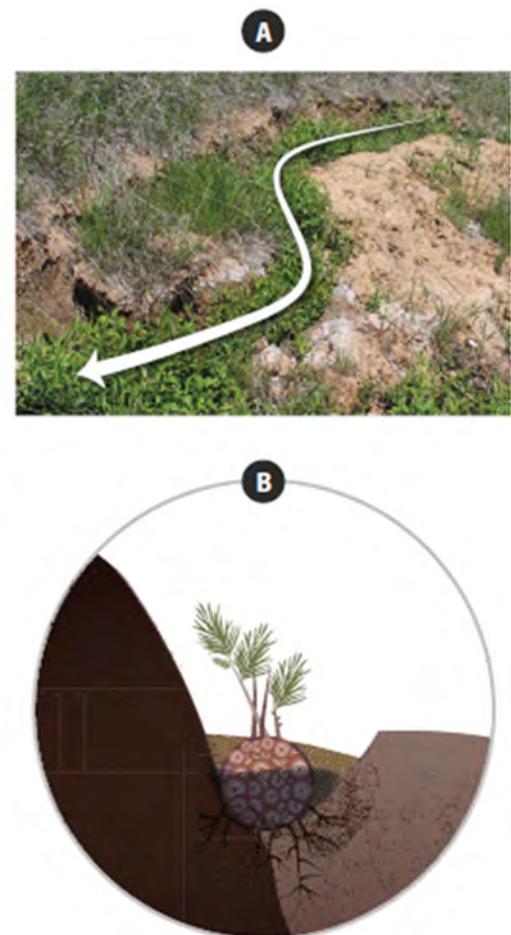
## Mitigating for Restrictive Layers

**Tillage**—If restrictive layers are within several feet of the soil surface, site treatments that break up portions of this layer can increase permeability, which will increase stability. Site treatments that accomplish this are bucket tillage, deep ripping, spading, and fill cut (benching and backfilling), which are discussed in Section 5.2.2. The drawback to tillage is that it will reduce soil strength in the short term until roots occupy the soil and increase soil strength. Temporary irrigation systems have been installed on sites where quick establishment of grass cover during summer and fall is essential for slope stabilization prior to winter precipitation (Hogan 2007). Tillage techniques that leave a deep uneven subsurface profile reduce the potential for downslope debris slides.

**Organic Amendments**—Where possible, the incorporation of organic matter will help keep the restrictive layer from returning to its original soil density and allow more time for roots to establish (Section 5.2.5). As described above, soil strength is reduced until vegetation becomes established, but the negative effects of reduced soil strength are offset by the increased permeability and reduced soil weight of the organic amendments.

**Key-In Surface Material Installations**—Installation of straw wattles, staked in place on contour, prior to installation of topsoil or compost can hold permeable surface layers in place long enough for plant root systems to knit the layers together. Consider using fully biodegradable wattles.

**Live Pole Drains**—Live pole drains are constructed to intercept water from seepage areas and remove it through a system of interconnecting willow bundles to more stable areas, such as draws, ditches, or other waterways (Figure 3-70). The interception and flow of water encourages the establishment and growth of willow cuttings along the length of the live pole drain.



**Figure 3-70 | Live pole drains**

Slumps are characterized by scarps, cracks, and benches. Water collects on benches and in cracks where it is transmitted into the slide mass, creating continued instability. Slopes can be stabilized by removing water through a series of hand-dug surface ditches. Using the cracks as guides for the location of ditches, they are filled in with soil and dug wide enough for a willow fascine (Section 5.4.3, see Live Fascines) to be placed in the bottom (A). Called “live pole drains” (Polster 1997), these structures not only quickly move surface water from the slide to more stable areas, but the willow cuttings in the fascines, encouraged by the presence of high soil moisture, grow into dense vegetation (B) that stabilize the slide through the deep rooting and dewatering.

*Photo credit: David Steinfeld*

## Water Input

Water, which is the driving force behind most landslides, comes through rainfall, groundwater flow, snow melt, or diverted from other areas through road drainage. Landslides often occur after a series of strong winter storms have delivered a high amount of rainfall over a short period of time. Under these conditions, soils have not had sufficient time to drain before the next storms arrive and water in the soil builds up to a very high pressure (Figure 3-67). With additional storms the combination of increased pore water pressure and additional weight of water in the slopes eventually leads to slope failure. Landslides can occur where water from road ditches or other road features drain water onto marginally unstable slopes. Major site factors that affect water input are rainfall duration and intensity, rain-on-snow events, and road drainage.

**How to Assess Water Input**—See Section 3.8.1 for how to assess water input.

## Mitigating for High Water Input

**Proper Surface Drainage**—Increased water is the driving force behind slumps and debris slides. Designing proper water drainage is probably the most important measure to implement for slope stability (Gedney and Weber 1978). Where road water is inadvertently routed into potentially unstable areas, there is a greater potential for slope failure (Fredricksen and Harr 1981). Road projects that are designed to move storm water away from or out of unstable slopes as quickly as possible through road and slope drainage structures increase slope stability. In some cases, installing a curb at the top of the cut slope will effectively move water away from unstable slopes below. Coordinate with engineers to ensure that surface runoff, during construction of phased projects, is addressed prior to completion of final storm water system.

**Species Selection**—Select species that have adapted to wet soils. These include sedges (*Carex* spp.), rushes (*Juncus* spp.), bulrushes (*Scirpus* spp.), willows (*Salix* spp.), cottonwoods (*Populus* spp.), and cedars (*Thuja* spp. and *Chamaecyparis* spp.). Because each species has a unique way, or strategy, of modifying the moisture regime of a site, planting a mixture of species is a way of ensuring that all strategies are represented on the site. For instance, willows establish quickly and can draw large quantities of moisture from the soil, but only when the willows have leaves. Cedars, on the other hand, are slower to establish, but longer lived. They can withdraw moisture from the soil during the winter, unlike deciduous species. When they are well established, they intercept large amounts of water in the crowns, preventing precipitation from reaching the soil. Trees have a great ability to significantly deplete moisture at considerable depths (Gray and Leiser 1982). Wetland species, such as rushes and sedges, unlike many tree and shrub species, grow well in saturated soils.

**Live Pole Drains**—The live pole drain (Polster 1997) is a biotechnical engineering technique where continuous willow bundles (Figure 3-70) are placed across a slope, much like an open drain, to redirect water to a more stable area, such as channels and draws. Where small slump failures have occurred during or after construction, live pole drains can be installed to increase slope drainage, add root strength, and remove soil moisture from the slide mass.

## Slope Length

Slope length is important for stability because the longer the slope, the more water concentrates in the lower portion of the slope. Increased water increases pore water pressure and soil weights, thereby decreasing the stability of the mid to lower sections of long slopes. This is one reason why slumps are often observed in the mid to lower slope

positions of longer fill slopes.

**How to Assess Slope Length**—Slope length can be obtained from road plans or measured directly in the field using a tape.

## Mitigation for Long Slopes

**Live Pole Drains**—Using live pole drains (discussed in the previous section), shortens the distance water is transmitted through the hill slope by intercepting surface and subsurface water in ditches at frequent slope intervals. Captured water is transmitted downslope through a system of continuous fascines to a stable channel (Figure 3-70).

**Create Benches**—Another method of reducing slope length can be the creation of a slope break, such as benches, steps, or trenches cut into the slope. The reduced gradient at these breaks slows the velocity of overland flow and collects sediments. However, unless the water is directed off the slope, total water input to the slope is not reduced.

## Slope Gradient

As slope gradient increases, the destabilizing gravitational forces acting on the slope become greater. On a level surface, the gravitational force stabilizes the soil mass but as gradients increase, the strength of the soil mass to resist sliding along the failure plane decreases.

**How to Assess Slope Gradient**—Assessing slope gradient is discussed in [Section 3.8.5](#) (see Slope Gradient).

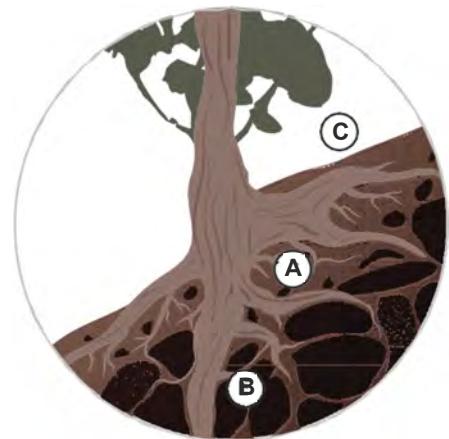
## Mitigating for Steep Slopes

Mitigating measures for steep slopes are discussed in [Section 3.8.5](#) (see Slope Gradient).

## Soil Strength

Soil strength (technically referred to as shear strength) is the characteristic of soil particles to resist downslope forces. Physical and biological factors play roles in soil strength. The major physical factors contributing to increased soil strength are reduced porosity (compacted soils), greater range of particle sizes (poorly graded soils), greater angularity of the soil particles, greater surface roughness, and the presence of silts and clays that add cohesive strength (Hall et al 1994).

The biological components of increased soil strength are the matrix of roots that reinforce the surface horizon, roots that anchor an unstable soil mantle to stable subsoils or rock, and stems (e.g., trunks of trees) that add support to the soil immediately uphill (Hall et al 1994) (Figure 3-71). The physical factors, unfortunately, do not always support the biological factors. For example, high porosity soils are of particular interest to the designer because of the role porosity plays in root growth. From an engineering standpoint, however, high porosity soils have lower soil strength because soil particles are not packed closely together and are less interlocking. Balancing the needs of creating a healthy soil for optimum vegetation while still maintaining slope stability until established vegetation adds root strength to the soil is a challenge to engineers and revegetation specialists. A road design that creates soils with high porosity will also decrease soil strength in the short term; however, if this practice increases the biological productivity of the site, the long-term result is a net increase in soil strength (Gray and Leiser 1982). The growth habits of native species can greatly influence slope stability because each species has a unique rooting pattern and root tensile strength. For instance, grass roots are very fibrous and abundant in the



**Figure 3-71 | Plant roots and slope stability**

Plant roots and stems increase slope stability by (A) reinforcing the surface horizon through a matrix of roots, (B) anchoring surface horizons to rock or subsoils, and (C) stems supporting the soil upslope.

surface horizon, adding surface stability when the grass cover is high. Grass and forb roots, however, add very little soil strength at deeper depths because their roots are not as strong and do not penetrate as deeply as tree roots (Gray and Leiser 1982). Alternatively, roots of shrub and tree species are long and deep rooted, with relatively high tensile strength (Gray and Leiser 1982). The main advantage of tree and shrub species is the long vertical roots (taproots) that can cross failure planes and bind the soil strata together (Figure 3-71).

**How to Assess Soil Strength**—The designer will probably not perform engineering tests for soil strength, yet knowing a little about these tests could be important, especially if the designer is making recommendations that soils on potentially unstable sites be deeply tilled or amended with organic matter.

A common method engineers use to estimate soil strength is to correlate soil classification (from sieve analysis and the characteristics of clay particles) with published literature values. Shear vanes or cone penetrometers are good methods to approximate the strength of fine-grained soils in the field, and published research is used to correlate these readings with laboratory shear strength test results.

The triaxial shear test is a more precise laboratory method to determine shear strength of soils. In this test, a long cylinder of the soil is placed in a latex membrane and submerged in a clear plastic cylinder filled with water. A vertical pressure is applied to the cylinder at a slow rate until the soil sample shears.

Very sensitive strain gauges measure the soil displacement, applied forces, and any pore water pressures that develop. Various water pressures are applied to the cylinder to simulate the confining pressure of soil depth (Brunsden and Prior 1984). This test can be used to determine the shear strength of soils that have been amended with organic materials.

## Mitigating for Low Soil Strength

**Biotechnical Engineering Techniques**—Many biotechnical slope stabilization techniques use vegetative cuttings from willows or other easy-to-root species to structurally reinforce the soil. As these materials root, they add further stabilization to slopes through interconnecting root systems and soil moisture withdrawal. These practices include stake planting, pole planting, joint planting, brush layers, and branch packing (Section 5.4.3).

**Shrub and Tree Seedlings**—On drier sites, where willow cuttings are less likely to survive and grow, shrub and tree seedlings can be used. While these species are slower growing, they usually have deeper root systems and persist longer once they are established. Grass and forb species can quickly establish on drier sites, but soil strength is limited to the surface of the soil profile where the roots are most abundant. For this reason, grasses and forbs do not provide as much stability (Figure 3-72). On potentially unstable sites, grasses grown between shrub and tree seedlings add soil strength to the



**Figure 3-72 | Effects of roots on slope stability**

This debris slide (noted by its shallow, steep appearance) took place two years after construction when the grass and forbs were fully established. Establishing shrub species, rather than grass and forb species, on steep, potentially unstable slopes, would be better for long term stability because shrub species are deeper rooting and have higher root tensile strength.

*Photo credit: David Steinfeld*

surface soil while tree and shrub species become established. On dry sites, however, grasses are excluded around seedlings or vegetative cuttings until the latter have become established. Because of the steep, shallow nature of many of these sites, planting seedlings is not always practical or successful. It is worthwhile to consider hydroseeding or hand sowing, covered by a surface mulch that will protect the surface from erosion for several years while the shrubs become established, should be considered.

**Soil Improvement**—Improving soil productivity will increase root densities and enhance slope stability (Halle et al 1994). Mitigating measures that improve water storage, organic matter, and nutrients should, with time, increase slope stability. Some practices, such as soil tillage, reduce soil strength in the short-term. However, once plants have become established more roots occupy the soil and slope stability is increased. Consider integrating tillage with practices that quickly reestablish vegetation to ensure that slope stability is not compromised in the short term. On slopes where root strength is critical for stability in the first year after construction, irrigation could be considered to quickly establish a dense vegetation cover.

**Temporary Soil Stabilization**—It is important to implement erosion control practices that stabilize the surface of steep slopes while vegetation develops a strong root system and top growth. Where trees are inappropriate because of clear zones and shrubs inappropriate because of sight lines, erosion control products work well to stabilize soils while grasses and forbs establish.

## 3.9 IDENTIFY FACTORS THAT AFFECT POLLINATORS

For projects where one of the objectives is to improve pollinator habitat, it is important to identify the core habitat elements supporting local pollinator populations. Similar to identifying limiting factors to plant establishment, as discussed in [Section 3.8](#), the designer evaluates a project area for how well it supports a diversity of pollinators prior to construction and identify those factors important for improving pollinator habitat on roadsides after project construction.

The Pollinator Habitat Assessment Checklist ([Chapter 7](#)) is a guide to assessing current and potential habitat conditions based on information collected during the field information planning phase ([Section 3.3](#) and [Section 3.6](#)). While the checklist provides habitat characteristics important for most pollinator habitats, the designer may want to modify the checklist to also include the climate, soils, vegetation, and pollinator species of interest specific to the project area and project objectives.

Once the designer has assessed the current condition of the pollinator habitat and identified parameters that are limiting to pollinator populations, a list of mitigating measures can be developed. Mitigating measures are site treatments that will improve pollinator habitat. For example, if breeding habitat is limiting, a mitigating measure is to include butterfly host plants.

There are usually several ways to mitigate each limiting factor and designers can select the measures that most suit their site conditions and objectives discussed in the following section.

### 3.9.1 NECTAR AND POLLEN SOURCES

Pollinators such as beetles, flies, wasps, moths, butterflies, bees, hummingbirds, and bats all forage for food on flowers. Nectar and pollen are sources of carbohydrates and protein, respectively, for pollinators. The act of pollination is an incidental effect of feeding or gathering food: animals visit flowers seeking sustenance, and in the process

transfer pollen grains which allow flowering plants to reproduce.

Flowering plants in roadsides are important sources of nectar and pollen for pollinators that reside within the roadside habitat (e.g., Munguira and Thomas 1992) as well as those that use the roadside as a partial habitat for foraging but reproduce or overwinter elsewhere (e.g., Ouin et al 2004). Adults of bees, butterflies, wasps, hummingbirds, and many species of flies, moths, beetles, and bats feed on nectar to maintain their energy levels. Some adult beetles and flies require the protein that pollen provides in order to reproduce. Female bees actively collect pollen to take back to their nests (Figure 3-73), where they provide for their offspring by leaving a supply of pollen moistened with nectar. The availability of pollen and nectar influences the abundance and diversity of pollinators found on roadsides (Saarinen et al 2005; Hopwood 2008).

## Attractiveness and Overlapping Bloom Times

It is important to include plants that are known to provide floral resources and attract pollinators. Native plants are more attractive to native pollinators than exotic plants, even when both plant types are present in sites (Hopwood 2008; Williams et al 2011; Morandin and Kremen 2013a). Some species of native plants are particularly attractive to a wide range of pollinators, offering either large quantities of nectar, high quality nectar, or pollen with high protein content. Sourcing plant material locally or within the ecoregion is best for pollinators because the phenology (flowering period) can also differ with the provenance of the plant material (Norcini et al 2001; Houseal and Smith 2000; Gustafson et al 2005) (Section 3.3.3 and Section 3.13.2). Bloom times of non-locally sourced plants have the potential to be out of sync with pollinators. This may be particularly problematic for bee species that are pollen specialists (e.g. *Melissodes desponsa*, a pollen specialist on thistles, Figure 3-73) and are reliant on the pollen from a small subset of plants and time their emergence from overwintering with the bloom time of their host plants.



Additionally, it is important to have flowers available to pollinators throughout the growing season. While pulses of bloom can provide critical resources to pollinators, sustained resources create functional habitat throughout the growing season to best support robust populations and communities. For example, solitary bees have distinct flight seasons that last 4 to 6 weeks. Bees that emerge early in the growing season can

**Figure 3-73 | Bees and other pollinators rely on flowering plants as sources of food**

In this picture, a female long horned thistle bee (*Melissodes desponsa*) collects pollen from a tall thistle (*Cirsium altissimum*) and stores it on her hind legs to transfer it back to her nest, where she will leave it for her young to eat.

*Photo credit: Jennifer Hopwood/Xerces Society*

### 3—PLANNING

begin to forage in February in warmer parts of the U.S., with late-season bees foraging through October. Social bee species have overlapping generations and require forage throughout the growing season.

Gaps in bloom early in the spring or late in the fall can be easily overlooked during the planning stages, but these are times when nectar and pollen are very important to the health of pollinators such as bumble bees, honey bees, and migrating monarch butterflies. Early-season pollen and nectar sources are important for those species that have flight seasons in the spring and will lead to greater reproduction of those species. Early season forage is also important for pollinators like bumble bees and honey bees that fly all season long but that are in need of floral resources early in the spring after overwintering. Late season flowers provide resources that ensure that queen bumble bees have ample food going into winter hibernation, and that honey bee colonies have enough food stores to last through the winter.

The availability of pollinator-friendly plant species and overlapping bloom times at a site prior to construction can be assessed by recording the plant species that are flowering and their percent cover every 2 to 4 weeks starting at the beginning of the growing season through the fall (February through October in California). These vegetation surveys would be conducted on both the project site and undisturbed reference sites ([Section 3.6.1](#)).

It is best to have a minimum of 3 to 5 species of trees, shrubs, or wildflowers blooming during each season (spring, summer, fall). Ideally there would be 5 to 10 species blooming per season, overlapping to ensure availability of resources to the entire pollinator community. Increasing the diversity of flowering plants in seed mixes and planting plans can help to prevent gaps in bloom. If flowering species have been recorded at reference sites during the growing season, these species can be used to develop a list of species with overlapping blooms for the revegetation project. When field surveys have not been conducted, a [species list using the ERA tool](#) can be used to generate lists of the recommended workhorse and pollinator-friendly plant species for all EPA Level III ecoregions in the continental United States. The ERA will also denote the bloom periods of the plant species and which general groups of pollinators the plant species will benefit.

## Floral Diversity and Cover

Whenever possible, it is best to increase the diversity of wildflowers and blooming shrubs or trees. Pollinator diversity increases with increasing flowering plant diversity (Potts et al 2003). Different pollinators have different floral needs and preferences, so including a diversity of plant species with different flower shapes, sizes, colors, and growth habits helps support the greatest abundance and number of species of pollinators (Ghazoul 2006; Ponisio et al 2015). Additionally, greater flowering species provide greater diversity of pollen and nectar sources for honey bees. Diversity in diet can help support honey bee immune system health (Alaux et al 2010; Di Pasquale et al 2013).

Floral cover is also important; having sparse resources will not sustain pollinator populations whereas high densities of blooms are more attractive and can support higher numbers of pollinators (Herrera 1998). For example, it is best for pollinator habitat to maximize the percentage of wildflowers included in revegetation projects involving herbaceous plants (Figure 3-74). The combination of grasses and wildflowers can effectively resist weed colonization and provide soil stabilization. A planting seed mix that consists of grass species at 50 percent or less of the seed mix volume can prevent the grasses from outcompeting the wildflowers. It is ideal to aim for 45 percent cover of flowering plant vegetation across a growing season. For example,

### 3—PLANNING

flowering plant cover could be spread out throughout the growing season, with 15 percent cover of vegetation in spring, 15 percent in summer, and 15 percent in autumn.

It can be helpful to emulate the species diversity found on reference sites located in nearby natural plant communities. Diverse plantings that resemble natural communities are the most self-sustaining and long-lasting plantings because they better resist weed invasions and pest outbreaks. A general walk through of the area may be enough to identify flowering species or a more intensive sampling can be conducted using monitoring procedures described in [Chapter 6](#). The Species Presence monitoring procedure ([Section 6.3.3](#)) can be used to record floral diversity and the Species Cover monitoring procedure ([Section 6.3.2](#)) can be used to record floral cover.

When working at sites with low diversity and floral cover, clumping single species together can benefit pollinators (Frankie et al 2002). For example, planting small clusters of single species of flowering plants to form patches of color when in bloom helps pollinators to spot the plants quickly. This is especially helpful for pollinators searching landscapes for limited resources and also helps pollinators to move quickly and efficiently between flowers to collect the resources they need. In larger plantings, clusters are less important as long as flowering plants are abundant.



**Figure 3-74 | High plant diversity and floral cover are important for pollinators**

This roadside through the Tonto National Forest in Arizona has high plant diversity and high floral cover, two characteristics that are valuable for pollinators.

*Photo credit: Arizona Department of Transportation*

Floral cover and diversity increases with site productivity. By improving soil characteristics, floral cover and diversity will increase. On projects where resources are limited, areas important for pollinator habitat can be identified and soil improvement conducted specifically in those areas.

### 3.9.2 BREEDING HABITAT

Pollinators have diverse habitat needs for breeding, in particular for their egg-laying sites. The habitat needs for insect pollinators are especially varied because insect pollinators select locations in which to lay their eggs based on the needs of their larvae. The main groups of insect pollinators, bees, beetles, flies, moths, butterflies, and wasps, all have four distinct life stages: egg, larva, pupa, and adult. Eating and growing are the primary functions of the larval stage, followed by the dormant pupal stage during which body tissues rearrange and transform into the adult stage. The primary goal of the adult stage is reproduction. The needs of pollinator larvae are often different than the needs of adults. For example, the eggs of some beetles and flies are laid near prey or vegetation or in the leaf litter so that their larvae, which are predatory, have access to the food resources they need. Butterfly and moth larvae (also known as caterpillars) consume vegetation and are found on the leaves and stems of plants, and their eggs are laid on certain species of host plants preferred by their larvae. Bees and predatory wasps construct nests in which to lay their eggs (Section 3.9.3). Roadsides can serve as breeding habitat for all of these and more.

#### Host Plants

Egg-laying sites for butterflies and moths consist of plants upon which the adult will lay eggs and the larvae will feed after hatching (Figure 3-75). Some butterflies and moths may rely on plants of a single species or genus for host plants. For example, caterpillars of the monarch butterfly feed only on species of milkweed (*Asclepias* spp.) and closely related genera. Some species are even more specialized; caterpillars of the Karner Blue butterfly (*Lycaeides melissa samuelis*) will only survive on lupine (*Lupinus perennis*). Other species may exploit a wide range of plants. The larvae of some swallowtails (*Papilio* spp.) can feed on a range of trees, shrubs, and wildflowers. Given this lifecycle pattern, establishing caterpillar host plants is recognized as a way to sustain butterfly and moth populations (Croxton et al 2005; Feber et al 1996).

Roadsides with host plants can support habitat generalist butterflies as well as habitat specialists and migrant species such as the monarch butterfly (e.g., Ries et al 2001; Kasten et al 2016).

There are two main considerations for assessing whether butterflies and moths are using the site for breeding: (1) if host plant species are present within the site and (2) if present, if eggs and/or caterpillars of butterflies and moths are also present at the site. If the goals of the project include supporting particular species of butterflies or moths, it is useful to monitor the presence of the host plants of the target pollinator species.

Section 6.4.3 includes a monitoring procedure for host plant abundance and population estimates of a target species (e.g. monarch butterfly) based on counts of eggs and caterpillars.

Many plants used in revegetation projects serve as butterfly and moth host plants, but it may be necessary to include additional species to support particular butterflies or moths. For example, planting milkweeds will contribute to the recovery of the monarch butterfly (Inset 3-5). The ERA tool includes information about the host plant needs of target butterfly and moth species. For additional information, see the HOSTS database of the world's Lepidopteran host plants created by the Natural History Museum, London. The database can be searched by either plant species or Lepidopteran species.



**Figure 3-75 | Caterpillars, butterflies, and moth larvae devour plant material**

This giant silkworm moth (*Antheraea polyphemus*) will feed on a number of host plant species, including maples (*Acer*) and plums (*Prunus*).

*Photo credit: MJ Hatfield*

## Egg-Laying Sites

The breeding habitat needs are less understood for groups of pollinators beyond butterflies and bees, though what is known suggests that egg-laying sites for beetles, flies, and other pollinators are usually close to food sources for their larvae. If larvae are carnivorous, as is the case for lady beetles, eggs are laid on vegetation near their aphid prey, while syrphid or flower fly species that consume detritus or other decomposing materials as larvae lay their eggs in leaf litter. Egg-laying sites can include moist soil, leaf litter or plant debris, on foliage near prey, or in crevices under bark or rocks (Table 3-11).

### **Inset 3-6 / Milkweeds and adjacent landowner concerns**

Increasing milkweed populations in North America is critical to the recovery of the monarch butterfly, but one obstacle to widespread inclusion of milkweeds in new plantings is the perception that milkweeds are in fact weeds. Concerns include:

- the potential for milkweeds to expand from the original planting site and encroach on adjacent land, and
- the chemical compounds present in milkweeds and their toxicity to livestock.

Although milkweed, the common name for plants in the genus *Asclepias*, implies that the plants are indeed weeds, milkweeds are a diverse group of native wildflowers with nearly 100 species in the U.S. that are not listed as noxious weeds at either the State or the federal level in the U.S. Milkweeds may be perceived as weeds because a few species will colonize disturbed areas and tend to reproduce vegetatively (in addition to reproduction by seed), sending up new shoots from roots that spread outward from the parent plant. This clonal reproduction allows their populations to expand over time, and plants may spread out of their original area. Common milkweed (*Asclepias syriaca*) exhibits the highest degree of clonal reproduction, and vegetative growth also occurs to a lesser degree in horsetail milkweed (*A. subverticillata*), narrowleaf milkweed (*A. fascicularis*), plains milkweed (*A. pumila*), prairie milkweed (*A. sullivantii*), showy milkweed (*A. speciosa*), and whorled milkweed (*A. verticillata*) (Borders and Lee-Mader 2014). Despite their vegetative growth, many of these species are unlikely to create an ongoing and unmanageable weed problem for roadside managers or adjacent landowners. However, the issue can also be avoided by selecting from the many milkweed species that do not reproduce clonally.

Another contributing factor to the perception of milkweeds as actual weeds is the presence of cardenolides, steroid plant compounds milkweeds use as a defense against herbivores. The amount of cardenolides present in plant tissue varies with the species of milkweed (it can also fluctuate seasonally) and can make the plants potentially toxic to livestock (Burrows and Tyrl 2013). Farmers and ranchers with livestock are often concerned about the presence and proximity of milkweeds to their stock. However, in properly managed rangeland and pasture, milkweed should pose no risk to livestock. Milkweeds are toxic only if consumed in large quantities, and milkweeds are highly unpalatable (Fulton 1972). Livestock, cattle in particular, will only consume milkweeds in the absence of other forage; milkweed can only poison a cow when it is in a pasture that is barren. Milkweed poisoning can be prevented by maintaining a sustainable stocking rate and by verifying that sources of hay are milkweed-free (milkweed retains its toxicity when dry and may increase in palatability).

Soil cover can potentially serve as a proxy for egg-laying sites for some pollinators. A monitoring procedure for measuring soil cover is presented in [Section 6.3.1](#). Visual scans of vegetation can be used to identify colonies of aphids, whiteflies, or other clusters of plant feeders that serve as egg-laying sites for pollinator species with predatory larval stages (e.g. flower flies).

### 3.9.3 NESTING HABITAT

Bees and predatory wasps provide for their young by constructing and provisioning nests in which their offspring develop. Bees provision their nests with pollen and nectar as food sources for their young, while predatory wasps supply their young with prey to consume. Some species of bees and predatory wasps nest underground, while others nest in tunnels and in cavities. Nesting is a critical factor affecting the ability of bees to persist within a site (Winfrey 2010; Menz et al 2011; Morandin and Kremen 2013b).

**Table 3-11 | Egg-laying sites for pollinators**

| Pollinator group      | Egg-laying sites  |
|-----------------------|---|
| Beetles               | In crevices, in the soil, on leaves, under plant debris or in leaf litter, on foliage near prey |
| Bees                  | In nests (See Section 3.9.3)  |
| Butterflies and moths | Host plants   |
| Flies                 | On foliage among prey, on larvae (for parasitic species), in leaf litter                        |
| Wasps                 | In nests (See Section 3.9.3)  |
| Bats                  | Breeding habitat unknown but bear live young in roosts such as caves, bridges, etc.             |
| Hummingbirds          | Eggs in nests, often in forks of branches of trees and shrubs                                   |

#### Soil as Pollinator Nesting Habitat

Most of North America's native bee species (about 70 percent) nest in the soil. Ground nesting bees provide for their young by excavating nests in the soil. Their offspring develop underground in cells (also known as brood chambers). Ground nesting bees need access to soil surfaces between vegetation to excavate and access their nests (Michener 2007). Some species will nest in a variety of soils while others have very specific requirements for the soil type, moisture, alkalinity, slope, and aspect (Cane 1991).



Many ground-nesting bees prefer to nest in sunny, bare patches of soil (Linsley 1958; Sardiñas and Kremen 2014). Such patches can be found under wildflower plantings where small areas of bare ground are exposed between plants (Figure 3-76A) or around the bases of native bunch grasses that tend to grow in dense bundles. Providing openings in scrub or forest habitat can promote ground-nesting bees (Figure 3-76B). Bunch grasses tend to provide better nesting habitat than sod-forming grass species, and roadsides with native bunch grasses have more nesting opportunities for ground-

**Figure 3-76 | Ground nesting bee nests**

The entrances of ground nesting bee nests often have small piles of excavated soil around them. Bees can nest in areas of bare ground (A) or amongst vegetation (B).

*Photo credits: Eric Lee-Mader/Xerces Society (A) and Matthew Shepherd/Xerces Society (B)*

### 3—PLANNING

nesting bees and, consequently, a greater abundance of ground-nesting bees (Hopwood 2008). Bunch grasses also allow establishment of other desirable species and prevents colonization from rhizomatous species. Undisturbed areas of bare ground can facilitate nesting activity (Potts et al 2005; Winfree et al 2009; Williams et al 2010; Roulston and Goodell 2011).

Assessment of patches of bare ground or patchy plant cover can be made through measurements of soil cover. A monitoring procedure for measuring soil cover is presented in [Section 6.3.1](#). Ideally, patches of non-erodible bare ground would be present and cover approximately 5 percent of the project site. If using thick mulch during establishment, it can benefit ground nesting bees and wasps if there are small patches of bare soil left without mulch. Another idea for creating a permanent area of bare soil is to have short sections of cut slopes or banks that are very steep and constantly raveling, preventing plants from establishing. These sections would need to be away from water so that soil would not erode into the stream system.

## Stems and Tunnels

A number of above-ground nesting bees nest in hollow stems or excavate pithy stems (e.g., elderberry or cane fruits) (Figure 3-77A). Many of these plants provide resources to other wildlife, such as berries (e.g., Salmonberry). Other native bees nest in tunnels in wood, such as abandoned beetle tunnels in logs, stumps, and snags (Michener 2007) (Figure 3-77B). Where site appropriate, planting native wildflowers with pithy stems, such as cupplant (*Silphium perfoliatum*), ironweeds (*Vernonia* spp.) and sunflowers (*Helianthus* spp.), along with shrubs such as wild rose (*Rosa* spp.), elderberry (*Sambucus* spp.), sumac (*Rhus* spp.), yucca (*Yucca* spp.) or agave (*Agave* spp.), will provide resources for stem-nesting bees.

**Figure 3-77 | Bee nest construction**

Bees can construct nests by excavating pithy stems (A) or in old beetle borer holes (B).

*Photo credits: Jennifer Hopwood/Xerces Society*



Assessment of potential tunnel nest sites can involve identifying woody plants (species and numbers) that support tunnel-nesting bees that can be planted at the project site. Additionally, because many tunnel nesting species create their nests in

tunnels originally built by other insects (e.g., beetles), in old wood, such as snags, or living tree trunks, it can be useful to quantify the amount of dead wood, snags, or wood with holes in it (e.g., fence posts) that already exist at the site or that can be established within the project site.

The best method for increasing nesting habitat for tunnel nesting bees (and predatory wasps with similar nesting habits) is to plant woody vegetation used by bees and to maintain snags or downed wood. A minimum of three species of woody plants or pithy stemmed plants is recommended, though a good rule-of-thumb is to include at least five species. The ERA tool provides information about plants that provide nesting resources for bees. Maintaining or importing dead wood, snags, or wood with holes in it (e.g., fence posts) within the project site will also benefit tunnel nesting bees.

## Cavities

Most native bees are solitary and nest either in the ground or in wood or pithy stems. Bumble bees, however, are social and have different nesting requirements. Bumble bees form social annual colonies founded by a single queen in the spring. A bumble bee colony can grow in size through the spring and summer as the bees work cooperatively to raise offspring and find food. At the end of summer, if the colony is healthy, new queens are produced. Queens find mates in the fall, their natal colonies die off, and queens find a place to overwinter, such as in leaf litter or under a shallow layer of soil. Bumble bee colonies need an insulated cavity in which to nest, such as underneath grass clumps (Svennson et al 2000), under the thatch of bunch grasses (Figure 3-78), or under rocks (Hatfield et al 2012). Bunch grasses have a tendency to “lodge” at maturity, meaning they create a gap between the grass and the soil surface that can provide an ideal nesting cavity for bumble bee colonies (Svennson et al 2000; Hatfield et al 2012). Mowing and grazing can have negative impacts on bumble bee colonies (Hatfield et al 2012). If a colony is destroyed via mowing mid-growing season, no queens will be produced, which can lower population levels the following year.

The nest entrances to bumble bee nests can sometimes be located in wood piles. In addition, wild bees may use holes in wood in woodpiles for nest entrances. Wood piles are also attractive to other wildlife, including birds. If non-native woody vegetation is being removed from a project site, leaving some of the wood piled at the edges of the habitat into one or more brush piles can provide nesting locations and also support populations of ground beetles.

Assessment of nesting habitat for bumble bees involves counting the number of different native bunch grass species present, calculating the relative area of the site bunch grasses cover, and noting if downed wood is present at the site.

If native bunch grasses are adapted to the site, it is best to have at least one native perennial bunch grass species present, covering at least 2 percent of site. Ideally there would be at least 2 different species in each project site. If woody vegetation is removed from the site, leave at least 1 wood or brush pile. It is also important to maintain areas of unmown bunch grasses throughout the growing season to support



**Figure 3-78 | Grass thatch is good nesting habitat**

There is not much to see from the surface but this grass thatch was home to a healthy colony of eastern bumble bees (*Bombus impatiens*).

*Photo credit: Jennifer Hopwood/Xerces Society*

bumble bee nests.

### 3.9.4 WATER SOURCES

Pollinators utilize water in multiple ways. Some directly consume water, while others use it to construct nests or for evaporative cooling. Pollinators can obtain water from edges of water sources, drops on vegetation, wet soil, puddles, or small pools on rocks.

#### Water, Mud, Minerals

Although pollinators typically obtain much of the water content they need by consuming nectar, water is also consumed directly on occasion or is collected by pollinators for hydration or evaporative cooling. On warm days it isn't unusual to see bumble bees gathered around the edges of puddles or perched on rocks in streams to sip water. Honey bees will collect water and transport it back to their hive in the heat of the summer to spread the water lightly over their brood and fan them with their wings to reduce the temperatures within the hive via evaporative cooling. Honey bees will also use water to dilute stores of crystallized honey within their hive. Water can also be used by ground nesting bees or wasps that carry it to the soil to moisten hard dry ground to make it easier to dig their nest tunnels.



**Figure 3-79 | Wasps build their nests from mud**

This female potter wasp is collecting mud in order to build her nest.

*Photo credit: Betsy Betros*

A number of wild native bees and wasps utilize mud in their nest construction (Figure 3-79). Blue orchard bees (*Osmia lignaria*) collect mud and use it to create nest chambers for their offspring. Blue orchard bees so rely on mud that the females will not nest in areas without a source. Potter wasps (*Hymenoptera: subfamily Eumeninae*) also use mud as nest partitions within an existing cavity, as well as free-standing mud structures. Mud-puddling is a behavior to gather liquid nutrients exhibited by butterflies and some other pollinators in which they congregate on wet soil to gather minerals and nutrients needed for their health.

Watersources for pollinators can include culvert outlets, ditches, draws, gullies, and intermittent streams. Water runoff from the edge of the pavement can also create a source of mud for pollinators.

It may not be practical to include a potential water source in every revegetation plan.

It also may not be a conservation priority for pollinators. As mobile animals,

Roadside Revegetation: An Integrated Approach to Establishing Native Plants and Pollinator Habitat

pollinators can fly to obtain the water or mud they need from surrounding land. However, projects that incorporate topographic enhancements into the roadside design will increase the potential for temporary surface water and mud sources ([Section 5.2.8](#)).

### 3.9.5 SHELTER AND OVERWINTERING

Like other animals, pollinators rely on various places that provide shelter during inclement weather. High winds, rain, cool temperatures, cloudy days, and other weather conditions all force pollinators to seek shelter. Pollinators are a diverse group of animals with a diverse set of needs for shelter and overwintering habitat (Table 3-12). Some rely on vegetation, such as the structure provided by grasses or the cover trees and shrubs provide. Others use rocks or crevices as shelter.

Pollinators also need a place in which to overwinter during the dormant season. Overwintering sites for most species are close at hand. Syrphid fly species and soldier beetles overwinter in roadside soil or litter (Schaffers et al 2012), and butterflies and moths also utilize roadsides as overwintering habitat (Schaffers et al 2012). However, a few species travel long distances to overwinter in warmer locations. Examples of migratory pollinators include monarch butterflies, painted lady butterflies, lesser long-nosed bats, and hummingbirds.

**Table 3-12 | Shelter and overwintering sites for pollinators**

| Pollinator group             | Egg-laying sites   |
|------------------------------|--|
| <b>Beetles</b>               | Larvae overwinter in loose soil or leaf litter Adults shelter under rocks, logs, brush   |
| <b>Bees</b>                  | Adults take shelter in bad weather under leaves on plants; overwintering of most species occurs. In nests ( <a href="#">Section 3.9.3</a> ), but queen bumble bees overwinter under shrubs or in shallow soil and leaf litter. |
| <b>Butterflies and moths</b> | Shelter in structure of vegetation; Overwinter in a protected site such as a tree, bush, tall grass, or a pile of leaves, sticks, or rocks   |
| <b>Flies</b>                 | Pupae and adults overwinter in soil or leaf litter   |
| <b>Wasps</b>                 | In nests ( <a href="#">Section 3.9.3</a> )   |
| <b>Nectar-feeding bats</b>   | Caves, mines, rock crevices, tree bark, under bridges and within bridge structures   |
| <b>Hummingbirds</b>          | Shelter in trees and shrubs; some are resident and some overwinter in southwest U.S., southern Mexico, and Central America   |

### Woody Vegetation and Grasses

Woody vegetation such as trees and shrubs can provide cover during the growing season that can serve as shelter for pollinators, and can also provide niches for overwintering. Some pollinators will overwinter under bark or in the soil just under shallow roots, or in piles of brush. Grasses can provide shelter for a variety of pollinators but notably for butterflies on roadsides (Saarinen et al 2005), and the root systems and grass thatch can also serve as overwintering habitat.

Assessment of shelter and overwintering habitat can involve identifying woody plants and grasses (species and numbers) that can be planted at the project site. It may be

### 3—PLANNING

useful to calculate the relative area of the site that grasses cover. Refer to the monitoring procedures for soil cover ([Section 6.3.1](#)) and species cover ([Section 6.3.2](#)) for additional information.

Including a diversity of types of plants in revegetation plans can help to ensure that vegetation structure that can act as shelter and overwintering habitat is present. Trees and shrubs may not be appropriate for every revegetation project; in those situations, including a diversity of grasses both cool and warm season grasses can increase vegetation structure.

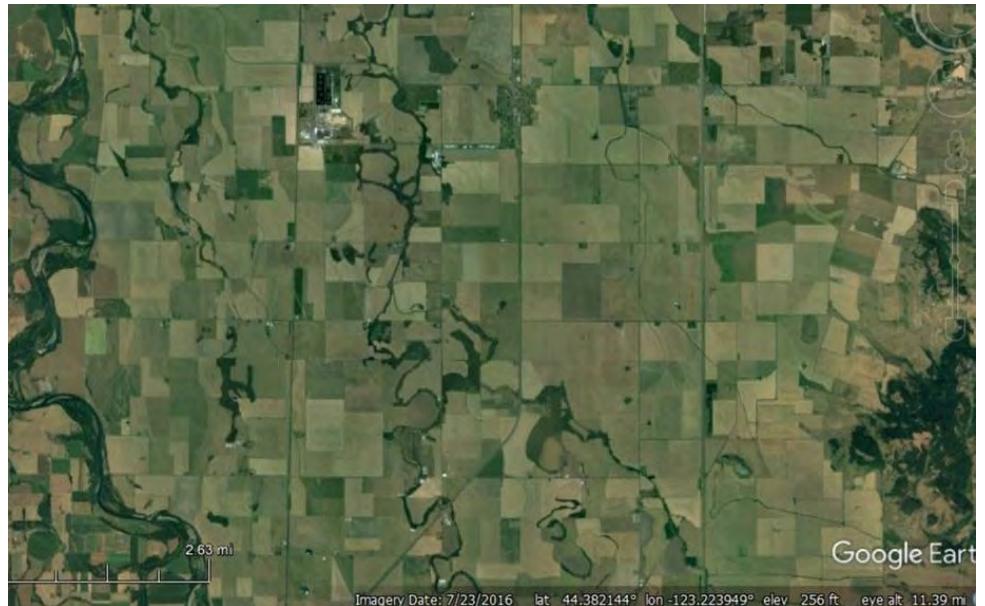
## 3.9.6 LANDSCAPE CONNECTIVITY

With habitat becoming increasingly fragmented (Saunders et al 1991), landscape connectivity is important for the populations of many species, including pollinators (Haddad 1999; Haddad and Baum 1999). Roadsides have the potential to act as corridors, strips of habitat or patches of habitat that serve as stepping stones that connect larger patches of habitat, facilitate movement of organisms between habitat fragments, aid in establishing or maintaining populations, and increase species diversity within isolated areas (Tewksbury et al 2002; Ottewell et al 2009).

Roadsides extend across a variety of landscapes and often contain greater plant diversity than adjacent lands. The linear shape and connectivity of roadsides may help pollinators move through the landscape (Soderstrom and Hedblom 2007), either for daily foraging or for dispersal between larger habitat patches.

Evidence suggests that pollinators use roadsides as corridors to facilitate movement through the landscape in search of food or in pursuit of new habitat (Lövei et al 1998; Ries et al 2001; Valtonen and Saarinen 2005; Hopwood et al 2010). Additionally, there is evidence of several pollinator species expanding their ranges along roadsides (Dirig and Cryan 1991; Brunzel et al 2004). Corridors like roadsides and other linear strips of vegetation may provide habitat resilience as changes in climate drive species range contractions or expansions.

It is important to consider the landscape context of the revegetation project. Developing a landscape connectivity map of the project area and adjacent lands, using information collected during the vegetation assessment ([Section 3.6.1](#)), that locates areas of high, medium, and low pollinator habitat qualities, can be a base for designing a revegetation plan that improves pollinator habitat within a larger landscape connectivity context (Figure 3-80). If resources are limited, it can be helpful to prioritize high floral diversity and cover and other factors that influence pollinators for projects that link or act as stepping stones



**Figure 3-80 / Roadsides can connect patches of habitat**

Landscapes with isolated or fragmented natural habitats make it difficult for pollinators to move from one natural area to another. The landscape shown in this photograph appears poor connectivity between natural areas (dark green riparian and forested areas) because of agricultural land use, however, the high road density creates opportunities to connect isolated natural areas and create migratory pathways by implementing roadside revegetation projects favorable to pollinators.

*Image from Google Earth*

between existing habitats on private, municipal, state, or federal lands. It is also important to prioritize those projects that can increase the connectivity of existing roadside habitat. If remnant roadside habitat exists, for example, it would be very beneficial if revegetation projects with high plant diversity were installed adjacent to or nearby.

### 3.9.7 ROAD MORTALITY

Roads can pose specific threats to pollinators. Roads can be a source of mortality for pollinators due to collisions with vehicles (e.g., Munguira and Thomas 1992). Roads may act as barriers to pollinator movement (e.g., Valtonen and Saarinen 2005). The prevalence of invasive and nonnative species on roadsides reduces pollinator abundance and diversity (e.g., Hopwood 2008). Finally, roadside vegetation is exposed to pollution from vehicles, which may impact pollinators (e.g., Jablonski et al 1995).

#### Vehicle Mortality

Collisions with vehicles are a source of mortality for pollinators that utilize resources in roadsides as well as pollinators travelling through the landscape. Mortality rates can be estimated by counting roadkill (McKenna et al 2001; Baxter-Gilbert et al 2016) but provide incomplete information. Estimates of the population sizes of pollinators using roadsides, in addition to counts of those killed by vehicles, provides a mortality rate that includes context for the proportion of the population impacted. Mortality rates of butterflies using roadside vegetation range from 0.6 percent to 7 percent of the population (Munguira and Thomas 1992) or 6.8 percent of the butterflies found on the roadside (Skorka et al 2013). Of the butterflies observed crossing roads, 2.8 percent were hit by vehicles (Ries et al 2001).

Rates of mortality due to collisions with vehicles are influenced by a number of factors. Some species of pollinators are more vulnerable to collisions than others due to their behavior or biology; species that are strong fliers, for example, appear to have lower rates of mortality than those that are not (Munguira and Thomas 1992; Ries et al 2001; Skorka et al 2013). Volume of traffic may influence rates of mortality (Skorka et al 2013); however, several studies have found that traffic volume does not consistently influence observed mortality (McKenna et al 2001; Saarinen et al 2005). The width of road or overall density of roads in the landscape may also influence butterfly response to roads. Wider roads may increase mortality rates (Skorka et al 2013) and some butterflies have decreased movement within a dense network of roads (Valtonen and Saarinen 2005).

Vegetation quality can also influence pollinator mortality: roadsides with more species of plants had fewer butterflies killed by traffic (Skorka et al 2013) (Figure 3-81). The frequency of mowing is also linked to a higher proportion of butterflies killed on roads; butterflies that had to disperse to find new habitat after roadsides were mowed may have had a greater likelihood of collisions with



**Figure 3-81 | Roadsides with high plant diversity have fewer butterflies killed by vehicles**

Research indicates that fewer butterflies are killed by vehicles when roadside vegetation has high plant diversity and floral cover.

*Photo credit: Maria Uribe/Iowa Living Roadway Trust Fund*

vehicles (Skórka et al 2013).

If quality roadside habitat is present, it may reduce the amount of pollinators killed by vehicles by providing pollinators with necessary habitat and less need to disperse elsewhere. It is important to consider increasing plant diversity of the roadside vegetation and reducing mowing beyond the clear zone (the strip of low growing or routinely mowed vegetation, or vegetation-free area, directly adjacent to the pavement) in order to reduce mortality rates due to vehicles. In areas with high traffic density, it may be helpful to increase the width of the clear zone, which is not typically used by pollinators as habitat, to increase the distance between pollinators foraging in the habitat and vehicles on the roadway. When developing the landscape connectivity map, it may also be helpful to prioritize locations for high quality revegetation projects with goals of supporting pollinators, focusing first on sites that are not in areas of high road density.

## Invasive Species

Many invasive and noxious plants can be present in roadsides (Tyser and Worley 1992; Gelbard and Belnap 2003) due to favorable conditions for plant introductions and invasions (Hansen and Clevenger 2005; Von der Lippe and Kowarik 2007). Nonnative plants can decrease the quality of roadside habitat for pollinators (Hopwood 2008; Valtonen et al 2006). Nonnative plants compete with native plants for resources as well as alter habitat composition, and some cause significant reductions in the abundance and diversity of pollinators and other herbivorous insects (Samways et al 1996; Kearns et al 1998; Spira 2001; Memmott and Wasser 2002; Zuefle et al 2008; Burghardt et al 2009; Tallamy and Shropshire 2009; Wu et al 2009; Hanula and Horn 2011; Fiedler et al 2012). There is also evidence that native pollinator insects prefer native plants (Burghardt et al 2009; Wu et al 2009; Williams et al 2011; Morandin and Kremen 2013a), even though many native insects will feed on nonnative plants when few natives are available (Zuefle et al 2008; Burghardt et al 2009; Wu et al 2009; Williams et al 2011).

To reduce invasive species and nonnative plants, it can be helpful to control weeds before and during construction ([Section 3.11.5](#)), as well as during the establishment phase following revegetation ([Section 3.11.6](#)) and maintaining a weed-resistant roadside following construction ([Section 3.11.4](#)). Salvage topsoil whenever possible, and source the imported project mulch, compost, and other inputs carefully, specifying testing certificates and/or certified weed free products where possible, in order to avoid introducing and spreading weed seeds. Additionally, purchasing seeds for the revegetation project within seed transfer zones can help to curb the spread of weed seed contamination of seed mixes.

## Roadside Contamination

Routine vehicle use and maintenance of roads contribute to roadside contamination by depositing pollutants, including vehicle exhaust and de-icing materials. Roadside soils and vegetation can be contaminated with heavy metals such as lead, iron, zinc, copper, cadmium, nickel, et al deposited from tire rubber, brake dust and gasoline and diesel combustion products (Gjessing et al 1984; Oberts 1986; Araratyan and Zakharyan 1988). Vehicle-de- rived contamination is proportional to traffic levels (Leharne et al 1992). In general, plant and soil contamination is greatest adjacent to the road and decreases with distance from the road (Quarles et al 1974; Dale and Freedman 1982; Jablonski et al 1995; Swaileh et al 2004). Contamination tends to decline within 20 meters but can still be present at high levels up to 200 meters from the road (Spellerberg 1998; Trombulak and Frissell 2000). Pollen and nectar contamination is also greatest nearest to the road (Jablonski et al 1995).

### 3—PLANNING

Heavy metals can be harmful to pollinators directly (Nieminen et. al 2001; Moroń et al 2010; Perugini et al 2011) or indirectly by weakening vegetation (Mulder et al 2005). However, few studies have explicitly examined the impacts to pollinators of heavy metal exposure in roadsides. Ozone, nitrates, and other exhaust gases may also have an impact on roadside vegetation and pollinators. Ozone and nitrates can inhibit floral scent, which reduces a pollinator's ability to detect flowers and in turn may reduce reproductive output of both pollinators and plants (McFrederick et al 2008). Dry deposition of particles with nitrogen derived from fuel combustion can create a strip of "fertilized" soil along roadsides, particularly in more arid regions (Gade 2013).

De-icing salts used on roads alter roadside soil chemistry by increasing sodium levels in plant tissues significantly (Snell-Rood et al 2014) and can damage plants (Bogemans et al 1989), with probable indirect impacts on pollinators ([Section 3.11.9, see Deicing for Winter Safety](#)). Varying levels of sodium in butterfly host plants can affect development of caterpillars in both positive and negative ways. Sodium is an important micronutrient for butterflies and moderate levels of sodium can increase flight muscle and brain size of adults (Snell-Rood and other 2014). More data about the impacts of salts, heavy metals, and exhaust on pollinators is needed. Until that data is available, increasing the width of the clear zone, particularly in areas with high traffic and frequent use of road salts, may help reduce pollinator exposure to these contaminants.

#### 3.9.8 VEGETATION MANAGEMENT

The management of roadside vegetation can have a significant impact on pollinators. Mowing vegetation beyond the clear zone multiple times a growing season, for example, can cause direct mortality to pollinators in the egg or larval stages that cannot avoid the mower (Humbert et al 2010) (Figure 3-82), can deprive pollinators of sources of pollen and nectar, as well as host plants for caterpillars (Johst et al 2006) (Figure 3-83), and can destroy bumble bee colonies (Hatfield et al 2012). However, the timing and frequency of mowing can be adjusted to reduce the impacts to pollinators (e.g. Halbritter et al 2015). When designing revegetation plans, it is important to consult with the site's maintenance department. Maintenance departments have processes and practices in place for reasons that are not obvious to habitat designers, such as the timing of mowing. Discussions with maintenance departments may result in a willingness to alter maintenance practices that can facilitate pollinator habitat, but that determination cannot be made without the designer opening communication with roadside maintenance.

If maintenance practices cannot be adjusted in ways that support pollinators, there are other opportunities for pollinator habitat on land managed by DOTs that are not roadside rights of way. These areas include DOT "back 40" lots, stockpile lots, maintenance yards, under used DOT office land, special management areas, bicycle paths, scenic viewpoints or historic or geologic points of interest or roadside rest areas. Maintenance is often glad to put forthright uses to these underutilized areas. These areas also offer opportunities for collaboration with citizen groups because they are less exposed to the hazards of traffic and DOTs may be more willing to allow volunteer groups to install or manage pollinator gardens in these areas.



**Figure 3-82 / Mowing pattern can facilitate pollinator habitat**

Cutting the clear zone with well-defined edge looks groomed and provides safe run-off zone.

Photo credit: Magnus Bernhardt/ODOT

If one of the goals of the project is to support pollinators, it is important that the vegetation can be managed such that pollinators benefit. For additional information about vegetation management and strategies that can support pollinators, see [Chapter 7](#).

## 3.10 INVENTORY OF SITE RESOURCES

Most project sites contain resources that can be used to meet revegetation objectives. Identifying these potential resources early in the planning process is essential so that they are not inadvertently wasted. The more that local resources are used, the more cost-effective, efficient, and effective the revegetation efforts may be. Physical resources to inventory include topsoil, duff, litter, parent materials, woody materials, logs, plant materials (seeds, seedlings, and cuttings), large rocks, and water (seeps, springs, creeks). It is good practice to also consider intangibles, such as community cooperation and the local knowledge base.

### 3.10.1 TOPSOIL

One of the most important site resources for revegetation is topsoil. If considered early in the planning process, topsoil can be salvaged and reapplied to disturbed sites after construction. This is one of the best ways of increasing productivity on a disturbed site.

Topsoil is inventoried early in the planning process to evaluate topsoil quality and quantity, costs, and the feasibility of removal and storage. Topsoil recovery is an expensive operation requiring knowledge of basic soil attributes. For this reason, it is a good idea to conduct a soil survey or assessment of those locations that will be disturbed. An example of soil and site information commonly collected for topsoil recovery is shown in Table 3-13. The road in this example is planned through undisturbed forested lands. Soils data is collected every 50 meters (at road stations) due to the high variability of the soils in this area. Where soils are very uniform, distances between plots can be increased. Soil texture, rock fragments, and depth of the topsoil are measured in the field. At selected intervals, or on different soil types, a sample is collected for lab analysis.

During topsoil survey, note other site attributes that could affect the quality of topsoil, especially the locations of all noxious weeds. These weeds can be treated or removed prior to topsoil salvage or the weed-infested areas can be avoided to prevent the spread these weeds across the project area.

The outcome of the topsoil survey is a short report and map in the revegetation plan showing the areas and depth to salvage topsoil. The report discusses the fertility of the topsoil, how it should be stored, and whether the duff and litter are removed and stored separately. The report also identifies areas where topsoil should not be collected, such as areas of noxious weeds or high rock. The volume of topsoil can be calculated based on soil depth and area of the road prism.



**Figure 3-83 | Mowing can affect food sources**

If mowing occurs too frequently, plants will not be able to flower and pollinators will have fewer sources of food.

*Photo credit: Jennifer Hopwood/Xerces Society*

**Table 3-13 | Example of a form for collecting topsoil information**

|                           | Plot number                  |                              |                  |                  |                  |
|---------------------------|------------------------------|------------------------------|------------------|------------------|------------------|
|                           | C1                           | C2                           | C3               | C4               | C5               |
| <b>Location</b>           | 1 + 300                      | 1 + 350                      | 1 + 400          | 1 + 450          | 1 + 500          |
| <b>Site Condition</b>     | Undisturbed                  | Undisturbed                  | Undisturbed      | Undisturbed      | Undisturbed      |
| <b>% Slope</b>            | 30                           | 45                           | 35               | 45               | 40               |
| <b>Aspect</b>             | N                            | N                            | NE               | S                | S                |
| <b>Topsoil depth</b>      | 12"                          | 14"                          | 14"              | 6"               | 6"               |
| <b>Topsoil texture</b>    | Loam                         | Loam                         | Loam             | Loam             | Loam             |
| <b>Topsoil % rock</b>     | 30                           | 25                           | 20               | 40               | 45               |
| <b>Subsoil texture</b>    | Sandy loam                   | Sandy loam                   | Sandy loam       | Clay loam        | Clay loam        |
| <b>Subsoil % rock</b>     | 40                           | 35                           | 45               | 45               | 35               |
| <b>Total soil depth</b>   | > 60"                        | > 60"                        | > 60"            | > 40"            | > 40"            |
| <b>% Soil cover</b>       | 100                          | 100                          | 100              | 100              | 100              |
| <b>Soil surface cover</b> | Litter, duff                 | Litter, duff                 | Litter, duff     | Litter, duff     | Litter, duff     |
| <b>Depth of cover</b>     | 1"                           | 1"                           | 2"               | 0.25"            | 0.25"            |
| <b>Parent material</b>    | Granite                      | Granite                      | Granite          | Basalt           | Basalt           |
| <b>Fracturing</b>         | Massive with some fracturing | Massive with some fracturing | Highly fractured | Highly fractured | Highly fractured |
| <b>Sample depth</b>       | 0-14"                        | 0-14"                        | 0-14"            | 0-14"            | 0-14"            |

### 3.10.2 DUFF AND LITTER

Duff and litter are the dead plant materials that have accumulated on the surface of the soil. The level of decomposition differentiates litter from duff. Litter is the layer of recently fallen, undecomposed leaves, needles, and branches; duff (which occurs immediately below the litter layer) is litter that is decomposed beyond recognition. The duff layer is a dark, light-weight organic layer. It is a large reserve of nutrients and carbon, and has a high water-hold capacity. Litter and duff layers protect the soil from erosion by absorbing the energy of rainfall impact and reducing overland flow. Combined, the litter and duff layers can be very thick, ranging from 1 to 4 inches depending on the productivity and climate of the site.

Litter layers typically have viable seeds originating from the overstory vegetation. Under the right conditions, these seeds will germinate. If collected, stored and reapplied correctly, this natural seed bank can be used as a seed source. Litter and duff layers can also contain mycorrhizal inoculum.

Litter and duff layers can be assessed concurrently with topsoil surveys by measuring their depths using a ruler. Refer to [Section 5.2.3 \(see Litter and Duff\)](#), for a discussion of methods for collection and application.

### 3.10.3 SUBSOIL AND PARENT MATERIAL

Certain subsoils and parent materials can be salvaged during road construction and used to produce manufactured topsoil ([Section 5.2.4](#), see [Manufactured Topsoil](#)). Textures low in rock content, including sandy loams, silt loams, loam, and sandy clay loams, are often good materials for manufactured soils. These are often found in areas where the parent materials are derived from alluvial or windblown deposits. They include river sands, pumice, volcanic sands, and loess.

### 3.10.4 WOODY MATERIAL

Woody material consists of live and dead plant materials that are cleared in the early stages of road construction. This material includes tree boles, root wads, bark, and branches. During clearing and grubbing, these materials are often concentrated in piles and burned. All of these materials can be ground up as shredded wood and used as surface mulches ([Figure 3-84](#)) ([Section 5.2.3](#), see [Shredded Wood](#)) or soil amendments ([Section 5.2.5](#)). Large wood can be used for biotechnical engineering structures, obstacles for erosion control, or placed upright or on the ground for pollinator habitat and site productivity. Substituting shredded wood derived from these materials for composts and soil amendments can lower overall project costs.

**Figure 3-84 / Creating shredded wood for mulch**

Woody material from road clearing can be ground up into shredded wood and used as a mulch or soil amendment.

*Photo credit: David Steinfeld*



Suitable plant materials are often destroyed during road construction. These consist of seeds ([Section 5.3.1](#)), plants ([Section 5.3.3](#)), and cuttings ([Section 5.3.2](#)). If these materials are collected from the construction site prior to disturbance and stored correctly, they can be used, instead of propagated or purchased plant materials, to revegetate the project. Surveys of the project site prior to construction will indicate the location and abundance of appropriate plant materials.

## 3.11 DEVELOPING A VEGETATION MANAGEMENT STRATEGY DURING PROJECT DESIGN

### 3.11.1 INTRODUCTION

During the planning phase, it is important to consider how vegetation will be maintained after the road project is completed. The effects of road surface and roadside vegetation management can have unexpected and often, unwanted effects on the long-term outcome of the revegetation project. Developing a written maintenance strategy can assure that there will be a rational documented approach to the management of roadside vegetation long after the project is completed and can be used to gain acceptance of the strategy from the maintenance department.

The intent of a maintenance strategy is to consider how the results of a revegetation project will affect the management and maintenance of the roadside and to incorporate this understanding into the revegetation plan. The strategy also anticipates and mitigates for those biotic and abiotic factors that may affect the development of native plant communities and pollinator habitats. Ideally, the planning team or designer meets with local roadside maintenance personnel to discuss their current and future anticipated maintenance procedures in order to learn what problems can be expected in reestablishing roadsides with native plants and how these problems could be addressed. This is a time when maintenance personnel can raise concerns and then work collaboratively with the design team to develop solutions. The planning phase is a good opportunity to develop a plan that maintenance personnel will understand and support.

The intended audience for this section is the designer and design team because they will integrate road maintenance and operations into how to achieve the long-term revegetation objectives during the planning process. Chapter 7 also covers vegetation management but from a maintenance perspective and the intended audience for that chapter is maintenance and operations staff. Chapter 7 focuses on approaches to road surface and roadside maintenance that will meet revegetation objectives after the revegetation project is completed.

A vegetation maintenance strategy may cover some, or all of the following:

- Protection of areas currently free of invasive species
- An outline of an integrated vegetation management approach to weed control
- How existing weeds may be controlled prior and during construction
- How a weed-resistant road environment will be created
- How roadside vegetation maintenance objectives will be achieved after construction
- How the vegetation maintenance strategy is handed off to maintenance personnel
- How roadside disturbances will be treated during the maintenance phase

### 3.11.2 INTEGRATING ROAD MAINTENANCE OBJECTIVES INTO THE REVEGETATION PLAN

For a successful project, it is important that road surface and roadside maintenance objectives align with treatments and species outlined in the revegetation plan. Many states have developed IRVM (Integrated Roadside Vegetation Management) Plans that outline roadside maintenance objectives. When available, it is strongly

recommended that the designer refer to these individual plans while developing the revegetation plan. The IRVM plan is “an approach to right-of-way maintenance that combines an array of management techniques with sound ecological principles to establish and maintain safe, healthy and functional roadsides” (Brant et al 2015). It applies many of the Integrated Pest Management concepts developed for agriculture, horticulture, and forestry to roadside vegetation management. The IRVM elements include prevention, monitoring, action thresholds, pest treatments, and evaluation (NRVMA 1997). Most of these plans are available on the internet or by contacting the state DOT or the local maintenance agency.

A vegetation management strategy considers how road maintenance objectives are integrated into plant species selection, planting patterns, vegetation control, invasive species control, and site treatments. Typical road objectives are stated below, along with their possible effects on roadside vegetation. In the development of the vegetation management strategy, consider how each objective will affect vegetation and, in turn, how vegetation treatments and design will affect roadside objectives.

- **Maintain line of sight**—It is important to not plant masses of tall grasses, shrubs or trees in areas where line of sight is important. If they are existing to remain on a site that is being revegetated, they may need to be maintained or removed and replaced with appropriate smaller plant material so as not to reduce line of sight. Designing for safety is further discussed in [Section 3.11.7](#).
- **Maintain clear zones**—The FHWA (2017) defines the clear zone as “an unobstructed, traversable roadside area designed to enable driver to stop safely or regain control of a vehicle that has accidentally left the roadway ... and an effective strategy for prevention and mitigation of roadway departure crashes.” The publication further states that “trees are the single most commonly struck objects in serious roadside collisions. Therefore, it is important to integrate the selection of appropriate plant species with safety objectives in mind.” The integration of habitat features such as brush piles or dead wood are to be placed well beyond the clear zone.
- **Maintain road surface**—Encroaching vegetation can reduce the longevity of road surfaces. Selecting plant species that are least likely to do this can preserve the longevity of roads.
- **Reduce fire hazards**—One of the reasons for maintaining low growing roadside vegetation is to reduce the starting of fires by motorists and the spread of wildfires. In planning, choosing plants that are more fire resistant and encouraging plants that stay green in the summer (e.g. native perennials) by creating good optimum growing environment (e.g. good soil conditions). Mowing, herbicides, grazing, and fire are also measures that may affect pollinator habitat and need to be planned accordingly ([Section 7.5](#)).
- **Maintain stable roadside**—Cuts, fills, and drainage facilities need to be maintained so that they are stable. When they are not, they create road hazards and degrade water quality. Unstable material will need to be removed and replaced. The removal area and replacement material will need to be reviewed and revegetated. Areas that fail will also need to be repaired and revegetated. It is important that the maintenance provider have the native-sourced seeds or plants for the waste or disturbed areas ([Section 7.2.2](#)).
- **Maintain or increase water quality**—Maintaining water flow off pavements, down ditches, and into natural waterways is important for road maintenance but may affect water quality or plant growth. Cleaning out ditches affects plants and cut slopes. Poor flow of water off pavements can cause rill and gullies on fill slopes affecting plants. Poorly designed culvert placement can cause downcutting

of natural drainage channels.

- **Reduce or eliminate invasive species**—Controlling invasive species may also negatively affect non-target plants and pollinator species and need to be considered when developing treatments. ([Section 7.5](#)).
- **Maintain safe road surfaces during winter months**—Winter roadway surface treatments vary throughout the country. Gravels used in mountainous, heavy snowpack areas, coarse sand used in smaller urban areas, and the more prevalent deicing chemicals can be the most devastating treatments on roadside vegetation. Sweeping gravels off pavement during melt periods buries roadside plants. Reusing gravels from along roadsides can spread weeds. Blowing snows can create drifts that melt later in the year and may call for different plant species in those areas to withstand that condition. Sand accumulations in swales and drainage structures can create clogs, ponding, and spreading sand bars which can smother groundcover over time. Deicing with salts and other chemicals can kill or reduce growth on certain plant species. This is further discussed in [Section 3.11.9](#) (see [Deicing for Winter Safety](#)).
- **Reduce impacts of wildlife to motor safety**—The selection of plant species and where they are planted along roadsides can affect how wildlife moves through a road corridor and their potential hazards to motorists. Planning for wildlife is an important consideration when selecting plant species and planting patterns. Providing desirable browse or a refuge of thick vegetation on both sides of an identified animal crossing can draw wildlife to the safe crossing location. Existing vegetation near known unsafe animal crossing locations may be controlled through removal, thinning, and mowing to reduce cover and forage opportunities, and to increase visibility in an attempt to reduce animal-vehicle collisions. Vegetation types, patterns, and maintenance can encourage the movement of wildlife to crossings. Wildlife fencing that direct animals to grade separated crossings are very successful in reducing wildlife-vehicle collisions (WVC). Consider planning the fencing with wildlife biologists who are familiar with wildlife passage patterns.
- **Protecting utilities**—Encroaching vegetation can overgrow and hide utility structures, which may lead to accidental damage during maintenance procedures, may affect maintenance access, and may cause maintenance review to be missed. Utility service may also be damaged and disrupted by accumulation of heavy vines, wind-blown branches, and by falling trees or branches. It is important to that species that may interfere with utility access and protection are not planted near these structures.

The vegetation management strategy will ideally span the entire length of the project, into the operation and maintenance phase ([Chapter 7](#)). In the early stages of planning, weed sources are identified and treatments to control or eliminate often begin prior to construction ([Section 3.11.6](#)). Areas of functioning plant communities, relative free of invasive species are identified and plans for protection are developed ([Section 3.11.3](#)). During planning, road maintenance objectives are integrated into the revegetation treatments. As the project moves into construction, treatments to create a weed-resistant environment ([Section 3.11.4](#)) and measures to reduce the introduction of weed sources are implemented ([Section 3.11.5](#)). Once construction is completed, the project is handed off to the agency responsible for road maintenance. At this point, the responsible agency implements their vegetation maintenance plan which dovetails with the revegetation plan objectives ([Chapter 7](#)).

### 3.11.3 PROTECTING HEALTHY PLANT COMMUNITIES

Mature forests, prairies, and wetlands are often high quality, healthy, and diverse landscapes supportive of a wide range of native plant, pollinator, and wildlife communities. These healthy plant communities are typically more resistant to invasive plant and animal species, and contain flora and fauna capable of supporting high level predators, particularly in larger areas. For the purposes of this report, landscapes of healthy plant communities capable of providing quality habitat for large animals, are referred to as 'refugia.'

Many of the larger blocks of refugia habitat have been preserved by local, State, and National parks, but remnants remain on undeveloped public and private land across the country that are essential for the preservation of native flora and fauna. Migrating animals and those that require large ranges depend on refugia and safe connections between refugia areas.

Revegetated roadsides can offer safe connections for wildlife to under and over-crossings and to other areas of refugia.

Despite best efforts to prevent impacts to environmental assets during the planning and construction of roadway expansions, development of roadway systems often causes disturbances to refugia. Introduction of road rights-of-way near or through these natural areas often results in a slow degradation of the landscape and wildlife population through the removal of habitat, altering or cutting off of wildlife corridors, and introduction of construction and traffic noise, pollution, deicing chemicals, and invasive plant and animal species.

Roadside revegetation treatments can be designed to provide a measure of protection for uninfested areas through buffering of on-going pollutions and disturbances and by preventing encroachment of invasive plants and annual weeds. Further protections of uninfested areas can be achieved through the processes listed below and described in more detail in following subsections:

- 3.11.4 Creating a Weed-Resistant Roadside Environment
- 3.11.5 Keeping Weed Sources from Entering the Project
- 3.11.6 Controlling Unwanted Vegetation
- 3.11.7 Designing for Safety and Utility Protection
- 3.11.8 Designing to Isolate Wildlife from Vehicles
- 3.11.9 Designing for Disturbances

### 3.11.4 CREATING A WEED-RESISTANT ROADSIDE ENVIRONMENT

The foundation of long-term weed control is to create a growing environment that encourages the development of a healthy native plant community resistant to the introduction and spread of weeds. For most weeds to become established, there usually is first an opening, or space, in the native plant community for plants to grow. These conditions occur in disturbed areas associated with recent construction, landslides, bladed ditches, gullies, waste areas and other areas where native plants and soil have been removed or disrupted. Secondly, the disturbed environment is usually favorable for weeds to take hold and thrive (e.g., optimal soil and climate for the weed species).. Third, the weed species has an advantage over native species. If a weed is more resilient or robust in its growth habits in both adjacent lands and disturbed sites, then it is more likely to become established.

Some mitigating measures that can be taken to limit the environment where weeds can gain a foothold are:

- **Minimize ground disturbance**—Most weed seeds need bare soil to germinate and become established. Reducing the footprint of the project results in less area being disturbed and exposed to possible weed invasion. Avoid ill-timed blading, mowing, dredging, and other ground-disturbing activities (Harper-Lore et al 2013).
- **Develop optimum environments for quick native plant recovery**—Establishing a healthy native plant community after construction greatly reduces the possibility for weed invasion. Optimum native plant establishment is dependent on the site’s soils and climate. Site factors restricting the downward root growth of native perennial plant roots (e.g., compacted soil) give the advantage to weed species. For example, a site with loose, reapplied topsoil is much more likely to support a native plant community than a compacted site lacking topsoil and organic matter. Unlike perennial plants, the roots of most annual weed species occur in the surface layer (Jackson et al 1988; Claassen et al 1995) and do not require deeper soils to survive and grow. Annual species therefore have an advantage over perennials on compacted or shallow soils. Developing site treatments that create an optimum environment for native perennial plant recovery will limit the establishment and spread of weeds (Sheley 2005).
- **Develop a weed-resistant seed mix**—Where appropriate, apply a seed mix that resists weed establishment. Using a seed mix composed of a diversity of species can reduce the potential for weed establishment by creating a plant community that fills niches that otherwise would have been occupied by weeds. Sowing native seeds at high rates can also flood bare soil with the seeds of desirable species, reducing the germination sites where weed species can become established. The objective of this strategy is to fill the openings first with desirable species to crowd out unwanted species ([Section 5.4.1](#)).
- **Establish desirable non-native plants if establishment of natives is not feasible**—Due to surrounding site limitations, project parameters, timelines, or other circumstances, using locally adapted native plant materials may not always be possible. For example, if the project is surrounded largely by contiguous populations of cultivars, using locally adapted plant materials may not be worth the effort. If the surrounding area is primarily native species, careful selection of cultivars is important to consider. Many cultivars, both native and introduced, have the potential to cross with their locally adapted native equivalent genera ([Section 3.13.2](#)).
- **Avoid applying nitrogen fertilizers in the first year**—Many annual weeds out-compete native plants on soils that are high in nitrogen because they are more adapted to utilizing nitrogen during early plant establishment. High rates of nitrogen fertilizers can encourage the growth of weedy annuals at the expense of the establishment and growth of native perennials (McLendon and Redente 1992; Claassen and Marler 1998), whereas perennial grasses have a competitive advantage at lower nitrogen levels (Welker et al 1991). Application rates of up to 108 lb/ac N (121 kg/ha N) have been shown to promote the establishment of introduced grasses over less competitive native grasses (DePuit and Coenenberg 1979). High rates of nitrogen fertilizers can also affect revegetation efforts by decreasing species richness and increasing the presence of non-native species (Munshower 1994; Wedin and Tilman 1996). When nitrogen fertilizer is applied at seeding, root systems of native perennials establish in the upper portion of the soil. The decrease in deep roots gives the annual weed species a competitive growth advantage (Claassen et al 1995). Limiting the amount of nitrogen fertilizer used the first year (during vegetation establishment) will help force the roots of perennials deeper into the soil where

there is more moisture. The deeper root system increases the competitive advantage of the native perennials over the annual weeds. This strategy, however, does not preclude the need for fertilization. One option is not to apply fertilizer until a year after sowing when the native plants have become established (Section 5.2.1), though this may not be possible due to construction schedules and budget constraints.

- **Apply mulch**—Mulch can be applied to the surface of a disturbed soil to create a poor germination environment for weed seeds (Section 5.2.3). Quality mulch often has high void spaces (typical of long-fiber mulch materials) and low water-holding capacity, which is an unfavorable environment for seed germination. The deeper the mulch, the less weeds will become established. Depending on the weed species, 2 to 4 inches of mulch are necessary to effectively control the establishment of many weed species (Pellett and Heleba 1995; Ozores-Hampton 1998; Ozores-Hampton et al 2002; Penny and Neal 2003).
- **Apply high C:N materials to high nitrogen sites**—If soils are high in nitrogen, applying a high C:N material (e.g., wood products, coir mulch, or pine straw) to the surface of the soil, or mixed into the soil, is another strategy to reduce the amount of available nitrogen for annual weed growth (REAP 1991; St John 1999). Over time, the organic material will break down and release nitrogen for uptake by perennials.
- **Retain shade**—Most weeds need full or partial sunlight to thrive (Penny and Neal 2003). To reduce the vigor of undesirable plants, retain as much shade as possible.

### 3.11.5 KEEPING WEED SOURCES FROM ENTERING THE PROJECT

Weed species are often brought onto the site from outside sources such as construction equipment, mowing equipment, cars and trucks, shoes and socks, and in materials used on the project such as gravel and rock. All too often, weeds arrive during the revegetation efforts in contaminated mulches, topsoil, hydroseeding equipment, and uncertified seed sources. Making the effort to prevent the introduction of weed propagules onto the project is always the preferred strategy because it is easier and more economical to prevent the introduction than to control or eliminate weeds once they have become established. There are many possible entry points for weeds:

- **Vehicles and equipment**—Weeds seeds and plant parts can arrive during construction on vehicles and equipment. Portable wash stations are often set up at staging areas and/or designated site entrance/exit points to thoroughly clean tires, wheel wells, and chassis of vehicles and equipment to reduce the possibility that weed seeds are brought in from other projects or areas or transferred within a project area.
- **Erosion control seeding**—Rye grasses are non-native and commonly used for temporary or permanent erosion control seeding. Seeds of these species are also found in material used for wattles. These grasses persist in the landscape and are difficult to eradicate once introduced. Unless these species are required by jurisdiction on the project, other species, such as native grasses used in conjunction with sterile hybrid grass seeds would satisfy erosion control needs and offer greater benefits to the environment. Erosion control matting works well to block the establishment of weeds while preventing erosion and facilitating the growth native plants.
- **Hydroseeding tanks, range drills, and other seed delivery systems**—Seed delivery systems, such as hydroseeding tanks and range drills, brought in from other projects, can contain plant species that are not wanted on the project. A

Roadside Revegetation: An Integrated Approach to Establishing Native Plants and Pollinator Habitat



**Figure 3-85 | Hay often contains weed seeds**

Hay bales often contain seeds from unwanted species that will germinate once the hay is spread. Some states have certified “weed-free” programs, however, “weed-free” does not mean the hay will be seedless.

*Photo credit: David Steinfeld*

thorough cleaning and inspection of this equipment are essential to eliminate the potential introduction of unwanted seeds. It is recommended that hydroseeding tanks are washed out and range drills air-blown before this equipment is brought on the project site.

- **Seed sources**—Commercial grass and forb seed sources, whether native or non-native, can contain weed seeds that were harvested along with the native seeds. The quantity of this material is dependent on the weed control practices and seed cleaning technology implemented by the seed producer. A good means of eliminating the possibility of contamination of native seeds with weed seeds, is to ask for seed tests of the seed being considered for purchasing. The purity seed test identifies the contaminants in a seed lot, including weed seeds, other plant seeds, and inert material. It is important to ask for tests that determine the presence of state-listed noxious weeds and other crop species (check state and federal lists to determine if any local weed species should be added to the testing list). Testing is conducted through certified seed labs per the standards of the Association of Official Seed Analysts. If the seed is not cleaned and retested, the seed lot can be rejected..
- **Mulches**—Buy mulch that is free of weed seeds. Hay and straw mulches are of special concern (Figure 3-85). Some states certify hay or straw as “weed-free” which means that the material is free of noxious weeds but not necessarily free of all seeds. Some straw comes from the stubble left after a seed harvest of native and non-native grasses, which often includes unharvested seeds. These seeds are viable and will establish into plants when the straw is applied. A site visit or discussion with the vendor prior to purchase is a good way to assess if there are seeds of unwanted species in the material.
- **Compost sources**—Compost sources are not always free of weeds, especially if the materials were not composted properly. Compost has to reach lethal temperatures and remain there long enough for plant seeds to be rendered nonviable. Fresh, moist compost piles, where temperatures are maintained between 140 to 160 degrees F for at least several days, will kill most pathogens and weed seeds (Epstein 1997; Daugovish et al 2006). When obtaining compost, it is important to ensure that the supplier complies with standards that meet the time-temperature requirements to ensure destruction of weed seeds. Visiting the mulch sources or testing mulch for weed seeds may also be appropriate prior to purchase ([Section 5.2.5](#)).
- **Gravel, sand, and rock sources**—Prior to acquiring road building materials such as sand, gravel, and rock, determine if this material comes from a source that is free of undesirable weeds. A plant survey of the source area will identify whether the material is suitable.
- **Haul routes and waste areas**—Haul routes and waste areas can have weeds that may be transported around the project on vehicles and equipment. It is recommended that these areas be treated prior to construction or avoided during construction to reduce this risk.
- **Salvaged and purchased topsoil**—During the survey for salvaging topsoil, areas of weed populations are also typically identified ([Section 5.2.4](#)). To reduce the possibility that these weed populations are spread when topsoil is salvaged and reapplied, weed populations are avoided during topsoil excavation. Salvaged topsoil is stored in a manner that limits weeds from becoming established. For topsoil that is being considered for purchase, prior inspection of the piles will assure that the soil is free of weeds (Figure 3-86).

### 3—PLANNING

- **Sand, gravel, rock, or topsoil storage**—Inspect areas where topsoil, gravel, rock, sand, or other materials to be used on the construction project will be stockpiled. If there are weeds growing nearby, it is very likely that seeds will end up on these piles, especially if the piles are to be stored for over a year. Good practices are to remove these populations prior to stockpiling materials and to require stockpiles be maintained free of weeds.



**Figure 3-86 | Quality topsoil is low in weed seeds**

Know the origin and quality of the topsoil. Topsoil sources contain seeds of the species that grew on them prior to salvage. In this picture, the topsoil pile on the right (B) was salvaged from a nearby pasture and the pile on the left (A) from an undisturbed native forest site. The pasture topsoil pile revegetated quickly (within 3 months after stockpiling) because of the abundance of non-native seeds in the soil. The forest topsoil revegetated slower because there were less seeds. Application of the pasture topsoil (B) resulted in a site dominated by introduced pasture plant species.

*Photo credit: David Steinfeld*

- **Hedgerows and windbreaks**—Planting windbreaks with non-native species, such as autumn olive, privet, honeysuckle, buckthorn, and multiflora rose, introduces fast-growing, highly competitive species (Harper-Lore et al 2013). Use of native species is preferable.
- **Livestock grazing**—Cattle can bring weed seeds from offsite where they have been grazing. Until native vegetation has become established, it is important to keep livestock out of the area.

#### 3.11.6 CONTROLLING UNWANTED VEGETATION

##### Stratifying the Project by Weed Status

Locating weed populations early in the planning phase will help in developing a successful vegetation management strategy. During the initial vegetation assessment (Section 3.6.1) and topsoil surveys (Section 3.10.1), weed populations are typically identified and mapped to produce a weed status map.

The weed status map is typically composed of these 4 mapping units:

- **State-listed noxious weeds**—Areas where state-listed weeds are present and regulations require their control.
- **Species of concern**—Areas where species of concern are present. These are plants that are known to occur within the area, are not regulated for treatment by the State or County, but cause concern due to their population density, life strategies that provide them with a competitive advantage over desired native plants, dense growth in waterways, affinity for habitats specific to those required by desired specialist plant species, toxicity or ability to cause injury to

livestock, etc. If the resources are available, the implementation of a new project can often provide a great opportunity to treat these species.

- **Weed-free, non-functioning plant communities**—These are areas where there is no state listed noxious weeds or species of concern present, but the plant communities are composed of exotic species, monocultures, poor pollinator species, or have depauperate plant communities with a large proportion of bare ground.
- **Weed-free, functioning plant communities**—Areas where a suite of native plant species is present, soils are intact, and there are minimal noxious weeds or species of concern present. These are optimum habitats for pollinators and larger areas may be wildlife refugia as well ([Section 3.11.3](#)).

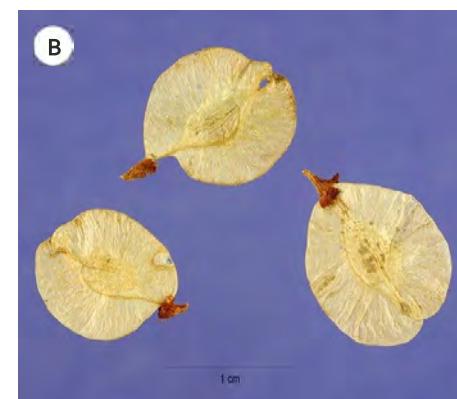
## Understanding the Life History

Understanding the life history and ecology of the regulated noxious weeds and species of concern of the project area is important in developing a weed control strategy. There are several resources to learn about specific noxious weeds or invasive species. The [USDA PLANTS](#) database describes each weed species and has PDF documents called Plant Guides that detail the life history of the weed and how it can be controlled. Another source of information is the [Invasive Species Assessment Protocol](#) (I-Rank) website and covers many of the nonnative plants in the United States. This website describes some of the important characteristics of the weed species and ranks each on its threat to native plant communities and its difficulty to control. Because these websites are frequently updated, it is important to check the most current lists. Local County Weed Boards and county and State road maintenance staff are also good sources of weed information, as they track and treat specific weed species, often within the project area.

The important information to have about each weed species in the project area includes:

- Life form
- How it reproduces (i.e. vegetatively, by seed, both)
- Viability and longevity of propagule
- Reproductive period
- Mechanism and distance of propagule dispersal (i.e., wind, animal/bird/insect, explosive seed capsules)
- Life/growth strategy
- Life cycle
- Limiting factors to establishment
- Importance to pollinators
- Treatment options

Based on the ecological requirements and life history information, weeds can be grouped into two treatment groups: (1) species that are treated prior to, and during, construction ([Section 3.11.6](#), see [Weed Control during Pre-Construction and Construction Activities](#)) and (2) species that are treated after construction ([3.11.6](#), see [Post-Construction Weed Control and Vegetation Management](#)).



**Figure 3-87 / Examples of plant seeds adapted for wind or gravity dispersal**

Yellow salsify (*Tragopogon dubius*) (A) is just one example of many plants within the Aster family that develop white, hairy pappus to facilitate airborne flight of seeds on the wind. The Siberian elm tree (*Ulmus pumila*) (B) develops a papery membrane around its seeds to assist their dispersal by gravity and wind.

Photo credits: (A) Jane Shelby Richardson, (B) Steve

## Weed Control during Pre-Construction and Construction Activities

Construction activities often occur during the time that seeds are ripening or beginning to disperse, which increases the potential that unwanted seeds will be scattered throughout the project area. To effectively treat some state or county listed weeds and species of concern, it is often optimal to treat prior to, and during, construction. These species include:

**Species with wide seed dispersal**—These are species that spread seeds long distances, which can be achieved by a number of methods. Some plants have structures that forcibly eject seeds when disturbed or at maturity employ ballistic dispersal (e.g., brooms [*Cytisis* spp.], bittercress (*Cardamine* spp.), jewelweeds (*Impatiens* spp.]). Ballistic seed dispersal can often provide propagation advantage to shorter plant species.

Additional seed dispersal mechanisms that allow transport are gravity and wind dispersal (Figure 3-87). In general, the taller the plant, the further its seeds are dispersed. Plants that utilize wind dispersal frequently have appendages on their seeds (more often this would be fruits) to aid movement via wind. Yellow salsify (*Tragopogon dubius*) is an example in the aster family that employs wind dispersal, while the elm (*Ulmus pumila*) is a well-known weedy tree species in many areas that utilizes gravity and wind dispersal.

Bird and insect assisted dispersal is another strategy that facilitates movement of plant propagules across considerable distances. Seeds or fruits may stick to the feet or body of an animal and be transported as they walk or fly. Seeds can also stick to the feet of humans, including restoration designers and implementers. Animals can consume the seeds of plants, depositing them at new locations once the seeds pass through their digestive systems. Animals and insects often actively move seeds or vegetative material from one location to another to utilize them as an immediate or stored source of food or nesting material. Humans move the seeds of many of these species when walking and driving as well.

Any non-native plant species that disperse their propagules substantial distances from their source are ones for which pre-construction treatment could be considered.

**Vine species**—Species that form vines [e.g., English ivy (*Hedera helix*) and kudzu (*Pueraria montana* var. *lobata*)] are of concern when they are attached to trees that might be transported from one location within the project to another. If the trees are felled and skidded for removal from the project site, or for use for wildlife habitat, stream restoration, erosion control, or other practices on site, pretreatment of the vines is essential. Unless the vining plants are effectively killed prior to felling, the movement of the tree will spread seeds and vegetative parts of the vine across the construction area. This often necessitates treatment for at least one season prior to construction.

**Species that propagate vegetatively**—Many species spread vegetatively from portions of roots, stems, or both [e.g., Himalayan blackberry (*Rubus armeniacus*)]. If there is no control prior to construction, whole plants or fragments can be moved during soil excavation, clearing and grubbing activities, and movement of equipment with the result that the species will establish in disturbed areas. Because of the potential spread and tenacity of some species, the objective is to eliminate the risk of spread by the end of construction. Possible treatments include:

**Applying herbicides**—State and local laws, including labeling laws, describe how herbicides are to be used. Additionally, site specific stakeholders might have policies or regulations for herbicide use and their consultation can be very helpful. Herbicides affect plant species through a number of mechanisms such as disruption of cell

division, regulation of growth, stopping photosynthesis, and many others. When possible, attacking the weed species through more than one mechanism will prove most effective. Most species that are considered weeds tend to spread readily and respond more vigorously following ground disturbance than many native species. Due to these factors, systemic herbicides are often desirable. The herbicide(s) used will be determined by regulations, the target treatment species, phenology of target species, tolerance for collateral damage to desired species, season of year, deadline for treatment, and more.

**Applying herbicides and mastication**—A strategy that has proven effective for monocultures of aggressive weedy species, such as Himalayan blackberry in the northwest, is to treat the plants with a combination spray of herbicide, followed by mastication or mowing, and then retreatment with herbicides. The herbicides used combine mechanisms of action as described above and are allowed to translocate for three to six weeks, depending on conditions and herbicides utilized. Once the herbicides travel through the plants' tissues, the above ground shoots are mowed to approximately two to four inches high. The shoots are then allowed to grow until sufficient leaves are developed so that another treatment of herbicide can be conducted. By doing this the plant has been attacked from a number of pathways: 1) multiple mechanisms of action through the herbicides used; 2) reduction of the photosynthetic material; 3) reduction of the seed source; and 4) eventual starvation of root tissue.

**Scraping and removing**—In cases when herbicide use is not desired or allowed, or when there is not sufficient time to conduct treatments, removal of the non-native weed species can be employed. The undesired plants can be scraped from the site during the clearing and grubbing process and buried at an agreed upon location or hauled off site to an approved facility. If an on-site spoils or waste area is to be used, it is important to include language in the special contract requirements to address the specific needs for containment. This often includes the prevention of further mixing or movement of soil once the weeds are placed, and installation of a "cap" of clean fill dirt on top of the weedy species.

**Hand removal**—If weed infestations are relatively isolated, have patchy distribution, or are in small populations they can often be removed by hand.

## Post-Construction Weed Control and Vegetation Management

Most unwanted vegetation can be treated after construction through revegetation contracts, however, long-term vegetation management is typically conducted by road maintenance personnel within a statewide Integrated Vegetation Management (IVM) plan or an Integrated Roadside Vegetation Management (IRVM) plan. Treatments, when they are selected through a decision-making process, include mowing, applying herbicides, mechanical removal, hand-pulling, grazing, fire, and biological control. A discussion of post-construction vegetation management is presented in [Chapter 7](#).

For areas that have been identified as weed-free, functioning plant communities ([Section 3.11.6, see Stratifying the Project by Weed Status](#)), it will be important to maintain these areas in a sustainable manner. An approach to maintaining these areas is outlined in [Section 7.2](#). Although hand removal can be labor intensive it is a great opportunity to involve community volunteers once construction activities have been completed.

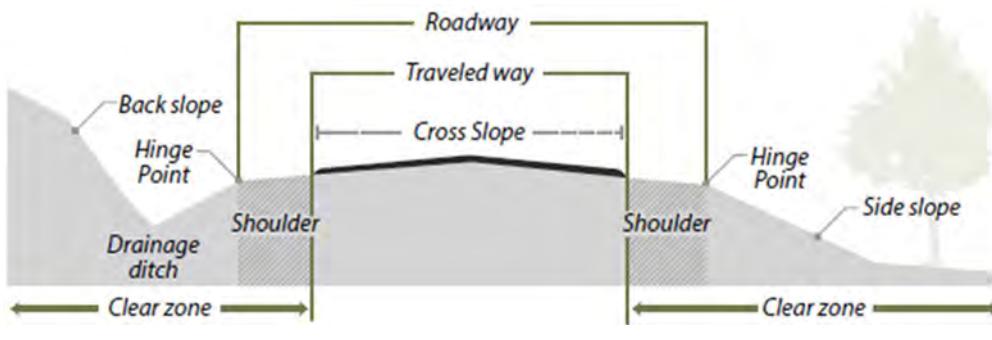
For weed-free, non-functioning plant communities, it is important to decide whether to enhance the current plant community or accept the existing conditions and conduct no additional work. If enhancement is desired, contract components can be included to

### 3—PLANNING

augment existing native plants to increase competition or to reset community succession, treat surrounding non-native plants, treat the soil to better support native plants, alter the light regimen to better support natives, and improve water drainage.

#### 3.11.7 DESIGNING FOR SAFETY AND UTILITY PROTECTION

Planting treatments along roadsides are limited by National Highway System (NHS) design standards and road development best practices guidelines provided by the American Association of State Highway and Transportation Officials (AASHTO) which are adopted by its member State departments of transportation (DOTs). Designers can find guidance on roadside revegetation treatments in key AASHTO guidelines that include the Roadside Design Guide, A Policy on Geometric Design of Highways and Streets (Green Book), Guidelines for Geometric Design of Very Low-Volume Local Roads, and A Guide to Achieving Flexibility in Highway Design. The design guidelines give minimum highway/rural roadside clear zones and urban streetscape horizontal clearances or operational offset distances recommended for motorist safety. The zone distances from edge of travel lane or edge of roadway pavement are determined based on traffic volume, road speed, vertical alignment, and roadside slope conditions. Clear Zones are defined as unobstructed, traversable roadside area that allows driver to stop safely, or regain control of a vehicle that has left the roadway (Figure 3-88) (FHWA 2008(B)).



**Figure 3-88 | Roadway clear zone illustration**

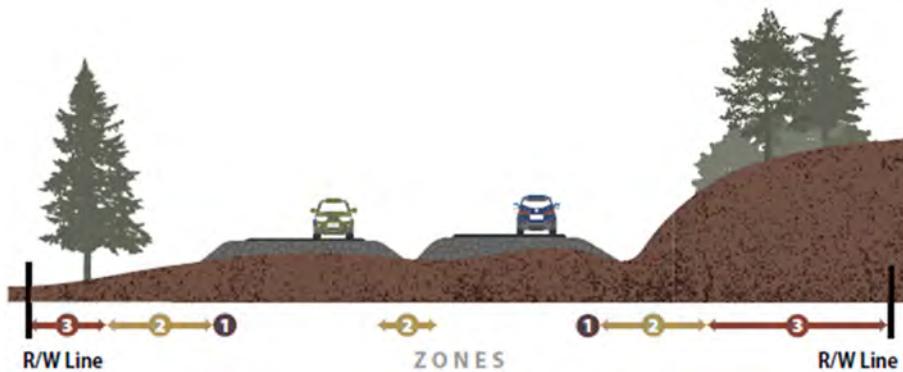
Trees, utility poles, fence posts, and other utility structures are examples of potential immovable objects that the guidelines recommend eliminating from clear zones or protecting with energy absorbing guardrails. Many DOTs recommend protecting existing trees and utility structures instead of removals where possible, based on cost-benefit data. Grasses and forb groundcover in the clear zones can provide a durable and forgiving surface for hazard-free motorist vehicle recovery use, as well as a low groundcover that provides open view of approaching large wildlife or traffic stopped around a curve.

**Hinge Point**—Point where the slope rate changes

**Clear Zone**—A traversable area that starts at the edge of the traffic lane, includes the shoulder, and extends laterally to sufficient distance to allow a driver to stop or return to the road before encountering a hazard or overturning.

Vegetation Treatment Zones (VTZ) are designations used by many DOTs to describe the vegetation coverage and maintenance requirements at certain distances off of the edge of roadway. VTZs typically correspond to highway clear zone distances. The VTZ dimensions, vegetation, and maintenance treatments vary by State and roadside conditions, but generally, DOTs have adopted a 3-zone treatment approach (Figure 3-89), (FHWA 2008(B)). Zone 1 generally extends from the edge of pavement to the drainage ditch along the roadway and is composed of low native grasses that are regularly mowed. Some DOTs mow this area less frequently and some choose to spray it with herbicide to eliminate plant growth entirely. Zone 2 extends across the drainage ditch and a few feet beyond, and is composed of grasses, forbs and low shrubs. Maintenance of zone 2 is focused on maintaining drainage and removal of tree species. Zone 3 extends from Zone 2 out to the edge of right-of-way and may be covered with grasses, forbs, shrubs and trees outside of prescribed safety distances. This area is generally not mowed and may contain large shrub and tree species outside of the clear zone. Maintenance for all zones consists of removal and control of invasive plant species and removal of tree species within safety distances.

**Figure 3-89 | Vegetation treatment zones**



**Clearance for “Line of Sight” and Safety**—Trees and shrubs are often thinned or removed in areas where roadway or roadside line-of-sight is impeded. Good communication with the government agency responsible for maintaining the road during the planning phase will help identify those areas not suitable for shrub and tree species. Native grasses, forbs, and low growing shrubs (3 feet tall or less) can provide durable groundcover for these areas and not affect line-of-sight or motorist safety.

**Trees Management Location**—Trees that reach 4 inch diameter at breast height (DBH) are typically considered immovable objects that can cause heavy damage, injuries, and loss of life in a vehicle crash. According to a study in 2005, more accidents occur between 0 and 12 feet from the travel lane with significantly less between 12 to 30 feet (Mok, Landphair, Naderi, 2006). Tree masses close to the roadway can provide cover for large mammals and can contribute to WVCs. DOTs typically will not allow tree species in clear zones, unless they are protected with guardrail. Best practices for WVC reduction includes full tree and shrub removals or strategic thinning out tree and shrub masses close to the roadway in order to remove desirable cover for ungulates and to open views for motorists to see potential wildlife hazards.

**Protection of Utilities**—Planting of large trees under power lines can lead to damage of utility lines during wind and ice events and can make line maintenance access difficult and dangerous. Coordination of the roadside revegetation plan with the utility agencies of jurisdiction can identify utility easements, planting requirements, utility maintenance access needs, and additional utility accommodations.

### 3.11.8 DESIGNING TO ISOLATE WILDLIFE FROM VEHICLES

According to a 2008 U.S. Department of Transportation study, “there are an estimated one to two million collisions between cars and large animals every year in the United States”...“commonly or typically...with deer (mule deer and white-tailed deer combined)...near forested cover and drainages.” (U.S. DOT, 2008). The collisions often kill the wildlife and can cause high damage expenses, injuries, and even loss of life for the driver. New roadway corridors inevitably intersect wildlife crossings. The analysis of the local wildlife population, their traffic patterns, proximity of their habitat fragments, and any data on area wildlife-vehicle collisions (WVCs) in the pre-budget planning stage can inform the designers on the roadway design features to consider in order to reduce the probability of future WVCs along the corridor. It is important to keep animals out of roadway corridors but to also plan for their safe crossing above or below the roadway (Figure 3-90). Preservation and enhancement of wildlife corridors under or over roadways can be an effective method for reducing WVCs. Natural wildlife corridors often occur along creeks, rivers, and along drainage swales, features that can be preserved through the use of natural bottom culverts or clear span roadway bridge structures. Design of wildlife crossings to accommodate the needs, preferences, and tendencies of the animals that may use the crossings can maximize the likelihood of use and increase safety for all involved.

The design considerations vary with each project location. The FHWA has produced two best practices manuals for designers of roadway corridors and wildlife crossings entitled, *Wildlife Vehicle Collision Reduction Study* (Huijser, 2008) and the *Wildlife Crossing Structure Handbook Design and Evaluation in North America* (Clevenger/Huijser, 2011). Each contain recommendations based on specific studies of wildlife interactions with various wildlife crossing structures, treatments, and conditions.

Wildlife crossing design for motorist safety will typically focus on accommodations for the largest most prevalent animals in the area, often deer. Designing for a variety of wildlife considerations may necessitate greater culvert or bridge clearance height and span length at each crossing, small tunnels for critter crossings, high-water crossings inside culverts (Figure 3-91), hundreds to thousands of feet of tall wildlife exclusion fencing or wildlife friendly/game fence that allows one-way animal pass through/pass back from the roadway (Huijser, Kociolek, McGowen, Cramer, and Venner, 2015), specific plant material to attract and guide animals to the crossing, or a combination of these measures. As the crossings directly impact the budget for the roadway project, and impact the safety of the public who will use the roadway, it is recommended that their evaluation and planning begin at the early planning stages of a new roadway alignment or bridge/culvert replacement project and that the revegetation expert be involved throughout the process.

Suitable habitat on both sides of the road is a necessary condition for all wildlife to cross, and areas with the highest quality habitat will often have the highest rates of crossing (Barnum 2003). Distance to cover is another factor that affects wildlife crossing use. Small animals prefer plentiful and consistent cover before and, if possible, through an undercrossing. Deer and elk tend to prefer crossing in open areas away from forest cover, especially during the winter (Clevenger and Walther 2005; Barnum, Rinehart, and Elbroch 2007). Other animals prefer a more balanced composition of cover and open space while most carnivores prefer a dense forest cover.

Roadside revegetation design provides the opportunity to support safe wildlife crossings by creating a roadside planting and maintenance plan that will reduce animal browsing within the ROW, and provide planted conditions that will attract wildlife to the crossing points. Open views to vegetation beyond the undercrossing will



**Figure 3-90 | Mule deer using an underpass**

Deer are frequently involved in WVCs. Early planning and thoughtful design of undercrossings and overpasses can encourage wildlife use and enhance safety for wildlife and motorists.



**Figure 3-91 | Culvert with high-water ledge for small mammal crossing**

Maintaining safe access to habitat, including during high water periods can enhance safety for wildlife and motorists and also keep the food chain functioning.

*Photo credit: Unknown*

### 3—PLANNING

encourage animal movement through the crossing to the other side, and continue on to wildlife corridors or refugia beyond. Ungulates, or hoofed mammals, especially deer, are browsers that prefer fresh new growth. Spring growth and new growth after cutting and mowing maintenance will attract ungulates and encourage them to linger and graze. Increasing mowing maintenance at wildlife undercrossings may increase more regrowth periods that can draw browsers to the crossing locations. Reducing the quantity of roadside mowing events away from undercrossings and strategically timing mowing operations to late fall or very early spring, can reduce the number of mass new growth events that draw ungulates to browse along the ROW.

Study results for vegetation control effect on WVCs indicate that flatter ROW side-slopes and lower vegetation along the roadway can improve driver safety and discourage use by large wildlife. Low plant material is less desirable for use as cover by ungulates, and opens up view-sheds so drivers can potentially see animals in the right-of-way (ROW) and allow time to react and slow down to avoid wildlife-vehicle collisions. Plant material 3 feet tall or lower, not counting seed head stalks that may rise taller later in the year is generally considered low. Elimination of large trees from roadway clear zones can remove desirable wildlife cover from the ROW and immovable tree trunks that can damage vehicles and injure motorists that leave the roadway.

Wildlife over-crossings are still rare in the United States but are preferred by ungulates and otherspeciesmore than under-crossings. Ungulatesand otherspecieswill use under-crossings if they appear open and free from predator hiding places. They generally prefer level open space leading up to the crossing, a generous tall and wide bridge span or box culvert, natural low vegetation or soil walking surface, and clear views to open space on the other side.

Easy access to wildlife crossing locations can increase their use. If an existing ROW fence exists beyond the wildlife crossing area, it is recommended that portions of fence be removed if possible. In the case of cattle or other livestock use outside of the ROW, a section of fencing may be replaced with a gate that remains open when the field is not occupied. Additionally, wildlife friendly adjustments may be made to the top and bottom strands of barbed wire, such as adjusting the wire spacing and/or replacing with smooth wire on the top and bottom to protect wildlife that jumps over or crawls under the fence.

High-use crossing locations and those with narrow rights-of-way may need the addition of protective traffic barrier and guardrail and/or lengths of tall animal fencing to help encourage animals back down to the under-crossing. Resources that provide wildlife control methods to consider for specific site conditions include Best Management Practices for Wildlife Corridors (Beier, 2008), Wildlife Vehicle Collision Reduction Study (Huijser, 2008), the Wildlife Crossing Structure Handbook Design and Evaluation in North America (Clevenger/Huijser, 2011), Construction Guidelines for Wildlife Fencing and Associated Escape and Lateral Access Control Measures (Huijser, et al., 2015), and Implementing Measures to Reduce Highway Impacts on Habitat Fragmentation (Louis Berger Group, 2011).

Experts recommend that wildlife crossings remain open and clear overnight, during the construction process if possible. They also recommend these areas be completed and planted as early as possible in order to reduce animal stress and to keep them from learning new less desirable places to cross the roadway. Supplemental hay and salt licks may also be used to encourage continued crossing use during construction; supplemental feeding may then taper off over a few months after construction.

Plant selection for wildlife crossing locations can be approached as seeded roadside pollinator habitat development supplemented with low native forbs and grasses that are preferred by wildlife for food and browse.

### 3.11.9 DESIGNING FOR DISTURBANCES

Disturbances that affect roadside vegetation often occur after plants have become established. While some of these disturbances are unforeseen, others can be expected. The designer may want to consider what disturbances can be expected and how they may be mitigated within the design of the roadsides and revegetation plan.

#### Deicing for Winter Safety

Approximately 70 percent of the roads in the US are in snow regions (FHWA 2012) that may require deicing practices to make them safe and passable during winter periods. Most deicing materials contain chloride-based salts which, when applied to road surfaces, lowers the freezing point and melts snow and ice. Solid salt ( $\text{NaCl}$ ) is the most common product used, followed by calcium chloride ( $\text{CaCl}_2$ ), magnesium chloride ( $\text{MgCl}_2$ ), potassium acetate ( $\text{KAc}$ ) and calcium magnesium acetate (CMA) flakes for bridges (AASHTO 2013). The suggested rate of  $\text{NaCl}$  is 100 to 300 pounds per lane-mile (Salt Institute 2008).

**Potential Impacts to soils and vegetation**—Deicing materials can pose a risk to soil properties and plant growth. Salt concentrations in roadside soils correlate positively with salt application rates (Jones et al 1992) and high levels of sodium disperse soil organic and inorganic particles, reducing soil permeability and increases runoff (AASHTO 2013a). The negative effects on plant growth, however, are often associated with road spray on plant foliage rather than presence of salts in the soil. Roads that are treated with deicing materials does not preclude that plants on adjacent roadsides will be affected. The effects of road salts on soils and plant depend on the:

- **Sensitivity of plant species to salt**—At very high salt levels in the soil, germination of native plant species can be reduced (Harrington and Meikle 1992, Fulbright 1988) or delayed (Ungar 1992) though there is considerable variability between native species. In general, shrubs and grasses tolerate salt concentrations better than trees (Sucoff 1975, Bryson and Barker 2002). And sensitivity among tree species ranges from sensitive to tolerant. In the Lake Tahoe Basin, for example, two and three needle pines (e.g. Jeffrey, ponderosa, and lodgepole pines) show salt damage more frequently than white and red fir (University of Nevada 2009).
- **Amount of salt applied annually**—The amount of salt damage is related to the total amount of salt applied during the winter. The higher the quantities of salt applied, the greater the effects to soil and plant quality. The University of Nevada (2009) found that the proportion of trees affected by salt damage coincided with the annual quantity of salts applied to the road surface.
- **Distance from the road**—The highest amount of salts occurs closest to the road and diminishes moving away from the road surface. Ninety percent of salt deposition from road spray often occurs within 65 feet of the roadside (Blomqvist et al 1999) and it is in this zone where foliar damage such as needle necrosis, twig dieback and bud kill on trees happens.
- **Type of salt**—The type of deicer may affect plant species differently. Trahan and Peterson (2008) found that  $\text{MgCl}_2$  was more damaging when directly applied to tree foliage than  $\text{NaCl}$ . Calcium magnesium acetate was found to be less toxic on certain grass species than sodium based salts (Robidoux and Delisle 2001) and may even improve soil properties by increasing permeability and providing calcium and magnesium for soil fertility (Fritzsche 1992).
- **Precipitation**—In areas of high precipitation, salts will become diluted and

move through the soil profile, reducing the potential negative effects to seed germination and plant growth. In areas of very low precipitation however, salts and sodium will remain at the soil surface and accumulate.

- **Soil type**—Soils with low pH values may benefit from some addition of road deicers. By raising soil pH, certain nutrients become more available ([Section 3.8.4](#)). Soils already high in sodium could become even more toxic to plant growth with NaCl additions.

**Assessing potential impacts of deicing practices**—During the planning phase, it may be important to assess the level of impact deicing practices have on native plant establishment and growth after construction. The type and amount of deicing material used on the highway project can be obtained from maintenance and operations records. If the quantities are considered high (Figure 3-47), then it may be beneficial to conduct a field survey of soils and vegetation along a stretch of road in or similar to the project area. The survey area can be stratified into zones parallel to the road alignment because the effects of salts on vegetation composition and health grade from most affected to least affected, moving away from the road. Specific monitoring procedures that may be helpful include Soil Cover ([Section 6.3.1](#)), Species Cover ([Section 6.3.2](#)), and Species Presence ([Section 6.3.3](#)), using a Rectilinear Sampling Area design ([Section 6.3.6](#), see [Rectilinear Areas](#)), because the narrow width of each sampling area is not conducive to using transects.

Comparing the differences in each zone may show the effects of deicing practices on vegetation. If there are no differences between each zone, then it can be assumed that deicing practices have no effect. If there are differences, then those differences are taken into consideration when developing seed mixes or soil improvement practices. It may be important to determine if these differences are due to deicing practices or due to other factors, such as mowing, surface contaminants, soils, air pollution, drought, or tree diseases or pests. One method to make this determination is to collect soil samples in each zone and measure soil conductivity using a pH meter ([Section 6.3.1](#)). If conductivity readings recorded in the zone next to the road are higher compared to the zone furthest away it would indicate that deicing practices maybe responsible. Refer to Figure 3-47 for interpreting conductivity values that affect plant growth.

**Mitigating for deicing practices**—If road deicing salts are determined to be detrimental to plant growth, the designer may want to select a species mix that has a higher tolerance for soluble salts. Selecting the tolerant plant species can be determined from information collected during the vegetation assessment ([Section 6.3.1](#)). Desirable species growing in the deicing zone are good species to consider in species mixes. The ERA tool may also give some guidance on those species most adapted to high salt environments.

## Gravelling for Winter Safety

In areas where gravels are frequently applied to road surfaces during snow or icy conditions, there can be a buildup of gravels on the road shoulders. Vegetation that has been established in these areas is often completely covered with gravel after snow melt. In addition, road maintenance often excavates these gravels to reuse and in the process, removes established plants growing in this zone. The designer will want to design revegetation treatments in these areas according to the expected disturbance. In areas where gravels are not salvaged, the designer may want to select plant species that survive and grow well in this unique growing environment. Some species respond favorably to being covered by gravels. These include species such as manzanita and willows that root from their stems when covered by soil or gravel. Tap-rooted species, such as lupines, can take advantage of such conditions because they can access moisture deep in the gravel deposits (Figure 3-92). A vegetation assessment ([Section 6.3.1](#)) of the road shoulders of the project area will identify species that have adapted to these conditions. In areas where gravels are annually removed, the designer will want to identify the width of this disturbance and remove the area from the revegetation plan.

### **Figure 3-92 / Graveling road surfaces can lead to burying roadside vegetation**

Gravel applied to road surfaces in winter for traction is swept or blown to the side, burying vegetation (A). Some species, such as *Lupinus* spp. (B), have adapted to these conditions and do well. Species such as pinemat manzanita (*Arctostaphylos nevadensis*) also do well when covered by gravel because the plant will produce roots from buried stems.

*Photo credits: David Steinfeld*



## Annual roadside maintenance

Ditches at the base of steep cut slopes are depositional areas for rock and soil that have moved down from the slopes above. This material fills ditches, disrupting the flow of water and creating potential road drainage problems during storms. Blading is the removal of material that has filled in the ditchline and is a normal maintenance procedure for erosive cut slopes. This operation not only removes plants that were established in the ditchline, but also destabilizes the surface slopes immediately above the ditch which can affect the revegetation of the entire slope. Designing cut slopes so that they are stable is one method of reducing the need to blade ditches. This includes reducing slope gradients near the ditches and establishing a good vegetative cover that resists slope movement.

## Recreational Activities

The road corridor is sometimes used for recreational purposes that can disturb established vegetation. This recreation is not usually sanctioned or intended, but it exists in certain areas nonetheless. Recreational disturbances include off-road vehicle travel, mountain bike use, trails to recreational sites, parking, and Christmas tree cutting.

It is important for the design team to identify the public's demand for recreational activities and to determine how these activities might affect short- and long-term vegetation goals. For example, abandoned roads that have been revegetated are often desirable places for off-road vehicles because they are open and flat. Roads bordering recreation destinations, such as favorite fishing spots, may have demands for access trails or scenic views that the public does not want blocked by tall vegetation. Public scoping often identifies these needs. There are several approaches to mitigating the effects of recreational impacts, most of which are forms of awareness, protection, and exclusion. Intruders can be excluded physically with barriers, such as ditches, fences, down trees, and large rocks outside of the clear zone. Communication is a good first option before these measures are put in place. For example, a sign explaining native revegetation efforts may help make potential users aware that they should take their activities elsewhere. Local residents are often great sources for ideas on how to approach these problems; off-road vehicle clubs are another. Educating the public on the purpose for the revegetation treatments can go a long way toward protection. Short paragraphs in the local newspapers or on the FHWA website for each project may help. Using local contractors to implement the revegetation work and engaging local residents brings ownership to the project. In addition to exclusion measures, designers can consider desire paths to accommodate foot traffic that is bound to occur. Incorporating and planning for such access can reduce the impact to the surrounding established vegetation and keep traffic confined as much as possible.

## Livestock

Damage to revegetation projects can be high in areas that are intensively grazed by cows or sheep. In areas with large livestock populations, planted tree seedlings can be injured by rubbing and trampling. Newly establishing native grass and forb cover can be harmed through grazing and by the high-pressure hoof marks tearing up the new roots and surface soil, leaving the site exposed to non-native annual species.

Restricting the entry of livestock for several years after planting or sowing, or until native grasses and forbs have established, is the best prevention measure. This is typically accomplished by fencing the entire area being revegetated. Fencing is most effective when it is installed prior to establishing native vegetation. For this reason, having the fence installed as part of the road contract will ensure that livestock is controlled prior to revegetation work. Working with the local USDA Forest Service or USDOI Bureau of Land Management range conservationist will be necessary to ensure that damage by livestock is kept to a minimum.

### 3.11.10 DESIGNING FOR CARBON SEQUESTRATION

Carbon sequestration is an important environmental and public health benefit that is a result of revegetating disturbed landscapes with native plants. It is often overlooked as a revegetation objective and as part of a vegetation management strategy. The "carbon sequestration capacity" is a quantifiable volume of carbon that can be estimated for existing roadside revegetation and compared to the proposed revegetation plan. Knowing the values in the planning stage can help the design team as they make decisions about design, implementation, and maintenance throughout the project.

development. Selection of plant material and the ongoing vegetation maintenance procedures have a dramatic effect on the carbon sequestration capacity of a revegetation project.

Carbon sequestration is a process in which CO<sub>2</sub> is transferred from the atmosphere into plants through photosynthesis and stored in long-term carbon pools. These pools consist of above-ground biomass (e.g. live trees, shrubs, grasses, and standing dead trees, branches, litter, and duff) and below-ground biomass (e.g. soil organic matter, roots, organisms). Roadside management practices that maintain or increase these carbon pools may reduce atmospheric concentrations of CO<sub>2</sub> and mitigate the effects of climate change (Proudfoot 2015). With the large land base of the US in roadsides, the current and potential capacity to capture CO<sub>2</sub> is considerable (Ament 2014). For example, roadsides along US highways and federal lands (10.5 percent of all public roads) currently capture nearly 2 percent of the total US transportation carbon emission (Lavelle 2014). Another way that atmospheric carbon can be reduced is by decreasing or changing roadside maintenance operations that generate greenhouse gas emissions such as mowing and mechanized pesticide applications. In combination, practices that reduce carbon emission and increase carbon pools can reduce atmospheric carbon while reducing maintenance costs (Proudfoot 2015).

## Minimizing Soil Disturbance

Soils contain large amounts of carbon fixed in soil organic matter. When soils are disturbed, CO<sub>2</sub> is released through the oxidation of organic matter. One of the best strategies for maintaining carbon in soils is to minimize soil disturbances. In planning road construction projects, this is accomplished by minimizing the footprint of the project. Another strategy is to create and maintain a resilient plant community that resists disturbances associated with soil erosion and landslides. When disturbances do occur, immediate action assures the quick recovery of native plants and carbon sequestration processes ([Section 3.11.10](#), see [Create a Good Growing Environment](#)).

## Revegetating with Trees and Shrubs

Establishing and maintaining trees and shrubs along roadsides can be a cost-effective means of capturing carbon (Brown 2010). Compared to other vegetation, trees sequester larger amounts of carbon for longer periods of time (an average of 120 years, FHWA 2010). Trees also shade ground surfaces, reducing the amount of heat generated from roadsides. Shrubs have less capacity to store carbon than trees but greater capacity than grasslands (Ament 2014).

There is an opportunity to use vegetative plantings to create or replace wind breaks, shelterbelts, and snow fences (see [Inset 3-3](#)). Using trees and shrubs for these purposes will not only reduce blowing and drifting snow, but can increase wildlife diversity, pollinator habitat, and capture carbon. Using a thick vegetative barrier composed of shrub species near the roadway may also have the added benefit of slowing out-of-control vehicles from roadsides impacts (Ament 2014).

It is important that trees and shrubs are planted in areas that meet road safety and maintenance objectives. When choosing tree and shrub species to plant, species that live longer and are larger at maturity have a greater capacity to store carbon than shorter lived, smaller plants (Proudfoot 2015). In addition, increasing the complexity of a forested site, by planting a multilayer of trees, shrubs, grasses and forbs, has the potential of increasing carbon sequestration (Ament 2014).

## Revegetate with Perennial Grass, Forb, and Wetland Species

Perennial grasses store more carbon in the soil than annual grasses (Cox et al 2006) and can sequester carbon for up to 50 years (FHWA 2010). In addition, perennial grasses

have greater ground cover than annual grasses which protect soils from surface erosion, water loss, and nutrient loss (Glover 2005), important for optimizing carbon capture. Wetland species capture more carbon than grasses and forbs because of the higher productivity of wetland sites. For this reason, wetland swales are preferable to dry swales (Bouchard 2013).

## Create and Maintain a Good Growing Environment

Revegetation planning that promotes healthy functioning plant communities is good for carbon capture and maintenance (Ament 2014). Restoring soils with organic amendments, tillage, mulch, and nutrients will increase the rooting depth and productivity of roadsides to store more carbon. Other practices that maximize slope stability and minimize surface runoff reduce the potential that soils are disturbed which maintains soil carbon. Where road sections are being abandoned or recontoured, restoring the soils and reestablishing perennial vegetation, such as shrubs and trees, have the potential to capture and store carbon while increasing pollinator habitat.

## Utilize Site Resources

Land clearing during road construction often creates woody material that is placed in piles and burned. This material can be processed and placed on constructed roadsides as a soil amendment or mulch. Unprocessed woody material, such as logs, can be used as wildlife structures in areas where they meet maintenance objectives. Depending on site factors, processed and unprocessed materials can last from years to decades, temporarily storing carbon before they decompose. How salvaged topsoil is removed and stored may also make a difference in how much soil carbon is oxidized during road construction. Removing topsoil when it is dry and keeping it dry during storage reduces the potential for oxidation of organic matter.

## Changing Mowing and Pesticide Practices

Changes in mowing and pesticide practices can directly reduce carbon emissions and increase carbon storage (Dunn 2013). A review of maintenance practices, when developing a vegetation maintenance strategy ([Section 3.11](#)) and an Integrated Vegetation Management plan ([Chapter 7](#)), can highlight areas where changes can be made. In addition to decreasing carbon emissions, changes to practices can also lower maintenance costs because of reduction in fuel and wages, and can be beneficial to pollinators ([Section 7.3.2](#)). Maintenance practices that can be adapted to reduce emissions and increase carbon capture include:

- **Mowing times**—Shifting mowing times from active growing periods (when mowing disrupts the flow of carbon to the soil) to times of the year when plants are more dormant (early spring, fall, and winter), will increase carbon capture (Dunn 2013).
- **Frequency of application**—Cutting back on the frequency of mowing or pesticide applications reduces carbon emissions.
- **Height of mowing equipment**—Raising mowing equipment several inches higher can save fuel costs and reduce the effects of carbon flow to the soil.
- **Treatment widths**—Reducing the widths of mowing and pesticide applications reduces travel time, amount of pesticide used, and carbon emissions.

## Reducing Road Salts

On road systems where applications of deicing salts are detrimental to roadside

vegetation ([Section 3.11.9](#), see [Deicing for Winter Safety](#)), minimizing the quantity of salt applied or the frequency of application, can reduce the effects on plant productivity and carbon sequestration (Ament 2014). These changes will also result in less carbon emission.

## Highway Carbon Sequestration Estimators

Numerous highway carbon sequestration estimators can help calculate the quantity of carbon being captured on a roadside and estimate the potential volume of carbon offsets. These tools do not necessarily provide estimates required for full project development and can only to provide a sense of scale (Proudfoot 2015). One such program is the Highway Carbon Sequestration Estimator. This tool is intended to help DOTs assess the return on investments for carbon sequestration practices based on state-specific considerations (FHWA 2010).

## 3.12 SELECT SITE IMPROVEMENT TREATMENTS

In this stage of planning, the treatments that will improve the site for plant growth (e.g. topsoil, compost) are selected. The selection of treatments begins with identifying the factors that limit plant growth. As the designer identifies limitations specific to the project site, a list of possible treatments that will mitigate or reduce the effects of the limiting factor is developed. Specific treatments to mitigate each limiting factor are presented in [Section 3.8](#). The case study presented in Inset 3-6 shows how a list of mitigating measures can be developed for specific limiting factors.

In narrowing down the possible treatments that will encourage plant success, the designer considers all possible resources on the project site that can be used in lieu of purchasing and transporting materials from offsite sources ([Section 3.10](#)). For example, in reviewing the plans of a road reconstruction project, it is found that a portion of the road is being realigned through a wooded area. For this section, the designer lists the potential site resources to include topsoil, large wood, tree branches and foliage, duff and litter. These resources can be used in a variety of ways to improve growing conditions and pollinator habitat.

At this point, the vegetation management strategy is revisited to ensure that the mitigating measures being considered in this process are compatible with road maintenance objectives ([Section 3.11.2](#)) or if they could present problems for long-term maintenance and if so, what modifications to the treatments can be made. The final selection of treatments is based on many factors, including project funding, project objectives, experience of the designer or contractor, and availability of resources and equipment. A Limiting Factor and Site Resource tables are available in this [Planning workbook](#).

**Inset 3-7 | Case Study—Defining limiting factors and selecting mitigating measures vegetation treatment zones**

**Site Inventory**—During the planning stage of a revegetation project in central Oregon, the soils assessment ([Section 3.6.2](#)) highlighted several limiting factors that would negatively affect plant growth.

**Limiting Factors**—From the list of limiting factors outlined in [Figure 3-11](#), factors affecting plant growth at this site were narrowed down to low precipitation, low water storage, high water loss, and low nutrients.

**Mitigating Measures**—For each of these factors, a list of possible mitigating measures (described for each limiting factor in [Section 3.8](#)) was developed.

**Site Resources**—A review of the site resources was made at this point to determine if there were any resources on the site that could be used in developing mitigating measures ([Section 3.10](#)). It was found that there were several areas where weed-free, high quality topsoil could be salvaged and reapplied. In addition, the project would produce a large amount of slash from cleared shrubs and trees that could be processed into shredded wood and used as a mulch. Other resources available were a local municipal waste treatment plant had Class A biosolids available for application in lieu of bringing in fertilizer, loam borrow from a source of pumice deposit, and an excavator that would be available for subsoiling.

**Maintenance**—The potential effects of these treatments on long-term maintenance was then considered ([Section 3.11](#)). It was determined that deep tillage was not compatible with safety in Zones 1 and 2 but would be done in Zone 3, otherwise there were no foreseen maintenance problems.

**Treatment Selection**—It was decided to salvage topsoil; however, the amount of topsoil would not cover all the project needs. It was determined that a manufactured topsoil would be created by using shredded wood processed the slash from road right-of-way clearing, loam borrow from excavated pumice, and biosolids from the local municipal waste treatment plant. To increase infiltration and rooting depth, it was decided that once the topsoil and manufactured topsoil was applied that it would be subsoiled only in subsoil Zone 3 but not in Zones 1 and 2. At planting, lupine would be included in the seed mix for additional nitrogen and inoculum would be applied to the seed mix. Because there would be additional shredded wood, it would be blown over the seed as a mulch.

| Critical plant factors                 | Parameters           | Limiting | Possible mitigating measures                     |
|--|----------------------|----------|--|
| <b>Water input</b>                     | Precipitation        | ☒        | Irrigate, water harvesting                       |
|  | Interception         |          |  |
|  | Infiltration         | ☒        | Tillage, organic matter, mulch, avoid compaction |
| <b>Water storage and accessibility</b> | Soil texture         | ☒        | Organic matter                                   |
|  | Rock fragments       |          |  |
|  | Soil structure       | ☒        | Tillage, organic matter, avoid compaction        |
|  | Rooting depth        | ☒        | Tillage, topsoil, planting islands/pockets       |
|  | Mycorrhizal fungi    | ☒        | Topsoil, inoculums                               |
| <b>Water loss</b>                      | Wind                 |          |  |
|  | Aspect               |          |  |
|  | Competing vegetation |          |  |
|  | Soil cover           | ☒        | Mulch  |
| <b>Nutrient cycling</b>                | Topsoil              | ☒        | Topsoil, planting islands                        |
|  | Site organic matter  | ☒        | Shredded wood, compost, litter, wood             |
|  | Nitrogen and carbon  | ☒        | Topsoil, compost, N-fixing plants, fertilizers   |
|  | Nutrients            | ☒        | Topsoil, compost, fertilizers, biosolids         |
|  | pH and salts         |          |  |
| <b>Surface stability</b>               | Rainfall             |          |  |
|  | Wind                 |          |  |
|  | Freeze/Thaw          | ☒        | Tillage, mulch                                   |
|  | Soil cover           |          |  |
|  | Surface strength     |          |  |
|  | Infiltration         |          |  |
|  | Slope gradient       |          |  |
|  | Surface roughness    |          |  |
|  | Slope length         |          |  |
| <b>Slope stability</b>                 | Permeability         |          |  |
|  | Restrictive layer    |          |  |
|  | Water input          |          |  |
|  | Slope length         |          |  |
|  | Slope gradient       |          |  |
|  | Soil strength        |          |  |

## 3.13 SELECTING PLANT SPECIES FOR PROPAGATION

Selecting plant species to propagate and install on a project is a multifaceted process. An understanding of the revegetation objectives ([Section 3.2](#)) is helpful in identifying the limitations of the site to support the species being considered ([Section 3.8](#)), identifying the factors that affect pollinator ([Section 3.9](#)), and determining if the plant species integrate into the vegetation management strategy ([Section 3.11](#)). This section covers how to use the comprehensive species list and seed zones and transfer guidelines to develop a potential plant species list.

### 3.13.1 DEVELOPING A POTENTIAL PLANT SPECIES LIST

From the comprehensive species spreadsheet developed in [Section 3.6.1](#), each species is evaluated for its potential to be used on the project. This is accomplished by sorting the spreadsheet using some or all of the following criteria:

- **Nativity**—If the revegetation objectives call for using native plants, then species on the comprehensive species list are first sorted by whether it is native or not.
- **Pollinator-friendly**—Based on the pollinator habitat assessment ([Section 3.6.3](#), see [Habitat Assessment](#)), a list of pollinator plant species to enhance site quality for pollinators can be developed from the ERA tool and the reference site plant species inventory.
- **Workhorse species**—The next sort is by workhorse species. Workhorse species is a term used to describe locally adapted native plants that: (1) have broad ecological amplitude, (2) high abundance, and (3) are relatively easy to propagate. A list of workhorse species for ecoregions (Level III) can be obtained using the ERA tool. Because these lists are still in development, some species may need to be evaluated for potential as a workhorse species based on the project objectives and needs. To determine if a species (not listed as a workhorse in ERA) is a potential workhorse species, sort the comprehensive species list by amplitude and abundance columns. Those species that have high amplitude and abundance are good candidates for workhorse species status. From these species, evaluate how easy they are to propagate. This includes the availability of the starter plant materials, how easy the species is to propagate in the nursery or seed production fields, how well the seeds store, the survival of the plant materials once they are installed on the project, and expense.
- **Availability of starter plant materials**—Seeds, plants, and cuttings often have to be collected in the wild and supplied to the nursery or seed producer for plant production, seed increase, or stooling beds. Species that are difficult to obtain or collect are not good candidates for workhorse species status ([Section 5.3.1](#) through [Section 5.3.3](#)).
- **Nursery and seed production**—Species that are difficult to propagate in the nursery, stooling beds, or seed production fields do not make good workhorse species ([Section 5.3.4](#) through [Section 5.3.6](#)). Because new techniques in propagating native species is constantly improving, talking to nursery managers or seed growers, in addition to referring to documented plant production protocols available on the internet, is important in maintaining a current workhorse species list.
- **Longevity**—Seeds that have a poor shelf life under seed standard storage practices (seed germination that drops significantly after one year in storage) are

often not good candidates (Section 5.3.4).

- **Field establishment**—The ease that a plant material will establish on a project site will determine if a species is a workhorse species (Section 5.3.3 and Section 5.5). Some species do not perform well because breaking seed dormancy and obtaining good germination may be difficult. Other species, planted as seedlings, experience unusually high transplant shock that significantly reduces plant survival.
- **Expense**—The total costs for establishing native plants on the project site is the easiest measure of whether a species is a good candidate for workhorse species status.
- **Working groups**—A working group is a mix of workhorse species developed for a specific ecological function or management objective. One of the best ways to develop working groups is to sort the comprehensive species list by ecological setting and succession. This will assemble species into groups that naturally occur together. From these groups, working groups are developed based on project objectives, such as pollinator habitat enhancement, weed control, visual enhancement, conservation management, and erosion control (Figure 3-93). The development of these working groups is often the basis for a project’s seed mixes and planting mixes for each revegetation unit.
- **Specialist species**—Species that are important for achieving project objectives yet do not meet the workhorse species criteria for seed propagation are called specialist species (Figure 3-94). These species may still be propagated; however, because there may be very little information about seed propagation, they may take more time and higher costs to develop. Projects that contain special microclimates or soils may require a unique mix of specialist species (e.g., a wetland working group), while other projects may require a specific species to meet a project objective (e.g., milkweed as host plant for monarch butterfly). If only a small quantity of specialist species is needed, then consider using methods other than seed propagation. These include collecting plant materials in the wild or growing seedlings in the nursery.

Once the spreadsheet is complete, the species and stocktypes to propagate can be selected.



**Figure 3-93 / Steep roadcuts require an erosion control working group**

Steep roadcuts on the North Umpqua Highway in Oregon required an erosion control working group that hold the slopes together and keep them from sloughing and eroding into road ditches. An erosion control working group composed of Roemer's fescue (*Festuca roemerii*), blue wildrye (*Elymus glaucus*), and California brome (*Bromus carinatus*) was applied with high rates of hydromulch and tackifier to the cut slopes. One year after application, cut slopes had a high cover of these native grass species that significantly reduced sloughing and erosion.

*Photo credit: David Steinfeld*

**Figure 3-94 | Example of a specialist species**

Aspen does not meet “workhorse” criteria because it is challenging to propagate (starter material is difficult to obtain and plants require special protection from browsing after outplanting). However, aspen is very important for ecological reasons, and therefore can be propagated as a specialist species for specific projects.

*Photo credit: Chris Jensen USFS*



**Table 3-14 | Selecting species to propagate**

The comprehensive species list developed in Table 3-6 can be used to determine the species that are most appropriate to use on the project. In this example, non-native species were removed from the list, leaving only native species. Then potential workhorse species were determined by their amplitude, abundance, and ease of propagation. A species such as *Achillea millefolium*, for instance, is considered a workhorse species because it fits all criteria—high abundance, high amplitude, and easy to propagate. Whether this species is used for this project depends on whether it is a member of a particular working group that meets a specific project objective. Because *Achillea millefolium* fits into the “visuals” working group, which is an important road objective, this species is selected for propagation. *Agastache urticifolia* is also showy and has a high amplitude and abundance. However, very little is known about the propagation of this species. While this species has the potential of being a workhorse species, it will be grown as a trial in small quantities. *Allium fibrillum* and *Allium macrum* are specialist species that occur together in a unique meadow habitat. Because this road project provides an opportunity to enhance meadow habitat, these two species are placed in a “conservation” working group. They are considered specialists because they have low abundance, low amplitude, and little is known about their propagation, yet they are an important component of the working group. These species will be selected for use on the project; however, because little is known about seed propagation, seeds will be sent to a nursery for plant or bulb propagation.

|                              |                           |      |      |       |          |        |           |     |              |        |
|------------------------------|---------------------------|------|------|-------|----------|--------|-----------|-----|--------------|--------|
| <i>Achillea millefolium</i>  | <u>common yarrow</u>      | High | High | Early | All      | Seeds  | Easy      | Yes | Visuals      | Yes    |
| <i>Agastache urticifolia</i> | <u>horsemint</u>          | High | High | Early | All      | Seeds  | Unknown   | No  | Visuals      | Trials |
| <i>Agoseris aurantiaca</i>   | <u>orange agoseris</u>    | High | Mod  | Early | All      | Seeds  | Unknown   | No  |              |        |
| <i>Agoseris glauca</i>       | <u>pale agoseris</u>      | High | Mod  | Early | All      | Seeds  | Unknown   | No  |              |        |
| <i>Agoseris grandiflora</i>  | <u>bigflower agoseris</u> | High | Mod  | Early | All      | Seeds  | Unknown   | No  |              |        |
| <i>Abies grandis</i>         | <u>grand fir</u>          | High | High | Late  | All      | Plants | Easy      | Yes |              | Yes    |
| <i>Abies lasiocarpa</i>      | <u>subalpine fir</u>      | High | Mod  | Late  | Cool     | Plants | Difficult | No  |              |        |
| <i>Allium acuminatum</i>     | <u>tapertip onion</u>     | Low  | Low  | Early | Wet      | Bulbs  | Difficult | No  |              |        |
| <i>Allium fibrillatum</i>    | <u>fringed onion</u>      | Low  | Low  | Early | Warm/Dry | Bulbs  | Difficult | No  | Conservation | Yes    |
| <i>Allium macrum</i>         | <u>rock onion</u>         | Low  | Low  | Early | Wet      | Bulbs  | Difficult | No  | Conservation | Yes    |
| <i>Allium madidum</i>        | <u>swamp onion</u>        | Low  | Mod  | Early | Wet      | Bulbs  | Difficult | No  |              |        |

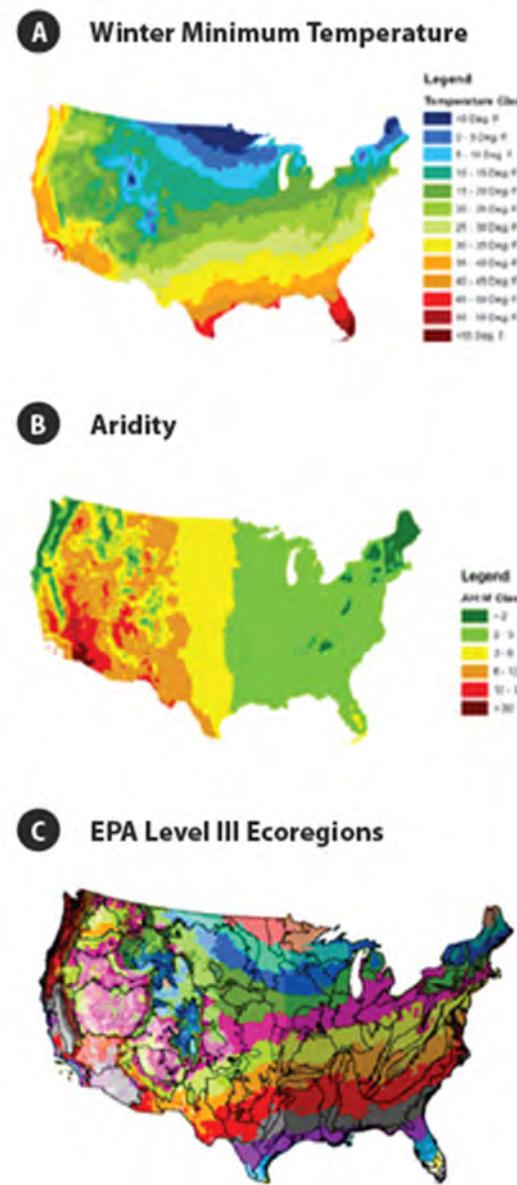
### 3.13.2 ENSURE LOCAL ADAPTATION AND MAINTAIN GENETIC DIVERSITY

#### Seed Zones and Transfer Guidelines

It is important to know the original collection source and genetic background of target plant materials to ensure better long-term adaptation to local conditions and protect plant-pollinator relationships and the genetic resources of local plant communities. Seed transfer guidelines (how far plant material can be transferred from point of origin to the project with minimal risk of maladaptation) were initially developed for commercially important forest tree species. This was the outcome of years of research that revealed that failures in tree planting establishment were sometimes the result of moving seeds too far from their source of origin. More recently, research has been completed or is under way to develop seed zones and transfer guidelines for grasses, forbs, and shrub species commonly used in revegetation activities, particularly in the western U.S. (e.g., Table 1 in Bower et al 2014; St. Clair et al 2013; Johnson et al 2013; Horning 2010).

Seed sources that originate within the specific seed zone where a planting site is located are likely to be well adapted, with improved survival, reproductive success, and resiliency in harsh sites and changing climate conditions. Genetic research indicates there is no fixed number determining the geographic distance that plants might be successfully moved (e.g., within a 50-mile radius). Rather, “local” is best defined in terms of the environment (local climate and soils) rather than absolute distance. Many factors contribute to the environmental conditions to which a plant species must adapt, including rainfall, aridity, maximum and minimum temperatures, aspect, soil drainage, and pH. The scale of adaptation also varies greatly among species. Some species (genetic generalists) can tolerate broader movement across environmental gradients than others (genetic specialists) and still be well adapted to local conditions and regions (Rehfeldt 1994; Johnson et al 2010). Thus, plant movement guidelines derived from empirical genetic studies are specific to the individual species and geographic area where the research was conducted.

In addition to improving the success of revegetation projects, seed zones can create efficiencies and economies of scale in commercial markets and seed banking programs and partnerships (Erickson 2008). This will reduce the cost of native plant material production and use, as well as increase the availability of genetically appropriate plant materials. Despite the numerous benefits, genetic guidelines for plant material movement are lacking for many native grass, forb, and shrub species used in roadside revegetation and pollinator habitat enhancement activities. In these cases, generalized provisional seed zones (Figure 3-95) (Bower et al 2014) may be useful in guiding seed movement and sourcing plant materials. The generalized guidelines are based on climate data (winter minimum temperatures and aridity) and boundaries of the Environmental Protection Agency (EPA) Level III ecoregions (Section 3.3.3, see [Ecoregions and Seed Zones](#)) to delineate areas that are similar climatically yet differ ecologically. These provisional seed zones can be considered a starting point for guidelines for seed transfer and can be used in conjunction with appropriate species-specific information as well as local knowledge of microsite differences. Inset 3-7 and Inset 3-8 provide further details on the provisional seed zones, as well as other information and tools to assist designers in choosing appropriate plant materials for revegetation and pollinator plantings. Provisional seed zones have been incorporated into the ERA online tool discussed in Section 3.3.3 (see [Ecoregional Revegetation Application \(ERA\)](#)) as an independent map layer.



**Figure 3-95 / Provisional seed zones**

Provisional seed zones for native plants are unique climatically delineated areas (A and B) nested within EPA Level III Ecoregion boundaries (C). The provisional zones can be used to guide seed sourcing decisions when species-specific genetic information is lacking (Bowers et al 2014).

## Genetic Variation

Another important issue in selecting native plant materials is maintaining genetic variation in the populations established in revegetation work. This is especially important to ensure resiliency in the context of a rapidly changing climate. Plant populations that are genetically variable can adapt and respond to changing stresses and climates. Collection and propagation procedures, as well as agronomic and nursery production methodologies, need to conserve sufficient genetic diversity to enhance revegetation success and buffer against environmental stresses and changes in both the short and long term ([Section 5.3.1](#) and [Section 5.3.4](#)). Additionally, a sufficient number of unrelated seed parents can be included to ensure that inbreeding does not become a problem in the future. Both issues come down to numbers—the more plants that contribute to the new population, the more genetic variation will be captured and the lower the likelihood that relatives will mate (less inbreeding). A good practice is for managers to consider these criteria, whether they are buying plant materials or collecting their own. When procuring seed on the commercial market, designers can consider consulting with seed producers and distributors and other reputable sources, including government websites and published literature, to determine the most appropriate available plant materials for a project area. Factors to consider include seed source origins relative to the project site, plant development methodologies, and certification class. In many states, the newer native species releases are certified as “Source Identified, Pre-Varietal Releases”, which originate from natural stands, seed production areas, seed fields, or orchards where no selection or genetic manipulation of the parent population has been conducted. The [Association of Official Seed Certifying Agencies](#) (AOSCA) has published certification standards and guidelines for the certification of Pre-Varietal Germplasm releases, however not all state certifying agencies have adopted these plant release types within their respective state laws and regulations.

### **Inset 3-8 / Locally adapted plant materials**

Choosing the right plant materials for a project is fundamental to revegetation success, both in the short and long term. With inappropriate seed mixes, projects may fail outright (e.g., low germination or high seedling mortality) or lead to more cryptic problems in the future, such as poor regeneration potential, phenological asynchrony with dependent pollinators, genetic degradation of surrounding plant communities, and loss of resiliency and adaptive capacity in responding to new stresses (e.g., invasive plant species or climate change).

A large number of studies have shown that locally derived and genetically diverse plant sources are likely to be best adapted to prevailing climatic and environmental conditions (Hufford and Mazer 2003; Savolainen et al 2007; Johnson et al 2010). This means that in addition to matching species assemblages to a project site, designers understand and consider the seed source origin and genetic diversity of available plant materials to be successful (McKay et al 2005; Crémieux et al 2010; Mijnsbrugge et al 2010; Schröder and Prasse 2013). Moreover, federal and state agencies are increasingly suggesting or requiring the use of locally adapted and regionally appropriate native plant materials in revegetation work based on site characteristics and ecological setting (see Appendix 1 in Johnson et al 2010).

Because restoration with native plants is still relatively new in the United States, the supporting research, infrastructure, and plant material development programs are in the early stages of development. Genetic guidelines for determining what is local are often lacking for many native species of interest (Erickson 2008; Johnson et al 2010). As a consequence, native plants of inappropriate or unknown origin are being sold and planted, including some that may originate well outside of the area targeted for planting.

#### **Provisional Seed Zones**

Seed zones help identify where plant materials originated and how far they can be moved. Empirical seed zones for individual species are developed through field trials in which a large number of seed sources from a wide range of source environments are evaluated for important adaptive traits, such as growth rate and vegetative and reproductive phenology. By relating measured traits to climate or other environmental variables, researchers are able to create maps and delineate areas (seed zones) that are relatively homogenous with respect to adaptive genetic variation. The seed zones represent areas within which seed and plant materials can be transferred with little risk of maladaptation or other adverse consequences (Campbell 1986; Sorensen 1992; Rehfeldt 1994; Erickson 2004; St. Clair et al 2005).

Generalized or “provisional” seed zones (Bower et al 2014) have been developed for the continental United States using minimum temperature and aridity variables in combination with EPA Level III Ecoregions (Omernik 1987). The resulting map (Figure 3-95) captures much of the variation existing in adaptive seed zones (Bower et al 2014; Kramer et al 2015). Therefore, the combined generalized or “provisional” seed zone and ecoregion mapping approach is a good starting place for species and geographic areas where empirical seed zones are unavailable.

Increasing the provisional seed zones, temperature minimum values were grouped into 13 discrete classes that reflect the temperature bands used in the USDA plant hardiness zone map (USDA Agricultural Resource Service 2012). The hardiness map is familiar to designers and land managers and has been widely used for decades. An annual heat:moisture index (AH:M) was used as a measure of aridity to distinguish areas that are warm and wet (low-moderate aridity), cold and wet (low aridity), warm and dry (high aridity), and cold and dry (moderate-high aridity). Index values were divided into six discrete classes, with higher values indicating more arid environments. Intersection of the minimum winter temperature with the AH:M layer created unique climatically delineated (temperature-aridity) zones. In the final map, EPA Level III Ecoregions were overlaid on climate zones to identify areas that differ ecologically although they may be similar climatically (Figure 3-95).

The provisional seed zones, along with empirical seed zones for some native plants, are available online in the **Seed Zone Mapper application**. Provisional seed zones have also been incorporated into the **ERA online tool**, discussed in **Section 3-3-3 (see Ecoregional Revegetation Application (ERA))**, as an independent map layer.

### **Inset 3-9 / What to do if there are no locally adapted native seed sources available**

Adapted from Erickson et al 2003; Aubry et al 2005

The volume of seeds needed for a revegetation project may not always be available in sufficient quantities, particularly when plans have changed or the revegetation specialist has not been involved until the latter stages of the project. In these instances, three choices are available to the revegetation specialist:

- wait several years until the appropriate seeds are available;
- use introduced species that are non-persistent, non-invasive, or sterile, or
- use non-native cultivars or non-local native cultivars.

#### **Defer Seeding**

If the appropriate species or seed sources are not available, then consider not seeding until the appropriate seeds become available. In the interim, consider using soil cover for erosion control.

#### **Introduced Species**

When appropriate seed sources are unavailable, sterile hybrids or annual/biennial/perennial introduced plant species that are non-persistent and non-invasive may be considered. Preferred non-native species are those that will not aggressively compete with the naturally occurring native plant community, invade plant communities outside the project area, persist in the ecosystem over the long term, or exchange genetic material with local native plant species. Some of these species include sterile hybrids, such as Regreen (a wheat x wheatgrass sterile hybrid) and annuals such as common oat (*Avena sativa*) and common wheat (*Triticum aestivum*). Consider avoiding exotic species that have not already been introduced into the area, or that have been found to be aggressive and/or persistent. Also, consider avoiding non-native species that were commonly used in the past, such as Kentucky bluegrass (*Poa pratensis*), smooth brome (*Bromus inermis*), crested wheatgrass (*Agropyron cristatum*), orchardgrass (*Dactylis glomerata*), yellow and white sweetclover (*Melilotus officinalis* and *M. albus*), alsike clover (*Trifolium hybridum*), and alfalfa (*Medicago sativa*), among others. These species are generally no longer recommended due to their highly aggressive nature, resulting in widespread displacement of native species and plant communities that are low in diversity and poor pollinator habitat.

#### **Non-Local Native Species**

Native species that do not occur naturally in the local ecosystem, or native plant material that does not originate from genetically local sources, may be considered. These types of plant materials may include commercial cultivars. A cultivar is “a distinct, often intentionally bred subset of a species that will behave uniformly and predictably when grown in an environment to which it is adapted” (Aubry et al 2005). These cultivars are generally not preferable for wildland use due to concerns over adaptability, genetic diversity level, and the potential for genetic contamination, or “swamping,” of local native gene pools, including those of threatened, endangered, and sensitive plants (Millar and Libby 1989; Knapp and Rice 1994; Linhart 1995; Montalvo et al 1997; Lesica and Allendorf 1999; Hufford and Mazer 2003). Because commercial cultivars are typically selected for agronomic traits, such as high fecundity, vegetative vigor, and competitive ability, their use may also adversely impact resident plant populations through direct competition and displacement. Cultivars bred for traits such as showiness may have little value to pollinators due to low pollen and nectar production. Plant-pollinator relationships could be disrupted if the growth and reproductive cycle of non-locally sourced plants is different or out of sync with pollinator needs (Norcini et al 2001; Houseal and Smith 2000; Gustafson et al 2005). This is especially a concern with specialist pollinators that are reliant on the nectar and pollen from a small subset of plant species and synchronize their annual emergence to the flowering time of their host plants. Cultivars of native species (and introduced look-alikes such as sheep fescue [*Festuca ovina*]) can also be problematic if they are difficult to distinguish from native germplasm. This could severely complicate efforts to collect and propagate local material and waste valuable economic resources.

Because of these numerous concerns, consider using cultivars sparingly or not at all, with project objectives clearly understood. Consult with the seed producer or distributor before buying seeds and ask for the most appropriate cultivar for the project area, where the source for the cultivar was collected (geographic location and elevation), and how many collections were made. The seeds will likely be certified with a certification tag attached to each seed bag. Consider obtaining tests for seed germination, purity, noxious weeds, and seeds per pound.

Failing to consider genetic variation when selecting plant materials could have significant consequences on the viability and sustainability of revegetation efforts. Yet it is easy to imagine how variability can be eroded. If plants are propagated from a very small and inadequate sampling of the population, genetic variation of the propagated plants will be greatly reduced.

Reproductive strategies vary widely among species. No single collection and propagation protocol will ensure the genetic integrity of all types of plants used in revegetation. However, the issue of genetic variation cannot be ignored. Consider accounting for the following when purchasing or collecting native plant materials:

- **Number of related individuals**—Identifying which plants in a population are likely to be related can be difficult without expensive genetic analyses, but there are ways to minimize the collection of related individuals. In general, avoid collecting plants growing very close to each other to minimize the risk of collecting siblings or even clones of the same plant (Vekemans and Hardy 2004; Rhodes et al 2014). It is recommended to collect plants growing throughout the whole site to ensure that the full diversity of the site is captured, especially plants growing along the edges of each population.
- **Number of parents**—Collecting seed or cuttings from a minimum of 50 unrelated parent plants will help ensure that most of the genetic variation in a population is captured. Additional plant material would be needed if contribution by parents (of seeds or cuttings) is unequal. For dioecious species, attention to male-female ratios is essential to ensure adequate representation of both sexes.
- **Source sites (stands)**—To represent the population of a seed zone well, consider collecting seeds or cuttings from multiple sites within the zone. Considering sampling a similar number of parents from each site. Seek out larger communities to help meet parent selection criteria.
- **Individual parents within a selected source**—Individual maternal parents (seed plants) are to be well separated from each other yet not isolated from other plants of the same species. This will allow cross pollination by numerous paternal parents, adding to diversity. A similar amount of seeds are to be collected from each parent. Collecting from plants throughout the entire site will also promote sampling the full range of diversity that is present.

Section 5.11, *Obtaining Plant Materials* provides additional guidance on ensuring genetic diversity when collecting seed and cuttings. Guidelines have also been developed to help designers work with seed producers and nurseries that follow practices for maintaining high genetic variability throughout the native plant material production process (e.g., Basey et al 2015).

## 3.14 SELECT PLANT ESTABLISHMENT METHODS

After compiling a list of species and genetic sources to use for revegetation, the next steps are to determine the optimal propagation methods for each species and to identify the most appropriate plant material sources for a particular site or revegetation objective.

A useful tool for selecting the plant materials best suited to the project is the Target Plant Concept (Landis 2009). The Target Plant Concept (Figure 3-96) is an integrated sequential process for evaluating plant material requirements within the context of project objectives and site characteristics that may influence the suitability of seed sources or stocktypes, as well as the timing and optimal method of out-planting. The concept is based on the premise that there is no such thing as an “ideal” all-purpose seed mix, genetic source, or stocktype that will always work in any situation. Instead, the fitness of the plant material is determined by its appropriateness to the site in which it will be out-planted. Because every site is unique, seed mix and sourcing decisions can be tailored to each site or project to the extent possible. Otherwise, time and investments in site preparation, plant propagation, and outplanting may be wasted.

At this phase in the overall planning process, two of the steps in the Target Plant Concept have previously been covered: the objectives for establishing vegetation (Section 3.2) and the factors that possibly play the largest role in limiting plant survival and growth on each revegetation unit (Section 3.8). The remaining target plant requirements to consider are as follows:

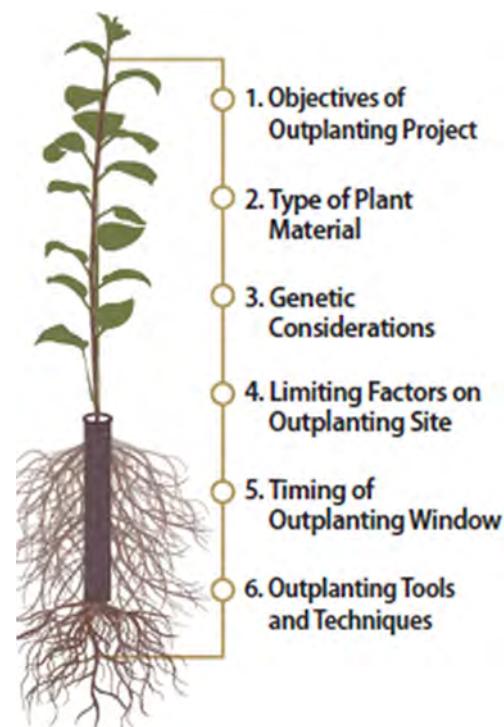
- Type of plant material to be used (seeds, cuttings, containerized or bareroot seedlings, or salvaged plants)
- Methods to be used to install plant materials and what post-installation plant care is appropriate
- Proper season for outplanting or seeding (the outplanting or seeding window)

Once species have been selected and genetically appropriate sources of plant materials have been identified, the next step is to determine the most appropriate plant materials for the project. Revegetating with native plants commonly involves multiple methods to reestablish vegetation on the project site. The following are possible considerations:

- Maximizing natural regeneration/recovery
- Salvage existing plants
- Direct seeding
- Outplanting nursery stock

### 3.14.1 SELECTING PLANT MATERIALS

In areas with relatively good soil stability that are bordered by healthy populations of viable native species, the existing seed bank and natural regeneration processes are key parts of re-establishing native vegetation on road sites. Minimizing the road footprint and damage from road construction are important aspects of any type of planning but are especially key if revegetation tactics involve supporting natural regeneration over more intensive revegetation interventions. In these cases, as long as topsoil is saved, the disturbance from road construction might serve to scarify the native



**Figure 3-96 / The Target Plant Concept**

The Target Plant Concept identifies six requirements for establishing native plants.

*Adapted from Landis 2009*

seed bank. Often the option of maximizing natural regeneration is not sufficient to fully revegetate a roadside environment; nevertheless, consider always anticipating the possibility. Salvaging and reapplying duff and litter layers to disturbed surfaces can aid in maximizing natural regeneration. The revegetation plan can acknowledge aspects of the revegetation process that are expected to develop naturally (Clewel et al, 2005).

However, if native seed regeneration is not sufficient to revegetate the site, additional plant materials will need to be obtained and established. Plant materials may include the following:

- Seeds
- Cuttings
- Plants

Determining which plant material to select for revegetation depends on the type of species being grown. For example, conifer trees have been shown to establish better and faster from plants than from seed or cuttings. Alternatively, grasses can be established from plants, but growing grass plants and planting them is very expensive compared to using seeds. Some species, however, do not produce reliable crops of seeds and therefore other plant materials, such as cuttings, will have to be used. Table 3-15 and Table 3-16 compare the advantages and disadvantages of different establishment methods and stocktypes. Various implementation guides provided in [Chapter 5](#) describe in more detail the process for obtaining plant materials.

## Seeds

Seeds are collected in the wild from native stands of grasses, forbs, shrubs, trees, and wetland plants. This plant material is used for seeding projects, such as hydroseeding of cut and fill slopes or other large areas of bare soil. Seeds of grass and forb species are best used for direct sowing, whereas seeds of shrubs and tree species are best used to grow nursery plants. If large amounts of grass or forb seeds are required for a project, seed collections can be increased through seed increase contracts. It can take up to three years to obtain enough seeds for a revegetation project—one year to collect the wild seeds and one to two years for seed increase. One of the advantages of direct seeding is that it can be an inexpensive method of reestablishing plants for a large area. Guides to collecting wild seeds, increasing seeds, and salvaging topsoil, duff, and litter are provided in [Chapter 5](#).

## Cuttings

Cuttings are taken from stems, roots, or other plant parts and directly planted on the project site or grown into rooted cuttings at a nursery for later outplanting. Only a few species, such as willow (*Salix* spp.) and cottonwood (*Populus* spp.), can be easily established from direct sticking of cuttings on a project site. Other species, such as quaking aspen (*Populus tremuloides*), can be established from cuttings in a controlled nursery environment but not in the field. Propagating plants from cuttings of most species is not possible under most growing conditions. Cuttings are collected in the wild in the winter and either stored or immediately planted on the project site. If large quantities of cuttings are required, they can be propagated by growing them in stooling beds for several years at a nursery or other growing facility. Guides to working with cuttings are presented in [Section 5.3](#) and [Section 5.4](#).

## Plants

Trees and shrubs are typically established using nursery stock rather than by direct seeding for several reasons. First, obtaining seeds from most tree and shrub species is expensive; in many years, they can be difficult to find or collect. Second, shrub and tree seeds germinate and grow into seedlings at a slower rate than grass and forb species, giving them a disadvantage on the sites where grasses and forbs are present. Starting shrubs and conifers from large plants instead of seeds gives them a competitive advantage over grasses and forbs because roots are often longer and better developed, allowing access to deeper soil moisture. Grass and forb species are seldom established from nursery-grown plants because of the high cost. Exceptions are when grass or forb seeds are rare or difficult to collect or increase (often referred to as “recalcitrant” species); if species are difficult to establish from seeds on disturbed sites; or when the project requires restoring threatened or sensitive species that are typically not considered workhorse species.

Plants are typically grown in a nursery or agricultural setting. However, for some projects, plants are salvaged from the construction site or adjacent areas. Sometimes salvaged plants are simply relocated quickly from one area to another. At other times, they may be transplanted into a nursery and replanted at a later time. Plants that are grown in a nursery need a lead time of one to two years from the time of ordering to availability. A variety of stocktypes are available from nurseries, including small to very large plants—plants in containers or those without soil around the roots (bareroot)—and plants grown in greenhouse environments or field-grown plants (Table 3-15). Selecting a stocktype will depend on the needs of the project, as there are multiple options for propagation and establishment, as well as many stocktypes to choose from.

**Table 3-15 | Comparison of plant material types for revegetation planning**

| Type   | Advantages   | Disadvantages  |
|--|--|--|
| <b>Balled-in-burlap</b><br><br>The plant is grown in the field, dug up with its roots and surrounding soil, and wrapped in a protective material such as burlap.   | Well-developed root systems increase chances of survival on site<br><br>Provide shade and earlier establishment of upper canopy on site  | Expensive<br><br>Large and heavy to transport  |
| <b>Bare-root</b><br><br>The plant is sold without any soil around its roots.   | Less expensive<br><br>Easier to transport to site; lightweight to carry around for planting<br><br>Roots have not been restricted by containers  | Requires care not to let root systems dry out before planting.<br><br>Difficult to establish in dry sites or sites with warm, sunny spring seasons.  |
| <b>Container</b><br><br>The plant is sold in a container of potting media or soil with drainage holes. Sizes and shapes of containers range from very small to very large.   | Well-established root systems with intact soil<br><br>Provide “instant” plants on site<br><br>Available in a variety of sizes; many are available year-round<br><br>Can be planted all year long   | Native soil not used in nursery; transplant shock may occur when roots try to move into native soil<br><br>Can be expensive<br><br>Can be difficult to transport to and around site if large numbers are used<br><br>Can be difficult to provide irrigation until established; may actually require more maintenance than plug |
| <b>Liners/Plugs</b><br><br>A small plant, rooted cutting, or seedling that is ready for transplanting. They are often used for herbaceous plants and grasses.  | Well-established root systems with intact soil.<br><br>Easy to transplant; plant material pops out of containers easily  | Same as above<br><br>Smaller plants may take longer to establish; may require more initial maintenance   |
| <b>Cuttings</b><br><br>A piece of branch, root, or leaf that is separated from a host plant and used to create a new plant. These may be placed in a rooting medium or stuck directly into the ground for planting.                        | Inexpensive to produce; cutting may easily be taken on site or from nearby site<br><br>Easy and light to transport; known to work well in rocky areas or areas difficult to access   | No established root systems<br><br>Timing of taking cuttings and planting them is important; varies among species and limited to dormant periods   |
| <b>Salvage</b><br><br>Native plants that are removed from a site (to a nursery, storage area, or directly to another field location) before ground disturbance at that site occurs. (Can also refer to salvaged cuttings or seed sources.) | Can use plant material that would otherwise be destroyed<br><br>Plant material could be local to site<br><br>Relatively inexpensive<br><br>Small or young salvage plants often adapt more readily to transplant than do mature specimens | Different native plants respond differently to being dug up; some loss could be expected<br><br>Requires fairly intensive measures to protect plants and ensure they have adequate irrigation  |

Adapted from Dorner 2002

## Potential disease and insect issues

Diseases can affect all native plant species used in revegetation, but assessing and mitigating for individual diseases is at times overlooked during revegetation projects. If a disease occurs, many times it is the result of trying to establish plant species in the wrong environment. For instance, species adapted to dry environments may be susceptible to certain diseases if planted in wet soils where a different set of root and other pathogens are present. While pathogens might be present on a site, this does not necessarily mean that plants will be affected. Much like humans, plants are always surrounded by a variety of pathogens, but it is not until they experience stress that they become more susceptible to diseases.

Planting appropriate species matched to sites where they have adapted to, using genetically appropriate stock, purchasing high quality plant materials from nurseries with good disease and insect management practices, improving the soil, and mitigating for climate extremes all play an essential role in creating healthy plants capable of resisting diseases or insect pests. If diseases are found on seedlings after planting, they may have previously become infected at the nursery. Appropriate sanitation procedures and practices a nursery might employ to help mitigate the risk of diseases include using only clean containers and potting media, starting with disease-free stock, and using only clean (non-recycled water). For more information on nursery practices that help prevent the spread of diseases or insect pests, the designer can find example guidelines in the [Native Revegetation Resource Library](#). There have been some cases when diseases originating in the nursery can spread to native plant populations with devastating effects. The recent spread of *Phytophthora ramorum* (the water mold pathogen that causes Sudden Oak Death, aka SOD) in California, Oregon and elsewhere serves as a dramatic example of the problems faced when pathogens are released into the landscape ([www.calphytos.org](http://www.calphytos.org)). Avoiding the purchase of damaged and infested stock and purchasing from growers that use best management practices can greatly reduce the risk of spreading diseases or insects.

A variety of insects can damage newly planted seedlings, as discussed briefly in this section. Because this is a specialized field, the designer is advised to consult technical or academic experts if insect problems are extensive on a revegetation project. Insect damage is grouped into four classes (Helgerson et al 1992), based on where insect feeding occurs:

- **Sap-suckers**—foliage
- **Root beetles**—root system
- **Terminal feeders**—terminal shoots
- **Secondary bark beetles**—stems

It may be difficult during the planning stages to determine if insects will be a limiting factor. However, designers can consult with entomologists to determine which insects might be known species of concern. Damage to seedlings by insects may be often overlooked because the injury usually occurs before the seedling shows visible signs of stress. By this time, the insect is no longer present, leaving only signs of previous activity. When dead or dying seedlings are discovered, it is important to systematically evaluate the seedling from the root system to the terminal bud for the presence of insects, insect damage, and diseases (discussed in the following section). If insects are found, consider collecting individuals for later identification. Designers can place insects in small glass or plastic containers until they can be examined by an entomologist. It is important to look under the leaves for aphids and scrape the entire

**For the Designer**  
Consult pathologists at extension offices, universities, or other agencies for expert input on disease issues.

**For the Designer**  
To prevent disease spread to native populations, designers should consider only working with nurseries that have practices to ensure only disease-free nursery stock is out-planted.

**For the Designer**  
Designers should consider consulting entomologists at extension offices, universities, or other agencies for expert input on insect issues.

seedling with a sharp knife to observe boring and tunneling. Damage caused by insects and diseases can occur under the bark of the stems and roots themselves. A hand lens is helpful when evaluating damage. The following discusses some of the most prominent insects that damage conifer seedlings. Non-conifer species will have their own unique associated pests. Nevertheless, classifying the insect into one of the four classes is a start in making a diagnosis.

- **Sap-Suckers**—Sap-suckers include the Cooley spruce gall adelgid (*Adelges cooleyi*) and the giant conifer aphids (*Cinara* spp.) that feed on succulent foliage of tree seedlings. While these can cause shoot deformity and foliage loss, aphids will normally not kill seedlings. The appearance of aphid ants (which cultivate aphid populations) on the leaves are indications that aphids are present.
- **Root Beetles**—The root bark beetles (*Hylastes* spp.) and root-collar weevil (*Steremnius carinatus*) girdle the roots and stems of conifer seedlings and will weaken or kill newly planted seedlings. The damage can be mistaken for herbivory by small mammals, but the lack of teeth marks and the below-ground location of the damage are indications that the damage is caused by root beetles.
- **Terminal Feeders**—The larvae of this group of insects feed on the terminal shoots of young conifers, killing much of the new growth. Continued annual attack by these insects can severely stunt conifer seedlings. The major insects include white pine weevil (*Pissodes strobi* [Peck]), ponderosa pine tip moth (*Rhyacionia zozana*), western pine shoot borer (*Eucosma sonomana*), and the cone worm (*Dioryctria* spp.).
- **Secondary Bark Beetles**—The Douglas-fir engraver beetle (*Scolytus unispinosus*) attacks the stems of stressed seedlings, creating galleries under the bark that weaken or kill Douglas-fir seedlings.

Some examples of ways to mitigate for insects include:

- **Plant a Variety of Species**—Insects are often host-specific, meaning they attack only one species. For this reason, a preventative measure is to plant a variety of species. If an insect infestation occurs, it may not affect all the seedlings planted.
- **Plant Healthy Stock**—Insects often attack weakened seedlings or seedlings that are stressed. Planting only healthy and vigorous seedlings, appropriate to the site, reduces the potential damage by insects.
- **Create a Healthy Soil Environment**—Seedlings grown on poor sites, or on sites outside of the species' environmental ranges, will be stressed and become susceptible to insect damage. Planting seedlings on optimal growing sites will produce healthy seedlings resistant to insect damage. Judicious use of fertilizers (that is, avoid over-fertilization) is also important for pest prevention. For example, sucking insects often attack plants that have been over-fertilized with nitrogen.
- **Install Bud Caps**—Some terminal feeders can be controlled through the use of bud caps that are placed over the terminal in the spring prior to shoot growth (Goheen 2005). Bud caps are materials made out of paper or fine cloth that temporarily cover the terminal and prevent adult insects from laying eggs on the bud.

For any stocktype, it is important to define various desired characteristics, including

age, size, likelihood of survival on the site, ability to compete with other vegetation and/or tolerate animal effects, and methods that will be necessary to out-plant and maintain the stocktype. For bareroot stocktypes, consider size, age, and potential survival rates. For cuttings, consider if the project requires containerized cuttings (with a root system in a container), heeled or bareroot cuttings (with roots but no container), or simply cuttings to stick directly in the ground on the project site. For cuttings, length, caliper, and conditions of storage are also important factors. For container plants, no standard terminology exists for describing the different types of container plants available (Landis et al 1992). They are usually described by their container type, referring to the volume and usually the shape of the container. Size and age of the plant are also described. When ordering container plants, consider age, caliper, height, and root size and depth, as well as any other special characteristics that might help the plant survive on the project site (Table 3-15).

Salvaging plants from the project area can be an important component of protecting native plant diversity on the project site. Sometimes salvaged plants are simply relocated instantly from one area to another. Other times, they may be transplanted or moved to a nursery area, cared for, and then re-planted at an appropriate time. As much native soil as possible is included when digging the salvaged plants, as this soil not only supports the health of the plants but also contains the native seedbank and root fragments of it and adjacent plants. “Salvage” may also involve harvesting cuttings or seeds from plants that are going to be removed during the road construction process.

Depending on the species, genetics, site limiting factors, and specific project objectives, some concept of the appropriate “target” stocktypes will already be defined. For example, some plants respond well to certain propagation methods; some will be salvaged (“wildlings”) from the site; and some are obvious candidates for direct seedling applications in order to facilitate fast regeneration. The needs and characteristics of a particular species will help determine if direct seeding, nursery propagation, or other methods are the more appropriate strategy. Many options can be considered, some of which are summarized in Table 3-16.

**Table 3-16 / Comparison of different plant establishment methods**

| Characteristic                                   | Wild | Cuttings | Seeds | Nursery plants |
|--|------|----------|-------|----------------|
| <b>Efficient use of seeds and cuttings</b>       | N/A  | No       | No    | Yes            |
| <b>Cost of establishment</b>                     | High | Moderate | Low   | Moderate       |
| <b>Ability to establish difficult species</b>    | Yes  | No       | No    | Yes            |
| <b>Option of using specific genotypes</b>        | No   | No       | Yes   | Yes            |
| <b>Precise scheduling of plant establishment</b> | Yes  | Yes      | No    | Yes            |
| <b>Control of stand composition and density</b>  | Yes  | Yes      | No    | Yes            |
| <b>Matching stocktypes to site conditions</b>    | No   | No       | No    | Yes            |
| <b>Depletion of adjacent plant stands</b>        | Yes  | Yes      | No    | No             |

### **For the Designer**

**Quality plant and seed availability at your desired planting time depends on early identification of plant material needs and purchase coordination with the Client and Contractor.**



**Figure 3-97 / Early planning for plant material procurement**

Early planning for plant material procurement is essential. Depending on plant species, stocktype, and quantity desired it can take up to three years or more to obtain sufficient plant material. Due to the biology of plants, a missed window of collection may result in a full year’s delay. Outplanting may occur across multiple growing seasons, requiring several delivery and installation events spanning a timeframe of years.

## Scheduling

Timing is a key factor for obtaining plant materials because many native plants or seeds are not widely available from nurseries and seed companies. Even if a species is available, the odds are that it is not from a local source that is genetically adapted to the project site. Therefore, it is essential to identify plant material needs early. It usually takes a lead time of two to four years to administer seed collection, seed increase, and seedling propagation contracts. Sometimes three years may be required to achieve sufficient quantities of seeds. Plants (seedlings, cuttings, and so on) ordered from nurseries take a great deal of advanced planning for both seed collection and for plant propagation. Failure to realize the lead time necessary for seed collection and propagation of appropriate native plant materials is one of the most common mistakes made in revegetation projects (Figure 3-97).

## Select Installation and Maintenance Methods

Many methods are available for installing plant materials on a project site. Seeds can be applied through hydroseeding equipment, disked or drilled into the soil surface, broadcast sown, imprinted, and/or covered with a variety of types of mulch. A variety of techniques are also available for installing cuttings and plants, including an expandable stinger (Figure 5-118), waterjet stinger, pot planter, auger, and a shovel. These methods are discussed in [Chapter 5](#).

Care is necessary after plant materials have been installed to ensure they become well-established. This includes protection from browsing animals, high temperatures, winds, competing vegetation, and drought. Measures that can mitigate for browsing animals include installing netting and tree shelters, over-planting to compensate for expected browse, as well as applying animal repellents. If netting or tree shelters are used, it is important to account for their removal in the implementation budget and contract. Seedlings can be protected from high temperatures and wind with shade cards, tree shelters, and large obstacles, such as logs. Competing vegetation is controlled through weeding, mulch, or herbicides. In extremely dry conditions, soil moisture can be supplemented by using irrigation systems.

### 3.14.2 DETERMINE OUTPLANTING WINDOWS

In addition to ensuring enough lead time to successfully carry out native revegetation goals, it is necessary to determine the optimum seasons for planting. There are advantages and disadvantages to carrying out operations at any time of the year, and determining the timeframe is based on the species, plant material, and site factors.

The optimum time of year to outplant for greatest plant survival is called the outplanting window. It is determined by graphing the annual precipitation, temperatures, and snow accumulation of a site. This information can be obtained from climate stations, as described in Phase One. Outplanting windows are also defined by the species, stocktype, and outplanting methods. For example, two different species of bunch grass might have very different survival rates depending on when they were outplanted. Figure 3-98 shows survival rates for bluebunch wheatgrass (*Pseudoroegneria spicata*), that survived better when planted in fall, and spreading needlegrass (*Achnatherum richardsonii*), that survived better when planted in spring.

Several factors are important for determining outplanting windows, including soil temperatures in late winter and early spring, precipitation during spring and early summer, soil temperatures in fall, precipitation in fall, and snow cover. Three examples of identifying outplanting windows are provided in Figure 3-99, Figure 3-100, and Figure 3-102. Applying the following set of guidelines to the climate data will help

clarify the optimum times to plant.

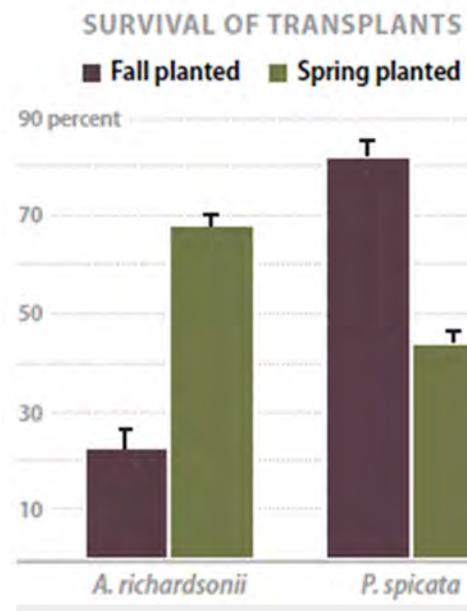
## Soil Temperatures in Late Winter and Spring

Seedlings have to develop new roots soon after they are planted to become established and survive the hot (and in many regions, dry) summer of the first growing season. For most species, new root growth occurs when soil temperatures exceed 42° F. Waiting for soil temperatures to warm to 42°F in the spring before outplanting, however, will shorten the period when soil moisture is available to the seedling. For this reason, a common restoration strategy is to outplant seedlings as soon after the last threat of a deep frost in the winter has passed, even if the soil temperatures are below 42° F.

## Precipitation in Spring and Summer

Much of the western United States experiences dry summers, with winters receiving most of the precipitation. Under this regime, soils lose their moisture in the summer and recharge from storms that occur in the fall and winter. Plants have adapted to this climate by growing new roots and leaves during spring, when soil moisture is high, and becoming dormant in summer, when soil moisture is depleted. The most critical factor for seedlings that are outplanted in the spring is the length of time available for the seedling to develop new roots before soil moisture is depleted. Sites with little or no precipitation in the spring or summer will dry out faster than sites with high spring and summer precipitation. A guideline is that an average monthly precipitation of less than

1.5 inches will do little to recharge a dry soil during late spring through fall. Figure 3-99, Figure 3-100, and Figure 3-101 show spring outplanting windows for three very different project sites. On low elevation sites west of the Cascade Mountains, the planting window can be fall and spring (Figure 3-99), whereas high elevation sites might have a planting window only in late spring (Figure 3-101).



**Figure 3-98 | Survivorship can vary between spring and fall plantings**

Outplanting is to be timed properly for best results given site conditions and species requirements

*Reproduced from Page and Bork 2005*

# 4—Revegetation Plan Example

## PREFACE

The following sample Revegetation Plan (the Plan) was written for an FHWA project that is currently in the planning stage. The Plan highlights actions to be taken by the USDA Forest Service Region 6 Restoration Services Team in association with FHWA. The Plan was previously submitted to FHWA and will be used in the permitting process by the Contractor and Contracting Officer's Representative during the implementation phase and will be referred to during the monitoring phase. Revegetation Plans are unique to each project. This sample plan is provided here as a stand-alone document in the larger manual and is meant only as a reference from which to develop a project-specific Revegetation Plan.

This Plan is unique to the Restoration Services Team in that it is the first to intentionally include elements in the planning, implementation, and monitoring phases with the specific goal of increasing plant-pollinator interactions. Areas that would support pollinator insects with moderate effort and alteration were identified as distinct revegetation units during the planning phase. Plant species lists were developed during the planning phase to include plants known to increase nectar availability and to provide nesting, breeding, and refuge benefits. Plant materials for these species were collected for seed increase and grow-out. Minor grading to create depressions for water retention, large woody debris placement to create microhabitats, and out-planting will occur during the implementation phase in an effort to increase pollinator habitat. Pollinator abundance and diversity information will be collected during the monitoring phase to better understand practices that increased pollinator use within the project area and to identify opportunities for improving these approaches on future projects.



FHWA's Historic Columbia River Highway State Trail project has a representative revegetation plan for the planning stage.

*Photo credit: Matt Horning, USFS*

Restoration Services Team- R6 USDA Forest Service  
63095 Deschutes Market Rd, Bend, OR 97701



---

# HISTORIC COLUMBIA RIVER HIGHWAY STATE TRAIL: SEGMENTS A-C FINAL REVEGETATION PLAN

June 2016

*Prepared for:*  
*Federal Highway Administration*  
***Western Federal Lands Highway Division***  
*610 East Fifth Street, Vancouver, WA 98661*

## TABLE OF CONTENTS

|   |    |
|---|----|
| Background_____   | 3  |
| Revegetation Plan_____  | 8  |
| Strategy_____   | 8  |
| Plant Species of Special Interest_____                          | 8  |
| Wyeth Trailhead and Parking Lot_____                            | 9  |
| Wyeth Campground Entrance and Gorton Creek Bridge_____          | 12 |
| Mitigation Site #2_____   | 14 |
| Stepped, Plantable MSE Wall_____                                | 17 |
| Summit Creek Viaduct_____                                       | 18 |
| HCRHST Embankments (Cuts and Fills) and Planned Idle Spots_____ | 19 |
| Staging Area(s)_____  | 21 |
| Non-Native Plant Control_____                                   | 22 |
| Monitoring Plan_____  | 23 |
| Success Criteria_____   | 24 |
| Approximate Timeline & Budget_____                              | 25 |
| Appendix One- Plant Species Suite_____                          | 31 |
| Appendix Two- Walker Macy Planting Plans_____                   | 32 |
| Appendix Three- Oregon Noxious Weed List_____                   | 41 |
| References_____   | 44 |

Prepared by

USDA Forest Service Region 6 Restoration Services Team  
63095 Deschutes Market Rd, Bend, OR 97701



Telephone: (541) 588-2695

Submitted June 8, 2016

## BACKGROUND

### Background

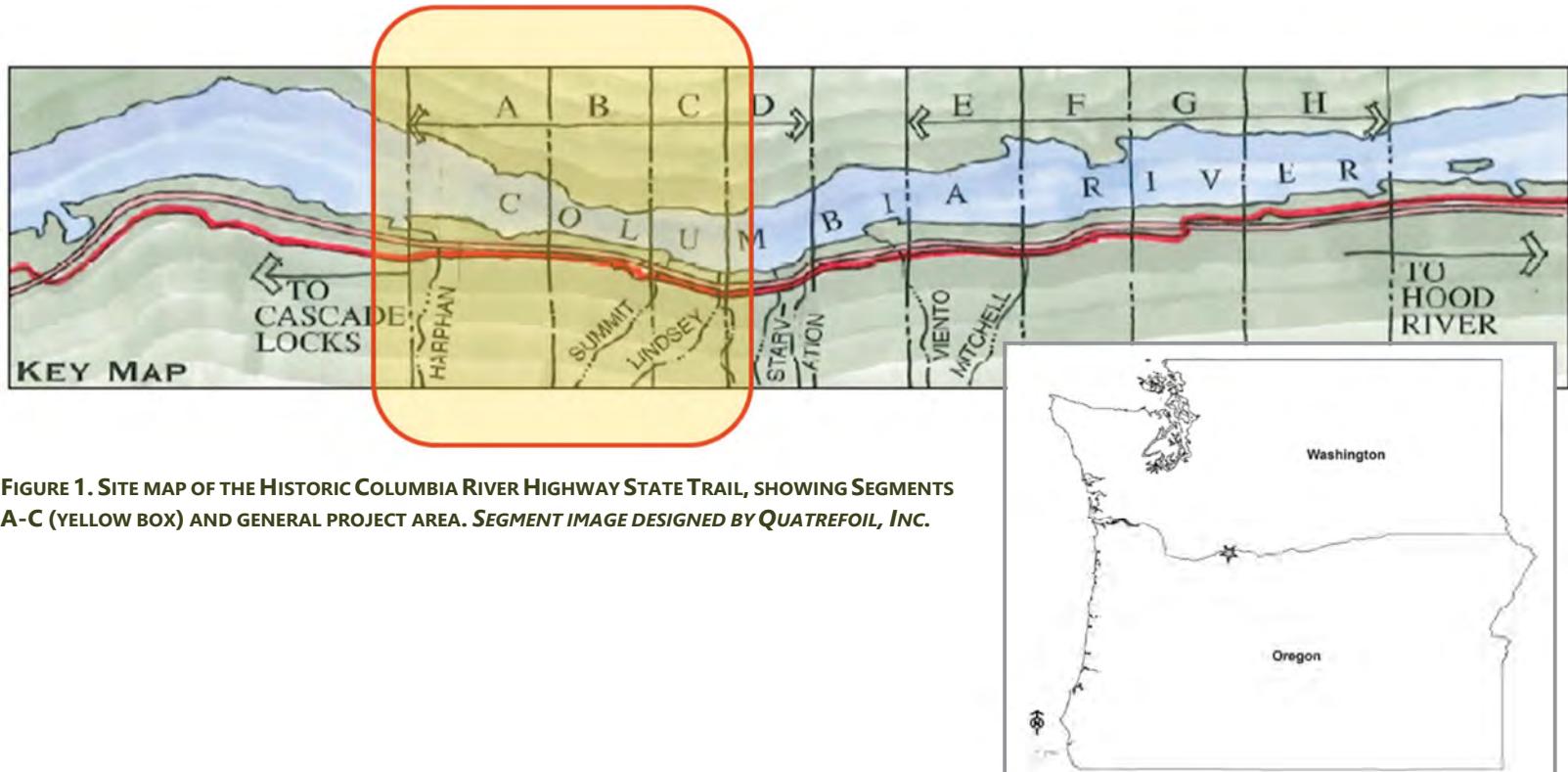
The Historic Columbia River Highway (HCRH), originally constructed between 1913 and 1922, took full advantage of the unparalleled beauty of the Columbia River Gorge. The highway dazzled tourists and locals alike as it meandered near breathtaking waterfalls, panoramic vistas, and spell binding geomorphology. Many of these scenic features such as nearly thirty named waterfalls, including Multnomah Falls and Hole-in-the-Walls, and the Crown Point Vista House still delight travelers today. The construction of Interstate 84 during the late 1940s and early 1950s disrupted the HCRH, leaving much of it in fragmented segments.

Directive was given to the State of Oregon in 1986 via the Columbia River Gorge National Scenic Area Act, to reconnect the fragments of the historic highway. Further direction was provided by the Oregon Legislature in 1987 to the Oregon Department of Transportation (ODOT) to facilitate the development of the Historic Columbia River Highway State Trail (hereafter HCRHST) by preserving and enhancing existing HCRH segments. Since then, multiple partner agencies including the Federal Highway Administration-Western Federal Lands Highway Division (WFLHD), ODOT, Oregon Parks and Recreation Department (OPRD), the U.S. Forest Service (USFS), the State Historic Preservation Office, and private entities have collaborated to reconnect the HCRH fragments. Sixty-two of 73 miles are currently open along the HCRHST, in the form of drivable motor vehicle roads and foot or bicycle paths. There are approximately eleven miles remaining of the original HCRH for which plans are in place to reconnect and incorporate into the HCRHST.

This document specifically addresses revegetation efforts planned by the U.S. Forest Service Region 6 Restoration Services Team (RST) within Segments A through C of the HCRHST. These segments cover from approximately one half mile west of the Wyeth Campground (west end of Segment A) to Lindsey Creek at the eastern end of Segment C, slightly less than ten miles west of the town of Hood River, Oregon (Figure 1). The combined length of Segments A-C is 3.08 miles and spans from station 499+89.18 at the west, to station 151+73.53 to the east.

Plans for Segments A-C call for developing a parking lot and trailhead on the west end, a mitigation area of approximately 5 acres, and new interpretive resting spots along the trail. Revegetation efforts will be closely coordinated with Walker Macy, a Portland Landscape Design firm.

## BACKGROUND



**FIGURE 1. SITE MAP OF THE HISTORIC COLUMBIA RIVER HIGHWAY STATE TRAIL, SHOWING SEGMENTS A-C (YELLOW BOX) AND GENERAL PROJECT AREA. SEGMENT IMAGE DESIGNED BY QUATREFOIL, INC.**

## BACKGROUND

### *Past Use*

The project area of the HCRHST has a long history of human utilization. Evidence including remnants of poured concrete foundations, door stoops, invasive plants, imported fill material, trails and roads are found throughout the project area. In addition, creek channels have been re-routed and straightened, culverts have been installed, and surface water flow patterns have been altered significantly. Many of the hardscaped artifacts will be preserved in place, while non-native vegetation will be eliminated.

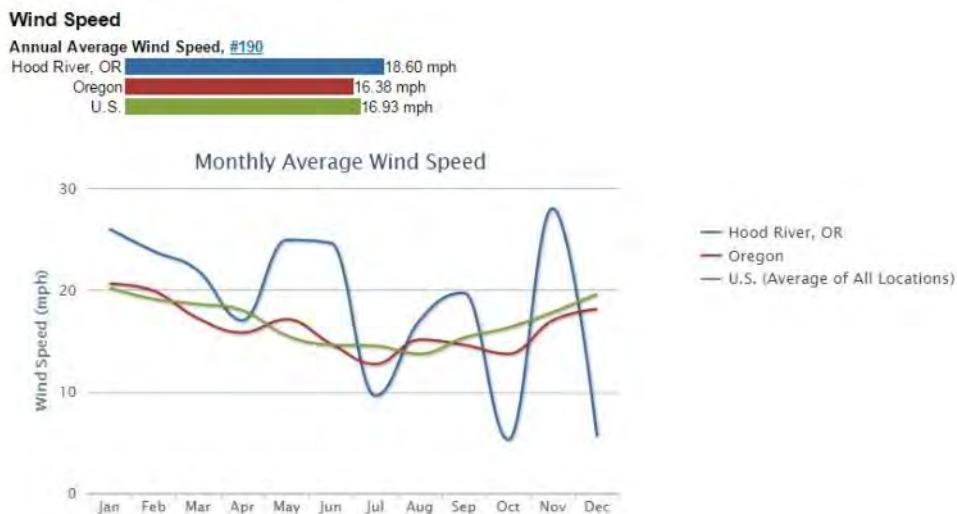
### *Soils*

Native soils in the majority of the project area are of the Wyeth Series. This soil type is well drained and consists of very cobbly loam, having formed from basalt colluvium. The project area experiences rapid runoff and moderate permeability.

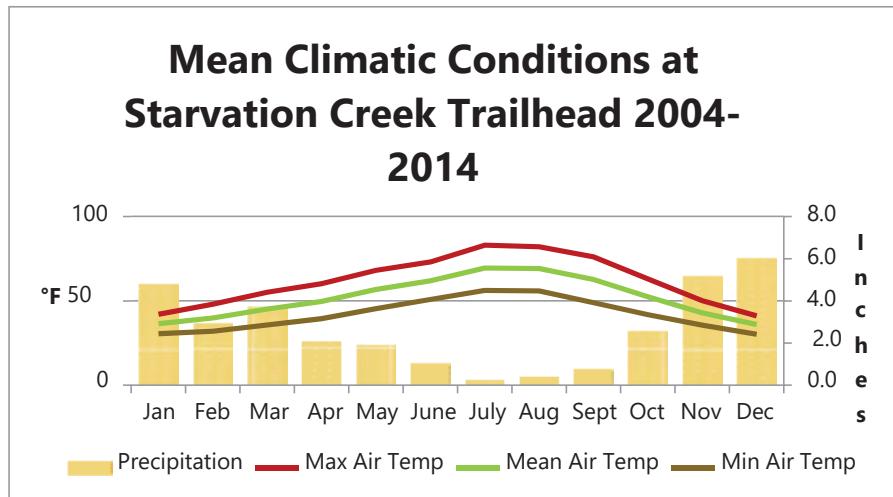
### *Climate*

The elevation is about 120 feet above mean sea level. The area is influenced by winds from the Columbia River Gorge (Fig. 2) and on average, the greatest precipitation occurs during the month of December with six inches (Fig. 3). The average growing season in nearby Hood River, Oregon occurs from approximately late April through mid- to late October, with about 183 frost free days. The maximum average temperatures are around 83°F in July, with the lowest minimums being around 30°F in December. Annual average humidity is highest in December at 84%, and lowest in July at 51% (2004-2014).

## BACKGROUND



**FIGURE 2. AVERAGE MONTHLY WIND SPEEDS IN HOOD RIVER, OREGON. DATA ARE AVERAGED FROM 1980-2010.**  
SOURCE: USA.COM HISTORIC WEATHER DATA



**FIGURE 3. THE AVERAGED MAXIMUM, MINIMUM, MEAN AIR TEMPERATURE AS WELL AS PRECIPITATION AT STARVATION CREEK TRAILHEAD. DATA ARE AVERAGED ACROSS 2004-2014. NOTE THE TWO DIFFERENT AXES.**  
SOURCE: PRISM TIME SERIES DATA FOR LATITUDE 45.7055N/LONGITUDE -121.5855W

## BACKGROUND

### *Trail Construction*

The trail is planned to be 12 feet wide, with two foot shoulders on either side. In some areas of constriction the trail width will be reduced to 10 feet. One exception is the planned pedestrian viaduct near Summit Creek that will be 14 feet wide. Grades will be limited to approximately five percent and when able, the trail surface will be sloped to the north. There will also be an additional five foot construction allowance on either side of the trail for temporary work access. In addition to the trail itself, plans call for a new trailhead with parking lot at the western terminus, a new motorist/pedestrian bridge over Gorton Creek, a pollinator meadow and mitigation area, a pedestrian viaduct near Summit Creek, and two new interpretive idle spots along the trail. Revegetation efforts will be proportional to the need for restoration and public visibility of each location.

# REVEGETATION PLAN

## Revegetation Plan

### Strategy

Locally sourced, genetically adapted plant materials will be utilized for this project. The canopy within segments A-C is quite varied; ranging from open meadow to partial forest canopy to dense woods. The mitigation site near the historic Wyeth trailhead is a mostly open area, surrounded by trees. However, there is a large infestation of invasive weeds (namely Himalayan blackberry (*Rubus armeniacus*) and Scotch broom (*Cytisus scoparius*)) that occupy greater than 40% cover. Once the Himalayan blackberry and Scotch broom are treated there will be very little to no canopy cover over the majority of the almost five acre site. This location is well suited to be left as an open meadow and as such has been designated as an area where we will focus revegetation efforts on creating a pollinator-supporting habitat (see Mitigation Site #2 discussion below). This emphasis on pollinator habitat for this area specifically addresses action items in the National Pollinator Strategy and the National Native Seed Strategy, in addition to other FHWA/WFLHD and Forest Service strategies.

Appendix 1 shows the suite of species that are planned for revegetation efforts. The plant species suite represents native plants that are present at the project site, at adjacent reference sites, or for which there are documented records of them having once been native and present but which have been outcompeted by introduced species. The suite is broad and not all species will be utilized at all sites. The most common container size used will be D27s. These containers are 2.5 inches wide by 7 inches long, and have a capacity of 27 cubic inches. This size pot accommodates a wide range of shrub and tree species and has proven reliable under similar out-planting conditions. Grass and forb seed will either be hydroseeded or hand broadcasted.

The planting goal of RST is to: 1) create good line-of-sight and facilitate public safety (as designed in Walker Macy planting plans); 2) create a natural appearance that is consistent with the surrounding environs; and 3) create habitat to support pollinators. RST will draw on the planting notes and design of Walker Macy (Appendix 2) but will incorporate multiple native species of the designated height classes into each planting area and blur the boundaries of the planting areas to create a random and natural appearing pollinator-friendly habitat.

### Plant Species of Special Interest

There are no known threatened, endangered, or sensitive plants within the area to be directly affected by this project (Robin Dobson, *pers. comm.*). RST is aware of only one observation of

## REVEGETATION PLAN

a sensitive plant species within the project vicinity, which occurs in a plant buffer of Segment B near station 105+50 (CH2M Hill 2014). This occurrence of long-bearded hawkweed (*Hieracium longiberbe*) is not expected to be affected. Although there are no known plants of concern within the area to be directly affected there are additional areas of potential habitat for long bearded hawkweed and Columbia kittentails (*Synthyris missurica* ssp. *stellata*). If RST Restoration Specialists observe a species of concern they will salvage plants and care for them ex situ at Dorena Genetic Resource Center, a USFS plant center in Cottage Grove, Oregon, or adjacent to the affected area in similar habitat (on or near the project site). The plants would then be resituated in their original location, or the nearest suitable location if the original is no longer available, once the potential for disturbance has passed.

### Wyeth Trailhead and Parking Lot

The planned parking lot at the Wyeth Trailhead incorporates a number of planting locations that encompass approximately one acre in area, including a drainage catch basin in the center and nine to twelve distinct planting pockets integrated into the outer curb. This area, more so than any other along the project, is designed with an eye toward more traditional landscape and horticultural practices. RST will work closely with Walker Macy to maintain public safety and line-of-sight while creating multi-seasonal interest.

RST will conduct weed control throughout the area prior to planting, especially near the entrance. There is currently a robust population of Scotch broom growing along Wyeth Road and continuing beyond the ownership boundary to the west. Although it would be ideal to treat the larger surrounding area (approximately four acres) to reduce future Scotch broom re-establishment, funding for this work is not currently available. Therefore, only the area referenced on the project plan sheets will receive treatment; less than one tenth of an acre. Once treated, the open area on the west side of the trailhead entrance will be planted with species that support pollinator-plant interactions (Table 1a). Efforts will be made to maintain visuals of the trailhead sign and entrance while blocking the view of the new vault toilet facility from the traveling public on Wyeth Road.

Contracting responsibility will be coordinated between WFLHD and RST. WFLHD will place 18" of weed-free topsoil or borrow material into all planting areas other than the planting pockets. The RST contactor will be responsible for providing weed-free topsoil to be combined with compost for use in the planting pockets and the two large planting beds on either side of the parking lot entrance. WFLHD will remove the non-native black locust trees currently on site.

# REVEGETATION PLAN

Table 1b depicts tree and shrub species to be incorporated into the parking lot landscape plan. Native trees will be installed in the curb planting pockets and will be approximately 1-2" caliper and 4-6' tall.

**TABLE 1A. PLANTS TO POTENTIALLY BE USED IN REVEGETATION EFFORTS NEAR THE ENTRANCE TO THE WYETH TRAILHEAD PARKING LOT. 'XX' DENOTES SEEDING RATHER THAN TRANSPLANTING TO BE UTILIZED. STOCK TYPE DEFINITIONS ARE AS FOLLOW: 'S' SEED; 'RL10' 10 CUBIC INCH CONTAINER VOLUME; 'D27' 27 CUBIC INCH CONTAINER VOLUME; 'D40' 40 CUBIC INCH CONTAINER VOLUME; '#1' ONE GALLON CONTAINER; '#5' FIVE GALLON CONTAINER; 'B&B' BALLED AND BURLAPPED.**

| Common                | Botanical   | Zone | Stock Type | Spacing<br>(on center) |
|-----------------------|---|------|------------|------------------------|
| common yarrow         | <i>Achillea millefolium</i> var.<br><i>occidentalis</i> | A    | S          | XX                     |
| pearly everlasting    | <i>Anaphalis margaritacea</i>                           | A    | S          | XX                     |
| narrow-leaf milkweed  | <i>Asclepias fascicularis</i>                           | A    | S, RL10    | 4"                     |
| showy milkweed        | <i>Asclepias speciosa</i>                               | A    | S, RL10    | 4"                     |
| aster                 | <i>Aster spp.</i>                                       | A    | S          | XX                     |
| mountain brome        | <i>Bromus carinatus</i>                                 | A    | S          | XX                     |
| Howell's reedgrass    | <i>Calamagrostis howellii</i>                           | A    | S          | XX                     |
| Pacific dogwood       | <i>Cornus nuttallii</i>                                 | B    | #1         | 15'                    |
| blue wildrye          | <i>Elymus glaucus</i> ssp. <i>glaucus</i>               | A    | S          | XX                     |
| smooth alumroot       | <i>Heuchera glabra</i>                                  | A    | S          | 4"                     |
| lupine                | <i>Lupinus latifolius</i>                               | A    | S          | 4"                     |
| creeping Oregon grape | <i>Mahonia repens</i>                                   | A    | D27        | 4'                     |
| Sandberg's bluegrass  | <i>Poa secunda</i>                                      | A    | S          | XX                     |
| self heal             | <i>Prunella vulgaris</i>                                | A    | S          | XX                     |
| checkermallow         | <i>Sidalcea spp.</i>                                    | A    | RL10       | XX                     |
| goldenrod             | <i>Solidago canadensis</i>                              | A    | S          | XX                     |
| fringecups            | <i>Tellima grandiflora</i>                              | A    | RL10       | 4"                     |
| western serviceberry  | <i>Amelanchier alnifolia</i>                            | B    | #1, #5     | 8'                     |
| Pacific dogwood       | <i>Cornus nuttallii</i>                                 | B    | #1         | 15'                    |
| oceanspray            | <i>Holodiscus discolor</i>                              | B    | D40        | 8'                     |
| Oregon grape          | <i>Mahonia aquifolium</i>                               | B    | D27, D40   | 4'                     |
| Douglas fir           | <i>Pseudotsuga menziesii</i>                            | B    | D27, B&B   | 15'                    |
| red flowering currant | <i>Ribes sanguineum</i>                                 | B    | #1, #5     | 4'                     |
| baldhip rose          | <i>Rosa gymnocarpa</i>                                  | B    | D27, B&B   | 4'                     |
| common snowberry      | <i>Symphoricarpos albus</i>                             | B    | D27, D40   | 4'                     |
| trailing snowberry    | <i>Symphoricarpos mollis</i>                            | B    | D27, D40   | 4'                     |
| western red cedar     | <i>Thuja plicata</i>                                    | B    | #1, B&B    | 15'                    |

# REVEGETATION PLAN

**TABLE 1B. PLANTS TO BE USED IN THE WYETH TRAILHEAD PARKING LOT, INCLUDING THE DRAINAGE BASIN. 'XX' DENOTES SEEDING RATHER THAN TRANSPLANTING TO BE UTILIZED. STOCK TYPE DEFINITIONS ARE AS FOLLOW: 'S' SEED; 'RL10' 10 CUBIC INCH CONTAINER VOLUME; 'D27' 27 CUBIC INCH CONTAINER VOLUME; 'D40' 40 CUBIC INCH CONTAINER VOLUME; '#1' ONE GALLON CONTAINER; '#5' FIVE GALLON CONTAINER; 'B&B' BALLED AND BURLAPPED.**

| Common                | Botanical   | Zone | Stock Type  | Spacing<br>(on center) |
|-----------------------|---|------|-------------|------------------------|
| common yarrow         | <i>Achillea millefolium</i> var.<br><i>occidentalis</i> | A    | S           | XX                     |
| narrow-leaf milkweed  | <i>Asclepias fascicularis</i>                           | A    | S, RL10     | 4"                     |
| showy milkweed        | <i>Asclepias speciosa</i>                               | A    | S, RL10     | 4"                     |
| creeping Oregon grape | <i>Mahonia repens</i>                                   | A    | D27, D40    | 4'                     |
| western sword fern    | <i>Polystichum munitum</i>                              | A    | Short 4, #1 | 4'                     |
| fringecups            | <i>Tellima grandiflora</i>                              | A    | RL10        | 4"                     |
| grand fir             | <i>Abies grandis</i>                                    | B    | #1, B&B     | 15'                    |
| vine maple            | <i>Acer circinatum</i>                                  | B    | #1, #5      | 8'                     |
| western serviceberry  | <i>Amelanchier alnifolia</i>                            | B    | #1, #5      | 8'                     |
| redosier dogwood      | <i>Cornus sericea</i> spp. <i>sericea</i>               | B    | D27, D40    | 8'                     |
| Oregon ash            | <i>Fraxinus latifolia</i>                               | B    | #1, #5      | 15'                    |
| oceanspray            | <i>Holodiscus discolor</i>                              | B    | D40         | 8'                     |
| Oregon grape          | <i>Mahonia aquifolium</i>                               | B    | D27, D40    | 4'                     |
| osoberry              | <i>Oemleria cerasiformis</i>                            | B    | #1, #5      | 8'                     |
| ninebark              | <i>Physocarpus capitatus</i>                            | B    | #1, #5      | 8'                     |
| Douglas fir           | <i>Pseudotsuga menziesii</i>                            | B    | D-27, B&B   | 15'                    |
| red flowering currant | <i>Ribes sanguineum</i>                                 | B    | #1, #5      | 4'                     |
| thimbleberry          | <i>Rubus parviflorus</i>                                | B    | D27, D40    | 4'                     |
| elderberry            | <i>Sambucus nigra</i>                                   | B    | D27         | 8'                     |
| common snowberry      | <i>Symphoricarpos albus</i>                             | B    | D27, D40    | 4'                     |
| trailing snowberry    | <i>Symphoricarpos mollis</i>                            | B    | D27, D40    | 4'                     |
| western red cedar     | <i>Thuja plicata</i>                                    | B    | #1, B&B     | 15'                    |

## REVEGETATION PLAN

### Wyeth Campground Entrance and Gorton Creek Bridge

The trail will cross the existing entrance to the Wyeth Campground and continue toward the east over the newly constructed Gorton Creek Bridge. WFLHD will install a water filling station near the Wyeth Campground entrance, adjacent to the grassy campground overflow area. They will also install a split-railed fence, positioned about halfway between the edge of the planting area and the edge of the trail along the length of the overflow area. At the request of the USFS (Columbia River Gorge National Scenic Area), RST will treat the planting area between the grassy overflow area and the trail with a mix of low growing shrubs, forbs, and clumping grasses. These will be planted sparsely and randomly in order to assimilate the surrounding environment, yet densely enough to provide visual and physical separation between the overflow area and the trail.

RST will install riparian plantings in disturbed ground at and near the new Gorton Creek Bridge. In an effort to maintain adequate water flow yet prevent backup, WFLHD has determined no live staking will occur in the rip-rapped areas. All other construction related areas near the creek will be planted. Table 2 below depicts the suite of species that is available for use in these areas.

# REVEGETATION PLAN

**TABLE 2. THE SUITE OF SPECIES AVAILABLE FOR PLANTING AT THE WYETH CAMPGROUND AND GORTON CREEK. STOCK TYPE DEFINITIONS ARE AS FOLLOW: 'S' SEED; 'RL10' 10 CUBIC INCH CONTAINER VOLUME; 'D27' 27 CUBIC INCH CONTAINER VOLUME; 'D40' 40 CUBIC INCH CONTAINER VOLUME; '#1' ONE GALLON CONTAINER; '#5' FIVE GALLON CONTAINER; 'B&B' BALLED AND BURLAPPED.**

| Common                | Botanical   | Zone | Stock Type  | Spacing<br>(on center) |
|-----------------------|---|------|-------------|------------------------|
| common yarrow         | <i>Achillea millefolium</i> var.<br><i>occidentalis</i> | A    | S           | XX                     |
| pearly everlasting    | <i>Anaphalis margaritacea</i>                           | A    | S           | XX                     |
| narrow-leaf milkweed  | <i>Asclepias fascicularis</i>                           | A    | S, RL10     | 4"                     |
| showy milkweed        | <i>Asclepias speciosa</i>                               | A    | S, RL10     | 4"                     |
| mountain brome        | <i>Bromus carinatus</i>                                 | A    | S           | XX                     |
| Howell's reedgrass    | <i>Calamagrostis howellii</i>                           | A    | S           | XX                     |
| blue wildrye          | <i>Elymus glaucus</i> ssp. <i>glaucus</i>               | A    | S           | XX                     |
| lupine                | <i>Lupinus latifolius</i>                               | A    | S           | 4"                     |
| creeping Oregon grape | <i>Mahonia repens</i>                                   | A    | D27, D40    | 4'                     |
| Sandberg's bluegrass  | <i>Poa secunda</i>                                      | A    | S           | XX                     |
| licorice fern         | <i>Polypodium glycyrrhiza</i>                           | A    | Short 4     | 2'                     |
| western sword fern    | <i>Polystichum munitum</i>                              | A    | Short 4, #1 | 4'                     |
| goldenrod             | <i>Solidago canadensis</i>                              | A    | S           | XX                     |
| fringecups            | <i>Tellima grandiflora</i>                              | A    | RL10        | 4"                     |
| vine maple            | <i>Acer circinatum</i>                                  | B    | #1, #5      | 8'                     |
| western serviceberry  | <i>Amelanchier alnifolia</i>                            | B    | #1, #5      | 8'                     |
| redosier dogwood      | <i>Cornus sericea</i> spp. <i>sericea</i>               | B    | D27, D40    | 8'                     |
| Oregon ash            | <i>Fraxinus latifolia</i>                               | B    | #1, #5      | 15'                    |
| oceanspray            | <i>Holodiscus discolor</i>                              | B    | D40         | 8'                     |
| Oregon grape          | <i>Mahonia aquifolium</i>                               | B    | D27, D40    | 4'                     |
| osoberry              | <i>Oemleria cerasiformis</i>                            | B    | #1, #5      | 8'                     |
| Douglas fir           | <i>Pseudotsuga menziesii</i>                            | B    | D27, B&B    | 15'                    |
| thimbleberry          | <i>Rubus parviflorus</i>                                | B    | D27, D40    | 4'                     |
| elderberry            | <i>Sambucus nigra</i>                                   | B    | D27         | 8'                     |
| common snowberry      | <i>Symphoricarpos albus</i>                             | B    | D27, D40    | 4'                     |
| trailing snowberry    | <i>Symphoricarpos mollis</i>                            | B    | D27, D40    | 4'                     |

# REVEGETATION PLAN

## Mitigation Site #2

The open area of approximately five acres, less than half a mile east of the Wyeth Trailhead and parking lot, is designated as a mitigation site. Restoration of this area will not only serve as mitigation for impacts incurred as a result of the project, but will also address Section 3 of the 2014 Presidential Memorandum “Creating a Federal Strategy to Promote the Health of Honey Bees and Other Pollinators” which provides directive to increase and improve pollinator habitat. Plant species that provide feeding, nesting, resting, and rearing (e.g., host plant) benefits to pollinating insects will be utilized to support and encourage plant-pollinator interactions. To this end, attention will be paid not only to nectar producing plants but also to plant architecture, the creation of micro-habitats and climates through the use of large and small woody material, depressions to retain moisture, areas of vegetative refuge, patches of bare ground (necessary for some ground nesting pollinator species), as well as other pollinator sustaining habitat characteristics.

Given the location of this project, two pollinator species of conservation concern will be supported explicitly. First, the imperiled western bumble bee (*Bombus occidentalis*) will be supported in this important part of its remaining range by providing a large and diverse resource of nectaring plants. Second, and more specifically, we will create breeding habitat to support the imperiled Monarch butterfly (*Danaus plexippus*). Milkweed is the only known plant genus to serve as host plant for the Monarchs in the lower 48 states. We will incorporate two species of milkweed (*Asclepias fascicularis* and *A. speciosa*) in the plantings at Mitigation Site #2, upon which Monarchs may lay their eggs. Critically, the local area which includes other nearby locations in the Columbia River Gorge, is a known Monarch migration and breeding area with observations of Monarchs close to the project area.

Mitigation Site #2 includes a pump house, a wellhead, and remnants of old buildings including steps, foundations, etc. These artifacts will remain in place, as is. There are also a number of old roads, the majority of which are gravel/crushed aggregate. WFLHD will remove the material from the old road prisms, spread shredded woody material over the entire mitigation site (4-6" deep), and incorporate the material to a depth of at least 18" by either ripping or using an excavator bucket.

Aesthetic augmentation will include the creation of an earthen berm by WFLHD to screen the pump house, and RST will select and install appropriate vegetation to enhance this screening. Topographic sculpting will create terrain variation to mimic existing conditions found elsewhere on the project site. Plants will be installed in the pollinator meadow in random

## REVEGETATION PLAN

groupings with some individual plants interspersed throughout, or broadcast seeded to create a natural appearing end meadow-woodland complex.

Because of the very different light and canopy regimes at the edge of the existing forest that surrounds the meadow, a different suite of species will be utilized in this area than those in the open habitat. There is a desire among the partners involved with this project to leave the area of Mitigation Site #2 somewhat open and create a meadow-like habitat to support the previously discussed pollinator initiatives. As such, plant species installed at this site will include more forbs, low shrubs, and native grasses than trees. It is expected that without regular maintenance the natural seed rain from the surrounding trees will eventually convert the open area into a treed area over time. This will not be facilitated through active revegetation efforts by RST, however.

The species suite to be utilized in pollinator meadow of Mitigation Site #2 will include some or all of the plants in Table 3a below. The mix used for the forested ecotone is reflected in Table 3b.

# REVEGETATION PLAN

**TABLE 3A. POTENTIAL PLANTS TO BE USED IN REVEGETATION EFFORTS TO ENHANCE POLLINATOR-PLANT INTERACTIONS AT THE OPEN HABITAT OF MITIGATION SITE #2. 'XX' DENOTES SEEDING RATHER THAN TRANSPLANTING TO BE UTILIZED. STOCK TYPE DEFINITIONS ARE AS FOLLOW: 'S' SEED; 'RL10' 10 CUBIC INCH CONTAINER VOLUME; 'D27' 27 CUBIC INCH CONTAINER VOLUME; 'D40' 40 CUBIC INCH CONTAINER VOLUME; '#1' ONE GALLON CONTAINER; '#5' FIVE GALLON CONTAINER; 'B&B' BALLED AND BURLAPPED.**

| Common                | Botanical   | Zone | Stock Type | Spacing<br>(on center) |
|-----------------------|---|------|------------|------------------------|
| common yarrow         | <i>Achillea millefolium</i> var.<br><i>occidentalis</i> | A    | S          | XX                     |
| pearly everlasting    | <i>Anaphalis margaritacea</i>                           | A    | S          | XX                     |
| narrow-leaf milkweed  | <i>Asclepias fascicularis</i>                           | A    | S, RL10    | 4"                     |
| showy milkweed        | <i>Asclepias speciosa</i>                               | A    | S, RL10    | 4"                     |
| Aster                 | <i>Aster spp.</i>                                       | A    | S          | XX                     |
| mountain brome        | <i>Bromus carinatus</i>                                 | A    | S          | XX                     |
| Howell's reedgrass    | <i>Calamagrostis howellii</i>                           | A    | S          | XX                     |
| Pacific dogwood       | <i>Cornus nuttallii</i>                                 | B    | #1         | 15'                    |
| blue wildrye          | <i>Elymus glaucus</i> ssp. <i>glaucus</i>               | A    | S          | XX                     |
| smooth alumroot       | <i>Heuchera glabra</i>                                  | A    | S          | 4"                     |
| Lupine                | <i>Lupinus latifolius</i>                               | A    | S          | 4"                     |
| creeping Oregon grape | <i>Mahonia repens</i>                                   | A    | D27        | 4'                     |
| Sandberg's bluegrass  | <i>Poa secunda</i>                                      | A    | S          | XX                     |
| self heal             | <i>Prunella vulgaris</i>                                | A    | S          | XX                     |
| checkermallow         | <i>Sidalcea spp.</i>                                    | A    | RL10       | XX                     |
| goldenrod             | <i>Solidago canadensis</i>                              | A    | S          | XX                     |
| fringecups            | <i>Tellima grandiflora</i>                              | A    | RL10       | 4"                     |
| western serviceberry  | <i>Amelanchier alnifolia</i>                            | B    | #1, #5     | 8'                     |
| Pacific dogwood       | <i>Cornus nuttallii</i>                                 | B    | #1         | 15'                    |
| oceanspray            | <i>Holodiscus discolor</i>                              | B    | D40        | 8'                     |
| Oregon grape          | <i>Mahonia aquifolium</i>                               | B    | D27, D40   | 4'                     |
| Douglas fir           | <i>Pseudotsuga menziesii</i>                            | B    | D27, B&B   | 15'                    |
| red flowering currant | <i>Ribes sanguineum</i>                                 | B    | #1, #5     | 4'                     |
| baldhip rose          | <i>Rosa gymnocarpa</i>                                  | B    | D27, B&B   | 4'                     |
| common snowberry      | <i>Symphoricarpos albus</i>                             | B    | D27, D40   | 4'                     |
| trailing snowberry    | <i>Symphoricarpos mollis</i>                            | B    | D27, D40   | 4'                     |
| western red cedar     | <i>Thuja plicata</i>                                    | B    | #1, B&B    | 15'                    |

# REVEGETATION PLAN

**TABLE 3B. PLANTS TO BE UTILIZED AT THE ECOTONE BETWEEN THE EXISTING FORESTED AREA AND THE NEWLY CREATED POLLINATOR MEADOW AT MITIGATION SITE #2. STOCK TYPE DEFINITIONS ARE AS FOLLOW: 'S' SEED; 'RL10' 10 CUBIC INCH CONTAINER VOLUME; 'D27' 27 CUBIC INCH CONTAINER VOLUME; 'D40' 40 CUBIC INCH CONTAINER VOLUME; '#1' ONE GALLON CONTAINER; '#5' FIVE GALLON CONTAINER; 'B&B' BALLED AND BURLAPPED.**

| Common                | Botanical                     | Zone | Stock Type  | Spacing<br>(on center) |
|-----------------------|-------------------------------|------|-------------|------------------------|
| creeping Oregon grape | <i>Mahonia repens</i>         | A    | D27, D40    | 4'                     |
| licorice fern         | <i>Polypodium glycyrrhiza</i> | A    | Short 4     | 2'                     |
| western sword fern    | <i>Polystichum munitum</i>    | A    | Short 4, #1 | 4'                     |
| grand fir             | <i>Abies grandis</i>          | B    | #1, B&B     | 15'                    |
| vine maple            | <i>Acer circinatum</i>        | B    | #1, #5      | 8'                     |
| western serviceberry  | <i>Amelanchier alnifolia</i>  | B    | #1, #5      | 8'                     |
| Pacific dogwood       | <i>Cornus nuttallii</i>       | B    | #1          | 15'                    |
| Oregon ash            | <i>Fraxinus latifolia</i>     | B    | #1, #5      | 15'                    |
| Oregon grape          | <i>Mahonia aquifolium</i>     | B    | D27, D40    | 4'                     |
| osoberry              | <i>Oemleria cerasiformis</i>  | B    | #1, #5      | 8'                     |
| western white pine    | <i>Pinus monticola</i>        | B    | D27         | 15'                    |
| Douglas fir           | <i>Pseudotsuga menziesii</i>  | B    | D27, B&B    | 15'                    |
| red flowering currant | <i>Ribes sanguineum</i>       | B    | #1, #5      | 4'                     |
| thimbleberry          | <i>Rubus parviflorus</i>      | B    | D27, D40    | 4'                     |
| elderberry            | <i>Sambucus nigra</i>         | B    | D27         | 8'                     |
| common snowberry      | <i>Symphoricarpos albus</i>   | B    | D27, D40    | 4'                     |

## Stepped, Plantable MSE Wall

There is a stepped, plantable mechanically stabilized earth (MSE) wall to be constructed from approximately ST 44+05 to ST 63+88. The height of this wall will vary from about 10-30 feet tall and it is planned to be 1,983 linear feet long. WFLHD will place topsoil along the entire base of the wall, burying the first two to three steps and creating a 3:1 slope (H:V). RST will place weed-free topsoil mixed with compost (1:1) over the remaining exposed wall to provide a continuous slope and effectively eliminating the appearance of steps. Shrubs and small trees will be randomly planted at the toe of the slope to provide vegetative screening, as well as sparsely planted along the newly created slope. RST will hydroseed the entire area with hydromulch, grass and forb seed, and tackifier. Irrigation will be an option item should the RST specialist determine it necessary for the survival of the plants. See Table 4 for the suite of potential restoration plants to be used on the plantable MSE wall.

# REVEGETATION PLAN

**TABLE 4. PLANT SUITE THAT MIGHT BE UTILIZED FOR VEGETATING THE STEPPED, PLANTABLE MSE WALL. 'XX' DENOTES SEEDING RATHER THAN TRANSPLANTING TO BE UTILIZED. STOCK TYPE DEFINITIONS ARE AS FOLLOW: 'S' SEED; 'RL10' 10 CUBIC INCH CONTAINER VOLUME; 'D27' 27 CUBIC INCH CONTAINER VOLUME; 'D40' 40 CUBIC INCH CONTAINER VOLUME; '#1' ONE GALLON CONTAINER; '#5' FIVE GALLON CONTAINER; 'B&B' BALLED AND BURLAPPED.**

| Common                | Botanical   | Zone | Stock Type  | Spacing<br>(on center) |
|-----------------------|---|------|-------------|------------------------|
| common yarrow         | <i>Achillea millefolium</i> var.<br><i>occidentalis</i> | A    | S           | XX                     |
| pearly everlasting    | <i>Anaphalis margaritacea</i>                           | A    | S           | XX                     |
| mountain brome        | <i>Bromus carinatus</i>                                 | A    | S           | XX                     |
| blue wildrye          | <i>Elymus glaucus</i> ssp. <i>glaucus</i>               | A    | S           | XX                     |
| smooth alumroot       | <i>Heuchera glabra</i>                                  | A    | RL10        | 4"                     |
| creeping Oregon grape | <i>Mahonia repens</i>                                   | A    | D27, D40    | 4'                     |
| Sandberg's bluegrass  | <i>Poa secunda</i>                                      | A    | S           | XX                     |
| licorice fern         | <i>Polypodium glycyrrhiza</i>                           | A    | Short 4     | 2'                     |
| western sword fern    | <i>Polystichum munitum</i>                              | A    | Short 4, #1 | 4'                     |
| fringecups            | <i>Tellima grandiflora</i>                              | A    | RL10        | 4"                     |
| western serviceberry  | <i>Amelanchier alnifolia</i>                            | B    | #1, #5      | 8'                     |
| Oregon grape          | <i>Mahonia aquifolium</i>                               | B    | D27, D40    | 4'                     |
| osoberry              | <i>Oemleria cerasiformis</i>                            | B    | #1, #5      | 8'                     |
| Douglas fir           | <i>Pseudotsuga menziesii</i>                            | B    | D27, B&B    | 15'                    |
| red flowering currant | <i>Ribes sanguineum</i>                                 | B    | #1, #5      | 4'                     |
| thimbleberry          | <i>Rubus parviflorus</i>                                | B    | D27, D40    | 4'                     |
| common snowberry      | <i>Symporicarpus albus</i>                              | B    | D27, D40    | 4'                     |
| trailing snowberry    | <i>Symporicarpus mollis</i>                             | B    | D27, D40    | 4'                     |

## Summit Creek Viaduct

The extensive pedestrian viaduct planned near Summit Creek will present specific challenges to revegetation efforts. The viaduct will cover approximately 0.16 acres and will prevent precipitation and direct sunlight from reaching the surface of the ground below the viaduct deck. Currently there are micro-habitats that have a variety of vegetation growing at the toe slope of rock walls. Soil accumulates at the junction of the rock wall and ground, and precipitation drips down the face of the rock; both contribute to the ability to support plant life. Revegetation efforts will take advantage of similar micro-habitats under the viaduct, utilize shade tolerant plant species, and incorporate organic matter into the soil to aid in

## REVEGETATION PLAN

moisture retention. The primary contractor will place topsoil conserved from the project at the viaduct footings to bring the soil up to existing native soil depths and provide planting medium. Please refer to Table 5 for candidate species for planting near the Summit Creek Viaduct.

**TABLE 5. PLANTS POTENTIALLY TO BE USED IN REVEGETATION EFFORTS AT THE SUMMIT CREEK VIADUCT. 'XX' DENOTES SEEDING RATHER THAN TRANSPLANTING TO BE UTILIZED. STOCK TYPE DEFINITIONS ARE AS FOLLOW: 'S' SEED; 'RL10' 10 CUBIC INCH CONTAINER VOLUME; 'D27' 27 CUBIC INCH CONTAINER VOLUME; 'D40' 40 CUBIC INCH CONTAINER VOLUME; '#1' ONE GALLON CONTAINER; '#5' FIVE GALLON CONTAINER; 'B&B' BALLED AND BURLAPPED.**

| Common                | Botanical   | Zone | Stock Type  | Spacing<br>(on center) |
|-----------------------|---|------|-------------|------------------------|
| common yarrow         | <i>Achillea millefolium</i> var.<br><i>occidentalis</i> | A    | S           | XX                     |
| pearly everlasting    | <i>Anaphalis margaritacea</i>                           | A    | S           | XX                     |
| smooth alumroot       | <i>Heuchera glabra</i>                                  | A    | RL10        | 4"                     |
| creeping Oregon grape | <i>Mahonia repens</i>                                   | A    | D27, D40    | 4'                     |
| licorice fern         | <i>Polypodium glycyrrhiza</i>                           | A    | Short 4     | 2'                     |
| western sword fern    | <i>Polystichum munitum</i>                              | A    | Short 4, #1 | 4'                     |
| fringecups            | <i>Tellima grandiflora</i>                              | A    | RL10        | 4"                     |
| western serviceberry  | <i>Amelanchier alnifolia</i>                            | B    | #1, #5      | 8'                     |
| redosier dogwood      | <i>Cornus sericea</i> spp. <i>sericea</i>               | B    | D27, D40    | 8'                     |
| oceanspray            | <i>Holodiscus discolor</i>                              | B    | D40         | 8'                     |
| Oregon grape          | <i>Mahonia aquifolium</i>                               | B    | D27, D40    | 4'                     |
| osoberry              | <i>Oemleria cerasiformis</i>                            | B    | #1, #5      | 8'                     |
| baldhip rose          | <i>Rosa gymnocarpa</i>                                  | B    | D27, D40    | 4'                     |
| thimbleberry          | <i>Rubus parviflorus</i>                                | B    | D27, D40    | 4'                     |
| common snowberry      | <i>Symporicarpus albus</i>                              | B    | D27, D40    | 4'                     |

### HCRHST Embankments (Cuts and Fills) and Planned Idle Spots

In coordination with WFLHD, RST will provide the primary contractor with a native grass and forb seed mix to use on exposed trail embankments during and post construction as a part of erosion control measures. RST will install native plants (Table 6) throughout the construction limits once construction activity ceases. Due to the effort to create a natural appearing setting, native shrubs and forbs will be planted in groupings, spread at varying intervals, and in a mix of species. Some areas, such as near Shell Rock Mountain and the new idle spots might not require any plant installation other than seed for erosion control efforts. The planting plans

## REVEGETATION PLAN

from Walker Macy will be referenced to maintain line-of-site and safety, utilizing shorter plants directly adjacent to the trail and creating a natural appearing visual experience.

Slopes of greater than average length or slope (1:2) will receive a mulch application of weed-free compost or shredded woody material, applied to a depth of 1-2 inches, and a final grass seed application. The addition of mulch and final seeding will assist to preserve slope integrity, decrease weed recruitment and establishment, and facilitate moisture retention. Practices that increase soil compaction are best avoided and it is ideal that slopes be left in a roughened condition (i.e., use a toothed rock bucket rather than smoothing bucket, avoid back blading, etc.).

**TABLE 6. POTENTIAL PLANT SPECIES TO BE UTILIZED ON CUT/FILL SLOPES AND PLANNED IDLE SPOTS OF HISTORIC COLUMBIA RIVER HIGHWAY STATE TRAIL. 'XX' DENOTES SEEDING RATHER THAN TRANSPLANTING TO BE UTILIZED. STOCK TYPE DEFINITIONS ARE AS FOLLOW: 'S' SEED; 'RL10' 10 CUBIC INCH CONTAINER VOLUME; 'D27' 27 CUBIC INCH CONTAINER VOLUME; 'D40' 40 CUBIC INCH CONTAINER VOLUME; '#1' ONE GALLON CONTAINER; '#5' FIVE GALLON CONTAINER; 'B&B' BALLED AND BURLAPPED.**

| Common                | Botanical   | Zone | Stock Type  | Spacing<br>(on center) |
|-----------------------|---|------|-------------|------------------------|
| common yarrow         | <i>Achillea millefolium</i> var.<br><i>occidentalis</i> | A    | S           | XX                     |
| pearly everlasting    | <i>Anaphalis margaritacea</i>                           | A    | S           | XX                     |
| mountain brome        | <i>Bromus carinatus</i>                                 | A    | S           | XX                     |
| Howell's reedgrass    | <i>Calamagrostis howellii</i>                           | A    | S           | XX                     |
| blue wildrye          | <i>Elymus glaucus</i> ssp. <i>glaucus</i>               | A    | S           | XX                     |
| lupine                | <i>Lupinus latifolius</i>                               | A    | S           | 4"                     |
| creeping Oregon grape | <i>Mahonia repens</i>                                   | A    | D27, D40    | 4'                     |
| Sandberg's bluegrass  | <i>Poa secunda</i>                                      | A    | S           | XX                     |
| licorice fern         | <i>Polypodium glycyrrhiza</i>                           | A    | Short 4     | 2'                     |
| western sword fern    | <i>Polystichum munitum</i>                              | A    | Short 4, #1 | 4'                     |
| self heal             | <i>Prunella vulgaris</i>                                | A    | S           | XX                     |
| fringecups            | <i>Tellima grandiflora</i>                              | A    | RL10        | 4"                     |
| Oregon grape          | <i>Mahonia aquifolium</i>                               | B    | D27, D40    | 4'                     |
| baldhip rose          | <i>Rosa gymnocarpa</i>                                  | B    | D27, D40    | 4'                     |
| thimbleberry          | <i>Rubus parviflorus</i>                                | B    | D27, D40    | 4'                     |
| common snowberry      | <i>Symporicarpos albus</i>                              | B    | D27, D40    | 4'                     |
| trailing snowberry    | <i>Symporicarpos mollis</i>                             | B    | D27, D40    | 4'                     |

# REVEGETATION PLAN

## Staging Area(s)

The potential suite of plant species for restoration of staging areas is diverse. The staging area(s) can encompass a variety of environments with differing light regimes, infiltration rates, and habitats. Other than weed control, most revegetation efforts will be delayed until after staging equipment and material are removed by WFLHD. Table 7 shows the species that may be used in revegetation efforts of staging areas.

**TABLE 7. PROPOSED REVEGETATION SPECIES FOR STAGING AREA(S) UTILIZED. 'XX' DENOTES SEEDING RATHER THAN TRANSPLANTING TO BE UTILIZED. STOCK TYPE DEFINITIONS ARE AS FOLLOW: 'S' SEED; 'RL10' 10 CUBIC INCH CONTAINER VOLUME; 'D27' 27 CUBIC INCH CONTAINER VOLUME; 'D40' 40 CUBIC INCH CONTAINER VOLUME; '#1' ONE GALLON CONTAINER; '#5' FIVE GALLON CONTAINER; 'B&B' BALLED AND BURLAPPED.**

| Common                | Botanical  | Zone | Stock Type  | Spacing (on center) |
|-----------------------|--|------|-------------|---------------------|
| common yarrow         | <i>Achillea millefolium</i> var. <i>occidentalis</i> | A    | S           | XX                  |
| pearly everlasting    | <i>Anaphalis margaritacea</i>                        | A    | S           | XX                  |
| mountain brome        | <i>Bromus carinatus</i>                              | A    | S           | XX                  |
| Howell's reedgrass    | <i>Calamagrostis howellii</i>                        | A    | S           | XX                  |
| blue wildrye          | <i>Elymus glaucus</i> ssp. <i>glaucus</i>            | A    | S           | XX                  |
| lupine                | <i>Lupinus latifolius</i>                            | A    | S           | 4"                  |
| creeping Oregon grape | <i>Mahonia repens</i>                                | A    | D27, D40    | 4'                  |
| Sandberg's bluegrass  | <i>Poa secunda</i>                                   | A    | S           | XX                  |
| licorice fern         | <i>Polypodium glycyrrhiza</i>                        | A    | Short 4     | 2'                  |
| western sword fern    | <i>Polystichum munitum</i>                           | A    | Short 4, #1 | 4'                  |
| goldenrod             | <i>Solidago canadensis</i>                           | A    | S           | XX                  |
| fringecups            | <i>Tellima grandiflora</i>                           | A    | RL10        | 4"                  |
| grand fir             | <i>Abies grandis</i>                                 | B    | #1, B&B     | 15'                 |
| vine maple            | <i>Acer circinatum</i>                               | B    | #1, #5      | 8'                  |
| western serviceberry  | <i>Amelanchier alnifolia</i>                         | B    | #1, #5      | 8'                  |
| Pacific dogwood       | <i>Cornus nuttallii</i>                              | B    | #1          | 15'                 |
| redosier dogwood      | <i>Cornus sericea</i> spp. <i>sericea</i>            | B    | D27, D40    | 8'                  |
| Oregon ash            | <i>Fraxinus latifolia</i>                            | B    | #1, #5      | 15'                 |
| oceanspray            | <i>Holodiscus discolor</i>                           | B    | D40         | 8'                  |
| Oregon grape          | <i>Mahonia aquifolium</i>                            | B    | D27, D40    | 4'                  |
| osoberry              | <i>Oemleria cerasiformis</i>                         | B    | #1, #5      | 8'                  |
| Douglas fir           | <i>Pseudotsuga menziesii</i>                         | B    | D27, B&B    | 15'                 |
| red flowering currant | <i>Ribes sanguineum</i>                              | B    | #1, #5      | 4'                  |
| thimbleberry          | <i>Rubus parviflorus</i>                             | B    | D27, D40    | 4'                  |
| elderberry            | <i>Sambucus nigra</i>                                | B    | D27         | 8'                  |
| common snowberry      | <i>Symporicarpus albus</i>                           | B    | D27, D40    | 4'                  |
| trailing snowberry    | <i>Symporicarpus mollis</i>                          | B    | D27, D40    | 4'                  |
| western red cedar     | <i>Thuja plicata</i>                                 | B    | #1, B&B     | 15'                 |

## NON-NATIVE PLANT CONTROL

### Non-Native Plant Control

#### Weed Control Plan

All Oregon Noxious Class 'A' and 'B' weeds will be treated within the construction limits (Appendix Three). The one exception to this is herb Robert (*Geranium robertianum*) which will not be targeted for treatment, as determined during site visits with WFLHD, ODOT, CH2M, CRGNSA, and RST. Of the targeted Class A and B species the following Class B species have been noted within Segments A-C: Canada thistle (*Cirsium arvense*), English ivy, Himalayan blackberry, Scotch broom, perennial peavine (*Lathyrus latifolius*), and St. Johnswort (*Hypericum perforatum*). Although not listed by the State, periwinkle (*Vinca minor*) will also be targeted for control, as will other species that appear to impede restoration efforts. Table 8 provides an estimated schedule of non-native plant treatment.

Only approved herbicides and surfactants will be used and all aquatic setbacks adhered to. Personnel from partnering agencies including the Mt. Hood National Forest, Columbia River Gorge National Scenic Area, Oregon Parks and Recreation Department, and Oregon Department of Transportation will be consulted regarding permissible herbicides and surfactants.

#### *Himalayan blackberry*

Himalayan blackberry will be sprayed with glyphosate or triclopyr + surfactant from a backpack sprayer or an ATV. In the area of Mitigation Site #2 the plants will then be mowed no sooner than 3 weeks after the first spray treatment, allowed to re-sprout, and chemically treated again. The area with the greatest concentration of Himalayan blackberry is Mitigation Site #2, but the plants do occur throughout the segments. RST will chemically treat these populations where they occur, but most likely will not utilize mowing.

#### *Scotch Broom*

Scotch broom populations are most robust at the Wyeth Trailhead/parking lot and at Mitigation Site #2. There are plants scattered throughout other areas of segments A-C, as well as along Wyeth Road near the proposed Wyeth Trailhead and parking lot. Treatments should include these plants that are in close proximity along Wyeth Road to prevent immediate recolonization of Wyeth parking lot. Plants less than three feet in length will receive herbicide treatment and left in place; those greater than three feet in length will be wrenched out, including as much root as is possible, and removed from the project site.

#### *Additional weeds*

In addition to the above, periwinkle has been observed scattered throughout segments A-C. Populations of these plants will be treated at the same time as the others previously discussed. Periwinkle will be sprayed with either glyphosate or triclopyr.

## MONITORING PLAN

### Monitoring Plan

The agreed upon parameters to evaluate native plant revegetative success for this project are percent cover and plant stem density (Table 8). Because of this, monitoring will be conducted at 1/100<sup>th</sup> acre plots distributed randomly throughout the project area. In an effort to ascertain unbiased data the plots will not be fixed and will occur in different locations from year to year. A minimum of ten (10) plots will be analyzed within the boundaries of Mitigation Site #2 and ten (10) additional plots will be randomly distributed throughout the project area. Native species cover and stem density will be recorded within each plot, as well as other variables.

Noxious weed success criteria is total percent cover. Prior to construction RST will conduct ocular estimation of non-native plant cover within Mitigation Site #2 and repeat annually. Percent cover of each non-native species will be noted within each plot described above; at Mitigation Site #2 as well as throughout the project area. Refer to Table 9 for an estimated monitoring timeline and Table 10 for overall costs associated with this project.

In addition to native and non-native plants, monitoring protocol for pollinating insects will be conducted at the newly created pollinator meadow of Mitigation Site #2. Of particular interest are native bees and Monarch butterflies. The streamlined bee monitoring procedure developed by the Xerces Society will be utilized twice per year after plant establishment. During the surveys all pollinators observed will be noted including wasps, flies, honey bees, native bees, Monarch butterflies, other butterflies, moths, ants, etc.

## SUCCESS CRITERIA

### Success Criteria

**TABLE 8. CRITERIA UPON WHICH NATIVE AND NON-NATIVE PLANT SUCCESS WILL BE MEASURED.**

| Item                             | Benchmark                                   | Timeframe            | Notes  |
|----------------------------------|---|----------------------|--|
| Noxious weed control             | ≤ 20% cover                                 | By year 3            | Oregon Noxious Weed List A and B   |
| Noxious weed control             | ≤ 10% cover                                 | By year 5            | Oregon Noxious Weed List A and B   |
| Planting density                 | ≥ 400 woody stems per acre                  | By year 5            |  |
| Native plant percent cover       | ≥ 20% cover                                 | By year 1            |  |
| Native plant percent cover       | ≥ 40% cover                                 | By year 3            |  |
| Native plant percent cover       | ≥ 80% cover                                 | By year 5            |  |
| Native plant diversity           | ≥ 5 woody species comprise ≥ 5% total cover | By year 5            |  |
| If performance standards not met | Use of adaptive management practices        | As early as detected | May include replanting, reseeding, noxious weed removal, improvement of soil quality, irrigation, mulch. |

## APPROXIMATE TIMELINE & BUDGET

### Approximate Timeline & Budget

**TABLE 9.** THE FOLLOWING TABLE DEPICTS THE ESTIMATED TIMELINE OF REVEGETATION EFFORTS PLANNED BY RST WITHIN SEGMENTS A-C OF THE HISTORIC COLUMBIA RIVER HIGHWAY STATE TRAIL.

| Task                       | 2016   |        |      |        | 2017   |        |      |        | 2018   |        |      |        | 2019   |        |      |        | 2020   |        |      |        | 2021   |        |      |        |  |
|----------------------------|--------|--------|------|--------|--------|--------|------|--------|--------|--------|------|--------|--------|--------|------|--------|--------|--------|------|--------|--------|--------|------|--------|--|
|                            | Spring | Summer | Fall | Winter |  |
| Weed treatment             |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |  |
| Collect plant propagules   |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |  |
| Grow-out plant propagules  |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |  |
| Install plants (as ready)  |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |  |
| Monitor non-native plants  |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |  |
| Monitor native plants      |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |  |
| Monitor pollinator species |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |        |        |      |        |  |

## APPENDIX ONE

### Appendix One- Plant Species Suite

POTENTIAL SUITE OF PLANT SPECIES TO BE USED IN REVEGETATION EFFORTS OF SEGMENTS A-C OF HCRHST. STOCK TYPE DEFINITIONS ARE AS FOLLOW: 'S' SEED; 'RL10' 10 CUBIC INCH CONTAINER VOLUME; 'D27' 27 CUBIC INCH CONTAINER VOLUME; 'D40' 40 CUBIC INCH CONTAINER VOLUME; '#1' ONE GALLON CONTAINER; '#5' FIVE GALLON CONTAINER; 'B&B' BALLED AND BURLAPPED.

| Common                | Botanical   | Zone | Stock Type  | Spacing<br>(on center) |
|-----------------------|---|------|-------------|------------------------|
| common yarrow         | <i>Achillea millefolium</i> var.<br><i>occidentalis</i> | A    | S           | XX                     |
| pearly everlasting    | <i>Anaphalis margaritacea</i>                           | A    | S           | XX                     |
| narrow-leaf milkweed  | <i>Asclepias fascicularis</i>                           | A    | S, RL10     | 4"                     |
| showy milkweed        | <i>Asclepias speciosa</i>                               | A    | S, RL10     | 4"                     |
| Aster                 | <i>Aster spp.</i>                                       | A    | S           | XX                     |
| mountain brome        | <i>Bromus carinatus</i>                                 | A    | S           | XX                     |
| Howell's reedgrass    | <i>Calamagrostis howellii</i>                           | A    | S           | XX                     |
| blue wildrye          | <i>Elymus glaucus</i> ssp. <i>glaucus</i>               | A    | S           | XX                     |
| smooth alumroot       | <i>Heuchera glabra</i>                                  | A    | RL10        | 4"                     |
| Lupine                | <i>Lupinus latifolius</i>                               | A    | S           | 4"                     |
| creeping Oregon grape | <i>Mahonia repens</i>                                   | A    | D27, D40    | 4'                     |
| Sandberg's bluegrass  | <i>Poa secunda</i>                                      | A    | S           | XX                     |
| licorice fern         | <i>Polypodium glycyrrhiza</i>                           | A    | Short 4     | 2'                     |
| western sword fern    | <i>Polystichum munitum</i>                              | A    | Short 4, #1 | 4'                     |
| self heal             | <i>Prunella vulgaris</i>                                | A    | S           | XX                     |
| checkermallow         | <i>Sidalcea spp.</i>                                    | A    | S           | XX                     |
| goldenrod             | <i>Solidago canadensis</i>                              | A    | S           | XX                     |
| fringecups            | <i>Tellima grandiflora</i>                              | A    | RL10        | 4"                     |
| grand fir             | <i>Abies grandis</i>                                    | B    | #1, B&B     | 15'                    |
| vine maple            | <i>Acer circinatum</i>                                  | B    | #1, #5      | 8'                     |
| western serviceberry  | <i>Amelanchier alnifolia</i>                            | B    | #1, #5      | 8'                     |
| Pacific dogwood       | <i>Cornus nuttallii</i>                                 | B    | #1          | 15'                    |
| redosier dogwood      | <i>Cornus sericea</i> spp. <i>sericea</i>               | B    | D27, D40    | 8'                     |
| Oregon ash            | <i>Fraxinus latifolia</i>                               | B    | #1, #5      | 15'                    |
| oceanspray            | <i>Holodiscus discolor</i>                              | B    | D40         | 8'                     |
| Oregon grape          | <i>Mahonia aquifolium</i>                               | B    | D27, D40    | 4'                     |
| osoberry              | <i>Oemleria cerasiformis</i>                            | B    | #1, #5      | 8'                     |
| ninebark              | <i>Physocarpus capitatus</i>                            | B    | #1, #5      | 8'                     |
| western white pine    | <i>Pinus monticola</i>                                  | B    | D27         | 15'                    |
| Douglas fir           | <i>Pseudotsuga menziesii</i>                            | B    | D27, B&B    | 15'                    |
| red flowering currant | <i>Ribes sanguineum</i>                                 | B    | #1, #5      | 4'                     |
| baldhip rose          | <i>Rosa gymnocarpa</i>                                  | B    | D27, D40    | 4'                     |
| thimbleberry          | <i>Rubus parviflorus</i>                                | B    | D27, D40    | 4'                     |
| elderberry            | <i>Sambucus nigra</i>                                   | B    | D27         | 8'                     |
| common snowberry      | <i>Symphoricarpos albus</i>                             | B    | D27, D40    | 4'                     |
| trailing snowberry    | <i>Symphoricarpos mollis</i>                            | B    | D27, D40    | 4'                     |
| western red cedar     | <i>Thuja plicata</i>                                    | B    | #1, B&B     | 15'                    |

## APPENDIX TWO

### Appendix Two- Walker Macy Planting Plans

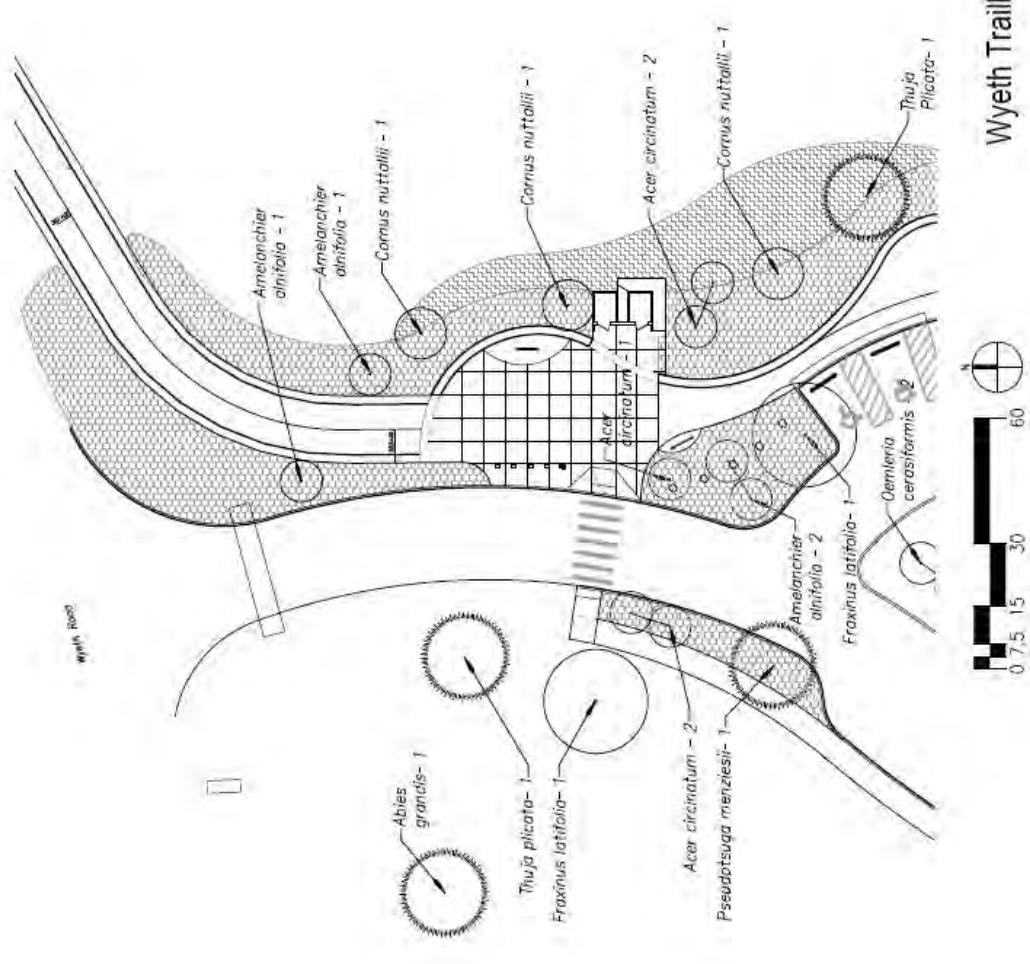
#### PLANT SCHEDULE

|                        | BOTANICAL NAME                            | COMMON NAME           | SIZE/CONDITION | SPACING     | COMMENTS  |
|------------------------|---|-----------------------|----------------|-------------|---|
| <b>TREES EVERGREEN</b> |   |                       |                |             |   |
|                        | <i>Abies grandis</i>                      | grand fir             | 5–6 ft.        | 15 ft. O.C. | Min. 3 ea. per cluster or as shown              |
|                        | <i>Pseudotsuga menziesii</i>              | Douglas fir           | 5–6 ft.        | 15 ft. O.C. | Min. 4 ea. per cluster or as shown              |
|                        | <i>Thuja plicata</i>                      | Western red cedar     | 5–6 ft.        | 20 ft. O.C. | Min. 3 ea. per cluster or as shown              |
| <b>TREES DECIDUOUS</b> |   |                       |                |             |   |
|                        | <i>Acer circinatum</i>                    | vine maple            | 5–6 ft.        | 6 ft. O.C.  | Min. 3 ea. per cluster or as shown; Multi trunk |
|                        | <i>Amelanchier alnifolia</i>              | Western serviceberry  | 3–4 ft.        | 6 ft. O.C.  | Min. 3 ea. per cluster or as shown; Multi trunk |
|                        | <i>Cornus nuttallii</i>                   | Pacific dogwood       | 5–6 ft.        | 12 ft. O.C. | Min. 3 ea. per cluster or as shown;             |
|                        | <i>Oemleria cerasiformis</i>              | osoberry              | 3–4 ft.        | 6 ft. O.C.  | Min. 3 ea. per cluster or as shown; Multi trunk |
|                        | <i>Fraxinus latifolia</i>                 | Oregon ash            | 1" Cal.        | 15 ft. O.C. | Min. 3 ea. per cluster or as shown              |
| <b>SHRUBS</b>          |   |                       |                |             |   |
|                        | <i>Cornus sericea</i> spp. <i>sericea</i> | redosier dogwood      | #2             | 4 ft. O.C.  | As shown  |
|                        | <i>Physocarpus capitatus</i>              | ninebark              | 4–5 ft.        | 6 ft. O.C.  | As shown  |
|                        | <i>Rubus parviflorus</i>                  | thimbleberry          | #2             | 30" O.C.    | As shown  |
|                        | <i>Sambucus nigra</i>                     | elderberry            | #2             | 8 ft. O.C.  | As shown  |
|                        | <i>Symporicarpus albus</i>                | common snowberry      | #2             | 30" O.C.    | As shown  |
|                        | mixture of forbs                          |                       |                |             | Min. 5 ea. per species cluster                  |
| <b>SHRUBS ZONE A</b>   |   |                       |                |             |   |
|                        | <i>Mahonia repens</i>                     | creeping Oregon grape | #2             | 24" O.C.    | Min. 12 ea. per cluster                         |
|                        | <i>Polystichum munitum</i>                | Western sword fern    | #2             | 30" O.C.    | Min. 9 ea. per cluster                          |
|                        | <i>Symporicarpus mollis</i>               | creeping snowberry    | #1             | 30" O.C.    | Min. 5 ea. per cluster                          |
|                        | mixture of forbs                          |                       |                |             | Min. 5 ea. per species cluster                  |
| <b>SHRUBS ZONE B</b>   |   |                       |                |             |   |
|                        | <i>Mahonia repens</i>                     | creeping Oregon grape | #2             | 36" O.C.    | Min. 7 ea. per cluster                          |
|                        | <i>Rosa gymnocarpa</i>                    | baldhip rose          | #1             | 36" O.C.    | Min. 5 ea. per cluster                          |
|                        | <i>Polystichum munitum</i>                | Western sword fern    | #2             | 30" O.C.    | Min. 5 ea. per cluster                          |
|                        | <i>Rubus parviflorus</i>                  | thimbleberry          | #1             | 30" O.C.    | Min. 5 ea. per cluster                          |
|                        | <i>Symporicarpus mollis</i>               | creeping snowberry    | #1             | 36" O.C.    | Min. 5 ea. per cluster                          |
|                        | <i>Ribes sanguineum</i>                   | red flowering currant | #2             | 48" O.C.    | Min. 3 ea. per cluster                          |
|                        | <i>Physocarpus capitatus</i>              | ninebark              | #2             | 48" O.C.    | Min. 3 ea. per cluster                          |
|                        | <i>Holodiscus discolor</i>                | oceanspray            | #2             | 60" O.C.    | Min. 1 ea. per cluster                          |
|                        | mixture of forbs                          |                       |                |             | Min. 5 ea. per species cluster                  |

#### PLANTING NOTES

1. Plant Zone B plants no less than 10 feet from the outside edge of pedestrian trails.
2. Shrubs to be planted within limit of disturbance or as shown.
3. In exposed clearings, plant clusters of deciduous trees in quantities and spacing as shown on plant legend.
4. Maintain clear line of sight from new trailheads to primary pedestrian trails whenever possible.
5. Shrubs and trees are to be planted no closer than  $\frac{1}{2}$  of their on-center spacing from outside edge of pedestrian trail or trailhead.

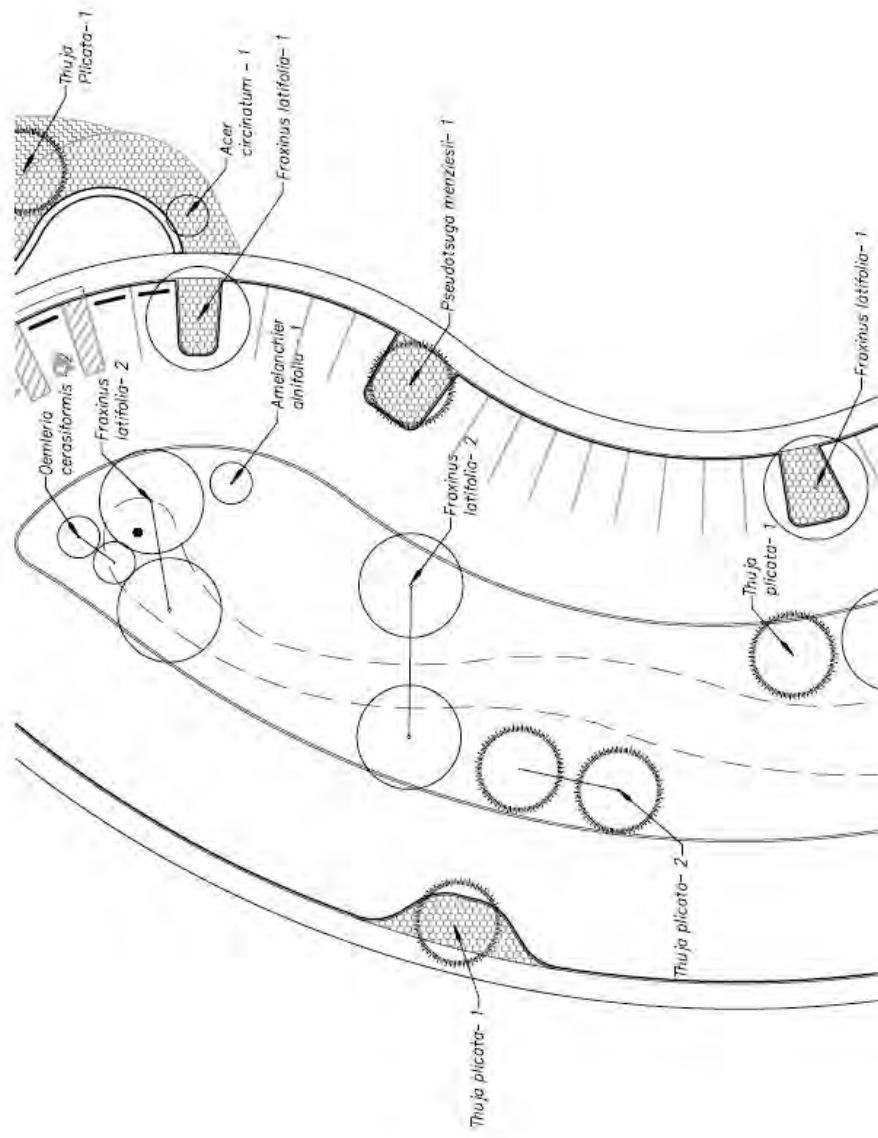
## APPENDIX TWO



Page 28

Historic Columbia River Highway State Trail, Segments A-C Final Revegetation Plan

## APPENDIX TWO

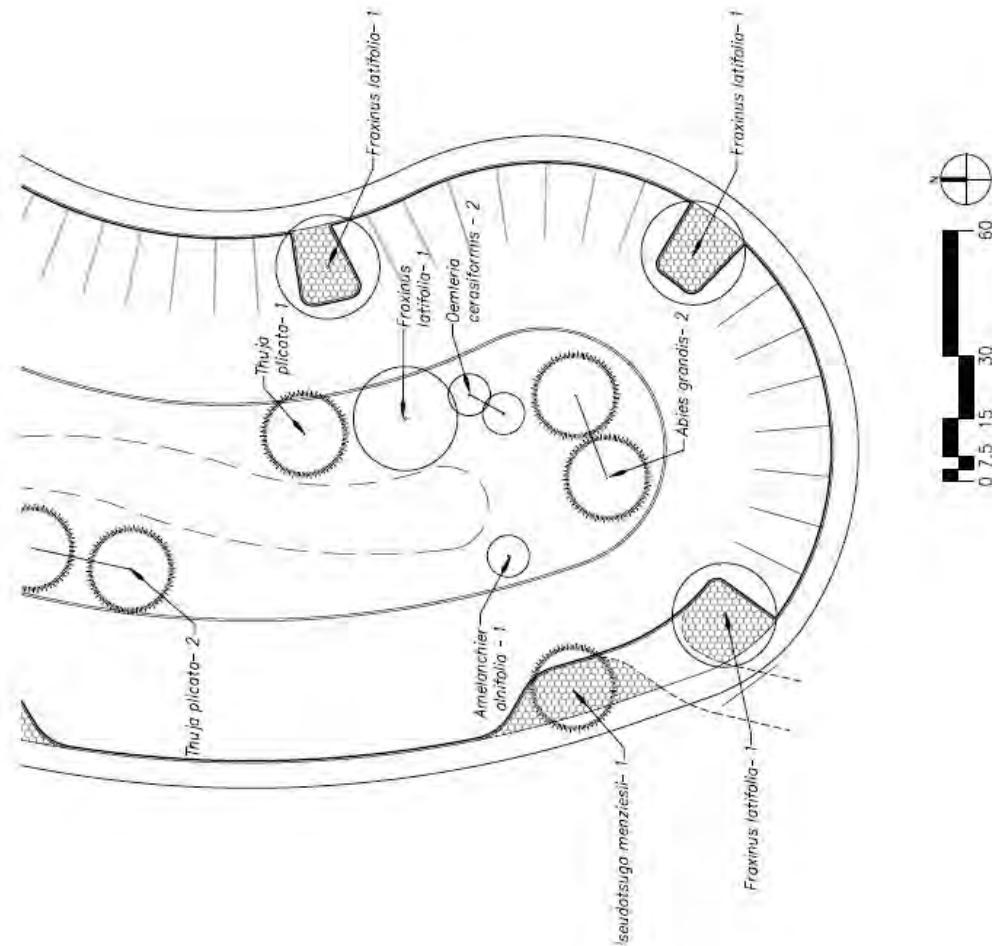


**Wyeth Trailhead**

Historic Columbia River Highway State Trail, Segments A-C Final Revegetation Plan

Page 29

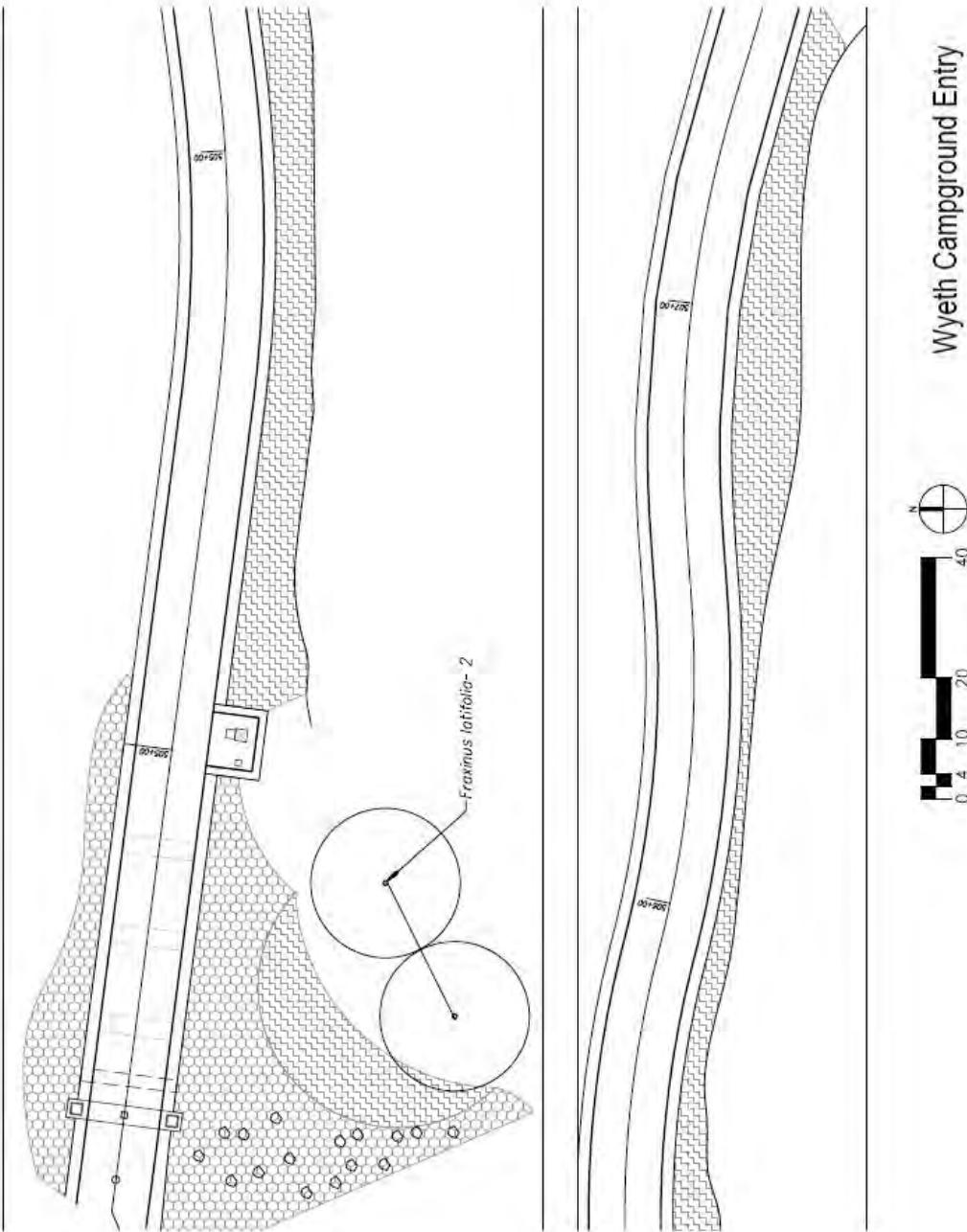
## APPENDIX TWO



Page 30

Historic Columbia River Highway State Trail, Segments A-C Final Revegetation Plan

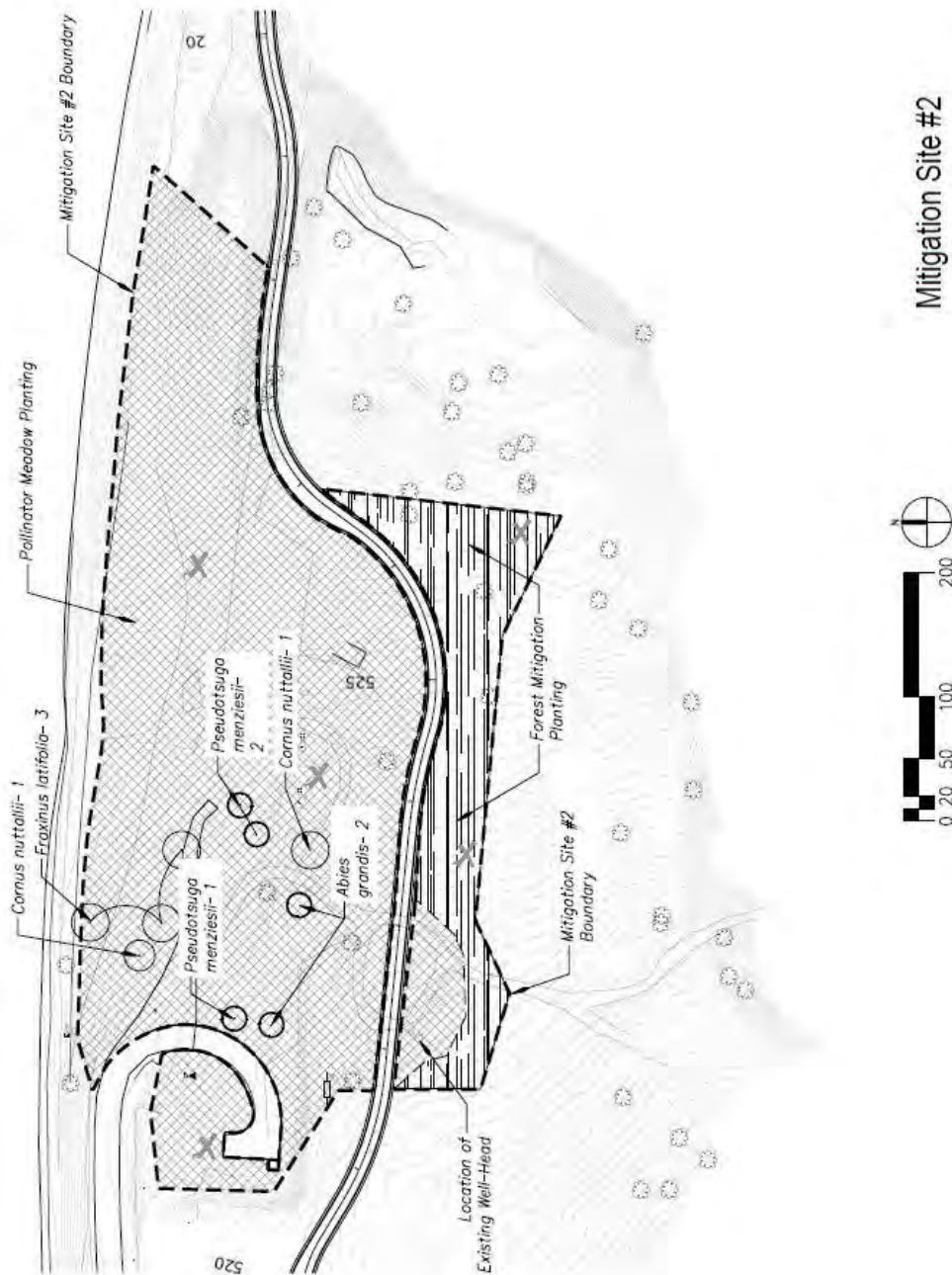
## APPENDIX TWO



Page 31

Historic Columbia River Highway State Trail, Segments A-C Final Revegetation Plan

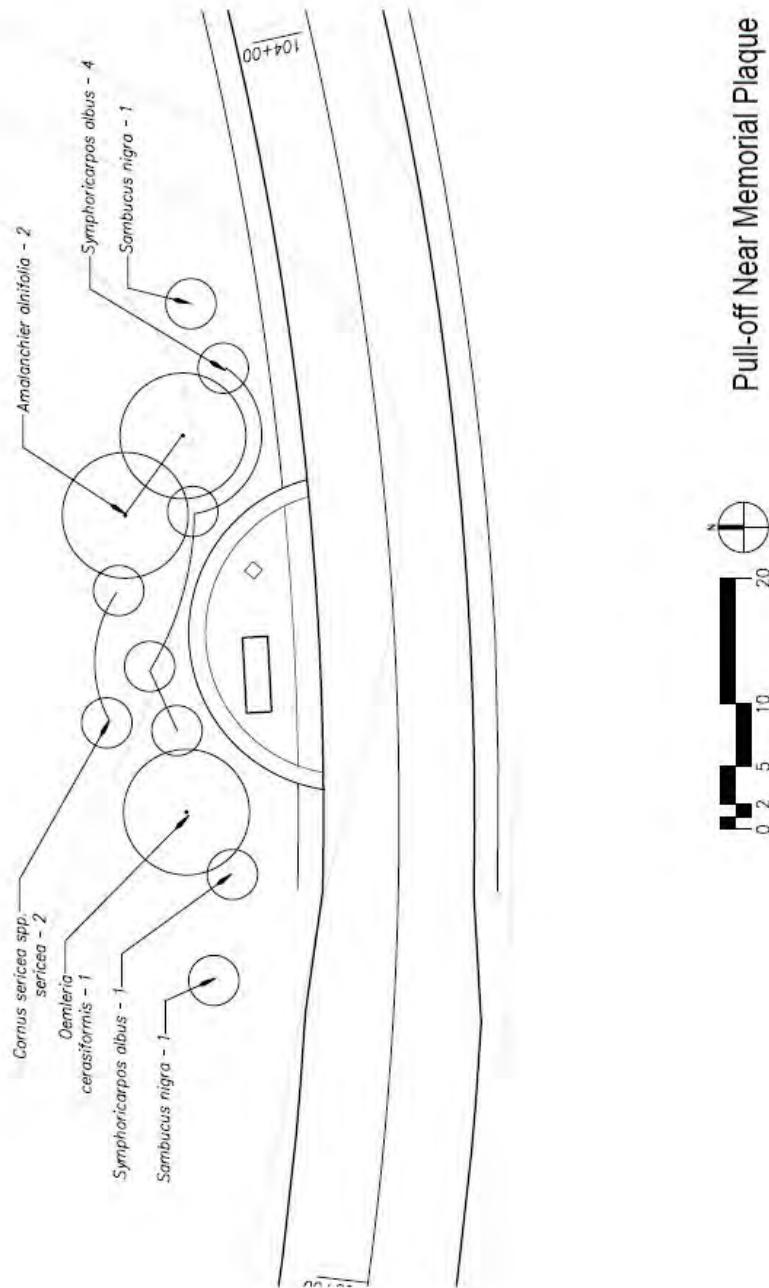
## APPENDIX TWO



Page 32

Historic Columbia River Highway State Trail, Segments A-C Final Revegetation Plan

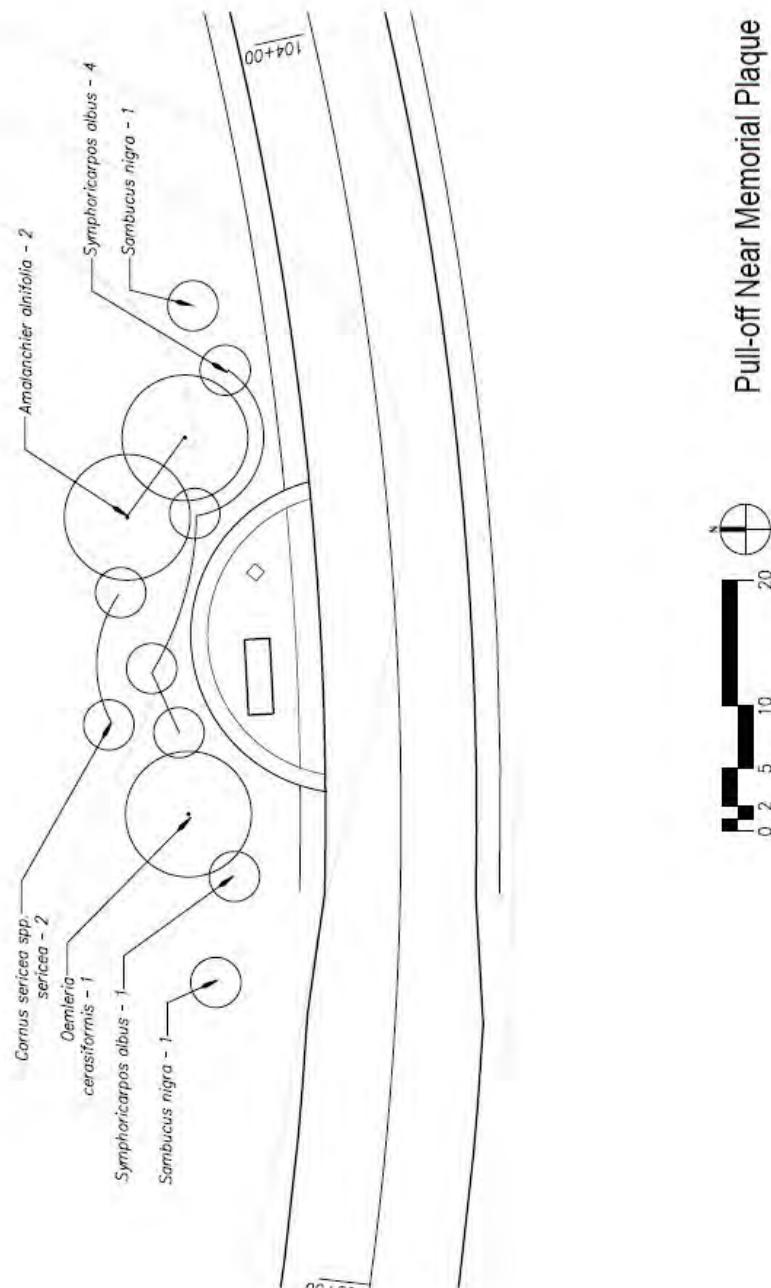
## APPENDIX TWO



Page 33

Historic Columbia River Highway State Trail, Segments A-C Final Revegetation Plan

## APPENDIX TWO

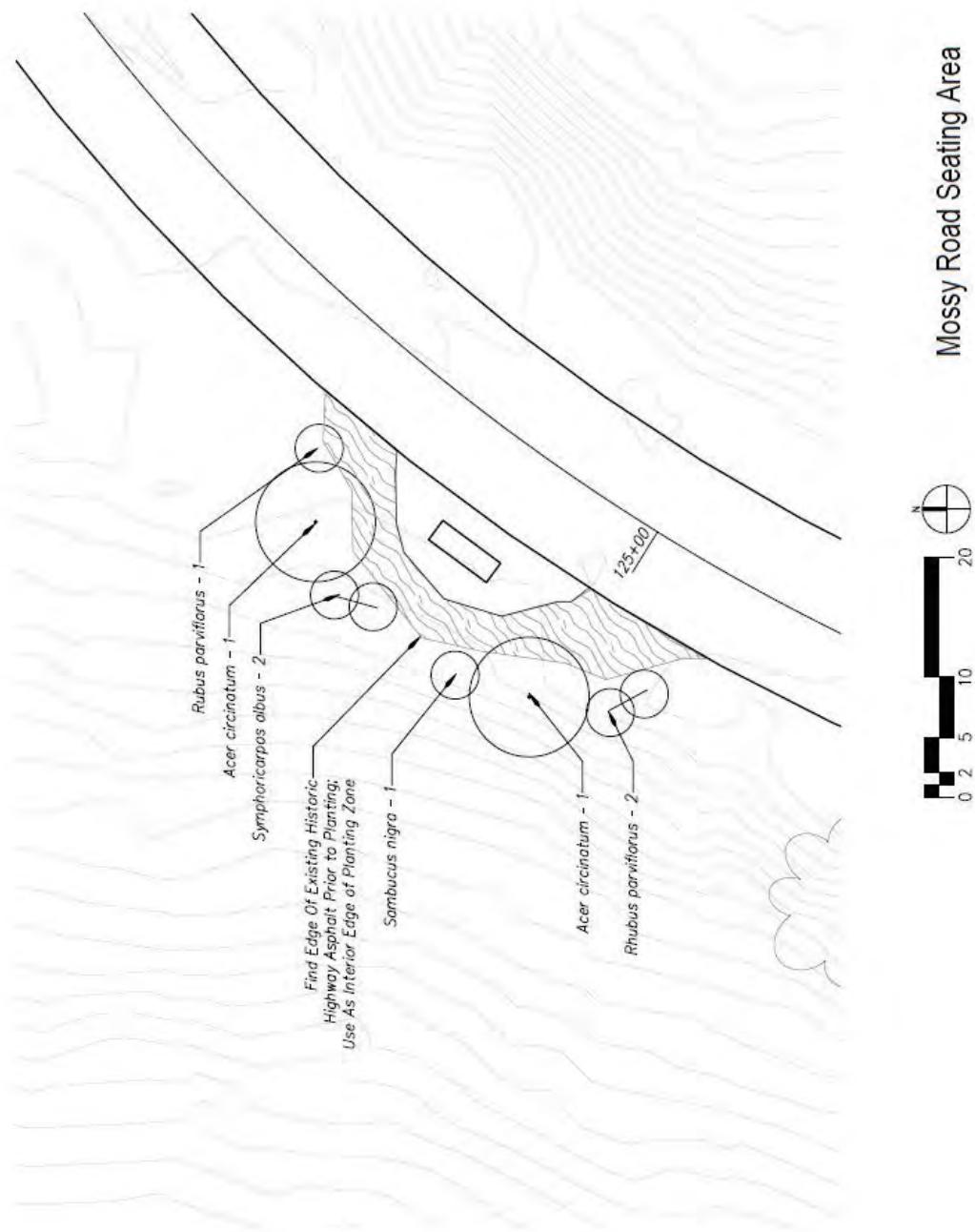


Historic Columbia River Highway State Trail, Segments A-C Final Revegetation Plan

Page 34

0 2 5 10 20  
Pull-off Near Memorial Plaque

## APPENDIX TWO



Page 35

Historic Columbia River Highway State Trail, Segments A-C Final Revegetation Plan

## APPENDIX THREE

### Appendix Three- Oregon Noxious Weed List

Table I: A Listed weeds

| Common name               | Scientific name                            |
|---------------------------|--|
| African rue (T)           | <i>Peganum harmala</i>                     |
| Cape-ivy                  | <i>Delairea odorata</i>                    |
| Camelthorn                | <i>Alhagi pseudalhagi</i>                  |
| Coltsfoot                 | <i>Tussilago farfara</i>                   |
| Cordgrass                 |  |
| Common (T)                | <i>Spartina anglica</i>                    |
| Dense-flowered (T)        | <i>Spartina densiflora</i>                 |
| Saltmeadow (T)            | <i>Spartina patens</i>                     |
| Smooth (T)                | <i>Spartina alterniflora</i>               |
| Common frogbit            | <i>Hydrocharis morsus-ranae</i>            |
| European water chestnut   | <i>Trapa natans</i>                        |
| Flowering rush (T)        | <i>Butomus umbellatus</i>                  |
| Garden yellow loosestrife | <i>Lysimachia vulgaris</i>                 |
| Giant hogweed (T)         | <i>Heracleum mantegazzianum</i>            |
| Goatgrass                 |  |
| Barbed (T)                | <i>Aegilops triuncialis</i>                |
| Ovate                     | <i>Aegilops ovata</i>                      |
| Goatsrue (T)              | <i>Galega officinalis</i>                  |
| Hawkweed                  |  |
| King-devil                | <i>Pilosella piloselloides (Hieracium)</i> |
| Mouse-ear (T)             | <i>Pilosella pilosella (Hieracium)</i>     |
| Orange (T)                | <i>Pilosella aurantiacum (Hieracium)</i>   |
| Yellow (T)                | <i>Pilosella floribundum (Hieracium)</i>   |
| Hoary alyssum (T)         | <i>Berteroa incana</i>                     |
| Hydrilla                  | <i>Hydrilla verticillata</i>               |
| Japanese dodder           | <i>Cuscuta japonica</i>                    |
| Kudzu (T)                 | <i>Pueraria lobata</i>                     |
| Matgrass (T)              | <i>Nardus stricta</i>                      |
| Oblong spurge (T)         | <i>Euphorbia oblongata</i>                 |
| Paterson's curse (T)      | <i>Echium plantagineum</i>                 |
| Purple nutsedge           | <i>Cyperus rotundus</i>                    |
| Ravennagrass              | <i>Saccharum ravennae</i>                  |
| Silverleaf nightshade     | <i>Solanum elaeagnifolium</i>              |
| West Indian spongeplant   | <i>Limnobium laevigatum</i>                |
| Squarrose knapweed (T)    | <i>Centaurea virgata</i>                   |
| Starthistle               |  |
| Iberian (T)               | <i>Centaurea iberica</i>                   |
| Purple (T)                | <i>Centaurea calcitrapa</i>                |
| Syrian bean-caper         | <i>Zygophyllum fabago</i>                  |
| Thistle                   |  |
| Plumeless (T)             | <i>Carduus acanthoides</i>                 |
| Smooth distaff            | <i>Carthamus baeticus</i>                  |
| Taurian (T)               | <i>Onopordum tauricum</i>                  |
| Woolly distaff (T)        | <i>Carthamus lanatus</i>                   |
| Water soldiers            | <i>Stratiotes aloides</i>                  |
| White bryonia             | <i>Bryonia alba</i>                        |
| Yellow floating heart (T) | <i>Nymphoides peltata</i>                  |
| Yellowtuft (T)            | <i>Alyssum murale, A. corsicum</i>         |

\* Indicates weeds targeted for biocontrol

(T) T designated species

## APPENDIX THREE

Table II: B listed weeds

| Common name                     | Scientific name   |
|---------------------------------|---|
| Armenian (Himalayan) blackberry | <i>Rubus armeniacus</i> ( <i>R. procerus</i> , <i>R. discolor</i> ) |
| Biddy-biddy                     | <i>Acaena novae-zelandiae</i>                                       |
| Broom                           |   |
| French*                         | <i>Genista monspessulana</i>  |
| Portuguese (T)                  | <i>Cytisus striatus</i>   |
| Scotch*                         | <i>Cytisus scoparius</i>  |
| Spanish                         | <i>Spartium junceum</i>   |
| Buffalobur                      | <i>Solanum rostratum</i>  |
| Butterfly bush                  | <i>Buddleja davidii</i> ( <i>B. variabilis</i> )                    |
| Common bugloss (T)              | <i>Anchusa officinalis</i>  |
| Common crupina                  | <i>Crupina vulgaris</i>   |
| Common reed                     | <i>Phragmites australis</i>   |
| Creeping yellow cress           | <i>Rorippa sylvestris</i>   |
| Cutleaf teasel                  | <i>Dipsacus laciniatus</i>  |
| Dodder                          | <i>Cuscuta</i> spp.   |
| Dyer's woad                     | <i>Isatis tinctoria</i>   |
| English ivy                     | <i>Hedera helix</i> ( <i>H. hibernica</i> )                         |
| Eurasian watermilfoil           | <i>Myriophyllum spicatum</i>  |
| False brome                     | <i>Brachypodium sylvaticum</i>                                      |
| Field bindweed* (T)             | <i>Convolvulus arvensis</i>   |
| Garlic mustard (T)              | <i>Alliaria petiolata</i>   |
| Geranium                        |   |
| Herb Robert                     | <i>Geranium robertianum</i>   |
| Shiny leaf geranium             | <i>Geranium lucidum</i>   |
| Gorse* (T)                      | <i>Ulex europeus</i>  |
| Halogeton                       | <i>Halogeton glomeratus</i>   |
| Houndstongue                    | <i>Cynoglossum officinale</i>                                       |
| Indigo bush                     | <i>Amorpha fruticosa</i>  |
| Johnsongrass                    | <i>Sorghum halepense</i>  |
| Jointed goatgrass               | <i>Aegilops cylindrica</i>  |
| Jubata grass                    | <i>Cortaderia jubata</i>  |
| Knapweeds                       |   |
| Diffuse*                        | <i>Centaurea diffusa</i>  |
| Meadow*                         | <i>Centaurea pratensis</i>  |
| Russian*                        | <i>Acroptilon repens</i>  |
| Spotted* (T)                    | <i>Centaurea stoebe</i> ( <i>C. maculosa</i> )                      |
| Knotweeds                       |   |
| Giant                           | <i>Fallopia sachalinensis</i> ( <i>Polygonum</i> )                  |
| Himalayan                       | <i>Polygonum polystachyum</i>                                       |
| Japanese                        | <i>Fallopia japonica</i> ( <i>Polygonum</i> )                       |
| Kochia                          | <i>Kochia scoparia</i>  |
| Lesser celandine                | <i>Ranunculus ficaria</i>   |
| Meadow hawkweed (T)             | <i>Pilosella caespitosa</i> ( <i>Hieracium</i> )                    |
| Mediterranean sage              | <i>Salvia aethiopis</i>   |
| Medusahead rye                  | <i>Taeniatherum caput-medusae</i>                                   |

\* Indicates weeds targeted for biocontrol      (T) T designated species

## APPENDIX THREE

| Continuation of B listed weeds   |  |
|--|--|
| Old man's beard  | <i>Clematis vitalba</i>                                  |
| Parrot feather   | <i>Myriophyllum aquaticum</i>                            |
| Perennial peavine  | <i>Lathyrus latifolius</i>                               |
| Perennial pepperweed (T)   | <i>Lepidium latifolium</i>                               |
| Pheasant's eye   | <i>Adonis aestivalis</i>                                 |
| Poison hemlock   | <i>Conium maculatum</i>                                  |
| Policeman's helmet   | <i>Impatiens glandulifera</i>                            |
| Puncturevine*  | <i>Tribulus terrestris</i>                               |
| Purple loosestrife*  | <i>Lythrum salicaria</i>                                 |
| Ragweed  | <i>Ambrosia artemisiifolia</i>                           |
| Ribbongrass (T)  | <i>Phalaris arundinacea var. <i>Picta</i></i>            |
| Rush skeletonweed* (T)   | <i>Chondrilla juncea</i>                                 |
| Saltcedar* (T)   | <i>Tamarix ramosissima</i>                               |
| Small broomrape  | <i>Orabanche minor</i>                                   |
| South American waterweed   | <i>Egeria densa (<i>Elodea</i>)</i>                      |
| Spanish heath  | <i>Erica lusitanica</i>                                  |
| Spikeweed  | <i>Hemizonia pungens</i>                                 |
| Spiny cocklebur  | <i>Xanthium spinosum</i>                                 |
| Spurge laurel  | <i>Daphne laureola</i>                                   |
| Spurge   |  |
| Leafy* (T)   | <i>Euphorbia esula</i>                                   |
| Myrtle   | <i>Euphorbia myrsinites</i>                              |
| St. Johnswort*   | <i>Hypericum perforatum</i>                              |
| Sulfur cinquefoil  | <i>Potentilla recta</i>                                  |
| Swainsonpea  | <i>Sphaerophysa salsula</i>                              |
| Tansy ragwort* (T)   | <i>Senecio jacobaea (<i>Jacobaea vulgaris</i>)</i>       |
| Thistles   |  |
| Bull*  | <i>Cirsium vulgare</i>                                   |
| Canada*  | <i>Cirsium arvense</i>                                   |
| Italian  | <i>Carduus pycnocephalus</i>                             |
| Milk*  | <i>Silybum marianum</i>                                  |
| Musk*  | <i>Carduus nutans</i>                                    |
| Scotch   | <i>Onopordum acanthium</i>                               |
| Slender-flowered*  | <i>Carduus tenuiflorus</i>                               |
| Toadflax   |  |
| Dalmatian* (T)   | <i>Linaria dalmatica</i>                                 |
| Yellow*  | <i>Linaria vulgaris</i>                                  |
| Tree of heaven   | <i>Ailanthus altissima</i>                               |
| Velvetleaf   | <i>Abutilon theophrasti</i>                              |
| Water primrose, floating primrose-willow, large-flower primrose willow (T) | <i>Ludwigia peploides, L. hexapetala, L. grandiflora</i> |
| Whitetop   |  |
| Hairy  | <i>Lepidium pubescens</i>                                |
| Lens-podded  | <i>Lepidium chalepensis</i>                              |
| Whitetop (hoary cress)   | <i>Lepidium draba</i>                                    |

\* Indicates weeds targeted for biocontrol     (T) T designated species

| Continuation of B listed weeds |                               |
|--------------------------------|-------------------------------|
| Yellow archangel               | <i>Lamiastrum galeobdolon</i> |
| Yellow flag iris               | <i>Iris pseudacorus</i>       |
| Yellow nut sedge               | <i>Cyperus esculentus</i>     |
| Yellow starthistle*            | <i>Centaurea solstitialis</i> |

\* Indicates weeds targeted for biocontrol     (T) T designated species

# RESOURCES

## References

The following resources were not cited directly, but were referenced in the development of this document.

- Bennett, Max. 2007. Managing Himalayan blackberry in western Oregon riparian areas. Oregon State University Extension Service, Oregon State University. Corvallis, Oregon.
- Bergerson, Terry. 2012. Visitor survey of day-use visitors at Starvation Creek State Park. Oregon Parks and Recreation Department.
- Biggerstaff, Matthew and Christopher Beck. 2007. Effects of method of English ivy removal and seed addition on regeneration of vegetation in a southeastern Piedmont forest. *The American Midland Naturalist* 158: 206-220.
- Binelli, Eliana K., Gholz, Henry L., and Mary L. Duryea. 2000. Chapter 4: Plant succession and disturbances in the urban forest ecosystem. In: Restoring the urban forest ecosystem. The School of Forest Resources and Conservation, Florida Cooperative Extension Service, Institute of Food and Agricultural Sciences, University of Florida.
- CH2MHill, Inc. 2014. Wetlands/Waters delineation report, Historic Columbia River Highway State Trail Segments A-D. Columbia River Gorge National Scenic Area Task Order No. T-13-001, Modification 004, Task 3.8 Multi-Discipline IDIQ Contract No. DTFH70-10-D-00020.
- CH2MHill, Inc. 2014. Mitigation Plan: OR DOT CRGNSA 100(1) Historic Columbia River Highway State Trail (Segments A-D) Columbia River Gorge National Scenic Area Task Order No. T-13-001, Modification 006, Task 3.9 Multi-Discipline IDIQ Contract No. DTFH70-10-D-00020.
- CH2MHill, Inc. 2015. Draft Design Acceptance Package, Historic Columbia River Highway State Trail: Gorton Creek to Lindsey Creek (Segments A-C). Prepared for Western Federal Lands Highway Division and Oregon Department of Transportation.
- Dlugosch, Katrina M. 2005. Understory community changes associated with English ivy invasions in Seattle's urban parks. *Northwest Science* 79(1): 52-59.
- Dobson, Robin. 2015. *Personal communication*. Email to author, April 20, 2015.
- Flory, Luke S. and Keith Clay. 2009. Invasive plant removal method determines native plant community responses. *Journal of Applied Ecology* 46: 434-442.

## RESOURCES

Green, George L. 1981. Soil survey of Hood River County area, Oregon. United States Department of Agriculture, Soil Conservation Service, in cooperation with the Oregon Agricultural Experiment Station.

Hood River County Chamber of Commerce. Hood River County, About Hood River.  
Hoodriver.org

Hopwood, Jennifer, Black, Scott, and Scott Fleury. 2016. Pollinators and roadsides: Best management practices for managers and decision makers. Prepared for the Federal Highway Administration.

Invasive plant species assessment working group. Invasive plant species fact sheet: Periwinkle *Vinca minor*. [www.invasivespecies.IN.gov](http://www.invasivespecies.IN.gov)

King County Noxious Weed Control Program. 2014. Himalyan blackberry best management Practices.

Michigan Department of Natural Resources. 2012. Invasive species- Best control practices: black locust (*Robinia pseudoacacia*).

Oregon Plant Atlas, Oregon Flora Project. Oregon State University. [www.oregflora.org/atlas.php](http://www.oregflora.org/atlas.php)

Pollinator Health Task Force. 2015. National strategy to promote the health of honey bees and other pollinators. The White House.

Quatrefoil, Inc. 2010. The Historic Columbia River Highway State Trail plan- Wyeth to Hood River.

Steinfeld, David E., Riley, Scott A., Wilkinson, Kim M., Landis, Tom D., Riley, Lee E.. 2007. Roadside revegetation: An integrated approach to establishing native plants. Federal Highway Administration, Western Federal Lands Highway Division report number FHWA-WFL/TD-07-005.

United States Department of Agriculture, Forest Service. 2009. Gifford Pinchot National Forest south zone invasive plant treatment 2009 implementation plan.

United States Department of the Interior, Bureau of Land Management. National seed strategy for rehabilitation and restoration.

<http://www.blm.gov/ut/st/en/prog/more/CPNPP/0/seedstrategy.html>

## RESOURCES

Ward, K., Cariveau, E., May, M., Roswell, M., Vaughan, N., Williams, R., Winfree, R., Isaacs, and K. Gill. 2014. Streamlined bee monitoring protocol for assessing pollinator habitat. Portland, OR: The Xerces Society for Invertebrate Conservation.

Wisconsin Department of Natural Resources. 2004. Black locust (*Robinia pseudoacacia*).

# 5—Implementation

- 5.1 Introduction**
- 5.2 Soil and Site Treatments**
- 5.3 Obtaining Plant Materials**
- 5.4 Installing Plant Materials**
- 5.5 Post-Installation Care of Plant Materials**



## 5.1 INTRODUCTION

A successful revegetation project involves not only good planning but effective implementation of the plan. The Initiation Phase of this report ([Chapter 2](#)) described the organizations, decision processes, and technical concepts involved in beginning a roadside revegetation project. The Planning Phase chapter ([Chapter 3](#)) outlined the steps in the revegetation planning process resulting in a revegetation plan. This chapter details how the revegetation plan is translated into action at the project site.

The shift from the planning phase to the implementation phase involves a change in approach, and often, personnel. The planning process tends to be orderly and systematic, with the design team able to take an idealized bird's-eye perspective of how the project might best proceed. In contrast, an implementation approach that is flexible and adaptable can help ensure the objectives of the plan are fulfilled while working with the sometimes-unpredictable nature of construction.

The tasks necessary to successfully implement a revegetation plan are:

- Developing contracts
- Developing an implementation schedule
- Maintaining a materials inventory
- Reviewing plans and coordinating with project engineer
- Assisting in implementing contracts

This chapter provides an overview of the implementation process, providing details on the key factors to consider, including soil and site treatments, obtaining plant materials, installing plant materials, and caring for plants after they have been installed.

### 5.1.1 DEVELOPING CONTRACTS

Revegetation tasks can be performed in-house, by agency personnel, or through contracts, either performed within the scope of road construction contracts or as separate revegetation contracts. The FHWA and most state DOTs have developed revegetation contract specifications within their standard specifications for construction of roads and bridges. These specifications are designed, in most cases, to be included directly into a road construction contract or revegetation contract. As a designer, it is important that the language in the contract specification accurately describes both the work to be done and the desired outcome, from a revegetation standpoint. Because every revegetation project is different in scope and environmental setting, it is essential that these factors are conveyed in the specification.

Contract specifications detail the *how*, not the *why*. To understand the background for a specification for revegetation, it may be useful to review the implementation guides presented in this chapter that pertain to the specific task or topic. With this information, a special provision or supplemental specification based on standard DOT or other agency specifications can be developed to respond to the specifics of the project and environmental surroundings. It may also be helpful to review contract specification examples developed by other agencies. Some revegetation specifications for the FHWA and several state DOT's can be found in the [Native Revegetation Resource Library](#) by selecting "Contract Specifications" under the Report Type search filter.

***For the Designer***  
**Special Provisions or Supplemental Specifications need agency review and approval and the process may take several months. Identification, development, and submittal of these specifications early in the project will reduce delay of final project review approvals for bidding.**

To evaluate the implementation of contracts, quality assurance standards are defined in the contract specifications. The contractor typically provides monitoring and inspection of these standards to ensure that tasks are carried out as contracted. For example, a hydroseeding contract may have a specification to apply 30 pounds of seeds, 1,000 pounds of wood fiber mulch, and 50 pounds of tackifier per acre. Quality control for this task might be to count the pounds of seeds, wood fiber, and tackifier going into the hydroseeder tank and measure the area to which the mix is applied. This assessment determines if that the rate of materials applied is within quality standards and, if not, alerts the contractor to make appropriate corrections. The written results of these assessments become “as-built” records which can be used by the designer when developing the monitoring report ([Section 6.2](#)).

Accurately calculating the contract area is essential in developing contracts. Table 5-1 is an example of how cross sections can be used to determine the contract area. When cross sections are not available, the contract area can be determined from the plan map. In this method, the area is measured and adjustments are made for slope gradient (Table 5-2). A third method is to physically measure the distances in the

**Table 5-1 | Calculating project area from road plans and cross sections**

In this example, cross sections are available for each station. The length of each cross section is measured and recorded in a spreadsheet. The slope length is multiplied by the distance between stations to calculate the area for that station. When all stations areas are calculated, the total contract area is summarized.

field.

| Station      | Slope length (m) | Distance between stations (m) | Area (m <sup>2</sup> ) |
|--------------|------------------|-------------------------------|------------------------|
| 20+1000      | 3                | 20                            | 60                     |
| 20+1020      | 4                | 20                            | 80                     |
| 20+1040      | 6                | 20                            | 120                    |
| 20+1060      | 7                | 20                            | 140                    |
| 20+1080      | 6                | 20                            | 120                    |
| 20+1100      | 4                | 20                            | 80                     |
| 20+1120      | 2                | 20                            | 40                     |
| <b>Total</b> |                  |                               | <b>640</b>             |
| Acres        |                  |                               | 0.16                   |

**Table 5-2 | Area computations adjusted for slope gradient**

When cross sections are not available, the plan map can be used to calculate area. On steeper terrain, the area may need to be adjusted by slope gradient based on the following factors.

| Slope (V:H) | Increase factor | Percent of increase |
|-------------|-----------------|---------------------|
| 1:1         | 1.41            | 41%                 |
| 1:1.5       | 1.20            | 20%                 |
| 1:2         | 1.12            | 12%                 |
| 1:2.5       | 1.08            | 8%                  |
| 1:3         | 1.05            | 5%                  |
| 1:4         | 1.03            | 3%                  |
| 1:5         | 1.02            | 2%                  |

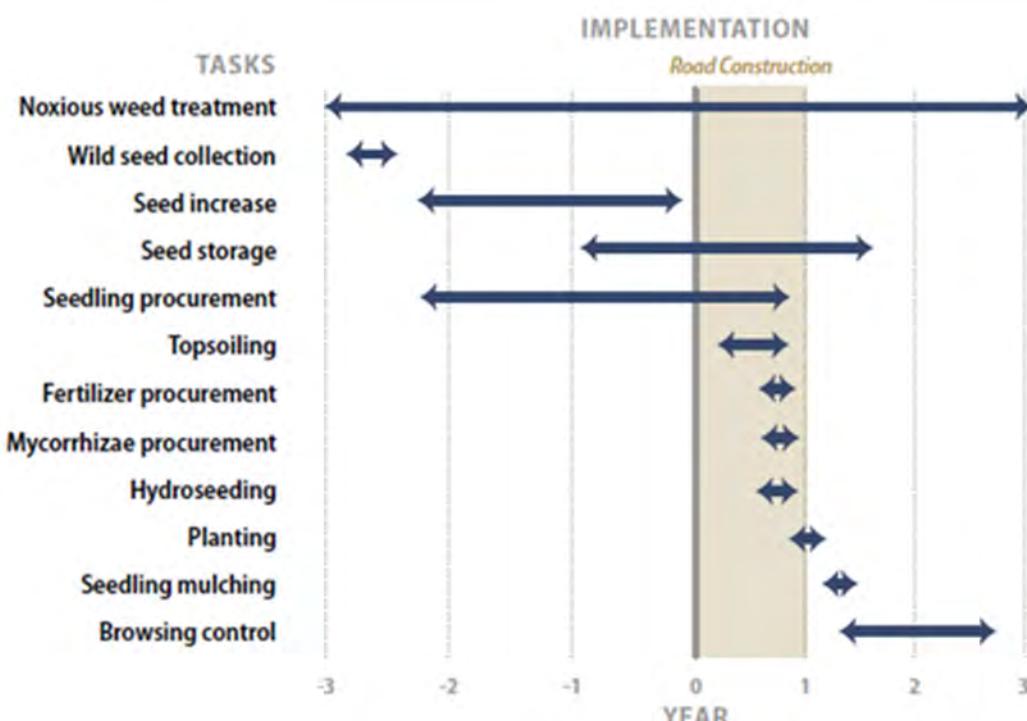
The contract specifies the work to be done by the contractor or agency personnel, with clear definitions of roles and schedules.

In general, a contract outlines the following:

- Supplies/services to be provided
- Scope of work (size, schedule, etc.)
- Project location
- Contractor obligations
- Revegetation specialist obligations
- Delivery details (who, how, when, including timelines and deadlines)
- Quality standards
- Contractor quality assurance plan (to be provided by contractor)
- Revegetation specialist quality assurance plan (to be provided by designer who will be inspecting the contractor's work)
- Price (including bid, unit prices, additive or deductive alternate items)
- Payment method (submission of invoices, approval of work)
- Contractor's designated representative (more efficient to only coordinate with one person)
- Safety plan
- Other terms and conditions (e.g., what to do in the event of changes)

### 5.1.2 MAINTAINING SCHEDULES AND MATERIALS INVENTORY

Revegetation contracts can begin up to three years before road construction. For example, it is not uncommon for a seed or plant procurement contract to begin during the planning stage of a road project and span into road construction. In addition, procuring plant materials and implementing mitigating treatments often involve separate schedules with different contractors. To keep these details straight, developing an implementation schedule can be helpful. Timelines are often developed during the planning stages and included in the revegetation plan and these become the basis of an implementation schedule.



**Figure 5-1 | Example implementation schedule**

In this implementation schedule overlapping tasks, some requiring different contractors, are displayed.

Most schedules have interdependent and time-linked tasks. In Figure 5-1, for example, the hydroseeding contract is dependent on the success of the seed increase contract. Before that, the seed increase contract requires wildland seeds to be collected by agency personnel or contractors. The success of the hydroseeding contract rests on the successful execution of two tasks that begin years before the actual seeding.

When creating schedules, it is important to incorporate additional time needed to issue contracts. This is especially crucial when ordering plant materials, which will likely require several years for production.

Material quantities for revegetation tasks change through the life of a road project. For example, a nursery may have succeeded in growing more seedlings for certain species, while producing less of others (due to poor germination, frosts, insects, disease, or other events). For plant material contracts, it is important to stay in touch with the growers to determine if there are any changes to the inventories, and make adjustments to the inventory as needed. Other material quantities that can change are topsoil and shredded wood for mulch.

### 5.1.3 COORDINATING WITH CONSTRUCTION ENGINEER

For some road projects, one engineer will oversee the project from inception through its completion. For more complex projects however, the design engineer may hand off the project to a construction engineer member of the project team or from the DOT or agency client to implement. The design engineer, in this case, ensures that tasks outlined in the revegetation plan are designed into the construction documents: plans, specifications, contracts, schedules, materials lists, and coordinated with erosion control plans. The construction engineer implements the road plan with guidance from the design team. Complex revegetation designs, such as vegetated retaining walls, planting pockets, and other biotechnical engineering structures, may require more involvement from the design team.

Before construction begins, a meeting is often scheduled with the construction engineer, road building contractor, and revegetation design team to discuss the revegetation plan. This meeting covers revegetation objectives and outlines the revegetation tasks that need to be coordinated with road construction activities. The construction engineer is made aware of the revegetation treatments and how they are to be implemented. After the initial meeting, the revegetation designer may keep in contact with the construction engineer to help implement the contract specifications. Timely field visits by the designer will help overcome difficulties in integrating revegetation treatments into construction design and schedule.

### 5.1.4 IMPLEMENTATION GUIDES

Implementation guides are presented in this chapter as background for developing contracts or procedures for revegetation tasks.

The following implementation guides are grouped into four subject areas:

- [Section 5.2 Soil and Site Treatments](#)
- [Section 5.3 Obtaining Plant Materials](#)
- [Section 5.4 Installing Plant Materials](#)
- [Section 5.5 Post-Installation Care of Plant Materials](#)

The eight guides in [Section 5.2](#) explain how to improve site and soil conditions prior  
Roadside Revegetation: An Integrated Approach to Establishing Native Plants and Pollinator Habitat

to the installation of plant materials. These guides cover the mitigating measures most often referenced in [Section 3.8](#).

[Section 5.3](#) includes six implementation guides that pertain to collecting and propagating plant materials. These guides describe how to take the species lists developed in [Section 3.13](#), and obtain the desired species in the wild as seed, cuttings, or seedlings. These guides also cover how to increase gathered wild collections at nurseries to ensure that the revegetation project has sufficient quantities of plant materials.

Once plant materials are obtained from the wild or from nurseries, they are installed on the project site. The four guides in [Section 5.4](#) cover the techniques for sowing seed, installing cuttings, and planting seedlings. They also cover how to determine the quality of the plant materials and how to care for them during storage and transportation.

[Section 5.5](#) outlines those practices that occur after the installation of plant materials. These practices help ensure that plants will become established. Practices include protecting seedlings from animal browsing, installing shade cards, irrigating, and installing tree shelters.

Other sources for implementation guides can be found in the [Native Revegetation Resource Library](#) by selecting “Implementation Guides” under the Report Type search filter.

## 5.2 SOIL AND SITE TREATMENTS

Most post-construction sites are in poor condition for plant growth and will likely require implementing a set of mitigation measures if good revegetation is expected. The following set of implementation guides cover the common mitigating measures for improving site conditions after construction.

- **Fertilizers**—Covers how to determine fertilizer quantity, type, and application method.
- **Tillage**—Describes the common practices of tilling the soil to improve water infiltration and root growing environment.
- **Mulches**—Seed germination, seedling survival, and surface erosion can be improved through the application of mulches.
- **Topsoil**—Outlines the removal, storage, and application of topsoil to reconstruct soils on highly disturbed sites. For sites where topsoil is not available or in short supply, organic matter can be applied to improve post-construction soils.
- **Organic Matter**—Discusses the types of organic matter, how to determine rates, and how it is applied. On some sites where the topsoil has been removed, pH levels may need to be raised to improve plant growth.
- **Lime Amendments**—Details materials, application methods, and how to determine liming rates.
- **Beneficial Microorganisms**—Many sites devoid of topsoil need mycorrhizae or nitrogen fixing plants to be introduced. This section covers how to obtain and apply the appropriate sources of these important biological organisms.
- **Topographic Enhancements**—Revegetation projects can be enhanced by integrating plants into bioengineering structures, water capture features, or planting islands or pockets.

## 5.2.1 FERTILIZERS

### Introduction

Fertilizers are used to bring soil nutrients up to levels essential for establishing and maintaining a desired plant community. When applied within a soil fertility strategy, fertilizer can be a good tool for revegetation but not necessarily needed for every project. In recent years, the use of fertilizers on roadsides for native plant establishment has come under greater public scrutiny and more restrictive water quality laws. Many roads are adjacent to streams, lakes, or residential areas that can be affected by runoff or leaching of inappropriately applied fertilizers. In some instances, fertilizers may not be recommended for establishing native vegetation (see Iowa Roadside Vegetation Management Handbook) because of the potential of encouraging invasive species over native plants. It is important for the designer to learn how to develop fertilizer prescriptions that integrate short- and long-term site fertility goals with water quality and native plant establishment objectives.

It is important to base a soil fertility strategy on the nutrient levels found in the reference soils when considering the application of topsoil, mulch, compost, wood waste, biosolids, and/or the planting of nitrogen-fixing species. In addition, using commercial fertilizer with other methods of raising nutrient levels, can result in a greater long-term nutrient management of the revegetation project (Section 3.8.4).

This section guides the designer through the steps necessary of developing a fertilizer prescription which is the instructions for ordering and applying fertilizers.

They include:

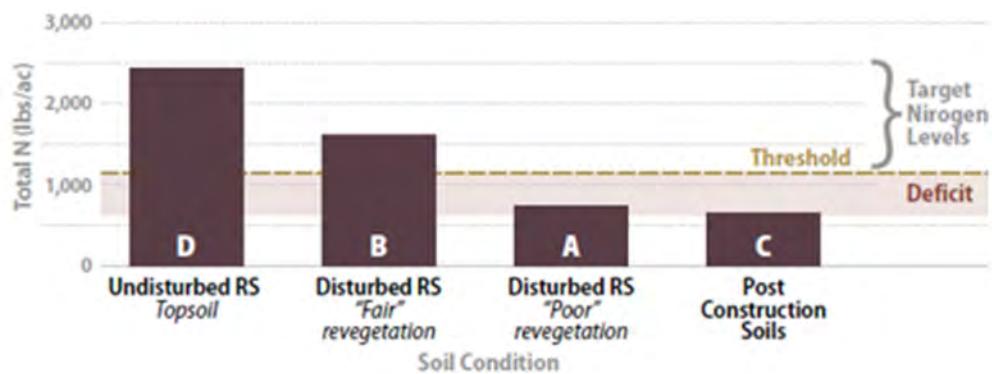
- Determine nutrient thresholds and deficits
- Delineate areas to be fertilized
- Select fertilizer analysis
- Select fertilizer release rates
- Determine application rates
- Determine timing and frequency
- Select application method

### Develop Nutrient Thresholds and Determine Deficits

All sites have a minimum, or threshold, level of nutrients that needs to be met for each plant community to become functioning and self-sustaining (Section 3.8.4). Threshold values can be determined by comparing soil tests from several disturbed and undisturbed reference sites (Section 3.5). Finding disturbed reference sites that range from poor success to good success provides a good understanding of nutrient levels and plant response. Based on nutrient values from good and poor revegetation sites, a target nitrogen range can be established between these values. Figure 5-2 provides an example of how a nitrogen threshold value was obtained by evaluating the total soil nitrogen levels from two disturbed reference sites, one considered "fair" revegetation and one considered "poor." The threshold was set between these two nitrogen levels. Threshold levels represent the minimum level of nutrients needed for a site. In this example, the target nitrogen range for establishing and maintaining the original plant community would be between the minimum nitrogen levels and the nitrogen levels found in the undisturbed reference sites.

#### *For the Designer*

Base fertilizer recommendations on soil tests and native plant needs.



**Figure 5-2 | Threshold values of nitrogen**

Threshold values are determined from reference sites. In this example, the threshold was established at 1,100 lb/ac, which was between the total N of a disturbed reference site with “poor” revegetation (A) and one with “fair” revegetation (B). Total N in post-construction soils was 650 lb/ac (C), making these soils deficient by 450 lb/ac. The undisturbed topsoil of reference sites showed a total N of 2,430 lb/ac (D), which sets the target nitrogen range between 1,100 and 2,430 lb/ac.

To determine whether nutrients are deficient, soil samples are collected from the post-construction sites and tested (Inset 3-2). The nutrient values obtained from these tests are compared against the target ranges to determine if a deficiency exists. By comparing post-construction nutrient values against threshold values, the nutrient deficit can be estimated for each nutrient. Figure 5-3 shows an example of how nitrogen deficits are calculated based on post-construction soil tests. In this example, total soil nitrogen is determined from soil tests. Because soil testing facilities report nutrients in a variety of rates, it is important to convert the rates to percentages then to pounds per acre. Values reported as gr/l, ppm, mg/kg, and ug/g are converted to percentages by multiplying by 10,000. Converting nutrient percentage to lb/ac of the nutrient is done by multiplying percent of nutrient, soil layer thickness, soil bulk density, and fine soil fraction together with a constant (Line E). The result is the pounds of nutrient in an acre of soil on the post-construction site. To determine if the nutrient is deficit, the pounds of nutrients per acre is subtracted from the threshold level (Line F). This value (Line G) represents the nitrogen deficiency and becomes the basis for determining fertilizer prescriptions.

|   |  |              |   |
|---|--|--------------|---|
| A | Total soil nitrogen (N)                    | 0.025%       | From soil test of post construction soils (gr/l, ppm, mg/kg, ug/g divide by 10,000 for %)                   |
| B | Thickness of soil layer                    | 0.5 feet     | The thickness of soil represented in A  |
| C | Soil bulk density                          | 1.4 gr/cc    | Unless known, use 1.5 for compacted subsoils, 1.3 for undisturbed soils, 0.9 for light soils such as pumice |
| D | Fine soil fraction                         | 70%          | 100% minus the rock fragment content – from estimates made from sieved soil prior to sending to lab         |
| E | N in soil layer<br>$A * B * C * D * 270 =$ | 331 lbs/ac   | Calculated amount of total nitrogen in soil layer. To convert to kg/ha: $E * 1.2$                           |
| F | Minimum or threshold N levels              | 1,100 lbs/ac | Determined from reference sites (see Figure 5-2)  |
| G | N deficit $F - E =$                        | 769 lbs/ac   | Minimum amount of N to apply to bring up the threshold  |

**Figure 5-3 | Determining nitrogen needs from soil tests**

Determining the amount of nitrogen (N) needed to bring soils up to a nitrogen threshold can be calculated from equations shown in this spreadsheet.

The availability of many nutrients is regulated by soil pH. As discussed in Section 3.8.4, many nutrients are tied up in low pH and high pH soils. Calcium and

magnesium are less available at low pH; phosphorus, iron, manganese, boron, zinc, and copper become unavailable in high pH soils. It is important to compare the pH of post-construction soils with reference site soils to determine if the pH is substantially different between the two. If the pH of post-construction soils is different, then taking measures to bring the pH closer to pre-disturbance values is important when developing a nutrient strategy (Section 5.2.6).

## Delineate Areas to be Fertilized

Because the post-construction site may differ in soil types and disturbance levels, it is important to delineated areas where fertilizer prescriptions may differ. These differences are usually based on post-construction soil type changes, topsoil salvage, organic amendment additions, or the species and plant material being grown. Areas adjacent to, or that feed into, live water are often delineated and treated with lower rates of fertilizer. Note: If seedlings of shrubs and trees are being planted, spot fertilization may be a more appropriate method than fertilizing the entire area (Inset 5-1).

### **Inset 5-1 / Spot-fertilizing seedlings**

Fertilizing shrub or tree seedlings is done by placing fertilizer in each seedling hole or on the soil surface after each seedling has been planted. This practice has some risks because fertilizers release salts that can damage roots. Studies have shown that placing fertilizers or liming materials in the planting hole or on the soil surface around seedlings at the time of planting can significantly decrease seedling survival, especially on droughty sites (Nursery Technical Cooperative 2004; Jacobs et al 2004; Walker 2002).

These practices may reduce the likelihood of seedling damage:

- Assess the need for fertilizer (do not apply if nutrient levels are adequate)
- Use slow release fertilizers with low salt indexes
- Use low rates of fertilizer if applying at the time of planting
- If applying in seedling hole at planting, use low fertilizer rates and place fertilizer to the side at least 3 inches away from the root system (see figure)
- Preferably broadcast fertilizers on the soil surface after seedlings are well established

When slow-release fertilizers are spread around well-established seedlings (several years after planting), seedlings often respond favorably, especially on highly disturbed sites. Walker (2005) found that slow-release fertilizers applied around the seedlings three years after they were planted increased stem diameter and shoot volume by 143 percent and 104 percent, respectively, over the control when they were measured five years later. In this study, rates of .05 grams of nitrogen per seedling showed the greatest response (at 8 percent nitrogen analysis, this would be over a half pound of bulk slow-release fertilizer per plant).



**21-5-20**

**All Purpose Fertilizer**

*Guaranteed Minimum Fertilizer*

|   |               |
|---|---------------|
| <b>Total nitrogen (N) .....</b>                               | <b>21.00%</b> |
| Ammonium nitrogen .....                                       | 6.50%         |
| Nitrate nitrogen.....   | 12.40%        |
| Urea nitrogen.....  | 2.10%         |
| <b>Available phosphate (P<sub>2</sub>O<sub>5</sub>) .....</b> | <b>5.00%</b>  |
| <b>Soluble potash (K<sub>2</sub>O) .....</b>                  | <b>20.00%</b> |
| Boron (B).....  | 0.02%         |
| Copper (Cu) .....   | 0.01%         |
| Iron (Fe).....  | 0.10%         |
| Water soluble manganese (Mn).....                             | 0.05%         |
| Molybdenum (Mo).....  | 0.01%         |
| <b>Water soluble zinc (Zn).....</b>                           | <b>0.07%</b>  |

**Figure 5-4 / Example of a fertilizer label for an “all purpose” fertilizer**

The three numbers (21-5-20) represent the percentage of nitrogen, phosphorus, and potassium respectively (21% N, 5% P<sub>2</sub>O<sub>5</sub>, and 20% K<sub>2</sub>O). The label may also contain percentages of other nutrients in the fertilizer. Multiplying these percentages (divided by 100) by the pounds of bulk fertilizer applied per acre will give the quantity of each nutrient applied per acre. In this analysis, applying 500 pounds of fertilizer to an acre would deliver 105 lbs N, 25 lbs P<sub>2</sub>O<sub>5</sub>, 100 lb K<sub>2</sub>O, 0.01 lbs B, 0.005 lbs Cu, etc.

## Select Fertilizer Analysis

A variety of commercially available fertilizers can be used for fertilizing disturbed sites associated with road construction. The composition, or makeup, of the fertilizer is called the fertilizer analysis. Each container of fertilizer will have a label with a stated “guaranteed analysis” that indicates the percentage of each nutrient contained in the

fertilizer (Figure 5-4). The label is the guide for determining which fertilizers to select and how much to apply. Table 5-3 and Table 5-4 provide analysis values for many common fertilizers. Labels can also be obtained from the manufacturer or fertilizer representatives.

Fertilizer labels report nutrients as a percentage. The example label for a 50 pound bag of fertilizer in Figure 5-4 shows 21 percent nitrogen (N), which indicates that 10.5 pounds of material in the bag is made up of nitrogen ( $50 * 21 / 100 = 10.5$ ). The bag also contains 0.02 percent boron (B), which indicates that there is 0.01 pounds of boron in the bag. Calculating the amount of phosphorous and potassium is different because the convention for reporting these nutrients is  $P_2O_5$  and  $K_2O$  instead of elemental P and K. To convert  $P_2O_5$  to P, the analysis for P is divided by 2.29. The percentage of P in the bag in Figure 5-4 is actually 2.2 percent, not 5 percent ( $5.0\% / 2.29 = 2.2$ ).  $K_2O$  is divided by 1.21 to obtain 1.6 percent K.

Fertilizers are selected based on whether they contain the nutrients that are deficient on the project site (Section 5.2.1, Develop Nutrient Thresholds and Determine Deficits). For example, if nitrogen, phosphorus, and boron are deficient, only fertilizers that contain these nutrients are considered. Most fertilizers contain more than one nutrient. For example, ammonium sulfate contains nitrogen and sulfur; and triple superphosphate contains phosphorus, sulfur, and calcium. Organic fertilizers contain a range of macro and micronutrients. Fertilizers containing more than one nutrient are used if the nutrients contained in these fertilizers are deficient in post-construction soils. Table 5-2 and Table 5-3 show the combination of nutrients that are available in some commercially available fertilizers.

**Table 5-3 | Analysis of common fertilizers**

| Available nutrients (typical percentages) |                             |    |                               |    |                  |    |    |    |    |      |    |    |   |    |    |
|---|-----------------------------|----|-------------------------------|----|------------------|----|----|----|----|------|----|----|---|----|----|
|   |                             | N  | P <sub>2</sub> O <sub>5</sub> | P  | K <sub>2</sub> O | K  | S  | Ca | Cu | Fe   | Mn | Zn | B | Mo | Mg |
| <b>Phosphorus</b>                         | Mono-ammonium phosphate     | 11 | 48                            | 21 |                  |    | 24 |    |    |      |    |    |   |    |    |
|   | Ammonium phosphate          | 82 |                               |    |                  |    |    |    |    |      |    |    |   |    |    |
|   | Diammonium phosphate        | 17 | 47                            | 21 |                  |    |    | 21 |    |      |    |    |   |    |    |
|   | Single superphosphate       | 19 |                               |    |                  |    | 12 | 20 |    |      |    |    |   |    |    |
|   | Triple superphosphate       |    | 45                            | 20 |                  |    | 1  | 13 |    |      |    |    |   |    |    |
|   | Phosphoric acid             |    | 53                            | 23 |                  |    |    |    |    |      |    |    |   |    |    |
|   | Dicalcium phosphate         |    |                               |    |                  |    |    |    |    |      |    |    |   |    |    |
|   | Soluble potassium phosphate |    |                               |    |                  |    |    |    |    |      |    |    |   |    |    |
| <b>Potassium</b>                          | Superphosphoric acid        | 80 | 35                            |    |                  |    |    |    |    |      |    |    |   |    |    |
|   | Potassium chloride          |    |                               |    | 61               | 50 |    |    |    |      |    |    |   |    |    |
|   | Potassium nitrate           | 13 |                               |    | 45               | 37 |    |    |    |      |    |    |   |    |    |
| <b>Micronutrients</b>                     | Potassium sulphate          |    |                               |    | 51               | 42 | 18 |    |    |      |    |    |   |    |    |
|   | EDTA                        |    |                               |    |                  |    |    | 10 | 10 | 10   | 10 |    |   |    |    |
|   | HEEDTA                      |    |                               |    |                  |    |    | 6  | 7  | 7    | 9  |    |   |    |    |
|   | NTA                         |    |                               |    |                  |    |    |    | 8  |      |    |    |   |    |    |
|   | DTPA                        |    |                               |    |                  |    |    |    | 10 |      |    |    |   |    |    |
|   | EDDHA                       |    |                               |    |                  |    |    | 6  |    |      |    |    |   |    |    |
|   | Granular borax              |    |                               |    |                  |    |    |    |    | 11.3 |    |    |   |    |    |
|   | Copper sulfate              |    |                               |    |                  |    | 25 |    |    |      |    |    |   |    |    |
|   | Ferrous sulfate             |    |                               |    |                  |    |    | 20 |    |      |    |    |   |    |    |
|   | Sodium molybdate            |    |                               |    |                  |    |    |    |    | 40   |    |    |   |    |    |
| <b>Ca &amp; Mg</b>                        | Zinc sulfate                |    |                               |    |                  |    |    |    |    | 36   |    |    |   |    |    |
|   | Zinc chelate                |    |                               |    |                  |    |    |    |    | 4    |    |    |   |    |    |
|   | Dolomitic limestone         |    |                               |    |                  |    | 21 |    |    |      |    | 11 |   |    |    |
|   | Magnesium sulfate           |    |                               |    |                  |    |    |    |    |      |    | 43 |   |    |    |
|   | Gypsum                      |    |                               |    |                  |    | 23 |    |    |      |    |    |   |    |    |
|   | Epsom salt (Epsogrow brand) |    |                               |    |                  |    | 13 |    |    |      |    | 10 |   |    |    |

**Table 5-4 | Estimated nitrogen release rates for commercially available fertilizers**

Nutrient release rates are obtained from lab testing but how they release on-site will vary from site to site, depending on temperature, moisture, and whether the fertilizer was placed on the surface or incorporated into the soil. If slow-release fertilizers are broadcast on the soil surface, release rates are slower than if incorporated into the soil where the conditions are better for break down. Arid sites have slower rates of release than sites with high moisture; cold sites take longer to release nutrients than warm sites. First-year nitrogen release rates for fertilizers are identified with an asterisk were adapted from Claassen and Hogan (1998).

Note: Non-asterisk fertilizers were based on best guess estimates.

| Source               | N %  | Available Nutrients    |                                 |                    |
|----------------------|------|------------------------|---------------------------------|--------------------|
|                      |      | 1st year N release (%) | % P <sub>2</sub> O <sub>5</sub> | % K <sub>2</sub> O |
| Ammonium nitrate     | 34   | 99 to 100              | 0                               | 0                  |
| Ammonium phosphate*  | 10   | 99 to 100              | 34                              | 0                  |
| Ammonium sulfate     | 21   | 99 to 100              | 0                               | 0                  |
| Anhydrous ammonia    | 82   | 99 to 100              | 0                               | 0                  |
| Calcium nitrate      | 15.5 | 99 to 100              | 0                               | 0                  |
| Diammonium phosphate | 18   | 99 to 100              | 46                              | 0                  |
| IBDU                 | 29   | 95 to 99               | 3                               | 10                 |
| Potassium nitrate    | 13   | 99 to 100              | 0                               | 45                 |
| Urea                 | 46   | 99 to 100              | 0                               | 0                  |

In selecting a fertilizer, the macronutrients (nitrogen, potassium, and phosphorus) that are deficient on a site are considered first because they are the most important for long-term site recovery. If these nutrients are not deficient, chances are that the remaining nutrients are not deficient either. On most highly disturbed sites, nitrogen is most likely to be deficient. Good practice is to consider using this nutrient when approaching fertilizer selection. Table 5-3 lists common nitrogen fertilizers with typical label analysis. It is common to apply more than one fertilizer to meet the various nutrient requirements of the soil. Nutrients other than nitrogen can be supplied by fertilizers shown in Table 5-2.

**Biosolids**—Biosolids are a nutrient-rich organic material produced from wastewater treatment sewage sludge that are high enough in macronutrients to be considered a fertilizer (Figure 5-5).

When applied to agricultural, forestry, reclamation, and landscaping sites it is a source of nutrients and organic matter. Biosolids improve roadside soils by increasing water-holding capacity, improving soil structure and infiltration, providing slow-release nutrients, and increasing soil biological activity (Sullivan et al 2007). Biosolids are sold as packaged fertilizers from commercial sources or directly in bulk from wastewater treatment centers.

Nitrogen is an important component of biosolids because this nutrient is limiting on most roadsides. Biosolids are basically composed of two forms of nitrogen:

- **Ammonium-nitrogen**—This form of nitrogen is readily available for plant uptake. If biosolids are surface applied, a portion of ammonium-N is lost as gas. Sprinkler irrigating or tilling biosolids into the soil immediately after application can reduce the amount of nitrogen that is lost (Sullivan et al 2007).
- **Organic-nitrogen**—Nitrogen in this form is bound to organic matter that has to be digested by soil microorganisms to be released. This process, called N mineralization, takes time. When first applied however, nitrogen release rates are rapid, but level off over the years, to supply a relatively consistent annual supply of nitrogen.

Biosolids are high in other macro nutrients including phosphorus, calcium, magnesium, and sulfur (Figure 5-5). Phosphorus however, is very low and may need to be added depending on the nutrient status of the roadside soils and the biosolids being applied. Phosphorus is present in biosolids at significant quantities yet the availability is around half of a commercial fertilizer because of the predominance of inorganic phosphorus (EPA 1995). Micronutrients, such as boron (B), copper (Cu), iron (Fe), manganese (Mn), molybdenum (Mo), and zinc (Zn) are present in varying amounts but may not be at the ratios found in a well-balanced commercial fertilizer and micronutrient fertilizers may need to be supplemented (EPA 1995).

Some biosolids are stabilized with alkaline materials which can raise the pH of roadside soils. Where soil pH is low, this can be an advantage, and biosolids can be used as a replacement for agricultural limestone (Sullivan et al 2007). However, on high pH soils, the addition of alkaline-stabilized biosolids can be detrimental to plant establishment and plant growth. For these reason, the pH of the roadside soils is important when considering applying an alkaline-stabilized biosolids.

Biosolids are typically applied on roadsides at higher rates than agricultural lands to improve soils and provide nutrients and organic matter capable of supporting native vegetation (EPA 1994). Rates range from 3 to 200 tons/acre but average around 50 tons (EPA 1995). biosolids are relatively light weight and can be half the weight by volume of soil. A 50 tons/acre application rate of biosolids is approximately 0.5-inch thick layer biosolids applied to the surface of a soil. Baxter and Stephan (2011) found that biosolids placed one inch deep on an abandoned timber haul road in Oregon and disked into the soil, was successful in obtaining a high cover of native grass species. On highway right-of-way plots in Rhode Island, Brown and Gorres (2011) found significant improvement of vegetative cover with a 2.0-inch application rate of biosolids compared to a 2.0-inch application of compost. The differences in plant response to compost and biosolids was due to the C:N ratios (biosolids – 7, compost – 64). In this study, biosolids had four times as much total N as the compost. Fava (2015) however, found that at a 1.0 inch application rate, vegetative cover did not differ from the control plots. The differences in findings of these studies point out the importance of knowing the composition of the biosolids and how they respond to the site they are being applied to. Administrative trials, where different rates of biosolids are applied in combination with compost or shredded wood, can will help develop appropriate rates and soil amendments to use to improve the roadside soils.

## Select Fertilizer Release Rates

Fertilizers are grouped by how quickly they break down and release nutrients to the soil. They are either fast-release or slow-release. Release rates are important because

|   |          |
|---|----------|
| <b>Organic matter</b>                           | 45-70%   |
| <b>Nitrogen</b>                                 | 3-8%     |
| <b>Phosphorous (P<sub>2</sub>O<sub>5</sub>)</b> | 3-8%     |
| <b>Potassium (K<sub>2</sub>O)</b>               | <1%      |
| <b>Sulfur (S)</b>                               | 0.6-1.3% |
| <b>Calcium (Ca)</b>                             | 1-4%     |
| <b>Magnesium</b>                                | <1%      |

**Figure 5-5 / Range of organic matter and nutrients in biosolids**

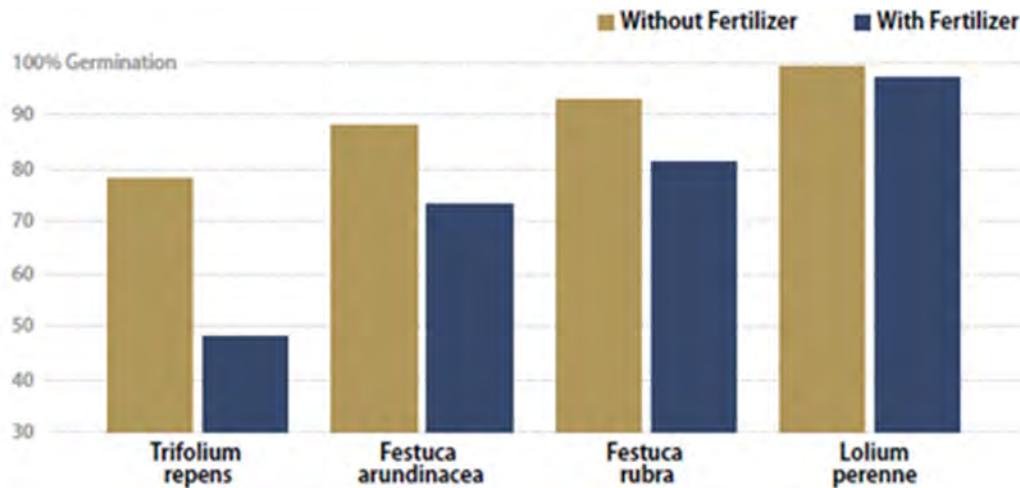
Biosolids are rich in nutrients and organic matter, often meeting the requirements to be classified as a fertilizer. This table shows the usual range of organic matter and nutrients by dry weight of material (from Sullivan and others 2007). Biosolids also provide micronutrients, including copper, boron, molybdenum, zinc, and iron.

they determine the rates at which nutrients become available to plants during the year. If nutrients are released during periods when vegetation cannot use them, some will be lost from the site through soil leaching. This is not only a waste of fertilizer but can be source of ground-water pollution.

**Fast-Release Fertilizers**—Fast-release fertilizers are highly soluble fertilizer salts that dissolve rapidly and move quickly into the soil during rainstorms or snowmelt. The fertilizer label gives an indication of how quickly nutrients are released. Terms such as “soluble,” “available,” or “water soluble” indicate that these nutrients are released relatively quickly. “Ammonium” and “nitrate” forms of nitrogen are also indications of fast-release fertilizers. The fertilizer label shown in Figure 5-4 indicates that the fertilizer contains a fast-release fertilizer and most of the nitrogen would be relatively mobile and available to plant growth within the first growing season. Ammonium nitrate, ammonium sulfate, potassium nitrate, and urea are several examples of fast-release fertilizers.

“Water soluble” or “available” nutrients do not always remain available or soluble after they are applied to the soil. Available forms of phosphorus, for example, react in the soil to form less soluble compounds; potassium gets bound up in soils with moderate to high proportions of clay; and many of the micronutrients (e.g., zinc, copper, and manganese) become unavailable when applied to soils with low pH (Section 3.8.4). Unless soils are sandy or rocky, it can be assumed that many of the nutrients stated as “available,” except for nitrogen and sulfur, will become somewhat immobile once they are applied. Over time, however, these nutrients will become available for plant uptake.

The advantages of fast-release fertilizers are they are relatively inexpensive, easy to handle, immediately available to the plant (especially nitrogen), and can be applied through a range of fertilizer-spreading equipment. Disadvantages are that some nutrients, such as nitrogen, will leach through the soil profile if they are not first taken up by plants or captured by soil microorganisms in the breakdown of carbon.



**Figure 5-6 | Reduced seed germination after exposure to fertilizer**

Germination for some species can be reduced following exposure to a 10-30-10 fertilizer solution at a rate of 750 pounds of fertilizer per 1,000 gallon hydroseeder (after Carr and Ballard 1979).

Nitrates from fast-release fertilizers have been found to leach through sandy soils to depths that are four times the rate of rainfall (Dancer 1975). For example, for sites with annual rainfall of 12 inches, nitrate could move to a depth of 4 feet if it is not taken up by plants or soil organisms. At this depth, nitrogen would be out of range of most establishing root systems.

Because fast-release fertilizers are salts, they have a potential to burn foliage and roots, especially when fertilizers are applied at high concentrations or when applied during dry weather (Section 3.8.4). High concentrations of fast-release fertilizers can also affect germination rates (Figure 5-6) because of the high soluble salt levels

(Brooks and Blaser 1964; Carr and Ballard 1979). Salt damage can be reduced by mixing fast-release fertilizers at lower concentrations or by applying them during rainy weather.

**Slow-Release Fertilizers**—These fertilizers are designed to release nutrients at a much slower rate. To be labeled slow-release fertilizer, some states specify the amount of nitrogen to be in a slow-release form. Forms of nitrogen shown on the label as “slowly available” or “water-insoluble” are good indicators that a fertilizer is in a slow-release form. The advantages of using slow-release fertilizers are as follows:

- Nutrients are supplied at a time when plants are potentially growing
- Less frequent applications
- Less potential for leaching into ground water
- Less potential to cause salt injury

The disadvantages are that many slow-release fertilizers are bulky, cost more to purchase and apply, and are limited by the type of fertilizer application equipment that can be used. Nevertheless, slow-release fertilizers have greater applicability for revegetating disturbed sites than fast-release fertilizers.

**Organic slow-release fertilizers**—These fertilizers come in either organic or inorganic forms. Organic fertilizers include animal manures (e.g. chicken, steer, and cow), bone meal, fish emulsion, composted sewage sludge (biosolids), and yard waste. Unprocessed organic fertilizers are hard to apply to roadside projects because they are bulky and high in moisture content. Many commercially available organic fertilizers have been processed to remove most of the water, which makes them easier to apply with most fertilizer-spreading equipment. For a good discussion on organic fertilizers, see [Landis \(2011\)](#).

The agents responsible for release of nutrients from organic fertilizers are decomposing soil bacteria. When soil bacteria are active, the release of nutrients is high; when dormant, the rate is low. The release of nutrients is therefore a function of moisture and temperature, which governs the rate of bacterial growth. Warm temperatures and high moisture, conditions conducive to plant growth, are also favorable for the breakdown of organic fertilizers. Because of this, the release of nutrients from the decomposition of organic fertilizers often coincides with the period when plants are growing (spring and fall) and the need for nutrients is greatest. The nutrient-release mechanism of slow-release organic fertilizers reduces the risk that highly mobile nutrients, such as nitrogen, will be released in the winter when plants are incapable of absorbing them and the potential for leaching is greatest.

Inorganic forms of slow-release fertilizers were developed for the horticulture and landscape industries where they have become an effective method of fertilizing nursery plants. These are expensive forms of fertilizer and have not been tested on roadside revegetation conditions. Nevertheless, they potentially have applicability for some native revegetation projects.

**Inorganic slow-release fertilizers**—These fertilizers include ureaform, nitroform, isobutylidene diurea (IBDU), sulfur-coated urea, and polymer-coated nitrogen, phosphorus, and potassium. These fertilizers have varying mechanisms for nutrient release. Fertilizer granules coated with materials that release nutrients only during warm, moist conditions ensure that nutrients are available during the period when plants are most likely to be growing. These coatings include sulfur (e.g., sulfur-coated urea) and polymers. Each fertilizer has its own formulated nutrient release rate, which varies from 3 months to 18 months. Release rates are available from the manufacturers for most inorganic, slow-release fertilizers. However, it is noteworthy

that these rates were developed for 70° F soil temperatures (Rose 2002), which are higher than soil temperatures in the western United States during the spring and fall when roots and foliage are growing. If roadside soils are colder than 70° F, nutrient release will take longer than what the manufacturer states.

## Determine Fertilizer Application Rates

Fertilizer rates are determined for each deficient nutrient, as shown in Figure 5-7. The calculation in this example was done to eliminate a nitrogen deficit of 769 lb/ac. Using a slow-release fertilizer with 8 percent nitrogen, the amount of bulk fertilizer necessary to bring nitrogen levels to minimum targets is 9,613 lb/ac ( $769 * 100 / 8 = 9,613$ ), which is a high rate of fertilizer to apply. With a release rate of approximately 40 percent the first year, 308 lbs N/ac would be available, far more than the establishing plant community could absorb. Alternatively, using a fast-release fertilizer with higher nitrogen analysis, such as ammonium nitrate (33 percent N), would reduce the amount of bulk fertilizer to 2,330 lbs/ac ( $769 * 100 / 33 = 2,330$ ). While there would be less weight with this more concentrated fertilizer, this is considered a dangerously high application rate because all of the nitrogen would be released in the first year resulting in a much greater potential that high amounts of nitrates are leached through the soil into the ground water and a higher risk that elevated salt levels would be toxic to plant growth. This example illustrates the difficulty in developing fertilizer prescriptions to meet long-term nutrient targets. How does the designer develop a fertilizer strategy to meet short-term and long-term plant needs without over- or under-fertilizing?

|   |  |              |   |
|---|--|--------------|---|
| A | Nitrogen (N) deficit   | 769 lbs/ac   | Calculated from example in <a href="#">Figure 5-3</a>       |
| B | N in fertilizer  | 8%           | From fertilizer label                                       |
| C | Total bulk fertilizer needed<br>$A * (100/B) =$  | 9,613 lbs/ac | To eliminate deficit  |
| D | Estimated first year N release rate of fertilizer  | 40%          | From <a href="#">Table 5-4</a> or obtain from manufacturers |
| E | Available N first year in fertilizer from first year application<br>$B * C * D / 10,000 =$ | 308 lbs/ac   | N available to plants and soil                              |
| F | Short-term N target (first year)   | 50 lbs/ac    | Depends on C:N ratio, plant cover, and age                  |
| G | Excess nitrogen<br>$E - F =$   | 258 lbs/ac   | Wasted N could leach from soils into water                  |
| H | Adjusted rates of fertilizer to add<br>$(F * 100/D) * (100/B) =$                           | 1,563 lbs/ac | To assure that N released first year is not wasted          |
| I | Remaining N deficit<br>$A - (H * B / 100) =$   | 644 lbs/ac   | Additional N needed as later applications of fertilizer     |

**Figure 5-7 | Example of calculating fertilizer application rates to reduce nitrogen**

This spreadsheet shows how a slow-release fertilizer would have to be applied at high quantities to reduce the nitrogen deficit. The problem with these high rates is that approximately 40 percent of the nitrogen (Line D) would be released the first year which is more than an establishing plant community could use, resulting in nitrogen leaching (Line G). Fertilizer rates can be lowered to meet just the first-year nitrogen needs (Line F) but not the long-term nitrogen needs of the site (Line I).

The approach presented in this section is based on meeting short-term nutrient needs of the establishing plant community while building a long-term nutrient capital. For example, applying fertilizer at the time of seeding requires very low rates of available nitrogen to meet the first-year needs of the establishing vegetation. Any extra fertilizer has the potential of being wasted. As the vegetation develops over the next few years, the ability of the plant community to take up more available nutrients increases. A strategy of applying low amounts of fertilizer the first year, followed up with higher rates in later years would supply the levels of nutrients needed for a developing plant communities without wasting fertilizers. This practice, however, is seldom employed in roadside revegetation projects. In fact, the typical fertilizer practice does just the opposite—high rates of fertilizers are applied with the seeds to a site that has no vegetation that could utilize the available nutrients. There is no return to the site in later years to assess whether additional applications of fertilizers might be essential for vegetation maintenance or growth. The approach advocated in this section is applying the appropriate mix of fertilizers to meet the annual needs of the vegetation while building long-term nutrient capital until the plant community is self-sustaining. This approach may require fertilizer application over a period of several years.

Because nitrogen is the key nutrient in establishing plant communities, this approach involves established short- and long-term nitrogen requirements of the plant community. Calculating long-term nitrogen target is addressed in [Section 5.2.1 \(see Develop Nutrient Thresholds and Determine Deficits\)](#). Short-term targets are more difficult to set because they change over time. They are governed by the following:

- Soil type
- Carbon-to-Nitrogen ratio (C:N ratio)
- Climate
- Amount of vegetative cover
- Type of vegetation
- Age of vegetation

Some general guides can be helpful in setting short-term nutrient targets for available nitrogen. Applying fertilizer at the time of sowing, for example, requires very low rates of available N because vegetation is not there to utilize it. Rates range from no fertilizer to up to 25 lb/ac of N when applying fertilizer with seeds. During plant establishment, available N can range from 25 to 50 lb/ac (Munshower 1994; Claassen and Hogan 1998). After plant establishment, rates can be increased to account for increased plant utilization above this amount. These suggested rates are adjusted upward on sites where high C:N soil amendments, such as shredded wood or straw, have been incorporated into the soil to compensate for nitrogen tie-up.

Calculating precise rates of supplemental nitrogen for incorporated organic amendments is very difficult. In nursery settings, rates of over 100 pounds of supplemental nitrogen have been recommended for incorporated straw, sawdust, and other high C:N materials (Rose et al 1995). However, applying supplemental rates on highly disturbed sites is to be done with caution, utilizing trials where possible to determine more precise fertilizer rates. Periodic soil analysis can provide the designer with a better understanding of the soil nitrogen status. To keep testing costs low, only available nitrogen and total nitrogen need to be tested ([Section 3.8.4](#), see [Mitigating for Lack of Topsoil](#)).

In determining how much fertilizer to apply, it is important to estimate how much nitrogen will be available the first and second years. Manufacturers have this information for most inorganic slow-release fertilizers. Claassen and Hogan (1998) have also performed tests on some slow-release organic fertilizers ([Table 5-4](#)). Release rate determinations are performed in the laboratory but the actual release rates will vary in the field by soil type and climate. In the example described in Figure 5-7, the first-year release rate of nitrogen from the slow-release organic fertilizer was estimated at 40 percent. This was a guess based on the manufacturer's estimates of 55 percent release, but because it was being applied to a semi-arid site where decomposition of the fertilizer would be slow, the rate was dropped to 40 percent (Line D). If 40 percent of the nitrogen became available the first year, 60 percent would remain for the following years (Line E). At this release rate, 308 lb/ac of nitrogen would become available the first year after application (Line F). While this is an extremely high rate, consider the application of ammonium nitrate at 100 percent first year release, which would supply 769 lb/ac (Line A) of immediately available nitrogen. Recalculating fertilizer rates using a more realistic rate of 50 lb/ac available nitrogen needed the first year after application (Line F), bulk fertilizer application rates would be 1,563 lb/ac (Line H). At this new rate, the site would have sufficient first- and second-year supplies of nitrogen but lack adequate nitrogen the following years. The remaining deficit to meet long-term nitrogen targets would be approximately 644 lb/ac, which could be supplied through later applications of fertilizer or other carriers of nitrogen (topsoil, compost, biosolids, wood waste, mulch, and nitrogen-fixing plants). In this example, nutrient strategy would be built upon treatments that would increase long-term nitrogen capital.

The process of calculating fertilizer application rates in Figure 5-7 can be used for other deficient nutrients, however, understanding the availability of these nutrients is problematic. Many nutrients become fixed in the soils and their availability is dependent on highly variable factors such as soil texture, pH, and placement in the soil. It is a reasonable assumption that unless the soils are sandy or very rocky, that all nutrients, aside from nitrate or ammonium forms of nitrogen, are relatively unavailable the first year after application. With time, however, they will slowly become available.

## Determine Timing and Frequency

The primary reason to fertilize is to supply nutrients during periods when plants can take them up for growth. The demand for nutrients changes throughout the year depending on the physiological state of each plant. In nursery settings, fertilizers are adjusted throughout the year at rates and formulations that correspond to plant requirements. While that capability is not possible in roadside environments, fertilizers can be used more wisely by applying an understanding of how the assortment of fertilizers function in meeting the nutrient requirements of plant communities. At least two plant growth phases are important in the timing of fertilizer application: (1) seed germination and plant establishment and (2) post-plant establishment.

**Seed Germination and Plant Establishment Phase**—Traditionally, fast-release fertilizers have often been applied at high rates in the fall in the northern United States during the seed-sowing operation. This practice is a quick and easy way to apply fertilizers. However, the timing can result in ineffective and wasteful use of fertilizers (Figure 5-8B) (Dancer 1975). In addition, application of fast-release fertilizers at this time can potentially pollute water sources. Slow-release fertilizers are more appropriate for seed sowing in the fall because much of the fertilizer, but not all, is expected to release nutrients the spring, not in the fall or winter (Figure 5-8D).

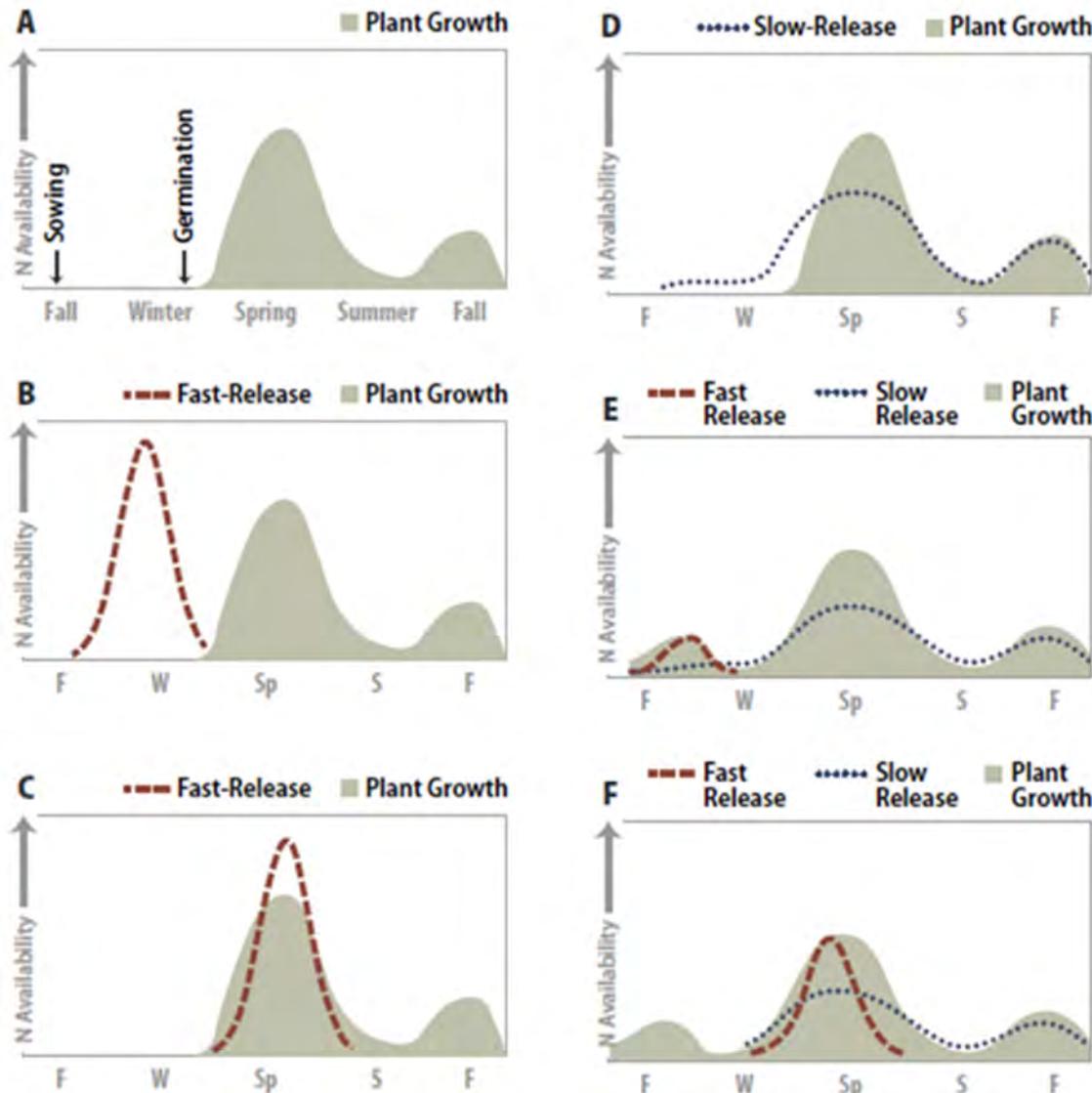
Perennial grasses and forbs do not require high levels of nitrogen for germination and early establishment (Reeder and Sabey 1987). In fact, an elevated level of available nitrogen can be a problem because it encourages the rapid establishment and growth of annual weed species over slower-growing perennial grass and forbs (McLendon and Redente 1992; Claassen and Marler 1998). It is important to calculate fertilizer quantities based on the plant requirements over time (Section 5.2.1, see Determine Fertilizer Application Rates).

One strategy is to apply little or no fertilizer during sowing and wait until seeds have germinated and grown into small seedlings before fertilizers are applied (Figure 5-8C). This strategy ensures that nutrients are available when the seedlings actually need them, not before. Fertilizers applied as slow-release form are preferred because they have less potential for causing salt damage when applied over emerging seedlings. Another strategy is to wait until the following fall (Figure 5-8E) or spring (Figure 5-8F) of the second year to fertilize.

**Post-Establishment**—Once vegetation is established (one or two years after sowing), fertilizers may be applied at higher rates with the assurance that nutrients will be taken up by the plants. It is important that the application rates are based on nutrient levels of the soil and the needs of the native plant species. On fertile soils, there may be no need to fertilize, whereas on soils without topsoil or low in organic matter, a post-establishment fertilizer application may be needed.

Slow- and fast-release fertilizers can be combined to provide short- and long-term nutrient requirements (Figure 5-8E and F). Spring applications of fast-release fertilizers are more effective than fall application because of the higher nutrient requirements of growing plants during that period (Figure 5-8F). In addition, spring applications may pose less risk of damaging vegetation through fertilizer salts on sites where precipitation in the spring is frequent enough to wash fertilizers from the foliage. The conductivity of a fertilizer solution being applied over existing vegetation can be measured with a conductivity meter to avoid salt damage (Section 3.8.4). If rates exceed 3,500 mS/cm, then diluting the solution or applying the fertilizer during rainy weather is advised. Fertilizer rates can also be adjusted based on the salt tolerance of

a plant species (see ERA).



**Figure 5-8 | Strategies for applying slow and fast release fertilizers**

Fertilizers should be applied during seasons and at rates and formulations that release nutrients when native plants can efficiently draw them from the soil. The following are strategies for applying slow and fast release fertilizers.

*Note: These examples are based on a Mediterranean climate and assumes that seeds germinate in the spring, not in the fall or winter.*

## Select Fertilizer Application Method

Because nutrients have varying degrees of mobility (nitrogen is highly mobile; phosphorus and many micronutrients are relatively immobile), how fertilizers are applied will determine how accessible nutrients are to the root systems of establishing vegetation. If nutrients are highly mobile, the easiest and least expensive method is broadcasting fertilizer to the soil surface to allow rainfall or snowmelt to release and move nutrients into the soil. A more difficult, yet more effective application method for immobile nutrients is to incorporate, or mix, fertilizers into the soil so fertilizer granules are uniformly distributed and accessible by root systems.

**Broadcast Fertilizer Application**—For fertilizers with highly mobile nutrients, such as nitrogen and sulfur, broadcast application on the soil surface is an appropriate practice. Broadcast fertilizer application is less effective, however, for immobile nutrients. These nutrients often become immobilized at the soil surface and are very slow to move into the rooting zone where they can be accessed. Depending on soil characteristics, such as pH and clay content, some immobile nutrients will take years to move only a few inches from the point of fertilizer placement.

A variety of dry fertilizer spreaders are available, from hand-operated to tractor-

mounted. Most equipment is limited to moderate slope gradients (less than 1V:2H). With all forms of spreaders, consider calibrating them before they are used to ensure that the correct rates are being applied.

Hydroseeding equipment can be used to apply fertilizer in the same operation with seeds, tackifiers, and hydromulch ([Section 5.4.2](#)). This equipment can also be used solely to apply fertilizers, especially after vegetation has become established. A great advantage to using hydroseeding equipment is that it can uniformly spread fertilizers on steep slopes and a variety of topographies. In addition, a combination of fertilizers can be easily mixed in the hydroseedertank and applied uniformly at relatively even proportions. This is especially useful for applying small quantities of fertilizer, such as micronutrients, which are difficult to spread evenly over large areas.

**Fertilizer Incorporation**—It is important that nutrients that are deficient and have low mobility be incorporated into the soil prior to sowing or planting. Incorporation is possible on gentle slopes but becomes very difficult with increasing slope gradients because of equipment limitations or slope stability. On sites where fertilizers containing immobile nutrients cannot be incorporated, an alternative is to create roughened soil surfaces ([Section 5.2.2](#)) prior to fertilizer application. Broadcast fertilizers will accumulate in the depressions of the surface. As soil gradually moves into the depressions over time through erosion (water, wind, or surface ravel), the broadcast fertilizers will become covered with soil. When this happens, immobile nutrients are accessible by roots and nutrient uptake is possible. Surface roughening also reduces the potential for fertilizers to move off-slope through erosion.

Some agricultural spreaders, called fertilizer banders or injectors, are designed to place fertilizer, or other soil amendments including mycorrhizae, at varying depths in the soil. Usually this equipment has a ripping shank or tine that loosens the soil, followed by a tube that drops the fertilizer, and coulters or rollers that close the furrow. As the bander is pulled through the soil, a line, or band, of fertilizer is created. Sowing and banding are often combined in one piece of equipment and applied at the same time. Fertilizer banders were developed for agricultural use and are limited by rock content and slope gradients, however, there are injectors that have been developed for wildland conditions (St. John 1995).

The most common approach to incorporating fertilizer is accomplished in two operations, broadcasting fertilizer on the soil surface and tilling it into the soil. In this approach, hydroseeders and broadcast fertilizer spreaders are used to apply fertilizers evenly over the soil, then the fertilizer is incorporated using tillage equipment outlined in [Section 5.2.2](#).

## 5.2.2 TILLAGE

### Introduction

Tillage is defined in this section as any mechanical action applied to the soil for the purposes of long-term control of soil erosion, reestablishment of native plant communities, and improve soil function. Most tillage equipment was developed for agricultural soils and has limited applicability for steep, rocky sites typically encountered in revegetation. This section describes the agricultural equipment that can be used for revegetating roadsides, as well as equipment specifically developed for extreme site conditions.

Among the reasons to use tillage in a revegetation project is to shatter compacted soils, incorporate soil amendments, and roughen soil surfaces. These objectives often overlap. For example, incorporating organic matter also loosens compacted soils and

roughens soil surfaces. Identifying the objectives for the project will lead to selecting and effectively using the appropriate equipment to achieve the desired soil conditions (Table 5-5).

## Shatter Compacted Soils

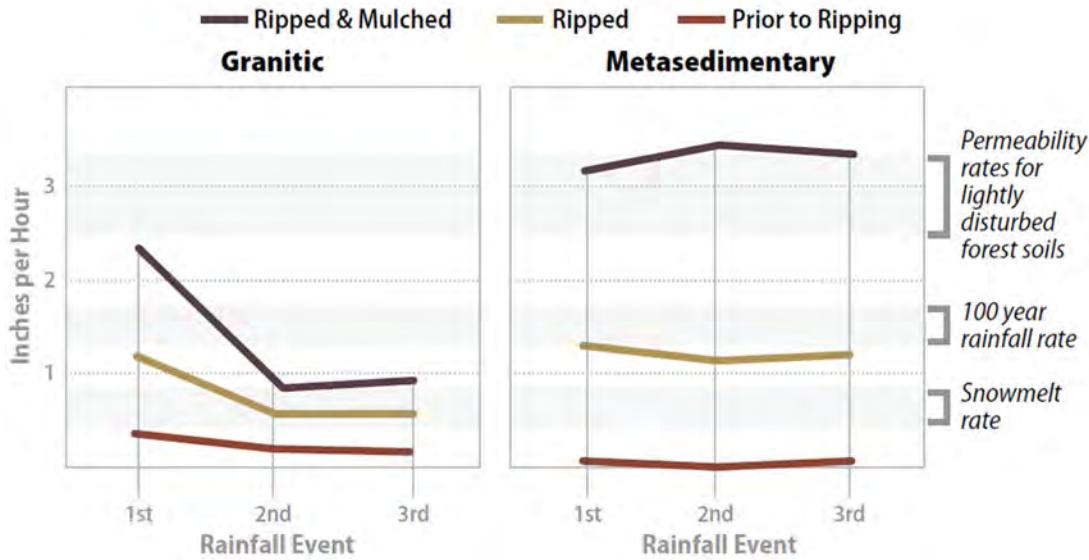
One of the primary purposes for tilling is to loosen compacted soils. When performed correctly, tillage can increase porosity in the rooting zone, increase infiltration rates, and increase surface roughness. For revegetation work associated with road construction and road obliteration, tillage to break up deep compaction is important for reestablishing plant communities. Shattering compaction at depths of at least 2 feet is essential for the healthy growth of most perennial plant species. Without this measure, it may take decades for deep compaction to recover its original bulk density (Wert and Thomas 1981; Froehlich et al 1983). In a review of tillage projects on rangeland soils, Gifford (1975) found that deep tillage greatly reduced runoff, while shallow tillage had little effect. Still, tillage alone may not return a soil to its original bulk density or hydrologic function, nor will the effects of tillage last indefinitely, especially in non-cohesive soils (Figure 5-9) (Onstad et al 1984). Many factors affect the return to bulk densities and infiltration rates typical of undisturbed reference sites. These include the type of tillage equipment used, tillage depth, soil moisture during tillage, soil texture, presence of topsoil, and organic matter content.

**Table 5-5 / Types of tillage and equipment**

The appropriate tillage equipment for the project depends on project objectives.

| Objectives             | Shattering             | Incorporating                       | Imprinting   |
|------------------------|------------------------|-------------------------------------|--|
|                        | Rippers and subsoilers | Disks, plows, excavator attachments | Dixon imprinter, excavator attachments, trackwalking |
| Loosen compacted soil  | Good                   | Good                                | Poor   |
| Incorporate amendments | Poor                   | Good                                | Poor   |
| Roughen surface        | Good                   | Good                                | Good   |

There are two fundamentally different equipment designs for reducing compaction. One design lifts and drops soil in place, shattering compacted soil in the process. This type of equipment includes rock rippers, subsoilers, and “winged” subsoilers. The second design churns and mixes the soil. Equipment that falls into this category includes disk harrows, plows, spaders, and attachments to excavators. This type of equipment can also incorporate soil amendments, such as organic matter or fertilizers, in the same operation, as described in Section 5.2.2 (see Incorporate Soil Amendments).



The terms “subsoiling” and “ripping” are used interchangeably to describe soil-shattering operations. Soil shattering involves pulling one tooth, or a set of teeth, at various depths through the soil to break up compaction created by equipment traffic. The rock ripper is a common tool found on most construction sites. When used to break up compaction, one or two large ripper tines are typically pulled behind a large bulldozer at 1 to 3-foot soil depths. While this equipment will break up compaction in portions of the soil where the ripper tines have been dragged, it does not effectively fracture the compacted soil between the ripper tine paths (Andrus and Froehlich 1983). The effectiveness of rippers can be increased by multiple passes through the soil or by adding tines to the toolbar. Even on small machines, up to five tines can be added to increase soil shatter.

Rippers have also been adapted to increase soil lift between tine paths by welding wide metal wings to the bottom of each tine. These wings are angled upwards so the soil between the tines has greater lift, and therefore greater shatter when the soil drops behind the wing. When two or more tines are placed together on a toolbar, they work in tandem to more effectively break up compaction. The resulting equipment is called a “winged” subsoiler (Figure 5-10). Andrus and Froehlich (1983) found that the winged subsoiler was a far more effective tool for breaking up compaction. This equipment fractured over 80 percent of the compaction in several operational tests, as compared to 18 to 43 percent for rock rippers and 38 percent for brush rakes. However, winged subsoilers are not practical in all soils, especially those with high rock fragments, buried wood, or slopes greater than 3H:1V gradients.

Achieving good shatter at deeper soil depths means that tillage equipment be adjusted for site-specific soil conditions, especially soil texture, soil moisture, and large rock content. Tines will slice through the soil, causing very little soil shatter, if soils are too moist during ripping. Subsoiling when soils are extremely dry can bring up large blocks of soils, especially when the soils are high in clays (cohesive soils).

It is important the winged subsoiler and rock ripper be adjusted to meet the soil conditions of the site. Making the proper adjustments can lead to greater shatter and more efficient use of tractor equipment. These adjustments include the following:

- Tine depth
- Tine spacing
- Number of tines

**Figure 5-9 | Benefits of ripping and mulching vary by soil type**

Short-term benefits of ripping (using a winged subsoiler) and mulching road surfaces vary by soil type, as shown in rainfall simulation tests on sites in northern Idaho. Granitic soils responded to ripping and mulching with increased permeability during the first storm, but permeability rates returned to near pre-treatments rates with successive rainfall events. Metamorphic soils reacted positively to both treatments and maintained high permeability rates after three rainfall events. Mulching improved permeability in both soil types. In fact, for metamorphic soils, the combination of ripping and mulching increased permeability to rates that were typical of lightly disturbed forest soils (adapted from Luce 1997).



**Figure 5-10 | Winged subsoiler**

Soil shattering becomes more effective when wings are mounted on subsoil tines. This equipment is called a winged subsoiler.

*Photo credit: Brent Roath*

- Wing width and angle (for winged subsoiler)

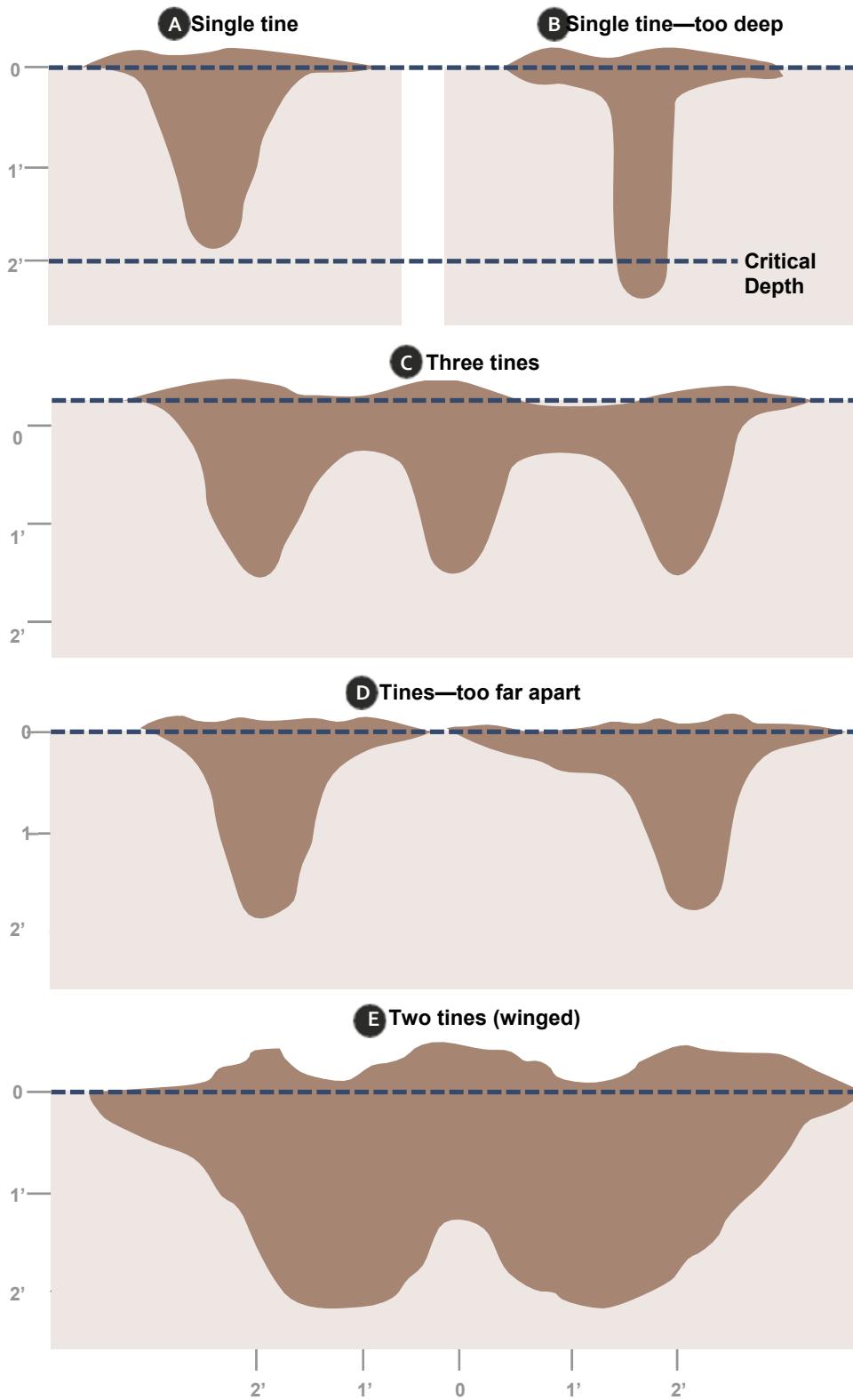
For optimum shatter, the depth of the tines is adjusted based on the soil properties and moisture conditions. The tines are adjusted to be above a “critical depth,” the point below which soils will not shatter effectively (Figure 5-11B). The critical depth changes for soil type and tine configuration. Soils high in clays with high soil moisture have shallower critical depths (Andrus and Froehlich 1983).

The closer the spacing of tines, the greater the shattering. The more tines that are placed on a toolbar, the more area of soil that can be shattered. However, where large rocks, slash, and roots are present, closely spaced tines will drag these materials out of the ground. Three to five tines are typically used for most soil types. Wing size, angle, and shape of the tines all play a role in breaking up compaction (refer to Inset 5-2 for specifications for winged subsoiler).

#### **Inset 5-2 / Contract specifications for a winged subsoiler**

A winged subsoiler consists of a self-drafting, winged subsoiler on a dolly mount, sized for use with a D-7 tractor. The unit consists of three-winged ripper tines capable of extending 12 to 34 inches below the draw bar. Wings are usually at least 20 inches wide with a 2-inch lift of the wings from horizontal. Tines have an individual tripping mechanism that automatically resets; tine spacing is adjustable, and individual tines removable. Various wing patterns are available and easily interchangeable. Good practice is to have an implement capable of achieving maximum fracture of compacted soils (minimum 24 inches) in one pass (adapted from Wenatchee National Forest contract specifications).

Typical settings for rock ripper and winged subsoiler equipment configurations are shown in Table 5-6. These are suggested settings to be applied after first monitoring the results of the equipment on the project soils. The most direct method for monitoring soil shatter is to measure the depth to the compacted soil with a soil penetrometer or shovel (Section 3.8.2, see [Soil Structure](#)). Immediately after a pass is made with the tillage equipment, the shovel or penetrometer is pushed into the soil and the depth to resistance (the compacted layer) is recorded. Measurements are taken every 6 inches across a small transect perpendicular to the direction of the tractor and spanning the width of the tillage disturbance. Plotting the depths to compaction on graph paper provides a cross-section of the shattering pattern (Figure 5-11 is an example of plotting soil shatter). If the shattering pattern is inadequate, adjustments can be made to the tine depth, tine spacing, and angle of the wing. If these adjustments fail to increase soil shatter, a second and perhaps third pass by the ripper or winged subsoiler can be considered. Successive passes made at 45 to 90 degrees angles from the first pass achieves the greatest benefit. Most soil-shattering equipment is attached to the tractor toolbar and is limited to slope gradients of 3H:1V or less. Subsoilers and rippers are best used for projects that consist of gentle terrain or obliterated road sections.



**Figure 5-11 |** Subsoiler tine and wing configurations determine effectiveness

The effectiveness of subsoiling or ripping equipment to shatter compacted soil is a function of tine depth, number of tines, distance between tines, and wing configuration. Pulling a single tine (A) above a critical depth does some soil shattering as compared to a single tine ripping deeper than a critical depth (B). Placing three or more tines together (C) can be more effective than one tine, but tines spaced too far apart will not effectively shatter the soil (D). Attaching wings to the tines is very effective in shattering compaction between the tines (E) (modified after Andrus and Froehlich 1983).

**Table 5-6 | Recommended design features for some tillage equipment**

Modified after Andrus and Froehlich 1983; Froehlich and Miles 1984

| Item                | Implement feature              | Recommended design                          |
|---------------------|--------------------------------|---|
| Disk harrow         | Disk diameter                  | 40-50 in.                                   |
|                     | Number of disks                | 6-12  |
|                     | Average disk weight            | >1,800 lbs                                  |
|                     | Disk arrangement               | Offset gangs, independent disks             |
|                     | Max slope (cross slope travel) | <5:1 H:V                                    |
| Brush blade         | Max slope (down slope travel)  | 5:1 H:V                                     |
|                     | Tine spacing                   | 22-26 in.                                   |
|                     | Tine depth                     | <20 in.                                     |
|                     | Max slope (cross slope travel) | <5:1 H:V                                    |
| Rock rippers        | Max slope (down slope travel)  | 3:1 H:V                                     |
|                     | Tine spacing                   | 24-30 in. (one pass) 40-48 in. (two passes) |
|                     | Ripping depth                  | 20-24 in.                                   |
|                     | Number of tines                | 5 (one pass)<br>3 (two passes)              |
|                     | Max slope (cross slope travel) | <5:1 H:V                                    |
| Wings of subsoilers | Max slope (down slope travel)  | 2.5:1 H:V                                   |
|                     | Ripping depth                  | 18-22 in.                                   |
|                     | Number of tines                | 3-4   |
|                     | Tine spacing                   | 30-40 in.                                   |
|                     | Wing width                     | 12-24 in.                                   |
|                     | Wing angle                     | 10-60°                                      |
|                     | Max slope (cross slope travel) | <5:1 H:V                                    |
|                     | Max slope (down slope travel)  | 2.5:1 H:V                                   |

The excavator is good piece of equipment on steeper slopes where its arm can reach 35 feet up and down slope and decompact targeted areas of compacted soil. In this operation, the bucket of the excavator is placed several feet into the soil, lifted and dropped in place (Figure 5-12). Special attachments, such as the “subsoiling grapple rake” have been developed for the excavator that can decompact, incorporate, and remove rock and slash in the same operation (Archuleta and Baxter 2008).

A general rule for tillage is to operate equipment on the contour to reduce the potential of water concentrating in the paths of the furrows and creating soil erosion and slope stability problems. This is especially important on steeper slopes where the potential for soil erosion and slope instability are greater. If several passes are made, it is important to make the last pass on the contour.

It is also important to consider that when cuts and fills are tilled, soil strength is reduced and the soils are less resistant to concentrated water. Improper road or slope drainage may result in rills and gullies on tilled soils, a situation that is less likely to occur when soils are compact. Therefore, on slopes that have been tilled, it is important that road water is directed away from fill slopes at least until vegetation has stabilized the slopes. It is important to discuss any tillage operation on slopes adjacent to roadways with the design engineer to ensure that slope stability and road objectives are not compromised.

### Incorporate Soil Amendments

Tilling is used to incorporate fertilizers, organic matter, lime, and other amendments that are placed on the soil surface and evenly distributed in the soil. Tilling with these objectives involves equipment that mixes soil, such as plows, tillers, disks, chisels, and soil spaders. These types of equipment are tractor-drawn and limited to gentle slope gradients (5H:1V or greater) and soils low in rock fragments. They are not designed to break up deep compaction. Under most disturbed soil conditions, the best that can be expected from this equipment is tillage to a depth of 8 inches.

The excavator is also another tool for incorporating soil amendments. It has the advantage over tractor based equipment in that it can incorporate soil amendments several feet into the soil. It can also be used to move topsoil or organic matter to concentrated

locations to create mounds or planting islands (Section 5.2.8). When islands are created for deep-rooted species, such as shrubs and trees, soil amendments are applied to the surface of the soil and incorporated several feet deep. Care taken on sites where natural soil horizons or soil layers have been placed (e.g., topsoil, litter, and

Roadside Revegetation: An Integrated Approach to Establishing Native Plants and Pollinator Habitat



**Figure 5-12 | Excavators decompacts and incorporates soil amendments**

The excavator incorporates and decompacts soils by lifting the soil and dropping in place.

*Photo credit: David Steinfeld*

duff) can help prevent mixing these layers together.

Rippers and subsoilers are less effective in incorporating materials such as fertilizers or organic matter into the soil. Nevertheless, spreading mulch on the soil surface prior to ripping or sub-soiling usually incorporates enough organic matter into the soil surface to enhance infiltration rates (Luce 1997). In the same manner, fertilizers applied to the soil surface, especially those containing immobile nutrients, will be mixed into the top several inches of soil and made available to surface roots.

## Roughen Soil Surfaces

Tilling is often done to roughen the soil surface for erosion control and to create a more optimum seedbed (Section 3.8.5, see Surface Roughness). The micro-topography of a roughened surface consists of discontinuous ridges and valleys where the valleys become the catch basins for seeds and surface runoff. Seeds have greater opportunities to germinate and become established in the micro-valleys because of increased moisture, higher humidity, protection from the wind, and shelter from the sun. Surface roughening is a side benefit of the incorporating and shattering operations described in Section 5.2.2 (see Shatter Compacted Soils) and Section 5.2.2 (see Incorporate Soil Amendments).

Roughening is also accomplished by either scarifying or imprinting operations. Scarification is the shallow loosening of the soil surface using brush blades, harrows, chains, disks, and chisels. It does not loosen compacted soils below the surface. Because it only loosens the soil surface several inches, the benefits for revegetation are only seen during seed germination and early seedling establishment. Once root systems hit a compacted layer, which is typically present several inches below the surface, growth is curtailed.

Imprinting is a form of surface tillage that leaves the soil with a pattern of ridges and valleys. The equipment applies a downward compressive force to a metal mold, leaving an impression on the soil surface. Specialized imprinters have been developed for rangeland restoration. For example, the "Dixon" imprinter was developed to restore perennial grasses for rangelands in Arizona and other arid states. It consists of a roller with large angular metal "teeth" that is pulled behind a tractor. The imprinter creates a pattern of V-shaped troughs, 4 to 7 inches deep, encompassing approximately 1 ft<sup>2</sup> area (Dixon and Carr 2001a, 2001b). This equipment also has a set of ripping shanks attached to the tractor that shatters deeper compaction before imprinting.

A common practice of imprinting is trackwalking (Figure 5-13). In this operation, a tractor is "walked" up and down cut and fill slopes, leaving a pattern of tractor cleat imprints on the soil surface, parallel to the slope contour, no deeper than several inches. These imprints are substantially shallower than those created by the Dixon imprinter, with less longevity. Because heavy equipment is used, trackwalking can compact soils. Compaction is not often considered when selecting trackwalking practices because soils of most roadside construction sites are already very compacted and trackwalking is unlikely to significantly increase compaction. This is one reason why trackwalking has been considered beneficial for erosion control and revegetation because it can create a somewhat better "short-term" growing environment and reduce surface erosion and sedimentation on very poor sites.

If the last operation on a construction site is to subsoil or rip soils 1.5 to 2 feet deep and leave the soil in a decompacted condition prior to revegetation, trackwalking would be more detrimental than beneficial on most soils in the long-term. The weight of the tractor used to create imprints would compact the tilled soil leaving the surface smoother (less rough) than if left alone. As noted before (Chapter 3), compaction



**Figure 5-13 / Trackwalking compacts soils**

Trackwalking creates imprints on the soil surface, but will also compact surface and subsurface soils.

*Photo credit: David Steinfeld*



**Figure 5-14 / Soil imprinting with modified excavator bucket**

An alternative form of imprinting road cuts and fills that does not compact soils is welding angle iron onto the bucket of an excavator. As the excavator pulls top-soil into place and contours the slope, it presses the face of the bucket into the soil surface to form surface imprints.

*Photo credit: David Steinfeld*

reduces infiltration and increases runoff; therefore, trackwalking has the potential to increase in soil erosion. Rainfall simulation tests can be run on sites near the construction project that have been trackwalked and compared with those that have been left in an uncompacted state to determine the effects on runoff and soil erosion (Hogan et al 2007).

An alternative to trackwalking is the use of the bucket of an excavator to pack and imprint the soil surface. Different patterns of steel "teeth" can be welded on the face of the bucket to achieve the desired surface micro-relief. Figure 5-14 shows an excavator bucket, with four strips of angle iron welded to its face, to create a pattern of 3-inch-deep impressions. The excavator, in this example, moves topsoil to the site, shapes the cut and fill slopes, and imprints the surface, all with one operation.

## 5.2.3 MULCHES

### Introduction

Mulch is defined as a protective material placed on the soil surface to prevent evaporation, moderate surface temperatures, prevent weed establishment, enrich the soil, and reduce erosion. Mulches, therefore, have many functions or roles in the recovery of native vegetation to a disturbed site. Confusion often arises around the use of mulches on revegetation projects unless the reasons for using them are clearly defined. In this discussion, mulches are grouped into four uses based on the revegetation objectives:

- Seed Covering
- Seedling Mulch
- Soil Improvement
- Seed Supply

For most projects, mulches are used to meet more than one objective but this is problematic when the methods for achieving more than one objectives are not compatible. For example, erosion-control products and practices that are effective for controlling surface erosion are not always optimal for establishing vegetation. For this reason, it is important to understand the objectives for mulching when selecting mulch types and application methods.

This section describes the objectives for applying mulches and the potential mulch sources. Many publications and much research are available on the effectiveness of mulches for erosion control and surface stabilization. This discussion focuses primarily on the characteristics of mulches for plant establishment.

### Seed Covering

One of the principal reasons for applying mulch is to enhance seed germination and early seedling establishment. During this critical period, desirable mulches will:

- Protect seeds and young seedlings from soil splash, sheet erosion, and freeze-thaw
- Keep seeds moist during germination
- Keep soils moist during seedling establishment
- Moderate surface temperatures during germination
- Prevent salts from wicking to the surface and harming germinating seedlings

The characteristics of mulch materials that make an ideal seed covering are those that

Roadside Revegetation: An Integrated Approach to Establishing Native Plants and Pollinator Habitat



**Figure 5-15 | Long-fibered mulches**

Long-fibered mulches, such as the wood strands shown below, create a good growing environment because seeds and seedlings are protected from excessive drying during germination and early seedling establishment. On sites where freeze-thaw is prevalent, long-fibered mulches can insulate the soil and protect emerging seedlings.

*Photo credit: David Steinfeld*

protect seeds from drying winds, solar radiation, high evapotranspiration rates, and surface erosion while still allowing seeds to germinate and grow through the mulch into healthy seedlings. Long-fibered mulches placed at the appropriate thickness, usually meet these characteristics. A long-fibered mulch is composed of particles that are long and thin (at least several inches long with a length to width ratios of greater than 4:1). Such materials include straw, wood strands, pine needles, and shredded wood (ground or chipped woody material).

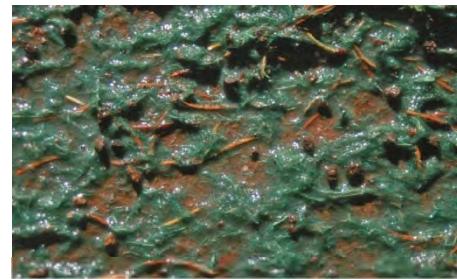
When applied correctly, the strands of long-fibered mulch loosely pack over each other, much like "pick-up-sticks," forming large air spaces or pores (Figure 5-15). Large pores function much like the air spaces in building insulation by moderating extreme temperatures. The bridging effect of the long particles also makes some of these materials more stable, and less prone to erosion or movement. Depending on the long-fibered material and erosivity of the site, it may not be necessary to apply a tackifier, though heavier materials, such as shredded woods or wood strands are less susceptible to wind displacement than lighter material like straw.

Long-fibered materials that are not recommended for mulch unless they are stabilized include shavings and materials less than two inches in length (Robichaud 2013).

Short-fibered mulches are much shorter in length and are typical of materials found in hydromulches. Comparing short-fibered mulches to long-fibered mulches, long-fibered mulches can be applied at greater thicknesses, which help maintain surface soil moisture and higher humidity around germinating seeds and emerging seedlings. In addition, long-fibered mulches can mitigate the effects of frost heaving at the soil surface (Kay 1978), significantly reduce high surface temperatures (Slick and Curtis 1985), and allow sunlight penetration, which enhances seed germination and seedling establishment. Large pores created by long-fibered mulches also allow better gas exchange between the soil and atmosphere (Borland 1990).

Short-fibered mulches usually have smaller pores and form denser seed covers. These materials are applied thinly (Figure 5-16); therefore, they offer less insulation than long-fibered mulches and have less value as a seed covering. Some researchers suggest that very fine textured mulches can increase surface evaporation by wicking moisture from the soil to the surface of the mulch (Slick and Curtis 1985; Borland 1990; Bainbridge et al 2001). These types of mulches are derived from several sources: paper fiber, which provides little cover or slope protection and more typically, wood fiber, which offers longer duration cover and slope protection. Short-fibered mulches are effective as an erosion-control cover when applied with a tackifier, but many are considered inferior to long-fibered mulches for germination and early seedling establishment (Kill and Foote 1971; Meyer et al 1971; Kay 1974, 1978, 1983; Racey and Raitanen 1983; Dyer 1984; Wolf et al 1984; Norland 2000). Recently developed products, like bonded-fiber-matrix (BFM) and High Performance Growth Media, are also applied hydraulically, but their fibers are mechanically kinked so that when applied at recommended rates provide loft and pore space suitable for better seed germination while also providing a high level of slope protection (Figure 5-17).

Compost are being used with increasing frequency on projects as a seed mulch. Composts are different from long-fibered mulches in that the fibers are much shorter in length (typically less than an inch) and composted. Because they have less loft, compared to long-fibered mulches, they do not perform as well, and may dry out faster, which could reduce germination rates on drier sites. When composts are applied to a slope with pneumatic blower or a high-speed conveyor to depths ranging from 1 to 2 inches, it is called a compost blanket. Seeding involves applying the compost blanket, then seeding and covering the seed with a  $\frac{1}{4}$ " to  $\frac{1}{2}$ " depth over the seed. Most pneumatic blowers can mix seed with the compost and apply it as a thin



**Figure 5-16 | Hydromulch**

Hydromulch with tackifier can stabilize the soil surface for up to a year but does not necessarily create an optimum environment for germinating seeds. The short-fibered textures typically form a covering that is too thin to maintain moisture around the seeds during germination when the weather is dry. The hydromulch (dyed with a green tracer) shown in this picture is applied at approximately 1,500 lb/ac. This low application rate may be acceptable on sites where soil erosion potential is low and surface soil moisture is high and seeds do not need to be covered for germination.

*Photo credit: David Steinfeld*



**Figure 5-17 | High performance growth media**

At 4,000 pounds per acre, High Performance Growth Media provides a quarter of an inch of loft which creates a better environment for germination than hydromulch.

*Photo credit: Profile Product*

layer over the compost blanket as a final pass. This is the easiest method, but if this is done, then increasing the seeding rate may be a good practice because some seeds will be near or on the surface of the compost, lowering the germination rate of the seed mix. Applying seeds with a pneumatic mulch blower is done by placing seeds in a "seed metering bin" attached to most mulch-blowing equipment. This equipment meters seeds into the mulch as it is being applied. The rate at which the metering system delivers seeds is calibrated prior to mulching to obtain the desired seed density (refer to [Section 5.4.1](#) for seed calibration methods).

The application of a tackifier is important because the medium-length fibers of compost do not provide enough weight and bridging between particles to provide protection from wind and water erosion. With tackifiers, composts can adhere to steep slopes in high rainfall areas. Compost blankets have a high moisture holding capacity and its dark color captures heat which aids germination. In addition, compost blankets provide nutrients to plants, which eliminates the need for fertilizer, and overtime increases soil organic matter. Compost derived from yard debris is readily available in some areas where it is produced to eliminate organic matter from the waste stream.

Erosion mats are another type of seed cover ([Section 5.2.3](#), see [Erosion Mats](#)). These materials come in rolls or sheets, which are laid out on disturbed soils and anchored in place after seeds have been sown. They are composed of such materials as polypropylene, straw, coconut, hay, wood excelsior, and jute. Good characteristics of erosion mats for seed germination and early seedling growth are those with enough loft, or porosity, to provide for optimum seed germination. Erosion mats create a micro-environment for seed germination while protecting the soil and allowing some sunlight to penetrate to the surface of the soil (Figure 5-18). It is important to recognize that the plastic netting in these mats degrade at varying rates depending on the climate and soil conditions. For example, on cool-dry sites, typical of high elevations, decomposition rates are slow and plastic may last for many years. Plastic netting that does not break down quickly can be a hazard to wildlife, entangling amphibians, reptiles, and birds. To reduce this hazard, many state DOTs are requiring that all erosion control mattings be fully biodegradable.

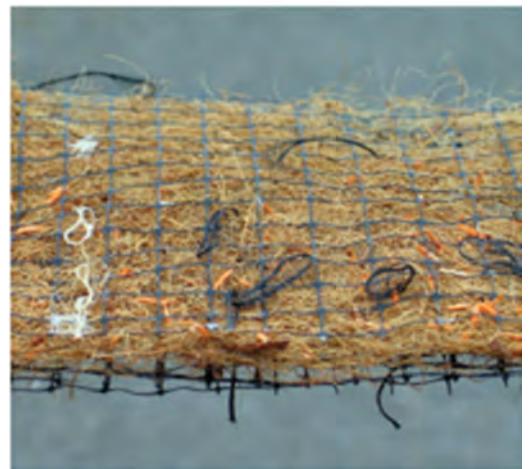
On sites where vegetation is expected to take several years to establish (e.g., arid, high elevation sites), it is important to apply a mulch with a longevity of more than one year. Materials with greatest longevity are most long-fibered wood mulches, as well as erosion mats made from coconut fiber. Straw, hay, and short-fibered products are less likely to be present after the first year however a High Performance Growth Media is available that is made of coconut fiber and it claims a 24-month functional life.

Mulching for seed covering is critical on sites that have high evapotranspiration rates during germination, unstable soil surfaces, susceptibility to freeze-thaw, or high soil pH. Mulching may be less important on sites where soil surfaces do not dry out during seed germination or on projects where seeds have been covered by soil.

## Seedling Mulch

Mulch is placed around newly planted or established plants to improve survival and growing conditions by:

- Reducing surface evaporation
- Preventing the establishment of competing vegetation
- Moderating surface temperatures
- Allowing water infiltration



**Figure 5-18 | Erosion mat**

Erosion mats can be good seed covers. Mats with the highest loft create the best microenvironment for seed germination while allowing some sunlight to penetrate to the surface of the soil. A consideration of the use of these products is how fast they decompose and the effects on wildlife.

*Photo credit: David Steinfeld*



**Figure 5-19 | Sheet mulch**

Sheet mulches come in a variety of materials, such as the paper/cardboard product shown in this picture. The size of the sheet mulch is large enough to keep competing vegetation away from the seedling. The 3-by-3-foot sheet mulch shown around this Pacific madrone (*Arbutus menziesii*) seedling is the minimum size for this site.

*Photo credit: David Steinfeld*

Studies have shown that survival and growth of young trees are significantly increased by applying mulches around seedlings at the time of planting (DeByle 1969; Lowenstein and Pitkin 1970; Davies 1988a, 1988b). Mulching around seedlings results in the greatest benefit on hot and dry sites (typically south and west aspects) and sites with aggressive competing vegetation. It is less important to apply mulch around seedlings on sites that have a low potential for establishing competing vegetation the first several years after planting or sites that have low evapotranspiration rates or high rainfall during the summer months.

Seedling mulches are applied either as an organic aggregate, rock, or as sheet mulches. Organic aggregate mulches consist of shredded or chipped wood derived from bark, wood, branches, sawdust, or lawn clippings. Rock aggregate mulches consist of a layer of rocks placed around each seedling. Sheet mulches are large pieces of non-permeable or slightly permeable materials made from translucent plastic, newspaper, coconut fiber, or geotextiles (woven fabrics) that are anchored around planted seedlings (Figure 5-19).

**Sheet Mulches**—A variety of sheet mulches are available commercially. These mulches are popular because of the relative ease of transport and installation. The effectiveness of sheet mulches increases with the size of the sheets. For most hot, dry sites, a 2.5-by-2.5-foot sheet is considered the minimum dimension (Cleary et al 1988). On harsher sites, 3-by-3-foot or even 4-by-4-foot sheets are necessary to control competing vegetation. When purchasing and installing sheet mulches, it is important to consider (after Davies 1988a, 1988b):

- **Selecting the right size**—The size of the mulch is based on site conditions and the type and amount of competing vegetation. For example, a hot, south-facing site with full cover of competing grasses will need a large sheet mulch; a north-facing slope with scattered forbs and grasses will suffice with a smaller size.
- **Ordering only opaque materials**—Translucent materials are not used as sheet mulches because weed seeds can germinate and grow under these materials. During summer, surface temperatures under translucent materials can be lethal to seedling roots.
- **Using sheet mulches with long life spans**—It often takes three to five years for seedlings to become established on hot, dry sites and for this reason it is recommended that the sheet mulch have a durability of three years (Cleary et al 1988). The use of fully biodegradable materials is recommended.
- **Weeding or scalp around seedlings prior to installation**—Sheet mulch cannot be installed properly without vegetation being completely removed.
- **Mulching immediately after planting**—In Mediterranean climates, waiting until later in the spring to mulch runs the risk that competing vegetation will have depleted soil moisture, thereby making the mulch ineffective during the first growing season. This can be avoided, by applying mulch during or immediately after seedlings are planted.
- **Staking or anchoring all corners of the mulch**—The sides of the mulch sheets can be easily detached by wind, animals, or competing vegetation growing under the mulch sheets. It is important that the corners are staked. Burying all edges of the sheets with soil provides the greatest effectiveness.
- **Visibility**—Sheet mulches can be very apparent in high visibility areas. Measures to reduce unsightliness of sheet mulches include covering with aggregate mulches such as hay, straw, or wood mulch, or selecting sheet colors that blend into the area.



**Figure 5-20 | Mulch conserves soil moisture and inhibits the establishment of unwanted vegetation**

This photograph was taken in late summer, months after adjacent soils had dried out. The lack of competing vegetation and the low surface evaporation resulting from the placement of 3 to 4 inches of coarse sawdust resulted in very high soil moisture. In addition, the high C:N ratio of the sawdust was believed to be a factor in inhibiting the establishment of unwanted vegetation.

*Photo credit: David Steinfeld*

**Organic Aggregate Mulch**—Organic aggregates are another group of materials that, when placed thickly around installed plants, will control the establishment of competing vegetation and reduce surface evaporation (Figure 5-20). These aggregates include hay, straw, or chipped and shredded wood materials. Organic aggregates are often used in highly visible areas because they are more aesthetic in appearance than sheet mulches. They are also used in planting islands for long-term control of competing vegetation.

The longevity of organic aggregate mulches is a function of the C:N ratio, texture, and depth. High C:N materials, such as uncomposted shredded wood, bark, or sawdust, will last longer than low C:N materials, such as composted yard materials, because these materials are in the initial stages of the decomposition cycle. Coarse-textured materials (Figure 5-21 A, B, C E, F), which include long-fibered mulches have greater longevity than finer-textured materials (Figure 5-21D) because coarser materials have less surface area per volume of material for microbial break down (Slick and Curtis 1985). Coarse-textured materials also tend to hold less moisture, which slows decomposition rates. The longevity of an organic aggregate mulch also depends on the application thickness—the thicker the layer of mulch, the longer it will last. In many climates, coarse-textured materials, such as shredded wood, can last for years on the surface of the soil.

The same factors that affect longevity (e.g., texture, C:N ratio, depth) also determine the effectiveness of aggregate mulches in deterring seed germination of unwanted vegetation around the seedling. Coarse-textured mulches are excellent because they hold very little moisture at the mulch surface and, therefore, are poor environments for seed germination of unwanted vegetation. Alternatively, fine-textured mulches create a more favorable environment for seed germination because they hold more moisture and are in closer contact with seeds. For this reason, many fine-textured materials, such as composts, are excellent growing media for weed seed germination and establishment. As discussed in Section 3.11.4, mulch materials with high C:N ratios discourage growth of weedy annuals because high C:N materials remove available nitrogen that would otherwise give these species a competitive advantage. The effectiveness of a mulch in discouraging the establishment of competing vegetation generally increases with the thickness with which it is placed on the soil surface (Baskin and Baskin 1989). The most effective mulch thicknesses are between 3 and 4 inches (Pellett and Heleba 1995; Ozores-Hampton 1998), but thicknesses as low as 1.5 inches are effective for some small-seeded weed species that need sunlight for germination (Penny and Neal 2003).

Organic aggregate mulches have several advantages over sheet mulches. First, organic mulches can be applied over a much larger area than sheet mulches. Some projects have organic mulches covering the entire site, while other projects concentrate it in strategic areas, such as planting islands. Second, organic aggregate mulches moderate surface soil temperatures, whereas sheet mulches can increase surface temperatures. Mulch thicknesses of 3 inches have been found to reduce soil temperatures below mulch layers by 8 to 10 degrees F (Slick and Curtis 1985; Steinfeld 2004), which can benefit the growth of seedlings on very hot sites. The insulative quality of mulches also affects the seasonal heating and cooling patterns in the soil. Soils under thick mulches take longer to warm in the spring, but in the fall, take longer to cool down. Depending on the temperature and rainfall patterns of a site, this could influence seedling establishment.

Mulch can create problems to planted seedlings if it is placed in contact with the plant stem. The high moisture around the stem can be conducive to pathogenic injury. On southern exposures, heat will build up at the surface of, and directly above, the mulch,



**Figure 5-21 / Different types and textures of organic aggregate mulches**

(A) freshly ground coarse shredded wood passing a 3-inch screen; (B) freshly ground coarse shredded wood passing a 1.5-inch screen; (C) freshly chipped wood; (D) composted mixtures of ground wood, biosolids, and yard wastes passing a 1.5-inch screen; (E) weathered straw; (F) ponderosa pine needles. All materials are coarse textured except compost (D).

creating extremely high temperatures on warm summer days. The high temperatures can cause heat damage to stems of young seedlings. It is important, therefore, to keep mulch several inches away from the stem of planted seedlings.

**Rock Mulch**—Rock can be an effective mulch cover around seedlings. It consists of placing any size rock fraction (e.g. gravels, cobbles, stones) around each seedling. Rock protects the surface from erosion, evaporation, and weed establishment and it may be naturally available on a project site. As with organic aggregate mulches, rock is placed in a 1 to 2 feet radius around and at least several inches deep. Rock is kept away from the stem of the seedling to avoid heat damage. Prior to installation, unwanted vegetation is removed. One of the disadvantages of using rock mulch is that rock fragments can move downslope on steeper slope gradients and bury seedlings. Applying rock on gentler slopes or creating small planting benches where mulch can be placed around the seedling are recommended options.

## Soil Improvement

Mulches are sometimes used specifically to increase the nutrient and organic matter status of a soil. Composted organic materials are used for these purposes and are characterized by having low C:N ratios, high nutrient levels, fine textures, and dark colors. While these materials are typically more effective when incorporated into the soil, they are sometimes applied to the surface of the soil where tillage is not feasible (steep and rocky) or tillage costs are unaffordable. Where composted organic materials are applied on the soil surface, the nutrient release rates will be much slower. Refer to [Section 5.2.5](#) for more information on composts.

## Seed Supply

The objectives for applying mulch on some projects are to spread materials that contain native seeds. Several mulch materials carry native seeds, including duff, litter, and straw from native seed production fields. When these materials are applied to the soil surface, seeds will germinate given favorable environmental conditions.

Litter and duff are organic layers that form on the surface of the soil under tree and shrub plant communities. They are years of accumulated leaves and needles at varying degrees of decomposition. Included in these layers are dormant seeds, many of which are still viable. When the duff and litter are collected and spread on disturbed sites, the environmental conditions for breaking the dormancy of the seeds present in the material may be met and seeds will germinate.

Litter and duff can be collected from adjacent forest- or shrub-dominated sites or salvaged prior to slope construction. Reapplying them to disturbed sites completes several operations at once: supplies native seeds, covers seeds, and increases long-term nutrient capital. Although this practice might seem expensive or impractical, when compared with purchasing and applying seeds, fertilizer, and mulch separately, the costs could be comparable. Refer to [Section 5.2.3 \(see Litter and Duff\)](#) for more information on litter and duff.

One of the byproducts of native grass seed production is the stubble that remains in the fields after seed harvest. This stubble contains varying quantities of unharvested seeds which eventually end up in bales. If bales are stored in dry conditions, seeds can remain viable for several years. When hay bales that contain the native seeds are spread as a mulch on disturbed sites, seeds come into contact with soil and eventually germinate. It is important when acquiring native hay that the species and the source of the seed lot is appropriate for the site. Refer to [Section 5.2.3 \(see Straw and Hay\)](#) for more information on straw and hay.

## Selecting the Appropriate Mulch Materials

A variety of materials can be used as mulches, including the following:

- Shredded wood
- Erosion mats
- Hay and straw
- Manufactured wood strands
- Duff and Litter
- Composts
- Hydromulch

The following sections describe these materials and how they are used in revegetation projects. Figure 5-21 shows examples of some of these mulches.

### Shredded Wood

Mulches produced from woody materials are used primarily for seed covering and seedling mulching. There is usually a readily available source of wood material from project sites situated in forested environments. Branches, stems, bark, and root wads are typical waste products from clearing and grubbing that can be shredded, chipped or mulched on site to produce various types of wood mulch. In the past, this material has been burned or hauled to waste areas for disposal. With greater burning restrictions and higher hauling costs, chipping these materials and returning them to disturbed sites as mulch are practices that are becoming more common.

**Shredded Wood Production**—Creating mulch from right-of-way clearing woody material involves planning and coordination. First the road contractor piles the woody right-of-way clearing debris into “slash piles.” These piles include tree boles, bark, branches, and stumps, but it cannot contain large rocks or other inert materials that can cause wear or damage to the equipment. When clearing and piling is completed, a company that specializes in processing wood waste is contracted (typically by the road contractor). In this operation, equipment is brought to each slash pile and materials in these piles are processed into mulch. The resulting wood mulch is either placed in piles adjacent to the slash piles or transported to designated storage sites (Figure 5-22). The timing of these operations may be limited by fire restrictions in the western United States from mid-summer through early fall.

If undesirable plant species are included in the slash piles, spread of these species is likely to occur when they are processed and applied as a mulch. This can be prevented by identifying these plant populations on site during the weed assessment (Section 3.11.6) and avoiding placing them into slash piles.



**Figure 5-22 | Shredded wood piles are kept below 12 feet for safety reasons**

Shredded wood piles generate heat as they compost (A). A 4-foot-long thermometer (B) can be used to determine internal pile temperatures. Some operations require piles to be turned when a specific temperature is reached.

*Photo credits: David Steinfeld*

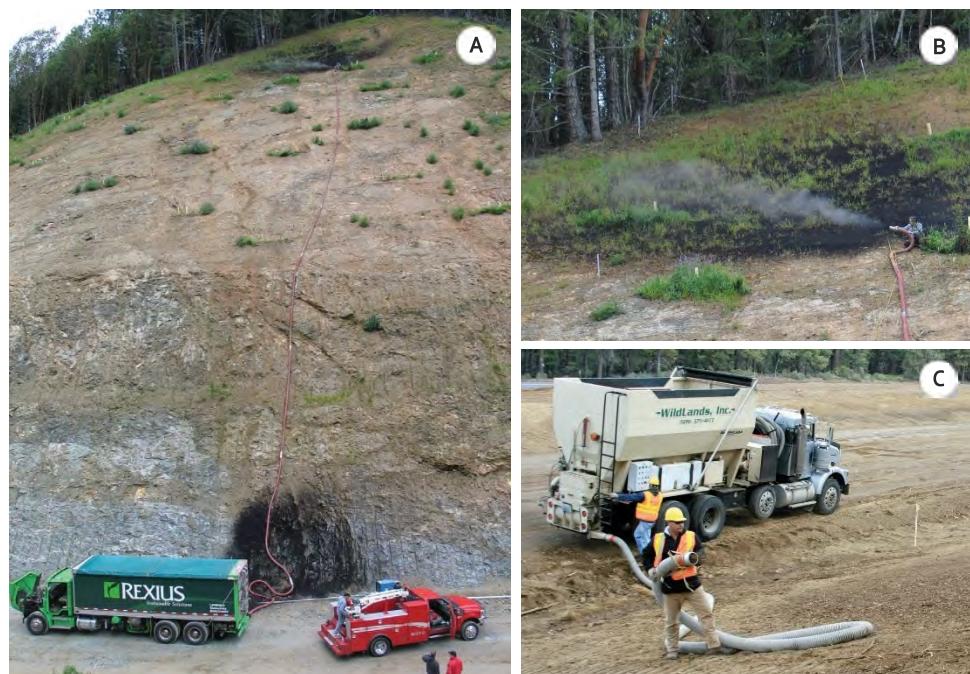
**Table 5-7 | General types of wood waste reduction equipment**

Modified from Re-Sourcing Associates and CPM Consultants 1997

| General equipment types | Examples                     | Feedstock                   | Particle geometry     |
|-------------------------|------------------------------|-----------------------------|-----------------------|
| Chippers                | Disc chippers, drum chippers | Whole logs, clean residuals | Clean edge, two-sided |

|                  |   |  |                                 |
|------------------|---|--|---------------------------------|
| <b>Hogs</b>      | Swing hammer, fixed hammer, punch and die, mass rotor | Wood waste, stumps, land clearing debris | Coarse, multi-surfaced, fibrous |
| <b>Shredders</b> | Low speed-high torque, high speed                     | Wood waste, stumps, land clearing debris | Coarse, multi-surfaced, fibrous |
| <b>Hybrids</b>   | Knife hogs, pan and disc                              | Wood waste, stumps, land clearing debris | Semi-coarse                     |

It is important to define the desired mulch characteristics prior to processing the piles. For example, the size and shape of the wood particles will determine their stability and propensity to move down slopes. Long strains of wood particles tend to be more stable than short strains because they knit together and have greater weight. Obtaining the proper size and dimension can be difficult because there is a variety of wood waste reduction equipment that produce different dimensions and fibrosity (the degree that wood fibers are separated). Obtaining the desired particle size and shape by specifying a screen size the material should pass does not always produce the desired material. Screens only sort for two dimensions, and not for length or fibrousness. Identifying the type of waste reduction equipment can narrow the type of mulch produced (Table 5-7). For example, mulch produced by shredders is long and fibrous (Figure 5-23 A and B), whereas mulch produced from chippers has sides that are of nearly equal lengths, with fibers still intact (Figure 5-23C). It is important that the correct equipment and screen sizes are used. It can be beneficial for the designer to request the contractor processing the material to send a sample of the different mulches that their equipment produces. In addition, being present at the beginning of the operations can assure that the proper material is being produced.



**Figure 5-23 | Blowing equipment is used to apply mulch on steep slopes**

Compost and shredded wood are applied on steeper slopes with mulch blowing equipment which is pneumatically delivered to the site through an application hose that can reach several hundred feet up steep slopes (A) with enough force for ample delivery of mulch (Image B is a close up of the application of mulch near the top of Image A). Large trucks can hold between 75 and 100 yards of mulch (A), while smaller trucks can hold up to 25 yards (C). A tackifier is added to compost to increase surface stability on steep slopes.

*Photo credits: David Steinfeld*

Generally, the coarser the size of the mulch, the less expensive the production costs because more mulch of coarser size can be produced in a given time frame than smaller textured mulch. Other factors, such as tree species, moisture content, and portion of tree processed, affect the characteristics of the mulch. If the wood is wet during processing, it is more likely to be shredded; if it is dry, it will be more chip-like. There is also variation in mulches based on species of origin. For example, ponderosa pine (*Pinus ponderosa*) and western juniper (*Juniperus occidentalis*) tend to create more

fibrous mulch than lodgepole pine (*Pinus contorta*). Processed root wads tend to be more fibrous than boles of trees. They may also contain large rocks which will damage processing equipment and need to be removed prior to processing or set aside and not used.

The shredded wood operation needs space to store the pre-processed woody debris, operate the processing equipment, and a storage site for the processed shredded wood. Depending on the operation, this could require a significant area. It is important to consider the size of the processed shredded wood piles when determining space requirements. Standards for pile heights of no greater than 12 feet are often used for fire and safety reasons. Piles larger than this height may be prone to spontaneous combustion. Long stemmed thermometers are available to monitor interior pile temperatures. One consideration for pile locations is the distance to the areas being mulched. Travel time between mulch piles and application areas can be as longer than the actual application of the material.

**Purchasing Shredded Wood**—On projects where waste materials are not available for mulching, purchasing shredded wood may be an option. Overall costs are much higher than processing waste wood on site because of the purchase of the material and transporting to the site. If commercial mulch sources are nearby, transportation costs are significantly reduced. Testing the shredded wood is important to obtain suitable material. Methods and specifications can be developed that are similar to those provided in [Section 5.2.5](#), or available in the [Native Revegetation Resource Library](#).

**Applying Shredded Wood**—Shredded wood is applied in several ways—pneumatic mulch-blowing equipment, adapted manure spreaders, and rock slingers. Pneumatic mulch blowing equipment ([Section 5.2.3](#), see [Seed Covering](#)) has varying transport capacities, ranging from 25 to 100 yards of material ([Figure 5-23](#)).

An application hose is positioned where mulch is to be applied and is pneumatically delivered from the mulch bins to the site. The amount of mulch applied depends on the revegetation objective. For seeding, application rates range from 100 yd<sup>3</sup>/ac (0.75-inch thickness) to 135 yd<sup>3</sup>/ac (1.0-inch thickness). For seedlings, mulch application ranges from 400 yd<sup>3</sup>/ac (3-inch thickness) to 540 yd<sup>3</sup>/ac (4-inch thickness). Note: The higher rates used for mulching seedlings are only used in close proximity to the plants. The remaining areas are mulched at a lower rate.

When applying shredded wood for erosion control objectives, the particle size of the material may be as important as the amount of material applied. On compacted soils, where overland flow is expected from heavy rainstorms, shredded wood lacking material smaller than an inch thick in length performs better in reducing sediment loss than material with finer wood. This coarse material can be applied at 50 to 60 percent ground coverage with good erosion control (Foltz and Wagenbrenner 2010).

Application rates depend on factors such as length and diameter of hose, blowing equipment, elevation rise, and dimensions of the area being covered. Rates of application typically range from 25 to 35 yd<sup>3</sup>/hr. If mulch is applied at a 1-inch depth (134 yd<sup>3</sup>/ac), it would take between 4 to 5 hours to cover an acre. These are optimum rates because they do not account for the time it takes to travel to the mulch source, load the mulch bin, and travel back to the application site. The time required to make these trips can sometimes be longer than the actual application time for mulching. Using larger transport capacities is one way to significantly reduce the time associated with refilling mulch bins.

Mulch blowing equipment can be used on any slope gradient that can be accessed by foot. By using ropes, slope gradients of up to 1H:1V can be accessed. Hose lengths can



**Figure 5-24 | A manure spreader adapted to side cast shredded wood**

Shredded wood can also be casted from adapted manure spreaders. On this project, the seed mix was sown using a hydroseeder, then 0.75 inches of shredded wood was applied over the seeds.

be attached to extend the delivery of mulch up to 400 feet. Because mulch is delivered through hoses, the system will plug if the size of the shredded wood exceeds the tube size. It is therefore important that mulch be free of large pieces of wood. Using a mulch blower is an excellent method for evenly applying shredded wood, but frequent monitoring by inspectors is important to ensure that the specified amount of mulch is being applied.

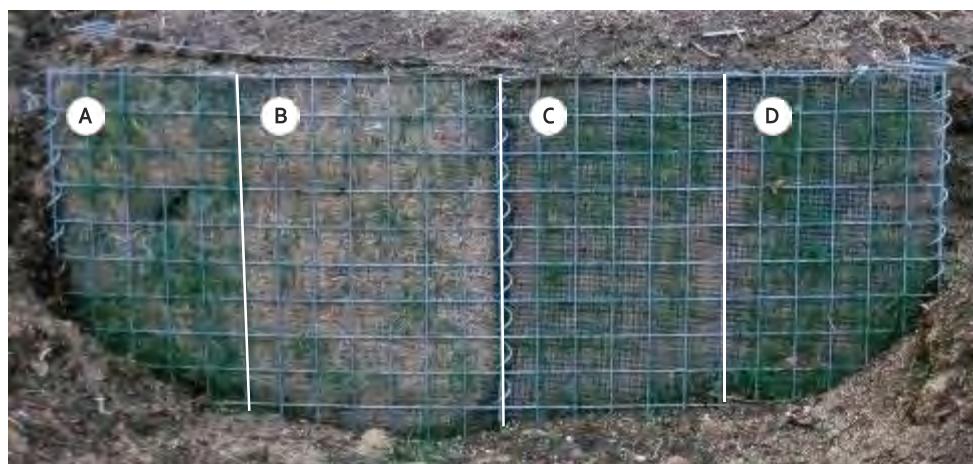
Shredded wood can also be applied with modified manure spreaders (Figure 5-24). Depending on the type of modifications, this equipment can apply shredded wood at a much faster rate than mulch blowing equipment, however this equipment generally is limited by the distance it can cast the material which is typically ranges from 35 to 75 feet depending if it is being applied upslope (less distance) or downslope (greater distances). Tackifiers are not typically applied to shredded wood after it is placed because this material is less susceptible to wind erosion than mulches that are finer textured and lighter.

**Seed Placement during Mulching**—Seeds can be sown prior to or during mulch operation. Seeds that are sown prior to mulching can be done with dryland sowing methods, hydroseeding, or through the pneumatic mulching equipment ([Section 5.2.3, see Seed Covering](#)). A thicker layer of shredded wood is applied in a second pass at a depth between 0.5 and 1.0 inches over the seed.

## Erosion Mats

Erosion mats (e.g. erosion control revegetation mats and rolled erosion control products) are manufactured blankets or mats designed to increase surface stability and control erosion. They are applied in direct contact to the soil surface for control of sheet erosion and to aid in the establishment of vegetation.

A multitude of products are on the market, with a range in design and costs. Determining which products will meet project objectives at the right price can be challenging. Several State Departments of Transportation periodically evaluate and compare the shear stress, soil erosion protection, longevity, and other characteristics for these products (Caltrans 2003), and these documents are available on the internet. State of Washington and State of Oregon DOTs are moving to require that all erosion control matting be fully biodegradable. Because the installed costs of erosion mats can be expensive, it is important that the job is done correctly. Taking the time to select the appropriate erosion mats, native species mix, and seed placement techniques is essential for ensuring that revegetation is successful. Small field trials using different species and erosion mats can help in these decisions (Figure 5-25).



The same characteristics that create an optimum environment for seed germination in other mulches are also important to consider when selecting erosion mats.

**Figure 5-25 / Field trials of species and materials**

Small field trials can help select the most appropriate species and materials for a project. The trial shown in this photograph compared straw mat (A and B) with a polywoven mat (C and D). It also compared the growth of blue wild rye (*Elymus glaucus*) and California fescue (*Festuca californica*) (A and C). The first-year results indicated that there was much better establishment of grasses on the polywoven erosion mat than the straw mat, yet no difference in species growth. Maintaining the trial for two years showed that California fescue outperformed blue wild rye. These results led to using California fescue and the polywoven erosion mat for the project described in Inset 5-3.

*Photo credit: David Steinfeld*

Typically, products that protect the seeds from drying out but allow light and space for germinating seedstogrow into seedlings will perform best for vegetation establishment. The thicker erosion mats with the most loft typically have better conditions for seedling establishment than thinner materials how- ever, if the mat is too thick, seedlings will not be able to emerge from the fabric.

The drawbacks to erosion mats are generally not in the product itself but in how it is applied to the site. Poorly applied erosion mats can result in sheet and rill erosion under the fabric. To avoid this problem, several important measures can be taken when installing erosion mats. First the surface of the soil is smoothed to a uniform fabric to prevent overland water flow from undercutting the mats.

Seeds are sown on the site prior to installing erosion mats. Because installation is commonly performed by the road building contractor, it is important that the designer works with the contractor to assure that seeds are being sown during the surface texture before the mat is placed. Landscape staples or pins are then installed at a specified spacing to hold the matting and ensure in intimate contact with the soil surface. The matting is trenched or keyed into the soil at the upper reaches of the seeding windows. Seeding can be done using any type of seeding method (e.g., hydroseed, slopes have been sown, care taken during and after mat installation can help avoid disturbing the seed. Unless the seeds are extremely small, sowing seeds over installed erosion mats is not recommended because larger seeds will hang up in the fabric. Small seeds can be applied over erosion mats if tackifiers are not used and if the timing is such that sufficient rain will move it through the erosion mat to the soil surface.

Some manufacturers offer erosion mats that are impregnated with seeds, eliminating the need for sowing. This method is advantageous on steep slopes or soil-faced gabion walls (Inset 5-3) where placing seed prior to mat installation is difficult. It is important to work directly with companies that provide these products by supplying them with source-specific seeds and specifying appropriate sowing rates. For successful germination, seeded erosion mats are installed so that seeds and fabric are in direct contact with the soil. This method works best in environments with reliable moisture during germination.

#### ***Inset 5-4 / Source identified straw bales***

A secondary product from the seed production contracts is the straw that remains after harvest. This material can be used for erosion control or seed covering. It can be used as a mulch and has the additional advantage of being a source of unharvested viable seed. This product is to be treated similarly to certify straw sources (**Section 5.2.3 (see Straw and Hay)**). No noxious or undesirable weed seed should be in the bales. A visit to the seed production fields prior to seed harvest will indicate if there are any unwanted species that will be present in the hay bales. Good practice is to keep bales of each seedlot separate from other sources to prevent mixing. If straw bales are stored for any length of time, good practice is to protect them from rain.

### **Inset 5-3 / Case Study—Erosion mats with native grasses and forbs**

Reconstruction of the Agness-Illahe Highway involved building long sections of gabion walls. Because this highway was visible from the Rogue River (a designated “wild and scenic” river in southwestern Oregon) and was heavily traveled by river runners and fishermen, it was important that the gabion walls be visually screened using native plants. Gabions were designed to hold 12 inches of compost-amended soil (topsoil was not available) on the face of the walls by wire mesh frames (Image A in the illustration below). Placement of seeds at the surface of the gabion wall was problematic. Several small plots using different erosion mats, seed mixes, seed rates, and seed-attaching methods were tested to determine how to best meet the revegetation objectives (Figure 5-25).

The results from these trials indicated that native grass and forb seeds could be attached to erosion mats using a tackifier (Image B). In 2003, the findings were applied to the construction project. Needing approximately 33,000 ft<sup>2</sup> of gabion wall facing, the erosion mats were prepared by rolling them out on a road surface, applying California fescue (*Festuca californica*), gluing the seeds to the mat, and re-rolling the erosion mats.

The seeds held tightly to the fabric during transportation and handling. At the construction site, seeded mats were attached to the wire mesh at the face of the wall (Image C) and compost-amended soil was placed behind the screen and lightly tamped. The gabion walls were built in the summer of 2003, but the seeds did not germinate until late fall after several rainstorms. Image D shows a close-up section of wall with newly germinating seedlings emerging through the erosion mat in late 2003, four months after wall construction. Image E shows 20- to 30-foot-high walls in July 2006, three years later, fully vegetated and effectively screening the walls from the road and river. For more detail, see the Native Revegetation Resource Library.

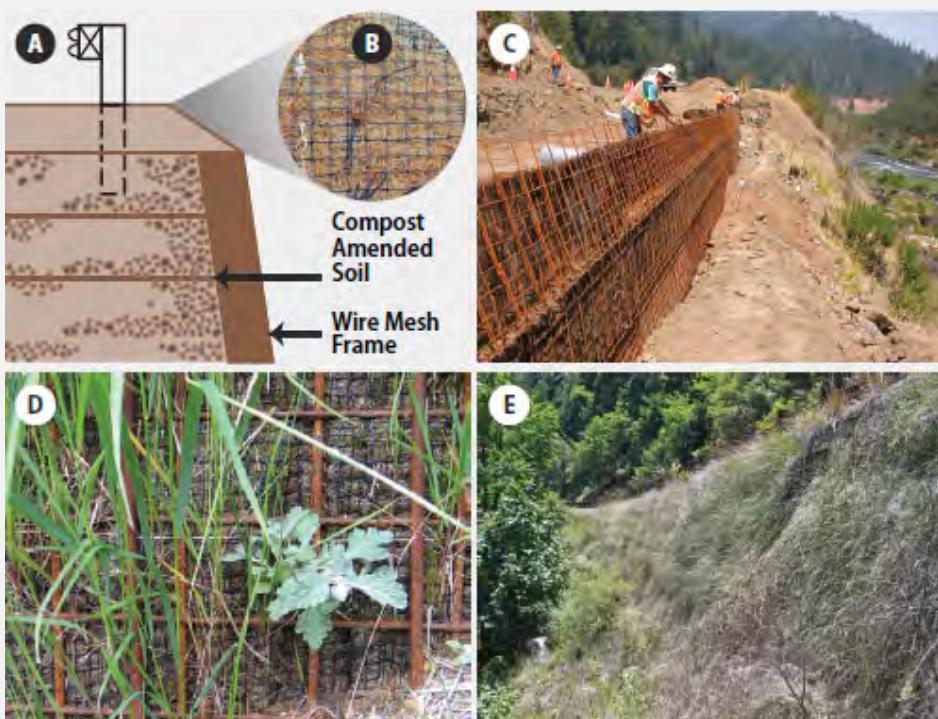


Photo credits: A, B, D and E by David Steinfeld, C by Scott Blower

## **Straw and Hay**

Straw and hay are long-fibered mulches used on many revegetation projects for seed cover and erosion control (Image E in the illustration in Inset 5-3). The terms “straw” and “hay” are often used interchangeably, however there are distinct differences—straw is the stubble left over after seeds have been harvested from commercial seed or grain crops while hay comes from grass/legume fields and usually the whole plant baled for stock feed. When hay is harvested, it usually contains seeds from a variety of pasture species.

Straw and hay are often used on revegetation projects because they are available, comparatively inexpensive, and generally successful in establishing grass and forb plants from seeds. The long stems of these materials create loft, or high porosity, that keeps moisture near the soil surface where seeds are germinating. This creates a

favorable environment for young seedlings by allowing sunlight to penetrate and protecting the young seedlings during the early stages of establishment.

Straw is often preferred over hay because it generally contains fewer undesirable seeds and the seeds that are contained in straw are not always inappropriate for use in revegetation projects. For example, straw from some grain production fields (e.g., rice straw) has seeds of species that are not adapted to many environments and therefore may not become established. Straw produced from native seed production fields may have desirable seeds if the fields are grown from identified genetic sources and used for projects within the appropriate seed transfer zones (Inset 5-4). Using source-identified straw not only act as a mulch when it is applied, but it also supplies extra native seeds to the site. Some native grass species grown for seed production are very difficult to harvest and clean. One strategy for these species is to allow the seeds to ripen in the fields and bale the seeds and stems together. The seeded bales can be applied directly to the site through straw blowing equipment, accomplishing seeding and mulching in one operation.

**Purchasing Straw and Hay**—The drawbacks to using straw and hay are that these materials can contain seeds from undesirable species, are susceptible to wind movement, may have limited application distance, and decompose in a relatively short time compared to other mulches. Introducing seeds of undesirable species from straw and hay sources is an important consideration when choosing a source. There are many examples where, in the urgency of erosion control, straw or hay from unknown sources has been applied, resulting in the introduction of weed species. The assumption is that short-term control of soil erosion and sediment production outweighs the long-term introduction of undesirable plant species. The possible results from these assumptions need to be understood and agreed upon prior to applying hay or straw from unknown sources. Good integration of erosion control and revegetation planning can eliminate the need for last-minute purchase and application of unknown or undesirable hay or straw sources.

Many states have certification programs that inspect fields and certify that the bales are “weed free” which means that the material is free of the noxious or listed weeds of the inspecting state. There can be other seeds in the bales, however, that may not be considered weeds by the certifying state, but unwanted on the project site. A conservative approach is to examine the fields that will produce the bales and observe which species are present before they are harvested. It can be assumed that if a species of grass or forb is present in a field, seeds from these plants will show up in the bales.

Some things to consider requiring when purchasing straw or hay are as follows:

- Does the source exceed State Certification Standards for “weed free”? Many states have straw certification programs but for states where no standard programs exist, acceptance can be based on seed crop inspection reports and/or visual field inspections prior to harvest. Standards may also be set by the Government and listed on individual task orders.
- Is straw or hay baled and secured? Generally, bales less than 100 pounds work best.
- Are bales wet after harvest or during storage prior to delivery? Not only is wet material harder to handle and less effective, mold is often present, which can pose respiratory risks to applicators.

The quality of straw and hay varies among grass species. Rice straw, for example, is wiry and does not readily shatter, which makes it more difficult to apply as compared to wheat, barley, or oats (Kay 1983; Jackson et al 1988). Native straw is generally longer and stronger than grain straw (Norland 2000), although some native species have better properties as mulches than others. Larger stemmed grasses, such as blue wildrye (*Elymus glaucus*), mountain brome (*Bromus marginatus*), and bluebunch wheatgrass (*Pseudoroegneria spicata*) make good mulches because of their large leaves and stems.

**Application**—Straw and hay can be spread by hand or with a straw blower. For large jobs, using a straw blower is the most practical application method. Many types of straw blowers are available, ranging from very small systems (Figure 5-26) that deliver from 30 to 180 bales per hour to large straw blowers that operate at rates up to 20 tons per hour. The distance that straw or hay can be blown depends on the hay-blowing equipment, wind conditions during application, straw characteristics, and whether the material is being applied upslope or downslope (cuts or fills). When wind is favorable, straw can be shot up to 100 feet. However, when wind is blowing against the direction mulch is being applied, the distance is reduced. Because of the limited application range, this equipment is limited to sites adjacent to roads. The upper portions of steep, extensive slopes are typically not reachable by straw blowers.

When straw is used as a seed mulch, it is important that the application rates are not so deep that a physical barrier is formed. A minimum depth that has been shown to control evaporation is 1 inch (Slick and Curtis 1985). Applying too much straw will restrict sunlight and growing space for establishing seedlings. A rule of thumb is that some surface soil (15 percent to 20 percent) is visible through the straw after application (Kay 1972, 1983; Jackson et al 1988). This equates to 1.5 to 2 tons per acre, depending on the type of straw and its moisture content. This rate can be adjusted based on the climate of the site.

Straw is susceptible to movement with moderate to high winds. Tackifiers are often applied over the straw to keep it in place (Kay 1978). Products such as guar and plantago are used with low quantities of hydromulch to bind straw together. Straw can also be crimped, rolled, or punched into the soil. A puncher is a roller with a set of straight studs, greater than 6 in long, that push straw into the soil. A crimper is similar but has serrated disks attached to a roller. This equipment stabilize the straw by burying portions of the stems into the soil and increase erosion protection because of the greater contact of straw with the soil surface. Some soil types may not be suitable for crimping, especially sandy soils that may not have the strength to hold straw in place. It is important to consider the potential for soil compaction that may result from heavy equipment on soils.



**Figure 5-26 | Straw blower**

Straw blowers range in size from machines that can apply 30 to 60 bales per hour (shown here) to very large straw blowers that can shoot up to 20 tons per hour.

*Photo credit: David Steinfeld*

## Wood Strands and Wood Wool

Wood strands and wood wool are commercially available wood products used for mulch. Wood strands are long, thin pieces of wood produced from wood waste veneer, whereas wood wool, known as excelsior, is wood slivers produced from aspen, spruce, and pine wood. These products are developed as an effective erosion-control alternative to straw and hay (Foltz and Dooley 2003). The advantages of wood strands and wood wool over straw are that they are free of seeds, have greater longevity, and are more resistant to wind.

Wood strands and wood wool form a stable surface cover with high porosity or loft; characteristics that are important for controlling soil moisture and temperature around the germinating seeds. The large spaces or pores created by the wood strands allow space, light, and protection for young emerging seedlings (Figure 5-15). Unlike straw, these materials keep their structure or porosity over time and do not compress with snow or lose fiber strength until they begin to decompose. The application rates for wood strands follow the guides for straw—at least 15 percent to 20 percent of the soil surface is visible. This may result in a lower application rate than recommended for erosion control. Installing small test plots of varying thicknesses of mulch is a good means to determine the appropriate thickness for optimum seed germination and erosion control. Wood strands are delivered in different size bales and applied by hand or through straw-blowing equipment (Figure 5-26). As with straw, this product is limited by the accessibility of the site by hay transportation and blowing equipment.

## Litter and Duff

Litter is the layer of fresh and partially decomposed needles and leaves that cover the surface of most forest and shrub plant community soils. Duff is the dark, decomposed layer directly below the litter layer (leaves and needles are not identifiable in the duff layer) that is high in nutrients and humus. In addition to providing soil protection and nutrients, litter and duff can contain dormant, yet viable, seeds from species that make up the forest or shrub plant communities (Section 5.2.3, see Seed Supply). When litter and duff are collected, matching them to the appropriate revegetation site is good practice. For example, litter and duff collected from cool, moist sites are not to be applied on hot, dry sites.

The depth that litter and duff accumulates will vary by species composition, age, and productivity of the plant community. The quality of the litter for erosion control and longevity varies by the dominant forest or shrub species. Pine needles provide the greatest benefit because the long needles interlock, reducing the potential of movement from rain or wind erosion (Inset 5-5) (Image F in Figure 5-21). Needles from species such as Douglas-fir are shorter and tend to compact, providing less surface stability. Litter from deciduous tree and shrub plant communities provide less protection because the leaves are less interlocking, provide less loft, and often form a mat that can be difficult for germinating seeds. Nevertheless, these materials can be

### **Inset 5-5 / Pine straw industry**

Using forest litter as a mulch is not a new concept — pine needles have been a popular landscaping mulch in the southern United States for the past 25 years. The “pine straw” industry, as it is referred to, has been established to harvest needles in a sustainable manner from young plantations of southern pine species to meet this demand.

a source of seeds and nutrients and not to be immediately overlooked.

In the western states, litter has typically been collected manually by raking. The collection of litter has been mechanized in the southern United States, and the pine straw industry is well developed in this part of the country. Baling equipment has been developed for this industry and could be applicable to the western United States.

Collection of needles and duff is done when the litter and duff are dry. If these materials are not used immediately, they are placed in small piles and covered with plastic to keep the materials dry. Excessive moisture can turn the piles into compost and possibly affect seed viability.

Litter and duff can be applied manually to disturbed sites. If the litter and duff is free of large materials, it can be applied in a variety of ways including pneumatic mulch blowers, straw blowers, and modified manure spreaders.

One method for determining the amount of viable seeds in a litter layer is to conduct a germination test at the project site over a several-year period by obtaining litter and duff from several potential collection areas and testing them on nearby sites. Dry litter and duff samples are collected from each plant communities and a known amount is weighed. The samples can either be spread in an area near the project (free of vegetation and topsoil) or on potting soil in a garden or greenhouse. In either case, the duff and litter is overwintered and a germination count is made in the spring. The numbers of seedlings per known volume or weight of litter and duff material is used to determine the rate of litter to be applied.

## 5.2.4 TOPSOIL

### Introduction

Topsoiling is the salvage, storage, and application of topsoil material to provide a suitable growing medium for plants and to enhance soil infiltration (Rauzi and Tresler 1978; Woodmansee et al 1978; Natural Resource Conservation Service (NRCS) 1994).

Topsoiling has been found to increase plant cover and biomass through an increase in nutrient availability, water-holding capacity, and microbial activity, including mycorrhizae (Claassen and Zasoski 1994).

While topsoiling has many benefits for revegetation, topsoiling cannot re-create the original undisturbed soil. In the process of removing and reapplying topsoil, soils undergo a loss of soil aggregation, organic nitrogen, arbuscular mycorrhizal fungi (AMF) inoculum, and microbial biomass carbon (Visser et al 1984). Minimizing topsoil disturbance is preferred to topsoiling, especially on sensitive soils, such as those derived from granitic and serpentine bedrock (Claassen et al 1995).

### Salvaging Topsoil

Salvaging topsoil is done in areas that will be severely disturbed during construction. These areas are usually identified early in the planning stages. Topsoil within these areas can be surveyed to determine the depth and quality of the topsoil that can be excavated. If weeds are observed during the field survey, it is a good possibility that the seeds of these species are present in the topsoil. These areas can be avoided or undesirable vegetation treated prior to topsoil removal. Ideally, only the topsoil, and not the subsoil, is removed in the excavation process. Mixing subsoil into the topsoil will dilute microbial biomass and mycorrhizal inoculum and reduce the quality of the topsoil.

During topsoil excavation, the litter and duff layers are removed together with the topsoil. These layers are sources of decomposed and partially decomposed organic

matter which will undergo some decomposition, releasing nutrients and organic matter to the soil during storage. There are circumstances where the duff/litter layer is removed and stored separately from the topsoil which is when the duff and litter are to be used as a native seed source or soil cover (Section 5.2.3, see Seed Supply).

Topsoil layers are typically removed with either the blade of a tractor or excavator bucket. Where precision is important, using an excavator bucket is the preferred tool. Thin topsoils, common on high elevation sites or soils where the subsoil cannot be mixed with the topsoils (e.g. subsoils with very high or low pH, high sodium, high salinity) can be scraped with the front of the excavator bucket to the desired depth. There is less depth control when using a tractor.

Topsoil is typically moved into berms at the bottom of fill slopes or the top of cut slopes and stored there until it is reapplied or transported to a storage area. This will likely require that the storage areas along the cut and fill slopes will have minimal amount of disturbance until the soil is reapplied. Typical berm dimensions are approximately 6 feet wide by 3 feet high. In areas where there is more topsoil than can be stored on-site, topsoil is trucked offsite to larger storage piles. When selecting off site storage areas, it is important to assess the site and surrounding area for unwanted vegetation because of the potential that this vegetation will become established on the topsoil piles. It is also important to plan salvaging operations so that there is a minimal amount of compaction prior to removal. This will likely require keeping large equipment travel on salvaged topsoil areas to a minimum.

To maintain optimum soil quality, it is best to excavate topsoil when soils are relatively dry. Under dry conditions, there is less potential to compact the soil, destroy soil aggregation, or oxidize organic matter. Dry topsoil store longer and maintain better viability than moist topsoil (Visser, Fujikawa, et al 1984). Restricting topsoil excavation operations to dry periods for large topsoil piles, or if topsoil is to remain in piles for more than one year, will increase the viability of the topsoil.

## Storing Topsoil

The question often raised around storing topsoil is how long it can remain piled before it loses its viability. Studies have shown that stored topsoil can remain viable from six months (Claassen and Zasoski 1994) to several years (Miller and May 1979; Visser, Fujikawa, et al 1984; Visser, Griffiths, et al 1984,) but will decrease in viability after five years (Miller and May 1979; Ross and Cairns 1981).

Viability of stored topsoil is a function of moisture, temperature, oxygen, nitrogen, and time. Stockpiled topsoil has been compared to "diffuse composting systems" (Visser, Fujikawa, et al 1984) because, under optimum conditions, organic material in the topsoil will compost. Decomposition of organic matter in stored topsoil will reduce microbial biomass essential for nitrogen cycling (Ross and Cairns 1981) and fine roots that store mycorrhizal inoculum (Miller and May 1979; Miller and Jastrow 1992). Optimum environments for decomposition include high moisture, warm temperatures, and available nutrients, all conditions present in most topsoil piles. Climates with lower moisture and temperatures can be more favorable to long-term storage. A study in Alberta, Canada, for example, revealed that topsoil had very little respiration or organic decomposition after three years in a stockpile due to the influence of the cold, dry climate (Visser, Fujikawa, et al 1984). Dry topsoils store longer and maintain greater populations of viable mycorrhizal fungi (Miller and Jastrow 1992). Topsoil piles that will be held over winter in areas of moderate to high rainfall will benefit by covering with plastic to keep the soil dry. This will also keep the piles protected from erosion and weed establishment.

The size of the pile can also affect the viability of the topsoil. The interior of large piles maintains higher temperatures and are usually anaerobic, which can be detrimental to soil microorganisms. Microbial biomass levels and mycorrhizal fungi have been found to be very low in the bottom of large stockpiles (Ross and Cairns 1981; Miller and Jastrow 1992). Most projects limit topsoil piles to 3 to 6 feet in height. This is not always possible, especially when topsoil storage space is limited. Under these circumstances, the size of the topsoil pile can be quite large. To reduce the negative effects associated with very large piles, topsoils can be salvaged dry and kept dry during storage. They can also be stored for as short a time as possible. In addition to maintaining the viability of the topsoil, minimizing storage time will reduce the risk of weed infestation.

Standard specifications often call for temporary seeding of topsoil piles. The benefits of this practice are erosion control and maintenance of mycorrhizae inoculum through the presence of live roots. This practice also runs the risk of introducing undesirable plant species that may be present in the seeding mix if a non-native species mix is used. Alternatives to this practice include hydromulching without seeds or covering with plastic however, both practices lose their effectiveness with time.

## Reapplying Topsoil

The depth of topsoil application is generally based on the amount of topsoil available and the desired productivity of the site after application. As a rule, the deeper that topsoil is applied, the greater the site productivity. If the objective is to restore a site to its original productivity, consider placing topsoil at a depth equal to or greater than the topsoil horizon of undisturbed reference sites. Sufficient topsoil quantities, however, are rarely available in the quantities needed to restore disturbed sites to their original topsoil depths. This often leads to applying topsoil too thinly across a project site. There may be a minimum topsoil depth below which the application of topsoil is not effective. Research on a northern California road construction site (Claassen and Zasoski 1994) suggests that a depth of 4 to 8 inches is necessary for an effective use of topsoil. On sites where the subsoil is unfavorable for plant establishment (e.g., very high or low pH, high sodium, high salinity), minimum depths of greater than 12 inches of topsoil may be considered (Bradshaw et al 1982).

Determining minimum topsoil application depths can be based on the minimum amount of N required to establish a self-maintaining plant community. A threshold of approximately 700 kg/ha (625 lb/ac) of total nitrogen in the topsoil has been suggested for sustaining a self-maintaining plant community in a temperate climate (Bradshaw et al 1982). Claassen and Hogan (1998) suggest higher rates, especially on granitic soils, of 1,100 lb/ac total nitrogen. Using total nitrogen levels from soil tests of topsoil in reference areas, the application thickness of topsoil can be determined using the calculations presented in Figure 5-27.

Some sites, like high elevation and desert environments, have very little topsoil to begin with. In these areas, salvage and application of topsoil may be less than 4 inches depending on the topsoil depth of the reference site. While this may not seem worth the effort, the addition of a thin layer of salvaged topsoil may provide soil organisms and seeds that are essential for restoring the site to original productivity and species composition.

|   |                 |  |
|---|-----------------|--|
| • Total soil nitrogen (or other nutrient of interest) in salvaged topsoil | 0.14%           | From soil test of post construction soils. Reported in gr/l, ppm, mg/kg, ug/g; divide by 10,000 for percentage |
| • Soil bulk density   | 1.1 gr/cc       | Unless known, use 1.5 for compacted subsoils, 1.3 for undisturbed soils, 0.9 for light soils such as pumice    |
| • Fine soil fraction  | 70%             | 100% minus the rock fragment content; from estimates made from sieved soil prior to sending to lab             |
| • Nitrogen for soil layer<br>• $A * B * C * 270 =$                        | 2,911 lbs/ac ft | Calculated amount of total nitrogen in 1 acre feet of soil   |
| • Minimum or threshold N levels   | 1,100 lbs/ac    | Determined from reference sites or minimum thresholds from literature  |
| • Minimum topsoil application<br>• $E/D * 12 =$                           | 4.5 inches      | The minimum thickness of topsoil to apply to meet minimum thresholds of nitrogen                               |

**Figure 5-27 | Minimum topsoil thicknesses can be calculated from nitrogen tests**

Soil testing of salvaged topsoil can be used to calculate the thickness to apply to meet minimum nitrogen levels.

Once the desired topsoil depth has been established, then it can be determined if there is enough salvaged topsoil available. The quantity of topsoil needed can be determined based on the depth of topsoil from Figure 5-28. For example, if an average of 10 inches of topsoil is needed on a project, approximately 1,350 yd<sup>3</sup>/ac topsoil will be necessary. If there is not enough topsoil to meet this depth, then an alternative approach may need to be taken, including strategic placement of topsoils, or areas where topsoil depths are reduced. The strategy might concentrate topsoil in areas such as planting islands, planting pockets, or in a mosaic pattern that blends with the natural vegetative community. In areas that have reduced or no topsoil, other mitigating measures could be used, such as incorporation of organic matter into the subsoil or mulching.

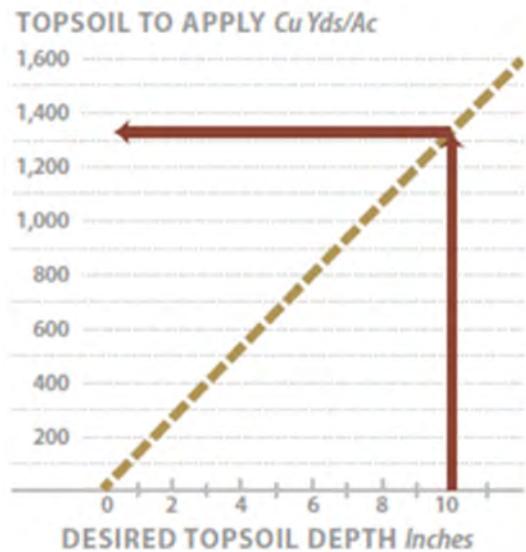
## Manufactured Topsoil

The term manufactured topsoil (also referred to as “engineered topsoil”) is used to define a soil created to perform like, or develop into, topsoil. It is usually manufactured offsite and transported to the area where it is applied. Manufactured topsoil is used in gabion walls, crib walls, or other bioengineered structures. It can also be used in planting pockets and planting islands.

Selection of the appropriate organic matter, soil texture, and soil amendments for manufactured topsoil will increase the success of the project. It is critical that manufactured topsoil be similar to the soils of the reference site with characteristics that will support the desired plant community. The basic components of a manufactured topsoil are the loam borrow, compost, and soil amendments.

**Loam Borrow**—Loam borrow is any material that is composed of mineral particles meeting a suitable texture class (Figure 5-29). Optimal loam borrow is low in coarse fragment content, is weed-free, and meets general soil quality specifications shown in Table 5-8.

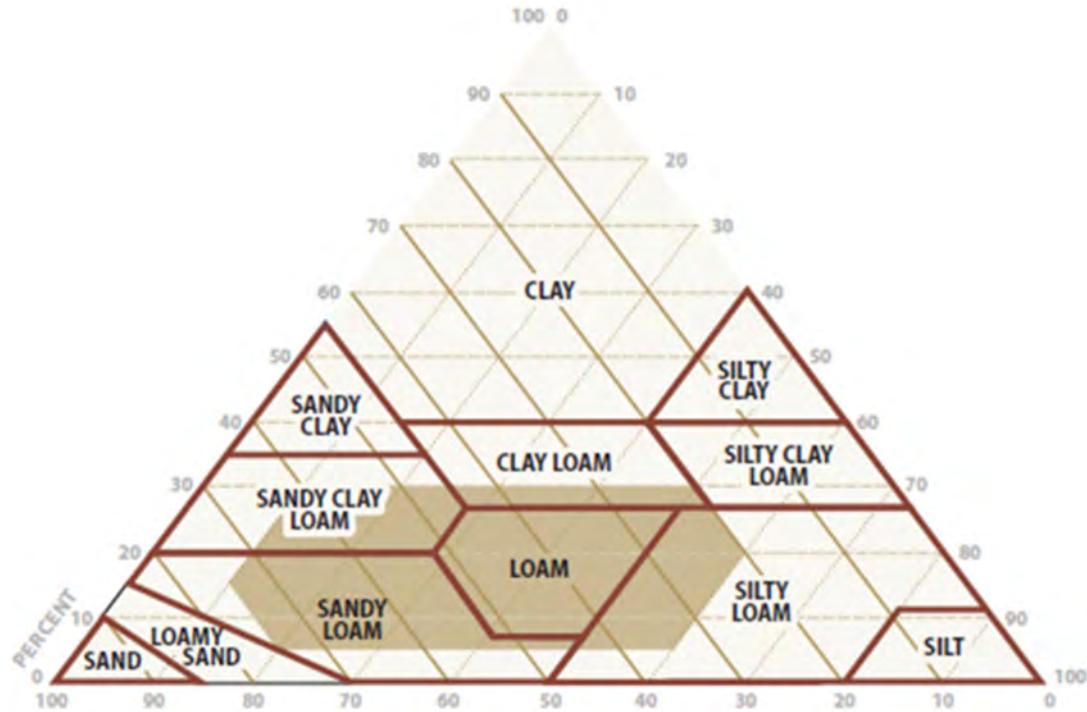
Testing loam borrow will determine if the material meets these general specifications. Loam borrow can come from many sources, such as subsoils, river sands, and terrace deposits. When loam borrow comes from subsoils or parent material, it can be assumed that beneficial soil micro-organisms, like mycorrhizae and nitrogen-fixing bacteria are not present. These microorganisms can be added to the loam borrow using inoculums (Section 5.2.7).



**Figure 5-28 | Determining the soil quantity needed for a specific topsoil depth**

The quantity of topsoil to apply to achieve a specified topsoil thickness can be estimated using this graph. For example, to create a topsoil, 10-inches deep, would require 1,350 yd<sup>3</sup> of topsoil.

**Compost**—The organic component of manufactured topsoil (Section 5.2.5, Organic Matter Amendments) is composted materials from a variety of materials, including yard waste materials (grass clippings, leaves, and shredded wood of trees and shrubs), sawdust, and biosolids. The heat generated during composting effectively reduces pathogens, weeds, and insects that may be hazardous to humans and detrimental to the reestablishment of vegetation. To reduce the potential for spreading weeds, compost specifications call for material that is free of weed seeds and vegetative



**Figure 5-29 | Soil textures suitable as loam borrow**

Soil textures that are suitable as “loam borrow” are shown in light brown on the USDA textural triangle.

material that may propagate weedy plants (e.g., blackberry canes). Specifications also call for well-composted, or stable, which can be indirectly determined by a respirometry test or calculated using C:N ratios obtained from laboratory testing of nitrogen and organic matter. A stable compost will have a low respirometry rate (<8 mg CO<sub>2</sub>-C per g organic matter per day) and low C:N (<25:1). A bioassay test that compares the rate of seedling emergence and seeds sown in compost to seeds sown in a control growing medium would be useful. Table 5-9 lists the laboratory tests for determining the optimum composts to use in manufactured topsoils.

**Table 5-8 | General specification ranges for loam borrow used in manufactured topsoil**

This table outlines the general specifications for loam borrow used in manufactured topsoil. It can be adapted depending on the soil characteristics of the compost and other soil amendments that will be used in the composition of the manufactured topsoil (modified after Alexander 2003b).

| Test parameters                                | Test methods   | Loam borrow                          |
|--|--|--------------------------------------|
| <b>Physical contaminants (man-made inerts)</b> | Man-made inert removal and classification (TMECC 0.308-C)                            | <1%                                  |
| <b>Trace contaminants</b>                      | Arsenic, Cadmium, Copper, Mercury, Manganese, Molybdenum, Nickel, Lead (TMECC 04.06) | Meets US EPA, 40 CFR 503 Regulations |
| <b>pH</b>                                      | 1:5 slurry pH (TMECC 04.11-A)  | 5.0-7.5                              |
| <b>Soluble salts</b>                           | Electrical conductivity using 1:5 slurry method (dS/m)                               | <5                                   |
| <b>Biosassay</b>                               | Percent seedling emergence and relative seedling vigor (TMECC 05.05-A)               | >80% of control                      |
| <b>C:N ratio</b>                               | (TMECC 05.02-A)  | <25                                  |

To ensure the delivery of high quality organic materials, compost can be obtained from a facility that participates in the Seal of Testing Assurance (STA) program (Inset 5-6). Upon request, these **compost facilities** send the latest lab results of the material of interest. If compost is from a STA composting facility is not available, other sources can be considered but testing the quality of the compost will be the responsibility of the designer. Compost testing by the designer involves sampling the compost piles and sending the samples to an STA lab (a listing can be found at the [Composting Council's website](#)).

At the same time that compost piles are tested, they can also be visually inspected to ensure that noxious weeds are not present on or near the composting facility.

The compost application rates typically range from 10 to 30 percent by volume of the loam borrow. [Section 5.2.5](#) describes how to determine specific organic matter rates.

**Soil Amendments**—Soil amendments, such as fertilizers, lime, and beneficial microorganisms, can be applied to the compost and loam borrow to bring the manufactured soil into acceptable ranges for pH, nutrients, and microbiological parameters. The type and amount of amendments to apply can be determined by conducting a lab analysis on a sample of the manufactured topsoil (e.g. the combination of loam borrow and compost at the specified mixing ratio). From the results of the soil analysis, a determination can be made for the application rates for fertilizers ([Section 5.2.1](#)), lime amendments ([Section 5.2.6](#)), and beneficial soil microorganisms ([Section 5.2.7](#)).

**Table 5-9 | General specification ranges for composted materials for composted materials in manufactured topsoil**

These generalized specifications for composts to use in manufactured topsoil can be adapted depending on the soil characteristics of the loam borrow and site conditions

Modified after Alexander 2003b

| Test parameters                                | Test methods  | Loam borrow   |
|--|---|---|
| <b>Physical contaminants (man-made inerts)</b> | Man-made inert removal and classification (TMECC 0.308-C)   | <1%   |
| <b>Trace contaminants</b>                      | Arsenic, Cadmium, Copper, Mercury, Manganese, Molybdenum, Nickel, Lead (TMECC 04.06)                | Meets US EPA, 40 CFR 503 Regulations  |
| <b>pH</b>                                      | 1:5 slurry pH (TMECC 04.11-A)   | 5.0-8.5   |
| <b>Soluble salts</b>                           | Electrical conductivity using 1:5 slurry method (dS/m)  | <5  |
| <b>Biossay</b>                                 | Percent seedling emergence and relative seedling vigor (TMECC 05.05-A)                              | >80% of control   |
| <b>% Moisture content</b>                      | Percent wet weight (TMECC 03.09-A)  | 30-60   |
| <b>Total organic matter</b>                    | Percent by dry weight, loss on ignition (TMECC 05.07-A)   | 25 to 60%   |
| <b>Stability</b>                               | Respirometry. Carbon dioxide evolution rate — mg CO <sub>2</sub> -C per g OM per day (TMECC 05.088) | <8  |
| <b>C:N ratio</b>                               | (TMECC 05.02-A)   | <25   |
| <b>Particle size</b>                           | Percent of compost by dry weight passing a selected mesh size, dry weight (TMECC 02.12- B)          | <ul style="list-style-type: none"> <li>• 3" (75mm): 100%</li> <li>• 1" (25MM): 90-100%</li> <li>• 3/4" (19mm): 65-100%</li> <li>• 1/4" (6.4mm): 0-75%</li> <li>• Maximum particle length of 6" (152mm)</li> </ul> |

Large quantities of manufactured topsoil can be mixed in a staging area. Using the bucket of a front-end loader or excavator, the compost and loam borrow can be measured out proportionately. For example, using a 5 yd<sup>3</sup> bucket, manufactured topsoil with a ratio of 25 percent compost and 75 percent loam borrow would have one scoop of compost applied to three scoops of loam borrow to produce 20 yd<sup>3</sup> of material. Additional amendments, such as fertilizers or lime materials, would be applied based on calculations for a 20 yd<sup>3</sup> pile. Once all the materials have been placed in the pile, it is thoroughly mixed using the front-end loader.

Purchasing loam borrow, compost, and topsoil will likely involve a set of contract specifications that ensure product quality. The specifications in Table 5-8 and Table 5-9 were developed for the U.S. Department of Transportation by the Composting Council Research and Education Foundation (Alexander 1993b). The tests are based on the Test Method for the Examination of Composting and Compost (TMECC) protocols. These are general quality guidelines and can be broadened or made more constraining depending on the specifics of the project. When considering purchasing these products, it is important to request the latest lab analysis. An STA facility (Inset 5-6) will have these reports available while others might not. It is important that these tests be run by STA laboratories

and that the sources be visited to determine whether there are undesirable or noxious weeds on or near the piles.

## 5.2.5 ORGANIC MATTER AMENDMENTS

### Background

High-quality topsoil is not always available in the quantities needed to meet the objectives of a revegetation project. In such cases, infertile subsoils can be augmented by the incorporation of organic matter amendments. This practice can be an important tool to begin the process of rebuilding a soil and reestablishing native vegetation.

One immediate effect of incorporating organic matter into infertile subsoils is increased infiltration. Water that would typically run off the soil surface during rainstorms now enters the soil. Amended subsoils also have greater permeability and often increased water storage. Changes in these factors can improve the overall hydrology of the site, making soil less susceptible to runoff and erosion.

Incorporation of organic matter can often improve plant establishment and growth rates, especially if composted organic matter is used. Composted organic materials increase soil nutrients and rooting depth, which can create better growing conditions for native plant establishment. The use of non-composted organic materials, such as shredded wood, can restrict plant growth for the first several years after incorporation because of the immobilization of nitrogen. Nevertheless, non-composted materials may be important to consider when composted sources are not available or too expensive, and there is readily available sources of material from clearing and grubbing of trees and shrubs. This material will have an immediate improved effect on infiltration and permeability, as well as speeding the long-term rehabilitation of the soil.

Incorporated organic matter becomes the primary source of energy for soil organisms and, whether fresh or composted, is the driving force behind soil development. In the process of decomposition, soil organisms turn cellulose into complex organic compounds while slowly releasing nutrients for plant growth. Some of the resulting compounds act similarly to glues, sticking soil particles together into aggregates, which ultimately create soil structure. The slow decomposition of organic matter delivers a steady supply of nutrients to the establishing plant community for many years.

The strategy behind many current revegetation projects is to obtain immediate cover with the use of seeds, fertilizers, and other amendments without considering what is needed for long-term site recovery. It is not uncommon to find good establishment of vegetative cover immediately (within a year) after revegetation work, but several years later find that it was not sustainable. Life expectancy of many revegetation projects has often been found to be very short (Claassen and Hogan 1998).

Incorporating organic matter takes a different approach. This strategy puts more emphasis on the development of the soils and less on the quick establishment of vegetation. It is based on the premise that creating healthy, functioning soils is the first step in reestablishing native vegetation. This cannot happen without basic minimum soil components, such as an organic source, nutrients, and good soil porosity. When these components are in place, a site can develop into a sustainable plant community. Incorporating organic matter into the soils of highly disturbed sites is an important component of meeting long-range revegetation objectives.

## Set Objectives

Section 5.2.3 described the use of organic matter as a mulch for covering seeds. When used as mulch, organic matter protects the soil surface from erosion, enhances seed germination, and with time, breaks down and improves surface soil properties. Applying organic matter on the soil surface may be far easier and more practical than incorporating it into the soil, however, the incorporation of organic matter into the soil offers the following additional advantages:

- Improves soils of “difficult” subsoils or parent materials
- Increases water-holding capacity
- Improves rooting depth
- Improves infiltration and drainage
- Encourages quicker release and availability of nutrients and carbon

Setting objectives clarifies why this mitigating measure is being considered and helps define the appropriate sources and application

### **Inset 5-6 / The Seal of Testing Assurance Program**

Just as many products at the local market have seals of approval (e.g., “Approved by the FDA” or “USDA Inspected”), the United States Composting Council operates an approval system for composting facilities. The Seal of Testing Assurance (STA) is a voluntary program that requires compost manufacturers to regularly test their composts using an approved third-party testing facility. The procedures for sampling and testing are outlined in the Test Methods for the Examination of Composting and Compost (TMECC) protocols. The STA program ensures that the company is reputable. Compost can be purchased from companies that do not participate in the STA program but this leaves the sampling and testing up to the buyer. This may involve visiting the composting site, collecting samples from the compost piles, sending them to a qualified lab to run TMECC tests, and interpreting the results when they are returned. STA laboratories can be found at [www.compostingcouncil.org](http://www.compostingcouncil.org). (after Alexander 2003).

rates for incorporated organic matter.

**Improving “Difficult” Parent Material**—The parent material from which soils are derived often plays an important role in how soils will respond to disturbances. Soils originating from granitic rock respond poorly to the removal of topsoil. Subsoils from this parent material have very high bulk densities and low permeability rates. They can “hardset” when dry, restricting root growth and increasing runoff (Claassen and Zasoski 1998). Any positive effects of deep tillage are often short-lived on granitic subsoils (Luce 1997) because they quickly return to high bulk densities and soil strengths. Granitic soils can benefit from the incorporation of organic matter, not only because of increased nutrients, but because the soil’s physical properties are improved. Organic amendments lower bulk densities and help to form pathways for water entry, soil drainage, and root growth. Organic matter can also increase the water-holding capacity of these soils.

Soils derived from serpentine rock have also been identified as difficult to revegetate because of heavy metals, low water-holding capacity, low nutrient levels, and a low calcium-to-magnesium ratio. When disturbed, these sites can take decades to revegetate, producing a continual output of sediments. In addition to using plant materials collected from similar serpentines sites, incorporating compost into serpentine soils can greatly improve revegetation success by increasing water-holding capacity (Curtis and Claassen 2005).

**Increasing Water-Holding Capacity**—The incorporation of organic matter can increase the water-holding capacity of soils with less than 9 percent available water capacity (Claassen 2006). These include soils with sand, loamy sand, and some sandy loam textures, as well as soils high in rock fragments. Incorporating organic matter to increase water-holding capacity can be critical on arid or semi-arid sites, or sites with very little summer rainfall. The increase in moisture-holding capacity depends on the type of organic matter and the degree of decom-position. Non-decomposed (fresh) organic sources, such as large wood chips, can decrease soil water-holding capacity because they act similarly to gravels and hold very little water. A test for determining how much soil moisture will be increased or decreased by the incorporation of organic matter is described in the [Native Revegetation Resource Library](#). The results from this test will help determine the source and quantity of organic matter to incorporate. If there is no increase in percent soil moisture with the additions of organic matter over the soil with no organic matter additions, it means that in the short term, applying organic matter will not improve water-holding capacity. For a more accurate assessment, contact analytical laboratories that perform soil moisture tests.

**Improving Rooting Depth**—For species that have deep rooting requirements, such as trees, shrubs, and some wildflowers, incorporating organic matter into the subsoil can increase rooting depth. This can be beneficial for plant establishment and long-term site recovery, especially when applied to soils derived from “difficult” parent materials. Soils that are compacted can also benefit from deep incorporation of organic matter. The incorporation of organic matter to deeper levels is often accomplished by applying a thick layer of organic matter to the surface and then incorporating it to the desired depths with an excavator or methods discussed in [Section 5.2.2](#). It is important that the organic matter is thoroughly mixed through the soil profile during incorporation.

**Improving Infiltration and Drainage**—The hydrology of most disturbed soils is improved with organic matter incorporation. The degree to which infiltration and permeability are improved depends on the size and shape of the organic source and application rates (as described in the following sections). Finer-textured soils (e.g., clays, clay loams, silty clay loams, sandy clay loams) will have better infiltration and permeability when organic matter is uniformly incorporated into the surface of the soil. This objective is important in areas where water quality issues are high.

**Improving Nutrients and Carbon**—Incorporating organic matter into the soil can ensure that

nutrients and carbon are available to decomposing soil microorganisms, which are essential for rebuilding a healthy soil. This objective is important for highly disturbed soils low in nutrients and carbon (e.g., sites lacking topsoil).

## Select Organic Materials

There is a range of organic material sources that are available. Developing a selection criteria based on the following characteristics can be helpful:

- Source of organic matter
- Level of decomposition
- C:N ratio
- Size and shape of material

Recognizing how these characteristics will help achieve the objectives for incorporating organic matter will guide the designer in making the appropriate decision for each project.

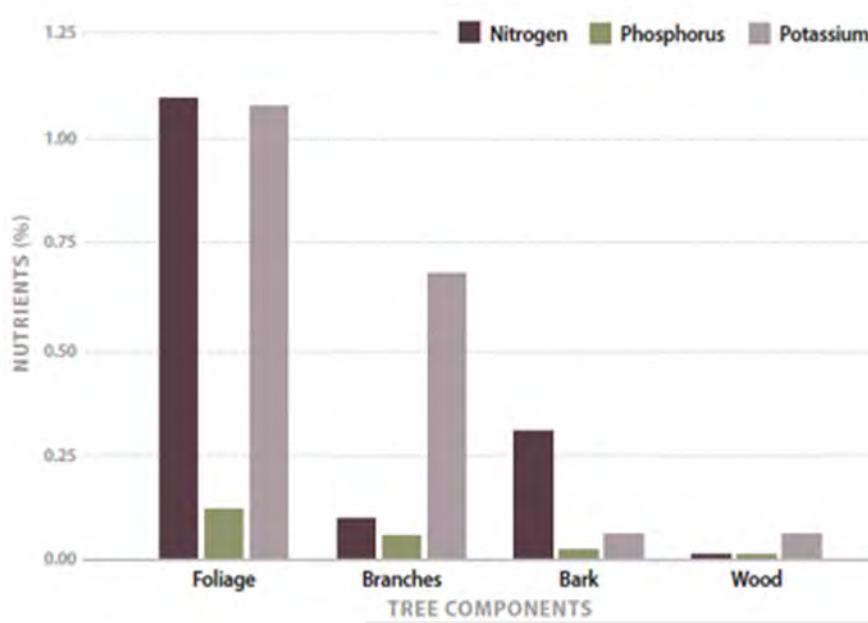
**Organic Sources**—Most available organic sources originate from waste byproducts of agriculture, forestry, and landscape maintenance. They include yard & household waste (lawn clippings, leaves, food waste), wood residues (sawdust, bark, branches, needles, roots, and boles of trees and shrubs from landscape maintenance, land clearing, logging operations, or mills), manures (poultry or cattle), agricultural waste products (fruits and vegetables), and biosolids (treated sewage sludge meeting U.S. Environmental Protection Agency regulations).

In the composting process, usually more than one source is used. A compost from one facility could include lawn clippings, leaves, yard waste, poultry manure, and shredded wood, while another facility could use yard waste, shredded wood, and biosolids.

Each organic source has a unique nutrient composition. Green alfalfa, for example, has a very different nutrient makeup than organic matter derived from wood residues of cleared right-of-way. Even within sources, there are very different nutrient compositions (Figure 5-30). For example, most of the nutrients of trees are concentrated in the foliage and branches, and very little in the bole of the tree. A chipped slash pile composed of higher proportions of branches and foliage will contain a greater level of nutrients than a pile composed primarily of tree boles and, as such, would probably be a more preferred organic source.

As organic sources decompose, they contribute their nutrients to the fertility of the soil. Knowing the source from which the organic matter was derived will give some indication of the amount of nutrients that might be supplied to the soil. This information is important to determine whether target long-term nutrient levels will be achieved.

Knowing the source of the organic matter might also identify contaminants that could be harmful to plant growth. Additions of materials, such as fly ash, feedlot, and municipal and factory waste



**Figure 5-30 | Nutrients in forest biomass**

Nutrients are held in different portions of the forest biomass, as shown for old growth Pacific silver fir. On a percentage basis, tree foliage is the storehouse for nutrients. Mulch derived from branches and foliage will have a greater nutrient content than mulch from bark and wood.

Adapted from Cole and Johnson 1981

products, could decrease the quality of an organic source. Testing these materials for contaminants, pH, soluble salts, and bioassay will identify potential problems.

**Level of Decomposition**—Organic matter can be in various stages of decomposition, from fresh organic matter with minimal decomposition to compost that has undergone extensive decomposition. The level of decomposition is an important consideration when selecting an organic amendment. Additions of relatively undecomposed organic matter to the soil can have negative short-term effects on plant establishment and growth.

**Fresh Organic Matter**—Organic matter is recently ground or chipped material that has undergone very little decomposition is considered “fresh organic matter.” These materials usually have very high C:N ratios and will immobilize soil nitrogen for months to several years after incorporation, depending on the characteristics of the organic source. Very slow establishment of plants can be expected when incorporating fresh organic matter unless a continuous source of supplemental nitrogen is applied (e.g., applications of slow release fertilizers, establishing nitrogen fixing plants). Nevertheless, applying fresh organic matter to highly disturbed soils can have a positive effect on slope hydrology and surface erosion. Fresh organic matter can increase infiltration and permeability in poorly structured soils by creating pathways for water flow.

Incorporating fresh organic matter is not generally practiced in roadside revegetation projects. However, considering the expense of purchasing and transporting composted materials to remote sites, as well as the availability and abundance of road right-of-way material that is typically burned for disposal, this is an option that may be worth considering (Inset 5-7). If shredded or chipped road right-of-way material is to be incorporated into the soil, it is best to allow it to age as long as possible in piles. Moving the piles several times to add oxygen will increase the rate of decomposition.

**Aged Organic Matter**—Some sources of organic matter have been stored in piles for long periods and are partially decomposed. They are darker in appearance than fresh sources, but the appearance of the original organic source can still be discerned (e.g., needles or leaves are still identifiable). These materials are sometimes referred to as “aged organic matter.” Because only partial decomposition has occurred, the C:N ratio is lower than with fresh organic matter. Nitrogen immobilization, however, will still occur for a significant period after incorporation. Aged organic sources have not typically undergone extensive heating, like composts, and they can contain seeds of undesirable weeds.

**Composted Organic Matter**—Compost is the result of controlled biological decomposition of organic material. During the early stages of composting, heat is generated at temperatures that are lethal to weed seeds, insects, and pathogens (Inset 5-8). Fresh, moist compost piles will usually generate heat in the first few days of composting, reaching 140 to 160 degrees F, which will kill most pathogens and weed seeds (Epstein 1997; Daugovish et al 2006). The resulting material is a relatively stable, sanitized product that is dark brown to black and beneficial to plant growth (Alexander 2003a, 2003b). Composts are very suitable materials for increasing the water-holding capacity of sandy soils, increasing nutrient supply, and enhancing soil infiltration and permeability rates.

**Biochar and Black Carbon**—Biochar is a carbon-based material, produced from biomass (e.g. wood chips and plant residues) in a low oxygen, high temperature environment in a process called pyrolysis. It is a type of “black carbon”—a continuum of charred residues that includes char, charcoal, bone char, carbon ash, carbon black, black carbon, carbonized carbon, coke, and soot—produced for the specific purpose of sequestering carbon (Spokas et al 2012). Biochar is very resistant to decomposition and lasts for hundreds of years in the soil, as compared to uncharred biomass which breaks down in

### Inset 5-7 / Highway 35

The Oregon State Highway 35 road was frequently affected by high intensity storm events that eroded portions of the roadsides and added sediment into the tributaries of Hood River. A road project was implemented in 2011 and 2012 to reconstruct portions of the highway and improve the hydrology of the roadsides to withstand road surface runoff. Shredded wood was applied at an average depth of 6 in and mixed into the soil approximately 18 to 24 inches deep. The purpose of incorporating shredded wood was to create pathways for water to flow through the soil and thereby increase infiltration and soil drainage. The slopes with shredded wood responded favorably to extreme surface runoff events (Image A). Runoff from a non-paved road moved quickly into the soil depositing road sediments over the surface of the fill slope. On slopes that were not amended, runoff was not absorbed into the soil and gullies were created, moving road and gully sediments off the project site (Image B). High rates of slow release fertilizers were applied to offset nitrogen tie-up due to the high C:N ratio shredded wood.

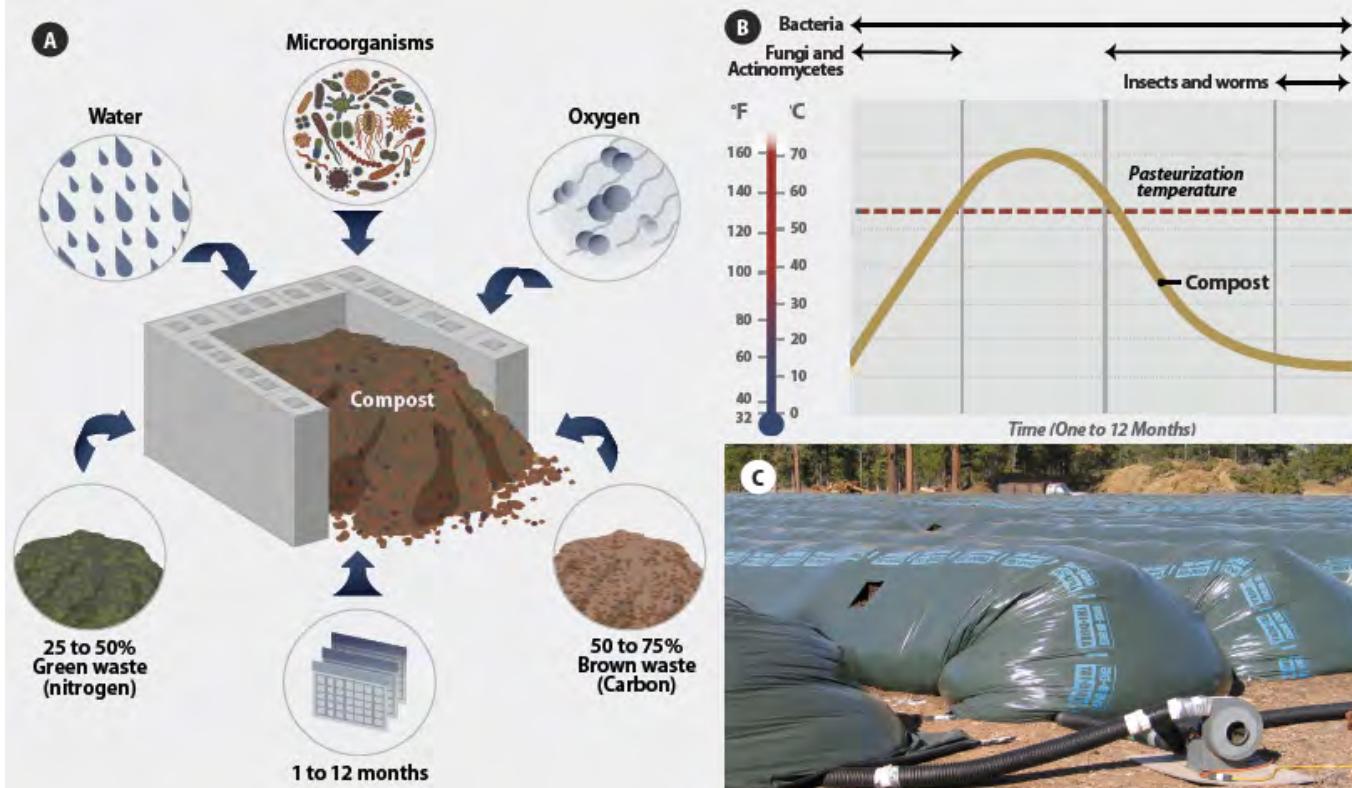
### Inset 5-8 / Compost production

The production and use of compost in the United States has flourished as a result of a ban in many of states on yard wastes in landfills. Since 1988, the number of yard waste composting facilities in the United States has expanded from less than 1,000 in 1988 to more than 3,500 in 1994. With the formation of the Composting Council in 1989, research in compost manufacturing has increased significantly.

Composting is the biological decomposition of organic matter under controlled aerobic conditions. To start the composting process, there should be organic matter, water, microorganisms, and oxygen (Image A in the illustration below). Heat is also needed but is created by the microorganisms as they proliferate. Temperatures exceed levels that kill most pathogens and weed species (Image B).

With time, in a controlled composting environment, microorganisms release carbon dioxide and water from the organic matter. The rate at which these are released, and ultimately the composting time, is a function of the type of material being composted and the composting method.

A variety of composting methods have been developed. State-of-the-art facilities and equipment that control and monitor oxygen, moisture, carbon dioxide, and temperature levels throughout the composting process produce relatively uniform products. The picture shown in Image C is of a composting system that pumps oxygen through a pipe centered in the wrapped piles of compost. Temperature and carbon dioxide are controlled through a venting system (adapted from Epstein 1997).



decades returning carbon to the atmosphere. It's high porosity and large surface area makes it very absorbent to nutrients, metals, and water and for this reason is of interest in roadside management as a soil amendment for plant growth and removing contaminants from road runoff. Biochar can be used on roadsides for (1) sequestering carbon, (2) improving soil characteristics, and (3) enhancing water quality.

Applying biochar to roadsides for carbon sequestration has been limited because of the high costs and availability of the material. Other methods of sequestering carbon on roadsides are more cost effective

such as mowing grasses less frequently, converting introduced annual grass and forb vegetation to perennial species, maintaining forested areas, using vegetation in lieu of traditional engineering solutions (e.g. living snow fences, slope stabilization), and converting open areas to shrub and forested species (FHWA 2010).

Depending on the soil type, incorporating biochar may immediately affect soil nutrition, water retention, or microbial activity which may improve soil productivity. In reviewing the literature, Spokas et al (2012) found that biomass yields increased in half of the studies where biochar or black carbon was added to the soil but either decreased or showed no significant differences in the other half. The reasons for conflicting study results are likely due to the variety of biochars used in these studies. Biochar properties range widely depending on type of material used (feedstock), temperatures (pyrolysis conditions), and how the biochar was stored. Differences in study findings may also be due to the soil type. Of the studies with positive results, a greater number of these studies were conducted on either weathered or degraded soils with limited soil fertility, similar to roadside conditions.

The large surface area and cation exchange capacity of biochars may immobilize plant nutrients, decreasing soil productivity and restricting plant growth. Combining biochar with compost, manure, fertilizer, and other amendments may mitigate these effects and be a good method for using biochar in soil remediation (Beesley et al 2011). It is important to understand the effects of applying biochar to a roadside project through trials or administrative studies. Applying biochar to roadsides for soil improvement has been done on a limited scale and research is needed to understand its value in improving these sites.

Biochar has the potential to be an effective sorbent for organic and inorganic contaminants in soil and water (Ahmad et al 2013). Used in storm water design, it holds promise in removing heavy metals and other road pollutants from road runoff, including brake lining dust, leaded gas, antifreeze, and herbicides. Mixed into the soil where road runoff concentrates (e.g. road shoulders, amended ditchlines, filter strips, bioretention swales, and constructed wetlands) biochars may capture contaminants before they enter live drainages. Mixing 4 percent biochar into a roadside filter strip, Brown (2016) greater infiltration rates, lower runoff levels, increase in soil moisture content nitrate reduction.

The origin of biochar may be critical in the type of contaminant being absorbed. Biochars produced under high temperatures have higher surface area and are more effective in capturing organic contaminants while lower temperature biochars are more suitable for removing inorganic contaminants such as metals (Ahmad et al 2013). The same absorptive characteristics that make biochars good for water quality may also capture nutrients important for plant growth. For this reason, if biochars are being used in road runoff structures, it is important to consider its impacts on plant growth. As stated above, using composts, manures, shredded wood, fertilizers, and other amendments with biochar, may increase soil productivity and plant growth. Laboratory or field testing of biochar with soil amendments prior to use on projects will identify the effectiveness of these mixes in removing road contaminants.

**Carbon-to-Nitrogen Ratio**—The C:N ratio is one of the most important characteristics to consider when selecting a source of organic matter. It is an indicator of whether nitrogen will be limiting or surplus (Section 3.8.4, see [Soil Nitrogen and Carbon](#)). The higher the C:N ratio, the greater the likelihood that nitrogen will be unavailable for plant uptake. When an organic source with a high C:N ratio is incorporated into the soil, carbon becomes available as an energy source for decomposing soil organisms. Soil microorganisms need available nitrogen to utilize the carbon source. Not only do microorganisms compete with plants for nitrogen, they store it in their cell walls, making it

unavailable for plant growth for long periods. As the carbon sources become depleted, the high populations of soil microorganisms die and nitrogen is released for plant growth (Figure 3-40).

When C:N is greater than 15:1, available nitrogen is immobilized but as ratios dip below 15:1, nitrogen becomes available for plant uptake. Most fresh and aged organic sources have C:N ratios greater than 15:1 (Table 5-10) and will immobilize nitrogen for some period of time when incorporated into the soil. When these same materials are composted, C:N ratios approach or even fall below 15:1 and provide a source of nitrogen to the soil. Co-composts, which are biosolids mixed into the compost, can have ratios between 9:1 and 11:1, indicating they are a ready source of available soil nitrogen. When materials with C:N ratios are below 10:1, they can be considered a fertilizer and labeled accordingly. The period of time that nitrogen remains immobilized in the soil is dependent on several factors:

- **Climate**—High moisture and warm temperatures are important for accelerating decomposition rates. For example, organic matter will decompose faster in Florida than in the mountains of Idaho.
- **Quantity of incorporated organic matter**—The more organic matter that is applied, the longer the immobilization. A small amount of incorporated sawdust will immobilize very little nitrogen as compared to several inches of the same material.
- **C:N ratio of organic amended soil**—The combined C:N ratio of soil and incorporated organic matter gives an indication of how the type and rate of incorporated organic matter will affect the soil C:N ratio. An amended soil with a high C:N ratio will have a longer immobilization period than a soil with a lower C:N ratio.
- **Depth of incorporation**—The depth to which high C:N materials are mixed into the soil will affect decomposition rates. For example, a layer of high C:N material near the surface of the soil will decompose slower than the same layer mixed to 12 inches deep because there would be less soil to organic matter contact.
- **Size and shape of organic matter**—The more surface area of the organic source, the faster decomposition will take place. A fine compost will decompose faster than a coarse, screened compost.
- **Nitrogen fertilization or fixation**—Nitrogen present in the soil or supplied from fertilizers or nitrogen-fixing plants will speed up decomposition rates.

It is difficult to predict how long nitrogen will be immobilized in a soil due to the incorporation of organic matter. The variety of available organic sources, unique soil types, and range of climates in the United States make this difficult. For practical purposes, it can be assumed that without supplemental additions of nitrogen (from fertilizers or nitrogen-fixing plants), the immobilization of nitrogen in soils with high C:N ratios will be in the order of months, if not years. To give some idea of decomposition rates, Claassen and Carey (2004) found that partially composted yard waste with a C:N of 18:1 took over a year for nitrogen to become available under aerobic incubation testing conditions.

The incorporation of materials with high C:N ratios may be beneficial to long-term soil aeration and water movement because the material will not break down as fast as materials with lower C:N ratios. For example, the incorporation of alfalfa hay (C:N = 13:1) will decompose quickly, and the effects on soil structure might be short-lived. Alternatively, pine needles (C:N = 110:1) or shredded Douglas-fir (1,200:1) will be effective for many years. High C:N materials are also a longer-term energy source to

**Table 5-10 | C:N ratios for common sources of organic matter**

From Rose and Boyer 1995; Epstein 1997; Claassen and Carey 2004; Claassen 2006

| Materials                            | C:N ratio          |
|--------------------------------------|--------------------|
| Wood: Ponderosa pine and Douglas-fir | 1,200:1 to 1,300:1 |
| Bark: Ponderosa pine and Douglas-fir | 400:1 to 500:1     |
| Wood: Red alder                      | 377:1              |
| Paper                                | 170:1              |
| Pine needles                         | 110:1              |
| Wheat straw                          | 80:1               |
| Bark: Red alder                      | 71:1               |
| Dry leaves                           | 60:1               |
| Dry hay                              | 40:1               |
| Leaves                               | 40:1 to 80:1       |
| Yard compost                         | 25:1 to 30:1       |
| Oat straw                            | 24:1               |
| Rotted manure                        | 20:1               |
| Alfalfa hay                          | 13:1               |
| Top soil                             | 10:1 to 12:1       |

From Rose and Boyer 1995; Epstein 1997; Claassen and Carey 2004; Claassen 2006

soil organisms that help create soil structure ([Inset 5-7](#)).

Nitrogen-based fertilizers can be applied to offset the effects of high C:N ratios on soil productivity by making nitrogen available for plant growth. [Section 5.2.1](#) describes fertilizer strategies for reducing the effects of high C:N soils.

**Material Size and Shape**—The range in sizes and shapes of organic matter plays a role in how quickly organic matter breaks down in the soil. Particles with greater surface area to volume ratios will decompose faster than particles with less surface area to volume under similar environments. Chipped wood, for example, has a low surface area to volume ratio and takes longer to break down than long strands of ground wood or fine screened sawdust, which have greater surfaces areas.

The particle size and shape of the organic source can also be important in slope hydrology by increasing infiltration and permeability rates. Long, shredded wood, for example, can create extended passageways for water movement. If applied at high enough rates, long fibers can overlap, creating continuous pores that will increase drainage. Wood chips applied at the same rates are less likely to form continuous routes for water drainage because of their shape.

Large undecomposed wood can significantly reduce soil water storage due to low water-holding capacity of the material. It is important to first test incorporating large, undecomposed woody organic matter into soils with low water-holding capacities to determine its effect ([Section 5.2.5, see Background](#)).

## Determine Application Rate

The objectives for organic matter incorporation help determine the rates for applying organic matter. Each objective described in [Section 5.2.5 \(see Set Objectives\)](#) yields a different application rate. For example, a project objective to increase permeability would likely require the addition of 6 inches of compost mixed into 24 inches of soil. This is a far greater quantity than if the objective was to increase nutrient supply, which would usually require 2 inches of compost added to the top 12 inches of soil.

Determining the rates of organic matter needed to improve nutrient status can follow the process outlined for calculating fertilizer rates in [Section 5.2.1 \(see Develop Nutrient Thresholds and Determine Deficits\)](#). If a nutrient, specifically nitrogen, is found to be deficient, the amount of organic material to apply needs to be determined. A nutrient analysis is necessary to make these determinations. Figure 5-31 provides an example of how to calculate the amount of organic matter to incorporate to meet minimum levels of nitrogen.

|   |   |                        |   |
|---|---|------------------------|---|
| A | Total nitrogen (or other nutrient of interest) in compost | 10 lbs/yd <sup>3</sup> | From laboratory report; most labs will report out nutrients in lbs/yd <sup>3</sup> of material  |
| B | Nitrogen deficit  | 769 lbs/ac             | Determine from reference sites or minimum thresholds from literature (see <a href="#">Figure 5-2</a> and <a href="#">Figure 5-3</a> ) |
| C | Minimum application rates<br>$B/A =$                      | 77 yd <sup>3</sup> /ac | Volume of compost to apply to the site to meet minimum thresholds   |
| D | Minimum application depth<br>$C/135 =$                    | 0.6 in                 | Thickness of compost to apply to the site to meet minimum thresholds  |

When organic matter is used to increase infiltration and permeability for water quality and soil erosion, a rate of 25 percent organic matter (by volume) to 75 percent soil (by volume) has been suggested by several researchers (Claassen 2006). This would likely require 4 inches of organic matter be incorporated for every 9 inches of soil ([Inset 5-7](#)). Actual field trials could be installed prior to

### Figure 5-31 | Determining application rates

The following calculations can be used to determine the amount of compost to apply to a site. They are based on laboratory test results of the compost and threshold nitrogen levels obtained from reference sites.

construction to measure the effects of soil amendments on infiltration. By incorporating several rates of organic matter on plots in disturbed reference sites near the project, the infiltration rates of each treatment could be determined using rainfall simulation equipment.

If the objective for incorporating organic matter is to increase the soil's available water-holding capacity, the rate of organic matter application might best be based on achieving a total available water-holding capacity for the desired vegetation of the project area (setting these targets is described in [Section 3.8.2](#)). [Inset 5-5](#) shows one method of determining relative water-holding capacity of amended soils.

## Ensure Product Quality

Purchasing compost involves a set of contract specifications that ensure product quality. [Table 5-10](#) is a "model specification" developed for the U.S. Department of Transportation by the Composting Council Research and Education Foundation (Alexander 1993b) for composts used as soil amendments on roadways. The tests are based on the TMECC protocols. These are general quality guidelines and can be broadened or made more constraining depending on the specifics of the project. When considering purchasing compost or any other type of organic matter from a manufacturer, it is important to request the latest lab analysis. An STA facility ([Inset 5-6](#)) will have these reports available while others might not. It is important that these tests be run by STA laboratories. It is a good practice to visit the location of the organic sources to determine whether there are undesirable or noxious weeds on or near the piles. If these species are present, consider not purchasing materials.

### 5.2.6 LIME AMENDMENTS

#### Introduction

Agricultural lime is used when soil pH of a disturbed site needs to be raised to improve plant survival and establishment ([Section 3.8.4](#)). Liming low pH soils improves plant growth by reducing aluminum toxicity, increasing phosphorus and micronutrient availability, favoring symbiotic and non-symbiotic nitrogen fixation, improving soil structure, and enhancing nitrification (Havlin et al 1999).

#### Set pH Targets

Each plant community has an optimal pH range. Plant communities dominated by conifers, for example, function well between pH 5.0 to 6.5, whereas grass-dominated plant communities in arid climates perform well between pH 6.5 and 8.0. It is important to set a realistic post-construction target pH when considering liming because large quantities of lime materials are needed for even small changes in soil pH. For example, raising the pH of a sandy loam soil from to pH 6.5 takes nearly twice the amount of lime necessary to raise it to pH 6.0. A half point pH difference in this case can result in an increase of more than 1,000 lb/ac in application rates.

As described in previous sections, it is important to understand the characteristics of the reference site topsoils and try to recreate these soil conditions after construction. On projects where topsoil is removed and not replaced, it is important to determine the difference between the pH of the reference site topsoil and the post-construction surface soil through soil testing. When the pH of the post-construction surface soil is significantly lower than the reference site topsoil, liming to raise subsoil pH to reference site topsoil levels (target levels) can be beneficial.

#### Select Liming Materials

There are several types of liming materials commercially available ([Table 5-11](#)) and selection of these

materials is typically based on costs, reactivity, effects on seed germination, and composition of the material. All liming materials will raise soil pH but not at the same level as pure limestone. To account for this, all commercially available liming materials are rated against pure limestone for neutralizing effects. The rating system is called calcium carbonate equivalent (CCE). Burnt lime ( $\text{CaO}$ ) for example, might have a CCE of 150, which means that it has a 50 percent greater neutralizing capacity than pure limestone and much less of this material needs to be applied to increase pH. A low CCE material, such as slag, might have a CCE of 60, which means that it has 40 percent less neutralizing capacity. Liming materials with high CCE, like  $\text{Ca}(\text{OH})_2$  (slaked lime, hydrated lime, or builders lime) and  $\text{CaO}$  (unslaked lime, burnt lime, or quicklime), can be caustic to germinating seeds. The particle size of the liming material determines how quickly the pH of a soil will increase. The finer the material, the faster the soil pH will increase. For example, a lime material passing a 100-mesh screen reacts faster and takes less quantity than material passing a 50-mesh screen. Finer lime materials are typically more expensive.

**Table 5-11 | Calcium carbonate equivalents**

Liming materials are rated by how well they neutralize the soil using pure limestone as the baseline of 100 percent. The rating system is called calcium carbonate equivalents (CCE). Values for some commercially available products are shown below.

*Campbell and others 1980; Havlin and others 1999*

| Material                                   | Chemical formula             | CCE     |
|--|------------------------------|---------|
| Slag                                       | $\text{CaSiO}_3$             | 60-90   |
| Agricultural limestones                    | $\text{CaCO}_3$              | 70-90   |
| Marl                                       | $\text{CaCO}_3$              | 70-90   |
| Pure limestone                             | $\text{CaCO}_3$              | 100     |
| Pure dolomite                              | $\text{CaMg}(\text{CO}_3)_2$ | 110     |
| Hydrated lime, slaked lime, builders' lime | $\text{Ca}(\text{OH})_2$     | 120-135 |
| Burned lime, unslaked lime, quicklime      | $\text{CaO}$                 | 150-175 |

## Determine Liming Rates

Determining how much liming material to apply is based on these factors:

- **Soil texture**—Soil texture plays an important role in lime requirements because the higher the clay content, the more lime that has to be added to the soil to raise the pH. A soil with a clay loam texture needs over three times more lime to raise the pH from 5.0 to 6.0 than a sandy soil. This is because finer-textured soils and organic matter have higher CEC (Inset 5-9).

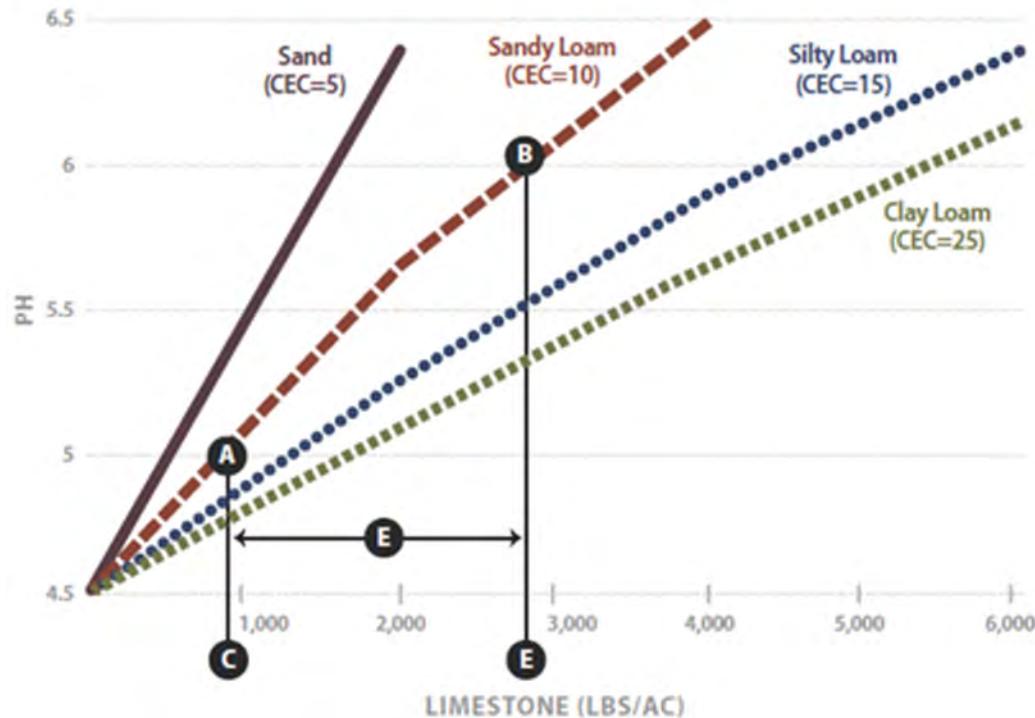
### **Inset 5-9 / Cation exchange capacity (CEC)**

The capacity of a soil to hold positive ions (referred to as bases or cations) is called the cation exchange capacity (CEC). A soil with a high CEC holds greater amount of cations, such as calcium and magnesium, than a soil with a low CEC. For this reason, a high CEC soil requires more liming material to raise it to the same pH level. Cation exchange capacity is directly related to the amount of clay and organic matter present in the soil—the higher the clay or organic matter content, the higher the CEC. Rock fragments have little or no CEC because they are massive in structure. Rates of liming on high coarse fragment soils are reduced proportionally.

- **Soil organic matter**—Soil organic matter (in humus form) has a high CEC and needs more lime material to raise pH.
- **Percentage of rock fragments**—Rock fragments have little to no CEC because they are massive and typically unweathered. Rocky soils will likely require less lime materials to raise pH.
- **Depth of liming material**—Lime materials are relatively insoluble and only change the pH of the soil around where they were placed. Liming rates are adjusted based on the soil depth to which the lime material is mixed.
- **Lime material composition**—Each liming material is rated by how well it neutralizes the soil. Less materials with high CCE ([Section 5.2.6, see Select Liming Materials](#)) are necessary as compared to low CCE materials.
- **Fineness of liming material**—The fineness of the liming material determines how quickly the pH will change. Very fine materials change pH quicker than coarse materials. Therefore, less quantity of finer-grade materials will likely be required for immediate pH soil change.

For a quick approximation of liming rates, refer to the Excel workbook titled “[Calculating Liming Rates Procedure](#)” in the [Native Revegetation Resource Library](#), which is based on the lime application rate curves shown in Figure 5-32. However, for a more accurate assessment of rates, the Shoemaker-McLean-Pratt (SMP) Buffer method is available. This test involves sampling of the post-construction surface soils be sent to a soils laboratory. Results are reported in a table that shows the quantity of lime needed to raise the soil sample to pH 7. The information can be graphed and used in a similar fashion to the example in Figure 5-32. The SMP test is well adapted for soils with pH values below 5.8 and containing less than 10 percent organic matter (McLean 1973). The future of the SMP test however, may be short-lived because of the hazardous chemicals that are used and other tests, such

as the Sikora Buffer method may be used in its place (Anderson et al 2013).



**Figure 5-32 / Approximate liming rates for disturbed soils**

This chart can be used to approximate the liming application rates for disturbed soils. The chart is based on measuring pH changes of four soil textural classes as limestone is incorporated into the surface 7 inches of soil (chart modified from Havlin and others 1999). For example, a sandy loam soil has an existing pH of 5.0 (A) and a target pH after liming of pH 6.0 (B). The amount of limestone to apply (E) is 1,900 lb/ac, which is calculated by subtracting 900 (C) from 2,800 (D). Refer to the [Calculating Liming Rates Procedure](#) workbook for a quick determination of rates and materials. More accurate lab results obtained from the SMP Buffer method for determining lime requirements can be substituted for values obtained in this graph.

## Apply Liming Materials

Limestone materials are commonly applied in powder form through fertilizer spreaders or hydroseeding equipment. Pelletized limestone, which is very finely ground material that has been processed into shot-sized particles, is easy to handle and can be used in fertilizer spreaders. Hydroseeding equipment, however, is probably the best method for spreading liming materials, especially very fine liming materials which can be difficult to apply through fertilizer spreaders.

Because liming materials are relatively insoluble in water, surface applications of lime, without some degree of soil mixing, renders the lime ineffective for immediate correction of soil acidity. Several studies have indicated that it can take more than a decade for surface-applied lime (not incorporated) to raise soil pH to a depth of 6 inches (Havlin et al 1999). It is important, therefore, to incorporate liming materials into the soil at the depth where the pH change is desired.

Incorporation can be accomplished on gentle slope gradients using tillage equipment, such as disks and harrows ([Section 5.2.2](#)). Liming materials can be mixed on steep slopes using an excavator. However, if equipment is not available for mixing on steep sites, applying very finely ground limestone through hydroseeding equipment is a possible way of raising the surface pH (Havlin et al 1999). This material raises pH faster and depending on the soil type and slope gradient, can increase the pH in the surface 3 or 4 inches of soil over time. The size specifications for very fine lime is 100 percent passing a 100-mesh sieve and 80 percent to 90 percent passing a 200-mesh sieve.

## 5.2.7 BENEFICIAL SOIL MICROORGANISMS

### Background

Beneficial microorganisms are naturally occurring bacteria, fungi, and other microbes that play a crucial role in plant productivity and health. Some types of beneficial microorganisms are called “microsymbionts” because they form a symbiotic (mutually beneficial) relationship with plants. In natural ecosystems, the root systems of successful plants have several microbial partnerships that allow them to survive and grow even in harsh conditions (Figure 5-33). Without their microsymbiont partners, plants become stunted and often die. Frequently, these failures are attributed to poor nursery stock or fertilization, when the real problem was the absence of the proper microorganism.

As described in [Section 5.2.7 \(see Sources and Application of Arbuscular Mycorrhizal Fungi\)](#) it is important to consider beneficial microorganisms as part of an overall strategy to conserve existing ecological resources on the site. These strategies include the following:

- Minimizing soil disturbance
- Conserving and reapplying topsoil and organic matter
- Leaving undisturbed islands or pockets on the project site
- Minimizing use of fast-release fertilizers

On projects where soil disturbance will be minimal or where topsoil is still present and contains functional communities of beneficial microorganisms, reintroducing the organisms will usually not be necessary. However, most road projects involve severe disturbances and, therefore, healthy populations of beneficial microorganisms may be depleted or even absent. Soil compaction and removal of topsoil, which is routine during road construction, is particularly detrimental to beneficial soil microorganisms. In addition, beneficial bacteria and fungi do not survive in soil for long periods of time in the absence of their host plants and may be killed during the topsoil storage period. Reintroducing beneficial microorganisms may be an important component in establishing and maintaining native vegetation and in restoring soils.

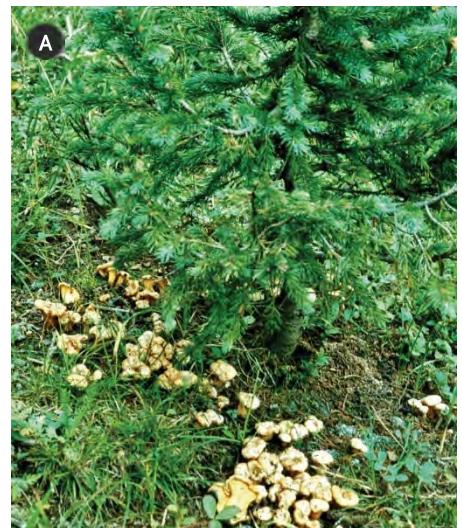
Appropriate beneficial microorganism can be reintroduced by inoculating seeds and plants with the beneficial microorganism, or introducing the microorganism in the planting hole. Applying inoculum however, does not always result in the colonization of the root system with the beneficial organism. As will be discussed, colonization depends on the quality of the inoculum and the soil environment. Plants with well colonized root systems may establish more quickly and with less water, fertilizer, and weed control, than non-colonized plants, thereby reducing installation costs.

The two most important microsymbionts for revegetation projects are mycorrhizal fungi and nitrogen-fixing bacteria.

### What are Mycorrhizae?

Mycorrhizae are one of the most fascinating symbiotic relationships in nature. “Myc” means “fungus” and “rhizae” means “root”; the word “mycorrhizae” means “fungus-roots.” The host plant roots provide a convenient substrate for the fungus and also supply food in the form of simple carbohydrates. In exchange for this free “room-and-board,” the mycorrhizal fungus offers benefits to the host plant:

**Increased Water and Nutrient Uptake**—Beneficial fungi help plants absorb mineral nutrients,



**Figure 5-33 | Symbiotic relationships of plants**

Many plants rely on symbiotic relationships to survive and grow in nature (A). The mushrooms under this spruce are the fruiting bodies of a beneficial fungus that has formed mycorrhizae on the roots (B).

*Photo credit: Thomas D. Landis*

especially phosphorus and micronutrients such as zinc and copper. Mycorrhizae increase the root surface area, and the fungal hyphae access water and nutrients beyond the roots (Figure 5-34). When plants lack mycorrhizae, they become stunted and sometimes chlorotic (yellow) in appearance (Figure 3-29 and Figure 5-34B).

**Stress and Disease Protection**—Mycorrhizal fungi protect the plant host in several ways. With some fungi, the mantle completely covers fragile root tips (Figure 5-34A) and acts as a physical barrier from drying, other pests, and toxic soil contaminants. Other fungal symbionts produce antibiotics that provide chemical protection.

**Increased Vigor and Growth**—Plants with mycorrhizal roots survive and grow better after they are planted on the project site. This effect is often difficult to demonstrate but can sometimes be seen in nurseries where soil fumigation has eliminated mycorrhizal fungi from seedbeds. After emergence, some plants become naturally inoculated by airborne spores and grow much larger and healthier than those that lack the fungal symbiont (Figure 5-34B). Mycorrhizal fungi form partnerships with most plant families, and three types of mycorrhizae are recognized:

- Ectomycorrhizal fungi (ECM) have relatively narrow host ranges and form partnerships with many temperate forest plants, especially pines, oaks, beeches, spruces, and firs.
- Arbuscular mycorrhizal fungi (AMF) are also known as endomycorrhizae or vesicular-arbuscular mycorrhizae. These fungi have wide host ranges and are found on most wild and cultivated grasses and annual crops, most tropical plants, and some temperate tree species including cedars, alders, and maples as well as flowering forbs used by pollinators.
- Ericoid mycorrhizal fungi form partnerships with the Epacridaceae, Empetraceae, and most of the Ericaceae; plants affected include blueberries, cranberries, crowberries, huckleberries, azaleas, rhododendrons, and sedges. Because these mycorrhizal associations involve unique species of fungi, few commercial inoculants are available and the best option is to use soil from around healthy plants.

For restoration purposes, the important thing to remember is that different plant species have specific fungal partners. ECM fungi are generally specific to one genus, whereas AMF fungi can colonize a wide range of genera. Conserving existing topsoil and organic matter is a key practice to protecting existing populations of beneficial microorganisms. Where disturbed soils are expected, practices may be necessary to reintroduce the key symbionts. Applying mycorrhizal inoculants is one option, which is based on the target host plants and the site condition. Identifying whether the host plant is endomycorrhizal, ectomycorrhizal, ericoid, or non-mycorrhizal is important in selecting inoculum. Conifer seedlings, for example, require very specific ectomycorrhizal fungi for successful inoculation. Endomycorrhizal species, on the other hand, are broad in range and therefore a general mix of several endomycorrhizal species can be used for a broader range of plant species.

## Sources and Application of Ectomycorrhizal Fungi

Three common sources of ECM inoculants are soil, spores, or pure culture vegetative inoculum.

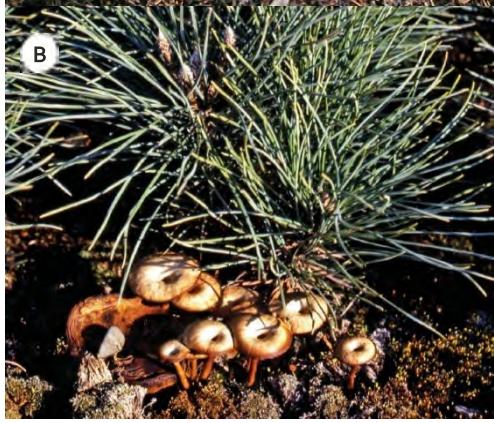
**Soil**—Topsoil, humus, or duff from beneath ECM host plants in or near the project can be used for inoculum if done properly (Figure 5-35A). Because disturbance and exposure to direct sunlight may kill these beneficial fungi, inoculation using these sources need to be done as quickly as possible. For the best results, small amounts topsoil, humus or duff are collected from several different



**Figure 5-34 | Mycorrhizal fungi benefits host plants**

Mycorrhizal fungi offer many benefits to the host plant. The fungal hyphae increase the area of absorption for water and mineral nutrients, whereas fungal mantle covers the root and protects it from desiccation and pathogens (A). Seedlings grown in nurseries soils that are low in mycorrhizae are often stunted as shown in this patchy nursery bed (B).

locations and mixed into the soil prior to planting.



**Figure 5-35 / Soil inoculum can be collected in the field**

Inoculum can be obtained by collecting topsoil or duff from areas with similar plant species (A) or from the spores of mushrooms (B), puffballs, or truffles (C) collected from around the host plant.

**Spores**—Spore suspensions are sometimes available from commercial suppliers. These spores are collected in the field from ripe fruiting bodies like mushrooms and truffles (Figures 5-35 B and C). The quality of commercial sources can be variable so it is important to verify the quality of the inoculum. It is also possible to make inoculum from spores by collecting ripe fruiting bodies of mushrooms, puffballs, or truffles from beneath healthy plants. They are then rinsed and pulverized in a blender for several minutes to make a slurry. Fungal spores do not have a long shelf life and benefit from being refrigerated and applied as soon as possible.

**Pure Culture Inoculum**—Mycorrhizal fungi are available commercially as pure cultures, usually in a peat-based carrier. Most commercial sources contain several different species of ECM. Because this type of inoculum is made from pure fungal cultures and does not store well, it is rarely available from suppliers.

Application rates and methods for ectomycorrhizal inoculums vary by species. Because these mycorrhizal fungi are very specific to their host species, it is important to work closely with company representatives when using ectomycorrhizal inoculum. Nurseries can inoculate plants with ECM and if this service is desired, it needs to be stated in the seedling-growing contract. However, as stated above, there is no guarantee that the plants that are inoculated will have colonized roots when they are planted at the project site.

Other ectomycorrhizal fungal inoculums are applied at the time of planting with the objective to get the inoculum in contact with the plant roots. Some formulations are mixed with water and the slurry is applied to the roots of nursery stock. However, the effectiveness of many of these applications has not been verified by research under roadside revegetation conditions.

**Verifying the Effectiveness of ECM Inoculation**—It is fairly easy to recognize ECM with the



**Figure 5-36 / Ectomycorrhizae fungi are visible on roots**

Ectomycorrhizae are visible on plant root systems as white or colored structures with a cottony or felt-like texture.

*Photo credit: Thomas D. Landis*

naked eye on the root system of a seedling. The short feeder roots of the seedling is covered with a cottony-white or a brightly colored mantle or sheath over the roots (Figure 5-36). Unlike pathogenic fungi, mycorrhizae will never show signs of root decay. Sometimes, mushrooms or other fruiting bodies will appear alongside their host plants which is an indicator of the species of mycorrhizae that is present. If mycorrhizae is not visible on the roots systems, plant samples can be sent to a laboratory where they analyzed for inoculation effectiveness.

## Sources and Application of Arbuscular Mycorrhizal Fungi

The two main sources of arbuscular mycorrhizal fungi inoculants include “pot culture,” (also known as “crude” inoculant), and commercially available pure cultures.

**Nursery Pot Culture**—With this option a specific AMF is acquired either commercially or from a field site as a starter culture, and then added to a sterile potting medium. A host plant such as corn, sorghum, clover, or an herbaceous native plant is grown in this substrate and as the host grows, the AMF spores multiply (Figure 5-37A). At maturity the shoots of host plants are removed and the substrate, now rich in roots, spores, and mycelium, is processed (Figure 5-37B). The resultant inoculum can be incorporated into growing media or planting beds before seeds are sown. This is a highly effective technique for propagating AMF in the nursery and could also be used on the planting site. For details, refer to *Arbuscular Mycorrhizas: Producing and Applying Arbuscular Mycorrhizal Inoculum* (Habte and Osorio 2001).



**Figure 5-37 | Arbuscular mycorrhizal fungi**

Because of their wide host range, AMF fungi can be raised on host plants like sorghum (A) producing spores and colonized roots that can be chopped-up for inoculum (B).

*Photo credits: Thomas D. Landis*

**Commercial Products**—Several brands of commercial AMF inoculants are available and usually contain a mix of several fungal species. Coarser-textured products are incorporated into soil or growing media, and finer-textured products are applied as wetable powders through sprayers or injected into irrigation systems. Inoculation effectiveness has been shown to vary considerably between products, so it is wise to install tests before purchasing large quantities of a specific product. Laboratories can provide a live spore count, which is the best measure of inoculum quality.

**Application of AMF Inoculants**—AMF inoculums typically come in a granular form with different grades of fineness. Coarse-grade products are mixed in the soil prior to sowing seed. Finer-grade inoculums, which are more expensive, may be mixed with water and applied directly onto seeds or as a

root dip. Use of fine-grade inoculum through hydroseeding equipment is another way to combine AMF with seeds as they are sown. There is little research on AMF inoculation effectiveness on roadside revegetation sites.

**Verifying the Effectiveness of AMF Inoculation**—Unlike ectomycorrhizal fungi, AMF are not visible to the unaided eye. To verify the effectiveness of AMF inoculation, roots are stained and examined under a microscope for the percent of the root system that is colonized by AMF.

## Management Considerations for Mycorrhizal Fungi

It may be helpful to work with a specialist to select the application rates and appropriate mycorrhizal partners for the plant species and outplanting sites. Some management modifications may be required to promote formation of mycorrhizal partnerships in the field. Fertilization is probably the most significant adjustment. Mycorrhizal fungi extend the plant's root system to extract nutrients and water from the soil. In some cases, fertilizer applications can be reduced by half or more due to the increased nutrient uptake by mycorrhizal fungi. Fertilizer type and form are also important. For instance, high levels of soluble fertilizers may inhibit mycorrhizae. An excessive amount of phosphorus in the fertilizer may inhibit the formation of the partnership and therefore could be reduced. If nitrogen is applied, ammonium-N is better used by the plant than nitrate-N (Landis 1989).

### **Inset 5-10 / Example of contract specifications for purchasing mycorrhizal inoculum**

**Purchase of Mycorrhizal Inoculum.** The mycorrhizal inoculum has a Statement of Claims that certifies:

- ✓ the date inoculum was produced
- ✓ mycorrhizal fungi species present in the inoculum
- ✓ number of propagules per pound of product, and
- ✓ the type and grade of carrier.

**Product Specifications.** Date of inoculum application will be within one year of production date. The storage, transportation and application temperatures of the mycorrhizae shall not exceed 90 degrees F. Inoculum must consist of at least 5 species of (choose endomycorrhizal, ectomycorrhizal or a combination of endo and ectomycorrhizal) fungi with no one species making up more than 25 percent of the propagules.

- ✓ The inoculum will contain these species: \_\_\_\_\_
- ✓ The inoculum will contain \_\_\_\_\_ live propagules per pound (Typical rates for endomycorrhizal inoculums average around 60,000 to 100,000 propagules per pound and 110,000,000 propagules per pound in ectomycorrhizal inoculums.)
- ✓ (*For applications to the soil surface only*) Live propagules must be smaller than 0.3mm.
- ✓ (*Optional*) A one ounce sample will be collected from each inoculum and sent to laboratory\_\_\_\_\_ for analysis using the \_\_\_\_\_standardized test to determine the number of propagules.

### **Application of Endomycorrhizal Inoculum to Soil Surface**

- ✓ Endomycorrhizal inoculum will be applied at a rate of \_\_\_\_\_ live propagules per acre (typical rates range from 1,000,000 to 3,600,000 live propagules per acre).
- ✓ Inoculum will be applied in the same operational period as seed application.
- ✓ If inoculum is applied through a hydroseeder, it needs to be applied within 45- minutes of being mixed in the hydroseeding tank.

### **Application of Mycorrhizal Fungi to Planting Holes**

- ✓ Mycorrhizal inoculum will be applied at a rate of \_\_\_\_\_ live propagules per seedling.

Controlled-release fertilizers are preferred because they release small doses of nutrients gradually, compared to the more rapid nutrient release from traditional products. Applications of certain herbicides, pesticides, insecticides, fungicides, and nematicides are detrimental to mycorrhizal fungi. See Inset 5-10 for an example of contract specifications for purchasing mycorrhizal inoculum.

## Nitrogen-Fixing Bacteria

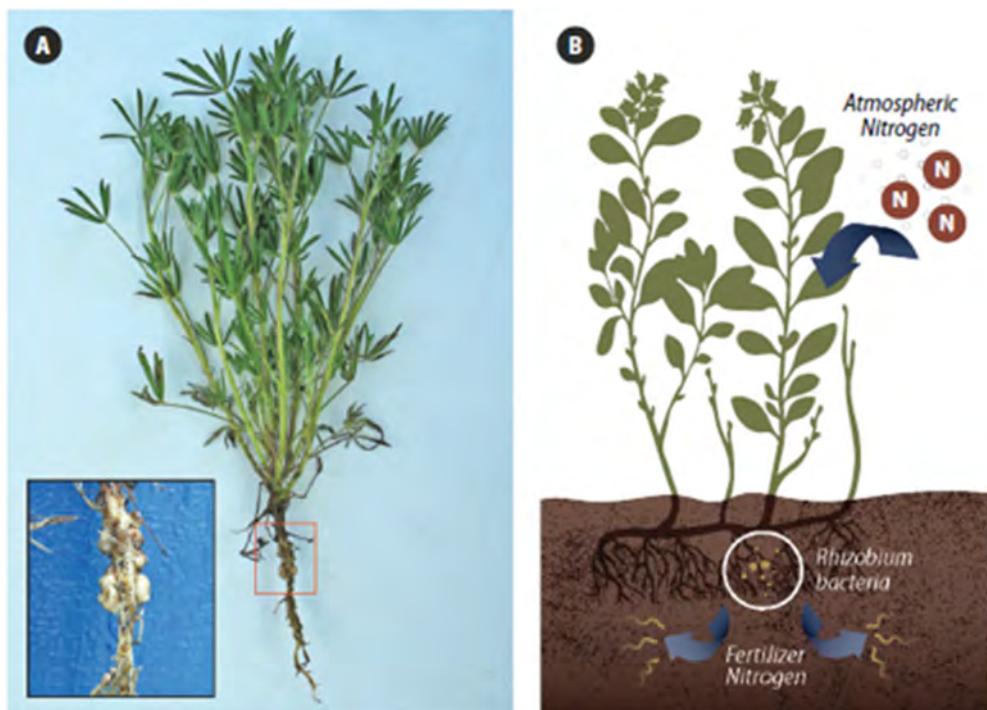
Nitrogen-fixing bacteria live in nodules on plant roots and accumulate (fix) nitrogen from the air and share it with their host plants (Inset 5-11). Unlike mycorrhizal fungi, which are found on most trees and other plants, only certain species of plants form symbiotic partnerships with nitrogen-fixing bacteria.

### **Inset 5-11 / How does biological nitrogen fixation work?**

The symbiotic partnership between plants and their nitrogen-fixing microsymbionts works this way:

Nitrogen fixing bacteria, present in the soil, are attracted to flavonoids secreted from the roots of host plants and invade the root cracks or deformed root hairs. In response, a root nodule appears and an environment for nitrogen bacteria is created. Within the low oxygen environment of each nodule, millions of bacteria convert atmospheric nitrogen to ammonia ( $\text{NH}_3$ ) through an enzyme call nitrogenase. This typically requires a relatively high amount of energy that the plant supplies to the bacteria. In return, the plant receives ammonia, which is converted to amino acids and used to synthesize proteins for plant growth. When the "nitrogen-fixing" plant sheds its leaves, or dies, the nitrogen stored in the plant's tissues is released into the soil and becomes available to other plants. This process, part of the nitrogen cycle, is the major source of nitrogen fertility in most natural ecosystems.

The role of nitrogen-fixing bacteria and their partner plants is important in revegetating roadsides. Because nitrogen-fixing plants are often pioneer species that are the first to colonize disturbed sites, they are ideal for revegetation or restoration projects. These species help restore fertility and organic matter to the project site. Nitrogen-fixing bacteria form nodules on roots of host plants (Figure 5-38A) and accumulate nitrogen from the air (Figure 5-38B). While plants that form this



**Figure 5-38 / Nitrogen-fixing bacteria**

Nitrogen-fixing plants, such as legumes, have symbiotic bacteria residing in nodules (blue inset) on their roots (A) which can chemically "fix" atmospheric nitrogen into forms that can be used by plants as fertilizer (B).

Photo credit: David Steinfeld

association are sometimes called “nitrogen-fixing plants,” the plant itself is not able to fix nitrogen by itself. It is only through the partnership with bacteria that these plants are able to obtain atmospheric nitrogen. In this partnership, the bacteria supply the plants nitrogen, and in exchange, the bacteria is given a place to grow and carbohydrates from the plant for energy. Without these bacterial partnerships, plants are not able to make direct use of atmospheric nitrogen.

Soil on restoration sites, however, may not contain the proper species of bacteria to form a symbiotic partnership with the plant. This is particularly true for compacted subsoils. Inoculating plants ensures that “nitrogen-fixing” plants form an effective partnership to fix nitrogen. Therefore, use of nitrogen-fixing plants can be an important part of accelerating rehabilitation of degraded land.

Two genera of nitrogen-fixing bacteria that are important in revegetation are *Rhizobium* and *Frankia*. *Rhizobium* grow with some members of the legume family (Figure 5-39A and B) and plants of the elm family. They form nodules on the roots and fix nitrogen for the plant. *Frankia* are a different kind of bacteria. *Frankia* partner with non-leguminous plants, such as casuarinas, alders, bitterbrush, and buffaloberry (Figure 5-39C and D) and more than 200 different plant species distributed over eight families. The species affected by *Frankia* are called “actinorhizal” plants

(Table 5-12).

**Table 5-12 | Nitrogen-fixing bacteria and their plants**

| Nitrogen-fixing bacteria | Family    | Subfamily       | % Nitrogen fixing plants | Common plant species                      |
|--------------------------|-----------|-----------------|--------------------------|---|
| <i>Rhizobium</i> spp.    | Legume    | Caesalpinoideae | 23                       | Redbud, honeylocust                       |
|                          | Legume    | Mimosoideae     | 90                       | Mesquite, acacia                          |
|                          | Legume    | Papilioideae    | 97                       | Lupine, milkvetch, black locust, clover   |
| <i>Frankia</i> spp.      | Birch     |                 |                          | Alder, birch                              |
|                          | Oleaster  |                 |                          | Silverberry, buffaloberry                 |
|                          | Myrtle    |                 |                          | Myrtle                                    |
|                          | Buckthorn |                 |                          | Cascara, snowbrush, deerbrush             |
|                          | Rose      |                 |                          | Mountain mahogany, cliffrose, bitterbrush |

## Uses for Nitrogen-Fixing Plants in Revegetation

Only a fraction of native species are nitrogen-fixing host plants. In the western United States, the most common are the lupines, vetch, bitterbrush, ceanothus, alder, and wax myrtle (Table 5-12). On nitrogen-poor sites, sowing or planting a higher proportion of these species can help a site to recover nitrogen fertility and organic matter (Figure 5-40A). The amount of nitrogen that is produced is related to the area of vegetative cover in nitrogen-fixing host plants, the productivity of the plants, and climate factors such as temperature and moisture. If percent cover of nitrogen-fixing host plants is low, then the amount of nitrogen supplied to the site will be correspondingly low (Figure 5-40B). Likewise, dry or cold conditions tend to result in slower accumulation of nitrogen. While many native and introduced nitrogen-fixing plants are attractive to pollinators, they are also attractive forage for large herbivores.

A plant survey of disturbed and undisturbed reference sites can indicate which nitrogen-fixing plants

will do well on a revegetation project. Observing the abundance of root nodules on these plants can provide some indication whether they are fixing nitrogen.

## Inoculating with Nitrogen-Fixing Bacteria

Nitrogen-fixing nursery stock with nodulated root systems generally exhibit faster early growth than seedlings than unnodulated root systems. Nursery inoculation can reduce costs in establishment and maintenance; several dollars' worth of inoculant applied in the nursery is inexpensive compared to applying nitrogen fertilizer. Faster establishment can also lead to greater herbaceous cover that can shade out unwanted vegetation. When nitrogen-fixing plants shed leaves or dies, the nitrogen stored in the plant's tissues is cycled into the soil and eventually taken up by adjacent plants. Early establishment of nitrogen-fixing plants accelerates natural nutrient cycling on disturbed sites and promotes the establishment of sustainable plant communities.

Note that in many cases, uninoculated seedlings may eventually form a partnership with some kind of Frankia or Rhizobium strain after they are outplanted. These may not be with optimal or highly productive bacterial partners, and it may take months or even years on highly disturbed sites. Until they become naturally inoculated, plants are dependent on nitrogen fertilizers and may become out-competed by weeds. Inoculating in the nursery ensures that plants form effective, productive partnerships in a timely fashion.

**Acquiring Nitrogen-Fixing Bacterial Inoculants**—Nitrogen-fixing bacteria are very specific; in other words, one inoculant cannot be used for all plants and a different inoculant strain for each nitrogen-fixing species is usually necessary. Superior strains can yield significant differences in productivity and growth rate of the host plant—in some cases over 40 percent better growth (Schmit, 2003).

Two forms of inoculant can be used: pure cultured inoculant and homemade (often called “crude”) inoculant. Pure cultured inoculant is purchased from commercial suppliers, seed banks, or sometimes, universities. Crude inoculant is made from nodules collected from roots of nitrogen-fixing plants of the same species to be inoculated. Whichever form is used, it is important to handle the inoculants with care because they are very perishable. Storing the inoculants in cool, moist conditions away from light will help maintain their viability.

Pure culture inoculants usually come in small packets of finely ground peat moss (Figure 5-41A). The inoculants are added to chlorine-free water to create a liquid slurry (allowing a bucket of tap water to stand uncovered for 24 hours is a good way to let chlorine evaporate). If a blender is available, using it to blend some inoculant in water is a good practice to ensure the bacteria will be evenly mixed in the solution. If a blender is not available, a mortar can be used. Five to ten grams (about 0.2–0.4 ounce) of manufactured inoculant can inoculate about 500 seedlings, usually exceeding the recommended 100,000 bacteria per seedling. Once seedlings begin to nodulate, nodules from their roots can serve as the basis for making crude inoculant as described below. This way, inoculant need only be purchased once for each plant species grown, and thereafter, crude inoculant can be made from nodules.

**Preparing Crude Inoculant**—Crude inoculant is made using nodules; each of which can house millions of bacteria. For Rhizobium, observing a brown, pink, or red color inside the nodule is usually a good indicator that the bacteria are actively fixing nitrogen. For Frankia, desirable nodules will be white or yellow inside. Grey or green nodules indicate that the



**Figure 5-39 | Nitrogen-fixing plants**

Nitrogen-fixing bacteria include Rhizobium that forms relationships with plants in the legume family including lupines (A) and clovers (B), and Frankia that forms relationships with other non-leguminous plants such as snowbrush ceanothus (C) and mountain-avens(D).

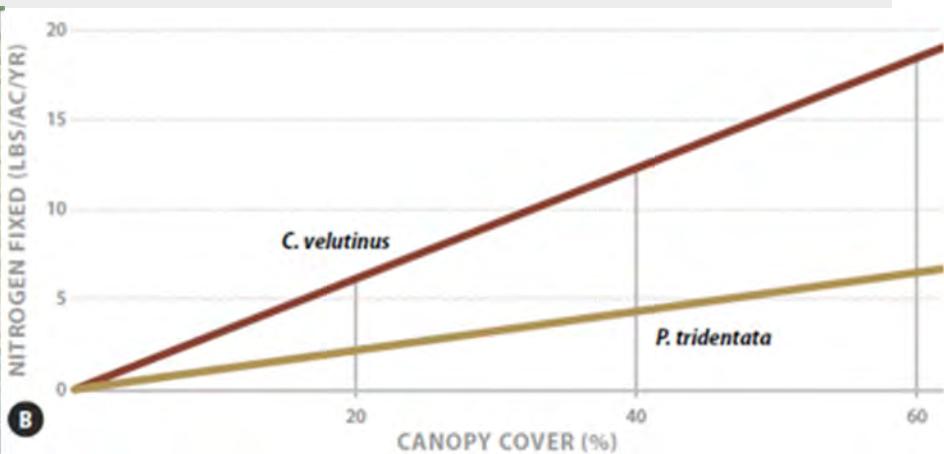
*Photo credits: Tara Luna*

structures are not viable.

#### Figure 5-40 | Amount of nitrogen is related to the cover of nitrogen-fixing plants

The accumulation of nitrogen by N-fixating bacteria is directly related to the cover of nitrogen-fixing host plants on a site. The large plants shown in this photograph are N-fixing lupines (A). The nitrogen-fixing potential of a 15-year-old stand of *Ceanothus velutinus* and *Purshia tridentata* was directly proportional to plant cover (B) (adapted after Busse 2000).

Photo credit: David Steinfeld



To make a crude inoculant, select healthy, vigorous plants of the same species as the plants to be inoculated. If available, choose seedlings that were inoculated with select bacteria. Search for nodules with the proper color and pick them off cleanly. If possible, collect nodules from several plants and place them in containers. As soon as possible after collection (within a few hours), put the nodules in a blender with clean, chlorine-free water. About 50 to 100 nodules blended in a liter of water are enough to inoculate about 500 seedlings. This solution is a homemade liquid inoculant, ready to apply in the same method as cultured inoculant as described below. If they are to be stored, place in refrigerated conditions to maintain viability.

**Applying Inoculant**—It is important to apply inoculant when seedlings are just emerging, usually within two weeks of sowing. This helps ensure successful nodulation and maximizes the benefits of using inoculants. One liter of liquefied inoculant made from either nodules or cultured inoculant as per the instructions above is diluted in water. For 500 seedlings, about 5 liters of chlorine-free water is used. This solution is then watered into the root system of each seedling using a watering can. In the field, for direct seeding applications, the slurry of commercial or crude inoculant can be added to the hydroseedertank along with the seed mix.

### Management Considerations for Nitrogen-Fixing Inoculations

**Verifying the Nitrogen-Fixing Partnership**—Allow two to six weeks for indications that seedlings have formed a symbiotic partnership with nitrogen-fixing bacteria. Signs include the following:

- Seedlings begin to grow well and are deep green despite the absence of added nitrogen fertilizer
- The root systems give off a faint but distinctive ammonia-like scent
- Nodules are usually visible on the root system after about four to six weeks (Figure 5-40A and Figure 5-42), and nodules are pink, red, or brown (for Rhizobium), or yellow



#### Figure 5-41 | Nitrogen-fixing bacteria are commercially available

Nitrogen-fixing bacteria are commercially available as pure culture inoculant, often in a carrier (A), or can be prepared by collecting nodules from roots of plants in the wild (B).

Photo credit: Tara Luna

or white (for Frankia)

**Post-Planting Care**—Several factors are of primary concern when using inoculants for nitrogen-fixing bacteria:

- **Fertilization**—The use of nitrogen-fixing bacterial inoculant likely requires some adjustments in fertilization. Excessive nitrogen fertilizer will inhibit formation of the partnership.
- **Water quality**—Excessive chlorine in water is detrimental to Rhizobium and Frankia. The water source may need to be tested and a chlorine filter used if excessive chlorine is a problem.
- **Micronutrients and soil quality**—Some nutrients are necessary to facilitate nodulation, including calcium, potassium, molybdenum, and iron. Excessively compacted soils, extremes of pH or temperature also inhibit nodulation.



**Figure 5-42 | Nitrogen-fixing bacteria will multiply as inoculated plants grow**

After successful inoculation, nitrogen-fixing bacteria will multiply on the root system as plants grow. The circle points to a visible Frankia nodule on an alnus seedling.

*Photo credit: Tara Luna*

## Other Beneficial Microorganisms

In nature, communities of bacteria, fungi, algae, protozoa, and other microorganisms in the soil make nutrients available to plants, create channels for water and air, maintain soil structure, and cycle nutrients and organic matter. A healthy population of soil microorganisms can also maintain ecological balance, preventing the onset of major problems from viruses or other pathogens that reside in the soil. The practice of protecting and reestablishing beneficial microorganisms is key for revegetation. As a science, however, the use of beneficial microorganisms is in its infancy. Although thousands of species of microorganisms have been recognized and named, the number of unknown species is estimated to be in the millions. Almost every time microbiologists examine a soil sample, they discover a previously unknown species (Margulis et al 1997). Conserving, maintaining, and creating healthy soils will help to create and sustain the natural populations of beneficial microorganisms.

### 5.2.8 TOPOGRAPHIC ENHANCEMENTS

#### Introduction

Topographic enhancements are alterations to the roadside landscape designed to improve the growing environment for plants. Topographic enhancements are important when site resources such as topsoil, organic matter, and water are limited (Section 3.10). It is often better to concentrate limited resources in key areas where resources can be most effective rather than spread them across the larger project area and dilute them to the point of having little benefit to reestablishing native vegetation. An additional benefit of topographic enhancement features is that most are designed to capture water related to road drainage. This improves water quality by reducing peak-flow water to watersheds and capturing sediments.

Topographic enhancement integrates three components into the roadside design: soil improvement, site stability, and water harvesting (Figure 5-43). Soil improvement can occur when limited topsoil and organic matter are strategically used to create growing areas with optimum rooting depth (Section 3.8.2, see Rooting Depth). Stable landforms are created that reduce surface erosion and increase slope stability (Section 3.8.5 and Section 3.8.6). Water harvesting can result when local topography is modified to capture runoff water and concentrate it in areas where it can be used by plants (Section 3.8.1, see Road Drainage) (for background on water harvesting, see Fidelibus and Bainbridge 2006). The integration of these three components will determine the

success of a topographic enhancement design.

Topographic enhancement strategies are considered during the road planning stage and the design of any features are in collaboration with the planning engineer. There are many types of topographic enhancement structures. This discussion is not exhaustive but is rather intended to introduce the designer to a variety of structures that can be installed during road construction to enhance the establishment of native plants.

## Planting Pockets and Microcatchments

When terraces are filled with growing media (topsoil or amended subsoil) and planted, they are referred to as planting pockets. Planting pockets are designed to have adequate soil depth and good water holding capacity to store intercepted water and support the establishment of planted seedlings. The surface of the planting pocket is in-sloped to capture water and sediment and the face of the pocket is protected from surface erosion using a mulch (Figure 3-15).

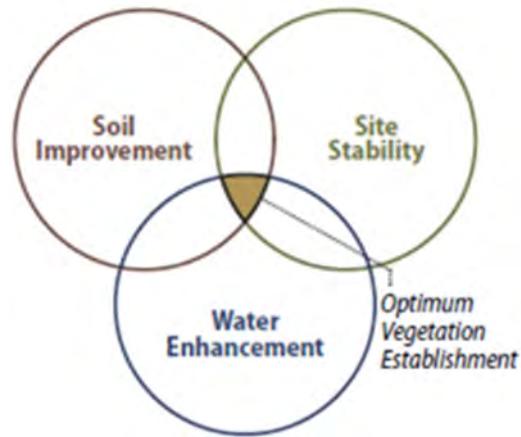
Fill slope microcatchments are structures that capture runoff from outsloped road surfaces and compacted shoulders into terraces and berms where it can be used for plant growth (Figure 3-14). Microcatchments include storage basins and berms. Berms are typically 4- to 8-inch-high obstacles placed on the contour. They are formed from soil, woody debris (logs), or manufactured products such as straw wattles or compost berms. Manufactured products and woody debris are “keyed” (partially buried) into the soil surface to prevent water from eroding under the structure. Compost berms are continuous mounds of compost that can slow water and filter sediments. Seedlings can be planted immediately above berms or obstacles to access captured water. Unless species that propagate vegetatively are used in these structures, it is important to avoid planting where sediment will bury the seedling.

## Planting Islands

Planting islands are used where resources, such as topsoil and organic amendments, are limited. They are designed into such revegetation projects as obliterated roads, view corridors, disposal sites, staging areas, and other highly disturbed sites. The strategy behind planting islands is to create areas of ideal growing environment for tree seedlings that replicates the natural topography and tree distribution observed in the surrounding landscape.



## TOPOGRAPHIC ENHANCEMENT



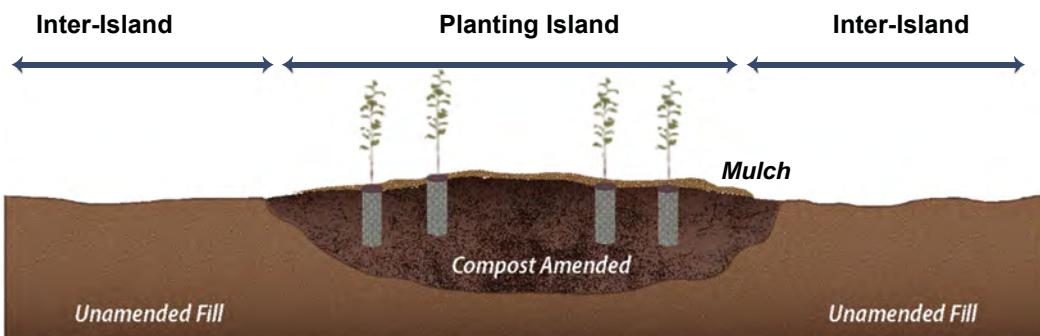
**Figure 5-43 | Topographic enhancement strategies**

Optimal topographic enhancement for establishing vegetation occurs when the roadside design includes soil improvement, site stability, and water capture treatments. It is important to consider topographic enhancement strategies early in project design.

**Figure 5-44 | Planting islands focus resources and work in concentrated areas**

Planting islands were created on this highly-compacted waste disposal site that lacked topsoil or organic matter. Spaced 15 to 20 feet apart, the planting islands were composed of 5 cu ft of compost/shredded wood mixed to a 2-ft depth using a backhoe (A). Two trees were planted at each island (B). The inter-island was seeded with a native grass and forb mix and covered with straw.

Islands can be created by excavating to a depth of several feet and backfilling with either topsoil or compost-amended material (Figure 5-44). Alternatively, compost and other soil amendments (including lime and fertilizers) can be spread over planting islands at the depth needed to amend the soil profile and mixed thoroughly through the soil with an excavator or backhoe. After planting, mulch can be applied across the surface of the entire island. Planting islands will generally occupy less than a quarter to a third of the entire site, leaving the remainder as "inter-island." The inter-islands will be much less productive than the planting islands. Grass and forb plant communities are therefore more suited to these areas (Figure 5-45).



**Figure 5-45 | Planting island cross section**

Illustration of a typical cross section of a planting island where soil depth is enhanced for tree establishment. Inter-island areas are planted to shrubs and grasses.

### Amended Ditches

The purpose of amended ditches is to treat road runoff before it reaches live channels. Amended ditches are designed for areas where there is not enough room along the roadside for vegetated swales but they differ in that they are not terraced, nor as wide. Because of their high infiltration rates, these structures retain more sediment and runoff as compared to typical ditches.



**Figure 5-46 | Amended ditches**

On the North Umpqua Highway project, amended ditches were created by mixing shredded wood and compost to a depth of 18 inches (A). The ditchlines were then formed using the bucket of the excavator (B). Surfaces were seeded, mulched, and lightly irrigated with a water truck being used for dust abatement. Within 6 weeks the ditches were stabilized with grasses and ready for the fall rainstorms (C).

*Photo credit: David Steinfeld*

Amended ditches are designed to have high infiltration rates and good water storage capacity. These soil characteristics allow road runoff water, originating from low to moderate rainstorm events, to be absorbed and stored in the soil below the ditch. As rainfall intensities increase, the amended ditches become saturated and convey water. To stabilize the channel surface from downcutting during high rainstorm events, the ditches are designed to support a continuous vegetative cover. The organic matter that is added to the ditches during construction, not only keeps the soils open and supports vegetative cover, has been shown to sequester road pollutants (Grismer et al 2011).



**Figure 5-47 | Vegetated retaining walls**

The MSE walls on the Blaine Road project required topsoil to be placed in one foot wide baskets faced with native seeded erosion mats. The 6 inch ledges between baskets were planted with native shrub seedlings (A) and within several years after planting, the walls were covered with shrubs and grasses (B).

*Photo credits: David Steinfeld*



One method of constructing an amended ditch is to place 6 inches by 3 feet layer mixture of shredded wood/compost along the ditchline and incorporating it into the soil to a depth of 18 inches with an excavator (Figure 5-46). The shredded wood/compost mixture is produced by mixing 3 parts shredded wood with 1 part compost. The shredded wood adds structure and drainage while compost adds slow release nutrients for establishing and maintaining vegetation. Compost reduces or eliminates the need for fertilizing which is important because fertilizers placed in ditchlines may enter stream courses and decrease water quality.

It is important to establish a vegetative cover on the ditches prior to when heavy precipitation periods are expected. Grass seeds, composed of species appropriate to the site, are applied to the amended ditches and covered with weed-free straw or shredded wood as soon as the ditches are constructed. If

seeding is done in the summer, in areas where rainfall is low, it may be necessary to irrigate to encourage the quick establishment of vegetation. The schedule of watering depends on the site however, a simple rule is to water when the surface of the soil, beneath the straw, just begins to dry out. Watering can be coordinated with the dust abatement operation.

## Biotechnical Engineering Structures

Topographic enhancement includes a variety of engineering structures that integrate vegetation into the design. These include vegetated retaining walls, brush layers, vegetated riprap, and live pole drains. Attention to the quality of topsoil is important in most biotechnical engineering structures. For instance, the quality of topsoil and how it is placed in vegetated retaining walls is critical for establishing vegetation (Figure 5-47). Where riprap is to be vegetated, it may require a deep layer of high quality composts cover placed over and between the riprap (Figure 5-48).

### Runoff Strips and Constructed Wetlands

Runoff strips are catchment structures constructed in areas where intermittent concentrated road drainage occurs. These are typically at the outlets of culverts or in road drainage dips. Runoff strips capture concentrated runoff into small ponds or catchment basins. These areas can be planted with riparian species, such as willows (*Salix* spp.) and cottonwoods (*Populus* spp.), or wetland species, such as rushes (*Juncus* spp.) and sedges (*Carex* spp.). Runoff strips are placed in draws or concave topography and are composed of engineered impoundment barriers, using riprap, logs, or gabion baskets, which store water from runoff events. The barriers have spillways (low points in the structure) and are keyed into the sides to ensure that concentrated water does not erode around its sides. Where runoff strips are on gentle gradients, constructed wetlands may possibly be developed (Figure 5-49).

## 5.3 OBTAINING PLANT MATERIALS

Obtaining the appropriate species, seed source, and stocktype for a revegetation project involves advanced planning and lead time. Obtaining genetically adapted materials, for example, may involve targeted collections of plant materials near or in the general geographic area of the project site several years prior to project implementation. The group of implementation guides in the following section focuses on three types of plant materials: seeds, cuttings, and plants. [Section 5.3.1](#) covers how to determine the amount of wild seed to collect, wild seed collection methods, cleaning



techniques, storage conditions, and quality testing. Methods for collecting the stems of willows and

**Figure 5-48 | Vegetated riprap**

Riprapped culvert outlets were reconstructed along the Nestucca River, an Oregon State Scenic Waterway, and required that they be revegetated with native riparian species. A compost blanket was installed over the riprap at a depth of 8 inches (A). It was planted with seedlings of a variety of riparian species shown in (B) a year later.

*Photo credits: David Steinfeld*

cottonwoods in the wild (as well as several other native species that propagate vegetatively) are described in [Section 5.3.2](#). Salvaging plants from the wild and replanting them on project sites are described in [Section 5.3.3](#).

For most projects, the collection of wild seeds, cuttings, or plants is not sufficient to meet project objectives. To increase plant materials, wild collections need to be sent to native plant nurseries for propagation. [Section 5.3.4](#) outlines the basic steps necessary to work with nurseries in establishing seed production beds for increasing seed banks of grass and forb species. Producing large quantities of cutting material of willow and cottonwood species can be accomplished by establishing stooling beds from wild collections at nurseries, covered in [Section 5.3.5](#) and [Section 5.3.6](#) describes how to work with nurseries to obtain high quality bareroot or containerized seedlings.

### 5.3.1 COLLECTING WILD SEEDS

#### Introduction

Wild seeds can be collected from native stands of grasses, forbs, shrubs, trees, and wetland plants found in or near project sites or other locations as determined via processes described in this report. The primary objective for wild seed collection is to obtain genetically appropriate, locally adapted seeds for starting nursery-grown plants ([Section 5.3.6](#)), nursery seed production ([Section 5.3.4](#)), and/or occasionally to sow directly on a disturbed site. Because seed and seedling propagation hinges on the availability of wild seeds, propagule collection is one of the first major tasks of developing a revegetation plan and can be initiated as early as possible while a project is in the planning stage. Depending on the purpose, the lead-time for collecting wild seeds could be up to three to four years before sowing or planting the project site (Figure 5-50). Seed collectors need to get approval from Project Engineer if harvesting is planned in ROW adjacent to an active roadway. Following safety practices near traffic). This lead-time is important because it requires multiple growing seasons to establish plants in the nursery or in grower's seed increase field.

Grass and forb species are often seeded directly onto disturbed sites. In order to obtain enough seeds for direct seeding, wild seed collections may be "increased" in nursery production or agricultural seed fields ([Section 5.3.4](#)) depending on the size of the revegetation project. Trees and shrub seeds, however, are not typically sown directly on disturbed sites but are instead sent to nurseries for seedling propagation, then outplanted one to three years later depending on the stocktype and seedling size specifications. Seeds from wetland genera, such as sedges (*Carex* spp.) and rushes (*Juncus* spp.), are often collected for both seed and seedling production purposes.

Revegetation plans are seldom finalized before wild seeds are collected. At a minimum, planning identifies revegetation units, describes reference areas, determines species to propagate, and involves completing a survey of the construction site to determine the amount of area to be revegetated. The quantity and location of wild seed collection are based on these early surveys.

Collecting wild seeds can be expensive. Multiple collection trips are often needed to monitor and collect populations because each species has a small ripening window, and most species do not ripen



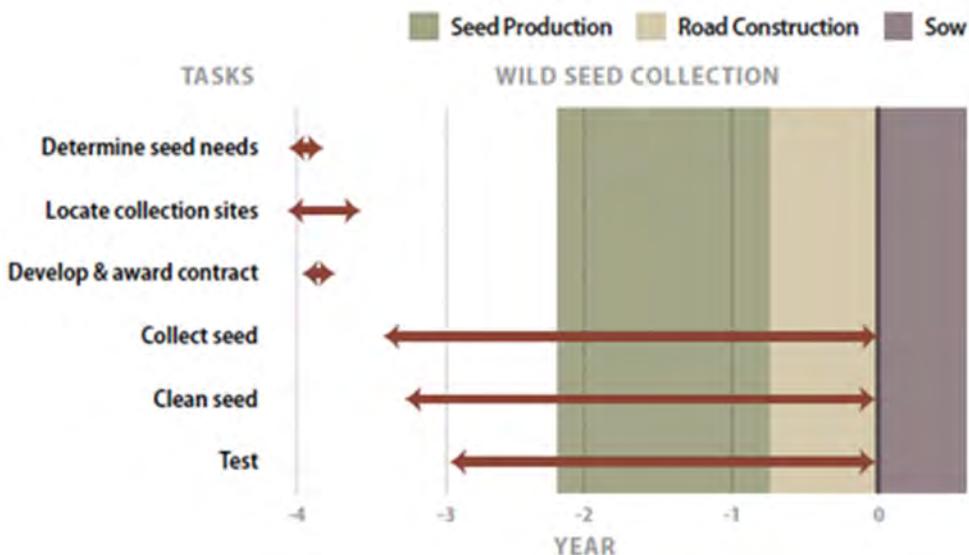
**Figure 5-49 | Constructed wetlands are effective at capturing runoff water**

Constructed wetlands capture water from roadside runoff and filter sediments before water enters perennial streams.

Constructed wetlands can create favorable habitat for unique flora and fauna.

*Photo credit: David Steinfeld*

at the same time. In addition, many species do not consistently produce seeds from year to year,



**Figure 5-50 / Early planning is essential when collecting wild seeds**

Wild seed collection is often one of the first contracts developed during planning. Seeds are needed to implement seed and seedling propagation contracts. A lead-time of three to four years is typically needed for wild seed collection. The collection time span for could be continuous up to the time of sowing depending on whether the collected seed is utilized for seed increase/ plant propagation or if the collected seed is directly sown on the project site.

requiring multiple years to generate adequate foundation collections.

Before collecting wild seeds or setting up collection contracts, it is advisable to the Natural Resources Conservation Service, determine if other genetically appropriate species seed sources are already available for the project. Often local agencies or commercial producers will have seeds in storage for many of the species growing near the project area, especially species used for reforestation.

## Develop Timeline

Wild seed collection is one of the first tasks to consider when beginning revegetation planning. Three to four years are often necessary in order to locate, collect, clean, and test wild seeds, and still allow the nursery or seed producer enough lead time for plant and seed production (Figure 5-50).

Seed collection contracts are typically awarded prior to the growing season to provide the contractor enough time to locate and assess the collection areas, source populations, and potential seed crop. Seeds are monitored periodically and collected when ripe. Wild seed harvests are cleaned, tested and certified, and stored prior to shipment to the nursery or grower for seed increase.

### Determine Wild Seed Needs for Seed Production

Wild seed collection and the nursery seed increase contracts are often developed simultaneously because the information needed for wild seed collection is based on the expected seed yields of the seed increase contract. This section describes how to calculate the amount of wild seeds to collect based on the amount of seeds expected from a commercial seed producer. The designer will most likely need to work directly with the contracted seed producer to better understand the potential seed yields of a specific crop grown in a specific location.

The amount of uncleaned wild seeds to collect for seed propagation contracts usually requires the following information (used in the calculations in Figure 5-51):

- Seed needs
- Years in seed production

- Sowing rates
- Annual seed yields
- “Cleaned-to-rough cleaned” seed percentage

The quantity of wild seeds to collect can be determined from this spreadsheet. Pearly everlasting (*Anaphalis margaritacea*) is used in this example.

|   |   |              |  |
|---|---|--------------|--|
| A | <b>Seed production needs</b>                | 22 lbs       | From seed needs plan   |
| B | <b>Years in production</b>                  | 2 years      | Seed production can span several years depending on lead time of project |
| C | <b>Sowing rates</b>                         | 1 lbs/ac     | Consult with seed producer or reference tables                           |
| D | <b>Annual seed yields</b>                   | 50 lbs/ac/yr | Consult with seed producer or reference tables                           |
| E | <b>A / B / D</b>                            | 0.22 ac      | Area seed producer needs to sow  |
| F | <b><math>E * C =</math></b>                 | 0.22 lbs     | Cleaned wild seeds that seed producer needs to sow                       |
| G | <b>Cleaned-to-rough- cleaned seed ratio</b> | 33%          | Estimated  |
| H | <b><math>(100 / G) * F =</math></b>         | 0.67 lbs     | Rough weight of seeds to collect   |

**Figure 5-51 | Determining wild seed needs**

The quantity of wild seeds to collect can be determined from this spreadsheet. Pearly everlasting (*Anaphalis margaritacea*) is used in this example. For sowing rates, annual seed yields, refer to [Table 5-15](#) and [Table 5-16](#), or discuss with the seed producer.

**Seed Needs**—The total seeds needed for each species on a revegetation project is based on the total planned revegetation acreage, seedlot characteristics (germination, purity, seeds per pound), site limitations (e.g., how well seeds will survive), and the desired seedling densities after seeds have germinated. Refer to [Section 5.3.4 \(see Determine Seed Needs for Seed Production\)](#) for methods to calculate how many seeds are needed for each species in a revegetation project.

**Years in Seed Production**—Every species has its own seed production characteristics. For example, species such as blue wildrye (*Elymus glaucus*) and California brome (*Bromus carinatus*) produce high seed quantities the first and second year, then level off or decline in years three and four. Species such as fescues (*Festuca* spp.) and Junegrass (*Koeleria* spp.) yield few seeds in the first year, but seed harvest levels increase to full production in the second or third year. For these species, consider scheduling a minimum of two years for seed production. Refer to [Section 5.3.4 \(see Determine Seed Needs for Seed Production\)](#) for discussion of first- and second-year yields for some commonly produced species.

Given proper storage conditions ([Section 5.3.4, see Seed Storage](#)), seeds of many species can be stored for several years or longer and still maintain good viability. For this reason, seed production does not have to occur all in one year. For projects that have several years lead time, maintaining production fields gives the designer more flexibility.

**Sowing Rates**—Commercial growers use a minimum amount of clean wild seeds (e.g., a foundation collection) to establish a seed increase field to produce a given quantity of seeds. While these rates differ somewhat between seed producers, general sowing rates for some commonly propagated

species in the western U.S. are shown in [Table 5-15](#) and [Table 5-16](#).



**Figure 5-52 | Wild collected seed needs to be cleaned**

Field-collected seeds include stems, chaff, flower parts, and seed attachments. Species such as cutleaf silverpuffs (*Microseris laciniata*) have a low “clean-to-rough” seed percentage and should be sent to a seed extractory for cleaning prior to sending to seed producers. The intended seeding method also strongly influences cleaning requirements.

*Photo credit: David Steinfeld*

**Annual Seed Production Yields**—The amount of seeds that are produced annually varies by species, geographic location of the fields, weather conditions, and experience of the seed producer.

Understanding what yields can be expected from seed producers will determine how many acres will be under production. Average seed yields for some species used in revegetation in the western U.S. are presented in [Table 5-15](#) and [Table 5-16](#).

**Cleaned-to-Rough-Cleaned Seed Ratio**—Foundation seed collections from the wild can include stems, chaff, and flower parts ([Figure 5-52](#)). It is a good practice for seed collectors to clean this material as much as possible before it is sent to a seed extractory for final cleaning. The amount of non-seed material can be a substantial part of the wild seed collection weight. “Cleaned-to-rough-cleaned” seed ratios ([Table 5-13](#)) can help calculate the extra weight of seeds to collect in the wild to compensate for seed cleaning. Dividing the desired amount of cleaned seeds by this ratio will yield the amount of wild seed that needs to be collected. This information can be provided by the seed extractory or specialists familiar with the cultivation practices of a certain species.

**Table 5-13 | Typical ranges of “cleaned-to-rough cleaned” seed recovery percentages**

To obtain the amounts of “rough” seeds to collect, divide the amount of cleaned seeds needed by the “cleaned-to-rough cleaned” factor. For example, if 5 pounds of cleaned seeds of prairie Junegrass (*Koeleria macrantha*) are needed, a minimum of 12.5 pounds of rough cleaned seeds should be collected ( $5 / 0.40 = 12.5$ ). The cleaned recovery percentage can vary greatly depending on seed collection method (e.g., stripping seed versus clipping seed heads which increases trash impurities).

Chart based on Pacific Northwest Region Forest Service seed collections data.

| Common name          | Scientific name                | Cleaned-to-rough cleaned factor |
|----------------------|--------------------------------|---------------------------------|
| Bluebunch Wheatgrass | <i>Pseudoroegneria spicata</i> | 0.25 to 0.33                    |
| Idaho Fescue         | <i>Festuca idahoensis</i>      | 0.33 to 0.50                    |
| Prairie Junegrass    | <i>Koeleria macrantha</i>      | 0.20 to 0.40                    |
| Squirreltail         | <i>Elymus elymoides</i>        | 0.20 to 0.25                    |
| Yarrow               | <i>Achillea spp.</i>           | 0.20 to 0.25                    |
| Sandberg Bluegrass   | <i>Poa secunda</i>             | 0.33 to 0.40                    |
| Blue Wildrye         | <i>Elymus glaucus</i>          | 0.50 to 0.65                    |

## Determine Seed Needs for Seedling Production

The quantity of wild seeds to collect for propagating seedlings at plant nurseries will be based on an estimate of the quantity of seedlings needed, the percent seed germination, the percent seed purity, seeds per pound, and the nursery factor. An estimate of germination, purity, and seeds per pound can be obtained through published sources, seed inventories, or from seed extractors managers. The nursery factor is a prediction of the percentage of viable seeds that will become “shippable” seedlings. Each nursery has developed a set of factors based on culturing experience and practices. It is good practice for nursery managers to supply nursery factors for each species or information on the amount of seeds to collect to meet the seedling order.

Nursery factors are often less than 50 percent.

Using the following equation, the amount of wild seed to collect can be estimated:

$$\frac{\text{Quantity of seedlings needed}}{[(\% \text{ of germ} / 100) * (\% \text{ purity} / 100) * (\text{seeds/pound}) * (\text{nursery factor} / 100)]}$$

## Locate Plants in the Wild

Potential collection areas are identified during the vegetation analysis phase (Section 3.6.1). General collection locations can be established by the designer with consult of a botanist or revegetation specialist familiar with the local vegetation. Contracts often require seed collectors to identify individual collection areas for approval prior to collection.

Effective wildland seed collections incorporate genetic principles and guidelines for ensuring local

adaptation and genetic diversity in the resulting seedlot. These include collecting seeds from multiple, well-distributed locations within an established seed zone or other biogeographically defined area. Genetic diversity may be enhanced by collecting seeds in approximately equal quantities from approved collection areas, and collecting from a large number of widely spaced or unrelated parent plants per area (over 50 is optimal). Plant populations growing in unusually harsh or challenging environment conditions (e.g., weedy sites) may be good candidates to consider due to natural selection pressures and genetic adaptations that may produce offspring with improved fitness and competitive ability (Strauss et al 2006; Leger 2008). Within any site, generally no more than 50 percent of the seed crop should be collected in a given year, and repeated collections in subsequent years are to be minimized or avoided to preserve viability of native plant population and dependent wildlife, including pollinators ([Section 3.13.2](#)).

Collection sites are kept free of any plants listed as noxious weeds by the State Agency or any species on the Federal Noxious Weed List because of the potential of seed contamination. Once located, the collection sites can be documented in GPS and/or marked with flagging at a point easily visible from the road used to access the site. It is expected that landowner or manager permission would be obtained when seed collection is to be conducted outside of the project area or agency-administered lands.

## Collect Seeds

Visually sound and sufficiently mature seeds are the only seeds ready for collection. Seeds are considered sound when the embryo has developed normally and there is no evidence of insect, disease, climatic, or other types of damage. Seed maturity in plants with fleshy fruits (many shrub and some tree species) often corresponds with changes in color (e.g., color changes from green to red, blue, purple, or white), taste (higher in sugars when mature), or hardness (fruit softens with maturity). Wind-dispersed seeds, which include many of the conifer species, usually change from green to brown when ripe. For grass species, a quick test for seed maturity can be determined by how seeds respond to being squeezed ([Inset 5-12](#)). A more effective method is to cut the seed and identify the development of the embryo under a hand lens or microscope. Because seed ripeness is influenced by the local weather and microclimate, determining seed ripeness often requires several monitoring trips to the field prior to collection.

### **Inset 5-12 / Stages of grass seed maturity**

For grasses, the stages of seed ripening can be determined by squeezing a seed between the thumb and forefinger. The stage of seed maturity is broadly defined by the following response:

- **Milk stage**—A milky substance is secreted, indicating an immature seed lacking viability.
- **Soft-dough stage**—Seed has a doughy texture, indicating it will have low germination and viability if collected.
- **Hard-dough stage**—No excretion of dough or milky substance when squeezed. Seeds are collected at this stage. Seeds can be collected at the transition between soft-dough and hard-dough stages. If collection occurs between these stages, seeds should not be stripped from the plant. Instead, seed heads should be cut and placed in collection bags where seeds will continue to mature.
- **Mature**—Seeds in this stage are usually too hard to bite. Collection should begin immediately because seeds can dislodge from the stem at any time.

### **For the Designer**

The seed collection methods and processes described in this section require a high level of expertise and familiarity with the target species. Designers should seek input from botanists, geneticists, and revegetation specialists as needed.



**Figure 5-53 | Seed ripens throughout the season for some plant species**

Species such as lupine (*Lupinus* spp.) have indeterminate inflorescence bloom and set seeds all summer. Seeds ripen first at the bottom of the stem and continue to ripen up the stalk as the season progresses.

*Photo credit: David Steinfeld*

Seed collection techniques are tailored to the species being collected. Grass and forb species, for example, can be hand-harvested by stripping or clipping stems just below the seed heads and placing them in collection bags or containers. Effective collection containers are made of materials that allow airflow, such as paper or fine mesh. Plastic bags or plastic containers are generally not effective. Other methods of collecting grass and forb seeds include mechanical flails and vacuums. While these methods can increase seed harvesting rates significantly, they are best done on nearly pure stands of a single species to avoid contaminating the seedlot with more than one species. Some forbs, such as lupine (*Lupinus* spp.), have indeterminate inflorescence, which means they continuously bloom, starting from the bottom of the flower head and progressing to the top (Figure 5-53). These species present a problem in seed collection because seeds ripen continuously through the growing season. Seeds from these species are often obtained by making multiple trips to the field and collecting seeds from the lower portions of the stem without disturbing the flowers or immature seeds above. Multiple collection trips may also be required if there is substantial variation in seed ripening among different type of microsites.

Seeds of many shrub species are often collected by holding a bag or tray under the plant and shaking the plant or flailing the branches with a stick or tennis racket. While the seeds of some shrub species ripen and remain on the seed head, others, such as *Ceanothus* spp., shatter when mature and are to be collected as soon as they ripen. Because multiple collection trips can be expensive, an alternative approach is to enclose the seed head of each plant in a mesh or paper bag before the seeds have begun to ripen. At the end of the season, ripened seeds will have dispersed into the bags which can be processed or transferred to other containers. It is good practice to avoid collecting seeds or seed-bearing fruits from the ground, although in some cases paper or cloth may be placed under the plant to catch the seeds (e.g., forbs and shrubs whose seeds shatter upon maturity). The seed collection contractor should specify the methods that will be used for collection.

Site location and other details of the collection can be documented on a form similar to the [NRCS data collection form](#). Key information to record includes the following:

- Species (scientific name)
- Geographic location information (GPS point) or latitude/longitude or UTM coordinates
- Date of collection
- Name of collector
- Number of populations collected
- Elevation
- Road project name

It is good practice to clearly identify each seed collection bag or container in the field using a label similar to the Forest Service identification tag shown in (Figure 5-54). These tags are generally available at Forest Service district offices or seed extractories. To ensure the identity of the seedlot in case the tag is accidentally removed during handling or shipping, it is a good idea to duplicate the tag and place it inside the collection bag. Consider grouping field collections into seedlots prior to sending these collections to the seed extractory for cleaning. Individual collections within a species are generally only maintained as separate seedlots if the objective is genetic testing or research. The expense of cleaning, packaging, and keeping records of a multitude of collections outweighs the necessity of storing them separately.

#### ***For the Designer***

State certification agencies can provide seed certification and other quality assurance services to agencies and other clients to help ensure that seed and plant products meet accepted standards. These services may be especially useful when an agency is unable to monitor all stages of plant material production from seed/cutting collection through seedling propagation and seed increase. Seed certification regulations and procedures are regulated by the [Association of Official Seed Certifying Agencies \(AOSCA\)](#).

## Clean and Test Seeds

A good practice is to clean wild seed collections to a standard that can be uniformly applied through seed sowing equipment for seedling production or seed increase. The method of application may determine the type of cleaning required, e.g., hand broadcast may need less processing than drill seeding. Seed extractories have the experience and equipment to clean wild seeds of many plant species that are frequently used in revegetation projects throughout the U.S. Seed cleaning is typically completed in two to three steps: (1) removing seeds from cones or seedpods (conifer species and some hardwood tree, shrub, and forb species), (2) detaching structures from seeds, and (3) removing all non-seed materials from collections. Removing seeds from most conifer cones involves using tumbling equipment to allow seeds to separate from scales. Some conifer species and many shrub and hardwood species require specialized equipment to break open the seedpod without damaging the seeds. Detaching seed structures involves the mechanical removal of awns (grasses) and fleshy structures, wings or other appendages. Once seed structures are detached, non-seed materials, including stems and chaff, can be removed from the collections, leaving high purity seeds (e.g., greater than 90 percent) or some user-specified level depending on the species and intended use of the particular seedlot. Seeds sown in seed increase fields with high precision equipment, for example, are generally cleaned to a higher purity level than seeds that will be hand sown or spread as mulch directly on a project site.

Seed extractories will dry, package, and store seeds, as well as test seeds on-site or send seed samples to a testing facility. Note that seed extractories cannot improve a poorly collected seedlot. Seed extractories cannot completely remove weed seeds, damaged seeds, or immature seeds from a collection, nor separate seeds from different crop species mixed in a seedlot. Removing contaminants from a seedlot becomes increasingly difficult and expensive as the size, shape, density, color characteristics of the undesirable material are more similar to the seed of the target species. Prior to collecting wild seeds, it is important to contact the seed extractory manager to discuss which species will be cleaned and for what purpose. Seed extractory managers are valuable sources of information on collection and care of a variety of native species seeds.

Cleaned seeds can be tested for germination, purity, seeds per pound, and presence of noxious weeds (Inset 5-13) by an approved seed testing laboratory (Inset 5-14). Testing involves collecting representative samples from each seedlot. Seeds are usually stored in large sealed drums or bags. If there are multiple containers per seedlot, samples from each container are to be drawn in proportion to the size of the container. Because the amount of seeds needed for testing may vary by species and laboratory, consider contacting seed testing facilities prior to submitting samples.

**For the Designer**  
See the list of Association of Official Seed Analysts (AOSA) members and their contact information.

### Forest Reproductive Material Identification Tag | U.S.D.A. Forest Service

**1) Identification Code:**

|         |        |              |            |       |   |      |      |       |
|---------|--------|--------------|------------|-------|---|------|------|-------|
| Species | Forest | BZ/Seed Zone | Type Coll. | Elev. | / | Band | Year | Cert. |
|---------|--------|--------------|------------|-------|---|------|------|-------|

**2) Number of Trees**

**7) Accession No.**

TI Collections: affix bar code tag if available

**3) Ranger District**

**8) Plant Association**

**4) Elevation**

Feet above sea level

**9) Alpha Code**

**6) Area of Collection**

T. R. S.

**10) Collection Date**

/ /

**11) Signature**

Person filling out tag

**Figure 5-54 | Recording seed collection information is imperative**

A plant material collection tag like the Forest Service example in this figure should be completed and attached to each collection bag sent to the seed extractory. A copy should also be placed inside the bag.

for special instructions. Ideally, seed sampling is conducted by a person who is trained and certified in this work, and preferably by a third party if contracts or purchasing documents are in place that contain seed quality requirements and specifications.

Seed viability usually decreases with longer durations in storage. It is good practice to conduct seed testing every few years, or at least the year before it is sown, to obtain the most accurate germination information. Copies of seed tests can be retained in contract files and on seed inventories.

### 5.3.2 COLLECTING WILD CUTTINGS

#### Introduction

Using cuttings can be a viable alternative to planting seedlings or sowing seeds to reestablish native vegetation. Vegetative material is collected from stems, roots, or other parts of donor plants and directly planted on the project site or sent to a nursery to produce rooted cuttings. The potential to produce roots from vegetative cuttings varies by species—from easy to propagate to extremely difficult. The most common species propagated from vegetative cuttings are shrubs typical of riparian ecosystems and some trees. Many deciduous species that grow well in riparian settings, such as willows (*Salix* spp.) and cottonwoods (*Populus* spp.), have a high success rate when propagated from cuttings. Shrubs with characteristics desirable to pollinators include ninebark (*Physocarpus* spp.) and twinberry honeysuckle (*Lonicera* spp.). Most temperate evergreen trees and shrubs, however, only root under very controlled environments with specialized propagation techniques.

The intent of this section is to provide the designer with a greater understanding of how to select and collect cuttings in the wild. The primary focus will be on the species in the genera *Salix* and *Populus*, because these are the most frequently used for direct planting of cuttings. With some exceptions, such as *Ribes* and *Cornus* species, most other temperate tree and shrub species can be sent to the nursery for the production of rooted cuttings before they are installed on project sites. In tropical and subtropical areas, a wider variety of species can be collected as wild cuttings. If temperate species other than willow and cottonwood are considered for propagation, nurseries can be contacted to determine the best methods for selecting, cutting, and handling the material.

Cuttings can be obtained from wild collections or from cultivated stands of donor plants, called stooling beds. Stooling beds are established at nurseries or other agricultural facilities from wild collections. In this section, the focus is on how to obtain cuttings from wild locations. The discussion of producing cuttings from stooling beds is provided in [Section 5.3.5](#).

### **Inset 5-13 / Seed tests**

Modified from Tanaka 1984

Seed testing is used to evaluate seedlot quality and provide information for determining sowing rates for seed and seedling production. Test results also provide a basis for determining seed costs when based on Pure Live Seeds (PLS), so are important for commercial seed trade, as well as for monitoring longevity and changes in germination or viability from harvest through cleaning and storage. Methods used for seed testing are based on rules of the Association of Official Seed Analysts (AOSA). A number of tests are normally conducted on each seedlot to evaluate physical and biological seed characteristics. See a [list of AOSA Labs](#) and their contact information.

#### **Physical Characteristics**

- **Purity**—Purity tests are used to determine the percentage by weight of four components: pure seeds of the desired species, seeds of other species, weed seeds, and inert matter, such as stems, chaff, scales, and small stones (Note: A purity test is based on 250 seeds and noxious seed on 2,500 seed sample). Graminoid seeds with more than 10 to 15 percent inert matter will be difficult to apply through a rotary seeder or rangeland drill. Purity tests verify the seedlot contains no “prohibited” noxious weed seeds and meets or exceeds standards for “restricted” or “other weed seeds” according to state standards for Certified Seed. Because each state has different lists of prohibited and restricted noxious weeds, it is important to request an “All-States Noxious Weed Exam.” While not prohibited or restricted by the state, some aggressive non-natives found through seed testing may still pose a threat to native plant communities.
- **Moisture content**—Seed moisture content for most species is determined by oven-drying. Seed samples are weighed and heated at 105° C (221° F) for 16 hours, then weighed again. Seed moisture is expressed as the percentage of the weight of the water lost over oven-dry weight. Electronic moisture meters are also frequently used but are not as accurate as the oven-drying method. They give rapid measurements when checking moisture in a large number of seedlots. Moisture tests are important for determining the storability of seeds. Typically, seed moistures for long-term storage are best at less than 10 percent.
- **Seeds per pound**—Seeds per pound is the weight of a given number of seeds of the desired species and does not include seeds of other species or weed seeds. The method weighs 100 seeds of 8 random samples and converts the values to number of seeds per pound.

#### **Biological Characteristics**

- **Germination**—germination test conducted in a controlled environment is the most reliable method for testing seeds. At least 400 seeds from the pure-seed component of the purity test are used in the test. Depending on the species, the seeds are usually divided into 4 replicates of 100 seeds each and chilled (stratified) for a pre-determined period and placed on trays in controlled germination chambers. At 7-day intervals, the number of seeds that have germinated (when all essential structures appear normal) are counted (AOSA 2002).
- **Tetrazolium staining**—Although controlled-environment germination tests are reliable, they are also time-consuming, particularly for those species requiring chilling. A rapid method of estimating viability is tetrazolium (TZ) staining. This test is preferred if results are needed immediately or if species to be tested have unknown chilling or germination requirements, which is often typical of many native species (Rauch 2006). The TZ test requires seeds to be immersed in 2,3,5-triphenyl tetrazolium chloride. Living cells stain red as the chemical is reduced by dehydrogenase enzymes to form a stable red triphenylformazan, which is insoluble in water. Seeds are cut and the embryos that are red-stained are counted as viable seeds. This test is useful for native species that produce seeds that are dormant and will not germinate without after-ripening (that is, seeds placed in an environment where they will continue to ripen) or without special germination enhancement treatments (stratification, scarification, gibberellic acid, etc.). In these cases, germination tests usually report out lower viability rates than actually exist. Because TZ tests measure the percentage of live embryos, they typically give a better indication of potential germination percentage. Parallel TZ and germination tests can be conducted to determine how many of the ungerminated seeds are viable or dead.
- **X-ray**—At least 400 seeds, divided into 4 replicates of 100, are X-rayed and evaluated for the presence of mature embryos, insect damage, filled seeds, damaged seeds, and other seed characteristics that could affect germination. X-raying is a fast and relatively inexpensive test, but not as accurate or informative as germination or TZ tests. It's also useful to conduct on a seed sample collected prior to harvesting to decisions on the timing of harvest initiation, or on post-harvest samples to evaluate the need for seed conditioning treatments to improve seed quality.



# 6—Monitoring

- 6.1 Introduction**
- 6.2 Developing a Monitoring Plan**
- 6.3 Plant and Soil Monitoring Procedures**
- 6.4 Pollinator Monitoring Procedures**
- 6.5 Photo Point Monitoring Procedures**
- 6.6 Developing a Monitoring Report**



## 6.1 INTRODUCTION

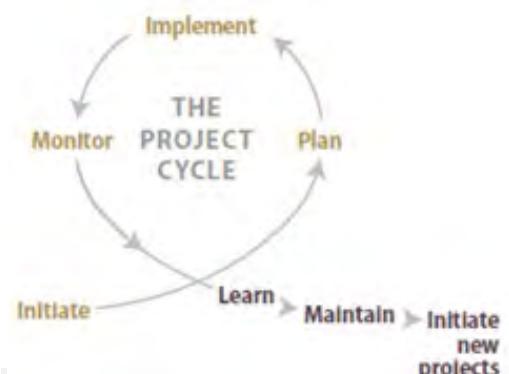
The goal of roadside revegetation is to establish plant communities along roadsides and other road-related disturbances that meet project objectives.<sup>1</sup> In practice, however, roadside revegetation projects rarely turn out exactly as planned. Therefore, regular visits to the project site to evaluate progress, and to intervene if necessary, are important parts of the revegetation process.

Monitoring is carried out for two reasons: (1) to correct, manage, and maintain the project effectively and (2) to learn what went right or wrong and apply this knowledge to future projects. Monitoring provides the answers to the following questions:

- Were revegetation goals and expectations met?
- Is native vegetation establishing appropriately or should corrective action be considered?
- Were desired future conditions (DFC) targets met?
- Are there differences in plant responses between different revegetation treatments?
- How are pollinators responding to revegetation treatments?
- Did revegetation result in a plant community capable of supporting a diverse pollinator population?
- Should the revegetation methods and techniques strategy be revised based upon monitoring data?

Answers to these questions may be obtained in a variety of ways depending on the purpose and needs of the project; however, a monitoring plan that can be applied to all revegetation projects does not exist. Instead, a monitoring plan is tailored to the objectives of each project and to the unique characteristics of the site. In developing a monitoring plan, it is important to determine the type and intensity of monitoring in advance. Simple field visits or photo point monitoring over time may provide sufficient information to determine whether objectives have been met, whereas statistically based procedures, outlined in [Section 6.3](#), may help determine if specific targets were achieved. Regardless of the intensity of the strategy, consistency and a long-term commitment are keys to successfully implement an informative monitoring strategy.

Monitoring completes the project cycle by providing feedback regarding the success or failure of a revegetation project. This information can be used to adapt and improve other revegetation projects that are in the initiation, planning, or even implementation phases (Figure 6-1). Information collected during monitoring can help determine if corrective actions, such as adjusting seed mixes or follow-up management activities (e.g., reseeding or replanting), are appropriate. Monitoring results may also be used to improve revegetation techniques for future projects.



**Figure 6-1 | Monitoring and the project cycle**

Monitoring completes the project cycle and the results are used to guide future management and maintenance work. Monitoring can also provide insights for improving future projects.

<sup>1</sup> This chapter uses abbreviated scientific names (plant symbols), from the [PLANTS Database](#) to conserve space in spreadsheets. The PLANTS Database includes scientific names, plant symbols, common names, distribution data, images, and species characteristics for vascular plants of the United States.

The monitoring phase may include the following steps:

- Determining what aspect of the project is to be monitored
- Developing a monitoring plan
- Collecting field data
- Analyzing field data
- Applying corrective measures
- Writing up findings

This chapter begins by describing monitoring plan development and moves through collecting, analyzing, and summarizing data in a monitoring report. A set of procedures are included that outline data collection for different plant, soil, and pollinator attributes.

## 6.2 DEVELOPING A MONITORING PLAN

Monitoring projects often fail because the purpose, goals, and monitoring methods are not clearly defined. The development of a monitoring plan is a process that clarifies what is to be considered prior to going into the field. Since field time is very expensive, development of a monitoring plan in the office can save valuable field time and improve the quality of the data being collected. The monitoring plan does not have to be lengthy; in fact, most can be written in a single page. The plan will be most useful if it addresses the following points (after Elzinga et al 1998):

- **Purpose**—Outlining the reason for monitoring, goals, and criteria for success
- **Intensity**—Determining the scope
- **Who**—Identifying expertise needed
- **What**—Determining parameters to monitor
- **When**—Determining frequency
- **Where**—Delineating where sampling will occur
- **How**—Selecting the monitoring procedures for data collection and analysis
- **Logistics**—Defining timeline, budget, data management, and equipment

These points become the backbone of the monitoring plan.

### 6.2.1 OUTLINING THE REASON FOR MONITORING (PURPOSE)

A common pitfall in many monitoring projects is the lack of clearly stated objectives or reasoning behind field visits and data collection. A good monitoring project is not defined by the amount of data collected, but whether the data adequately and effectively answered whether the objectives of the revegetation project were met. Successfully answering this question is only possible when the objectives for monitoring are clearly stated. Monitoring efforts not only link back to the original project objectives, but also to the specific DFCs developed in the planning phase ([Section 3.2](#) and [Section 3.7](#)). Objectives and DFCs set the targets (sometimes referred to as “performance standards,” “thresholds,” “success criteria,” or “indicators”) against which the project is evaluated (Clewel 2004).

A monitoring plan begins with a statement of purpose or reason for monitoring. This is often a restatement of the objectives and DFCs of the revegetation project. If the objective of a revegetation project is to improve pollinator habitat by increasing the cover

and diversity of native plants, then the monitoring plan would focus specifically on those elements pertaining to species diversity and plant cover and avoid collecting extraneous data. In addition, monitoring native bees and monarch butterflies would indicate how populations of insects have responded to the resulting plant composition. Time will be saved and the value of the data will be increased by streamlining data collection to include only the information needed to answer the question of whether the project met the objectives or DFCs stated in the plan.

DFCs are often stated in quantitative language. For example, the DFCs for a cut slope three years after construction could be stated in the revegetation plan as “less than 25 percent bare soil will be exposed” and “vegetative cover will be composed of greater than 70 percent native species.” These DFCs are very specific and can be used as target values, or thresholds, for determining whether the project was successful. Monitoring methods, or procedures, are developed specifically around each of these DFC targets. When monitoring is approached in this manner, only the information needed to determine whether targets were met is collected. Expensive, superfluous data collection is avoided because the DFC targets are clearly defined. The monitoring plan defines how this information will be measured and evaluated. After field data is collected, the DFC target values can be used to evaluate success or failure and whether corrective action is needed.

## 6.2.2 DETERMINING THE INTENSITY (INTENSITY)

The scope of data collection will depend on the size and importance of a revegetation project. Not all revegetation projects will involve the same intensity of monitoring to meet project goals. Some portions of the revegetation project might entail only a field review and a series of photographs, while other areas within the project will involve a statistically designed monitoring procedure. At a minimum, most projects call for annual visits and recorded observations or qualitative assessments.

Portions of revegetation projects may benefit from statistically based monitoring procedures. For example, if a project has a water quality goal for sediment control that road sections near a high-value fishery have no greater than 20 percent bare soil for sediment control the first year after construction, the importance of this project to fisheries and water quality would underscore the importance of high confidence in the accuracy of the data. This could involve more intensive data collection and statistical analysis to ensure a higher level of certainty. Alternatively, road sections that do not affect the stream system may not involve the same level of effort. In such cases, qualitative assessments, such as photo point monitoring, may suffice (Section 6.5). In general, the scope of monitoring reflects the importance of the revegetation objective, ecological sensitivity of the project area, and budgetary constraints. The levels of monitoring intensities include the following:

- **Low**—Site visits, field notes, photographs
- **Moderate**—Photo point monitoring
- **High**—Statistically based data collection and analysis

## 6.2.3 IDENTIFYING THE NEEDED EXPERTISE (WHO)

For efficiency and safety, a minimum team of two people is recommended for revegetation monitoring. The team is typically composed of the designer, or someone familiar with the revegetation project, and a person trained in plant identification. If parameters, such as soil characteristics are being monitored, then a person with soils

background would also be involved, or if pollinator surveys are being conducted, then the team would have a person knowledgeable in identifying pollinators.

It is important to have one person be responsible for all monitoring activities. This person develops and implements the monitoring plan, conducts data collection and analyses, and completes the final monitoring report. It is advantageous that the project designer or personnel who planned and implemented the revegetation project be involved with data collection and even be the person that oversees the monitoring activities. Monitoring is often delegated to others with less knowledge of the project, but a great opportunity for learning is lost when this occurs. Monitoring is the feedback loop for the designer and implementer to make improvements on the next project.

#### 6.2.4 DETERMINING MONITORING FREQUENCY (WHEN)

Specifying the timing of data collection in terms of years following project completion is important in determining if DFC targets have been met. Monitoring that occurs within a year after project completion is conducted to assess whether some areas need to be reseeded or replanted, and to determine efficacy of erosion-control devices (fabric, wattles, mulch, etc.). This low intensity monitoring is often conducted through site visits during which ocular estimates are made. Assessing the long-term success of a project is done three or more years after revegetation treatments have been completed. Some portions of a revegetation project may warrant more than one sampling visit. For example, a planting contract with a DFC of 400 live trees after the first year of planting and 300 trees alive after three years would be monitored the first and third years after planting.

Specifying the month or season when a project is monitored is also important. If the identification of individual plant species is the monitoring goal, monitoring is scheduled during the appropriate phenological window for plant identification. The bloom period is also the time when pollinators are sampled since this is when their populations are at their greatest. If outplanted stock is to be measured for survival or growth, the appropriate time to monitor is after seedlings have become established, which is typically 6 months to a year after planting. For climates with extended dry periods, common to the western United States, this monitoring is typically conducted in the fall. Alternatively, the collection of soil cover data for erosion control is done prior to intense rainstorm periods, which in New Mexico, Arizona, Colorado, Utah, and Texas is in the summer; for Gulf States, it is in the late summer and fall; and for the western United States, it is in fall and winter.

Bee populations vary seasonally and annually. For this reason, it is important for pollinator monitoring to sample a project more than once. For example, it may take several years for a newly seeded area to become established before there is an increase in pollinators. On these sites, monitoring pollinators the first year after seeding may not yield as much useful data as the second and third year, which may be the best years for monitoring. Pollinator surveys are best conducted when plants are flowering, which may warrant two or three surveys a year, especially if there is a variety of flowering species with different bloom times. Depending on the region, this can be in late spring/early summer, mid-summer, and late summer.

## 6.2.5 DELINEATING SAMPLING LOCATIONS (WHERE)

The sampling unit is the area in which a specific monitoring procedure will be used. Revegetation projects may be monitored as one sampling unit or several. A way to determine the locations of the sampling units is to revisit the revegetation unit map developed for the revegetation plan ([Section 3.4](#)). If the project is large or complex, some or all of the revegetation units may be treated as separate sampling units. Referring to the purpose for monitoring at the beginning of the monitoring plan helps identify the most important areas to monitor. For example, if the purpose for monitoring is to determine whether water quality objectives were met, only the areas adjacent to drainages and waterways would be delineated for high intensity monitoring, leaving the remainder of the project for low or moderate intensity monitoring. If the monitoring objective is to determine how well native plant species established, then setting up a sampling unit that would cover the entire revegetation project area may be appropriate. Comparing changes in plant or pollinator populations may benefit from locating and monitoring a nearby reference site.

## 6.2.6 DETERMINING PARAMETERS TO BE MONITORED (WHAT)

Clearly stating which data are essential to collect can increase the quality of the monitoring data and minimize field time. Referring to the reason for monitoring, outlined at the beginning of the monitoring plan, helps narrow the parameters for data collection. These parameters are discussed in detail in [Section 6.3](#).

## 6.2.7 SELECTING MONITORING PROCEDURES (HOW)

There are many vegetation and pollinator population monitoring methods, protocols, and procedures from which to select. Using an established set of monitoring procedures, tailored to the monitoring objectives, is important because they can save time and increase the quality of the data collected. The procedures outlined in this chapter were developed specifically for roadsides and road-related disturbances. A monitoring procedure, as defined in this report, provides information for conducting monitoring. These procedures are used for moderate and high intensity monitoring. Qualitative procedures, such as photo point monitoring, are used for moderate intensity monitoring. Statistically based procedures have been developed for high intensity monitoring.

For high intensity vegetation and pollinator monitoring, there are three sets of statistically based monitoring procedures that are used together. In the development of a monitoring plan, one procedure from each of the three procedure groups is selected for each sampling unit. These are discussed in [Section 6.3](#).

## 6.2.8 LOGISTICS

The monitoring plan also includes a timeline that shows the periods and completion dates for field monitoring, data analysis, and the monitoring report. Included in this section are the specialists involved in monitoring, their expertise, and the estimated time involved. A budget can be developed from this information. Finally, a list of equipment can be attached to the monitoring plan.

## 6.3 PLANT AND SOIL MONITORING PROCEDURES

This section describes how to develop and implement a set of statistically based monitoring procedures specific to the revegetation project. It outlines methods to record, summarize, and analyze data. Monitoring personnel can obtain spreadsheets, which include field data forms and analysis spreadsheets for each procedure, from the [Native Revegetation Resource Library](#). To obtain a list of Excel monitoring procedure workbooks referenced in this chapter, type “xls” in the Search field.

The following three questions are answered to develop a procedure for soil and plant monitoring for each sampling unit:

- What is the shape of the sampling unit?
- What are the monitoring objectives?
- What is being monitored?

The answer to these questions will lead the designer to draw from different sections of this chapter. Take, for example, a road cut that is being monitored to determine if soil cover targets conformed to the Storm Water Pollution Protection Plan. Using Figure 6-2, the Linear sampling area procedure would be used because the road cut is long and narrow. Since the objective is to determine if soil cover targets were met, the Compliance and the Soil Cover procedures would be used.

### Sampling Unit Shape

The shape of the sampling area is important for determining how transects and quadrats are laid out in a sampling unit. For roadside monitoring, there are three main shape categories to choose from, each with a corresponding procedure:

- **Linear**—This shape is long and narrow and is used to monitor cut slopes, fill slopes, and abandoned roads ([Section 6.3.6](#))
- **Rectilinear**—These sampling areas are typical of staging areas, material stock piles, or large wide areas associated with road construction ([Section 6.3.6](#))
- **Dispersed**—A series of discrete areas that include planting islands and planting pockets ([Section 3.8.2](#), [Section 5.2.8](#), [Section 6.3.6](#))

### Parameters to be Monitored

Parameters for monitoring revegetation projects may include the following, depending on monitoring objectives:

- **Soil cover**—The amount of exposed bare ground is used to determine if treatments produced adequate soil cover for erosion control ([Section 6.3.1](#))
- **Species cover**—Used to determine the percentage of aerial or basal cover occupied by individuals or groups of species to quantify species prominence ([Section 6.3.2](#))
- **Species presence**—Used to assess what species are on the site, how well species in a seed mix became established, or whether noxious, invasive, or undesirable species are present ([Section 6.3.3](#))
- **Plant density**—Used to assess how many seedlings or cuttings became established after outplanting or how much mortality has occurred ([Section 6.3.4](#))
- **Plant attributes**—Used to determine growth rates of outplanted seedlings or cuttings ([Section 6.3.5](#))

| What is the shape of the sampling area? | Section to read:        |
|---|-------------------------|
| Linear                                  | <a href="#">6.3.6.1</a> |
| Rectilinear                             | <a href="#">6.3.6.2</a> |
| Dispersed                               | <a href="#">6.7.6.3</a> |

| What is being monitored? |                       |
|--------------------------|-----------------------|
| Soil Cover               | <a href="#">6.3.1</a> |
| Vegetative Cover         | <a href="#">6.3.2</a> |
| Species Presence         | <a href="#">6.3.3</a> |
| Plant Density            | <a href="#">6.3.4</a> |
| Plant Attributes         | <a href="#">6.3.5</a> |

| What is the objective for monitoring? |                         |
|---------------------------------------|-------------------------|
| Compliance                            | <a href="#">6.3.7.1</a> |
| Treatment Differences                 | <a href="#">6.3.7.2</a> |
| Trends                                | <a href="#">6.3.7.3</a> |

### Figure 6-2 / Quick guide to high intensity monitoring procedures

For statistically based monitoring of plant and soil attributes, select a procedure that best answers each of these questions.

- **Pollinator abundance**—Used to assess quantities of honey bees, native bees, and other pollinators (Section 6.4)

## Monitoring Objectives

Monitoring objectives also guide how data are collected and statistically analyzed.

Procedures have been developed for three basic types of monitoring objectives (Section 6.3.7):

- **Compliance**—This is often the main purpose for monitoring. It is used to determine if project objectives were met or simply to summarize monitoring data
- **Treatment differences**—This monitoring objective examines the differences between revegetation treatments or revegetation areas. It is useful to evaluate the effects of different revegetation treatments (seed mixes, fertilizers, soil amendments etc.) on plant establishment
- **Trends**—This objective evaluates changes in the revegetated plant community in the disturbed area over time. It is used when it is important to understand how plant communities evolve and change

### 6.3.1 SOIL COVER

Reestablishing a soil cover on disturbed sites is important for erosion control. The following soil cover procedures can be used to determine the amount and type of cover existing on the soil surface after slopes have been constructed. The quadrat is the unit of area monitored for soil cover. Exact measurements are based on recording the surface soil condition at data points within each quadrat. Quadrats can be read in the field using a fixed frame or read later in the office from digital photographs of the quadrat taken in the field (Section 6.3.1).

#### Sampling Soil Cover Using a Fixed Frame

Soil cover is quantified at each quadrat based on readings from a 20-point fixed frame. This method is a point intercept method of estimating cover in a defined area (quadrat). Several types of frames have been developed. The most useful frames are lightweight, portable, and stable. One frame that meets these criteria uses a laser pointer to identify data points. The frame shown in Figure 6-3 has 20 slots to position a laser pointer. During monitoring, the laser pointer is moved to each of the 20 slots. Soil cover attributes are recorded where the laser hits the ground surface.

The fixed frame is located along the tape in a consistent manner throughout the sampling



**Figure 6-3 / Fixed frame for transect sampling**

A fixed frame can be adapted to allow for the positioning of a laser pointer at 20 points in the frame. Soil cover or plant cover attributes are recorded at each laser point. The frame in this example is 8 inches by 20 inches, with 2-inch spacing within rows and 4-inch spacing between rows. The laser is a Class IIIa red laser diode module that produces a 1-mm dot.

*Photo credit: David Steinfeld*

of a unit (Figure 6-4). For example, the frame might be placed on the right side of the tape with the lower corner of the frame at the predetermined distance on the tape. This procedure would be applied consistently across the entire sampling unit. The legs of the frame are positioned so that the surface of the frame is on the same plane as the ground surface.

Each data point is characterized from a set of predetermined label descriptors that include, but are not limited to, the following:

- Soil
- Gravel (2 mm to 3 inches)
- Rock (>3 inches)
- Applied mulch
- Live vegetation (grasses, forbs, lichens, mosses)
- Dead vegetation

Ideally, only cover in contact with the soil surface is considered effective ground cover for erosion control purposes; however, differentiating between plant leaves and stems that are in direct contact with the soil as opposed to simply in close proximity to the soil surface can be difficult and is not always feasible. In this procedure, the assumption is that if live or dead vegetation is within 1 inch of the soil surface, it is recorded as soil cover.

Vegetation often blocks the view of the soil surface and is therefore removed prior to data collection. When this is the case, the quadrat is clipped of standing vegetation (dead or alive) at a 1-inch height above the ground surface prior to taking the readings. The clipped vegetation is removed from the plot before the reading is made.

Data can be recorded on a field computer or field recording sheets. Field recording sheets bypass the need for electronic equipment in the field; however, data is entered into a spreadsheet at a later time for analysis. An Excel workbook with field entered forms and a data analysis package (see Figure 6-5) is available for this procedure in the [Native Revegetation Resource Library](#).

## **Sampling Soil Cover using Digital Photographs and the Cover Monitoring Assistant (CMA) Program**

Digital cameras are being used more frequently for monitoring plants and animal life. This technology allows the data recorder to spend most of the field time laying out transects and taking photographs of quadrats rather than collecting data, thereby reducing field time significantly. With recent developments in photo-imaging software, photographs can be assessed quickly in the office. Because these are permanently stored records, digital photographs have several advantages: photographs can be reviewed during the “off” season or “down time”; they are historical records that can be referenced years later; and they can be reviewed by supervisors for quality control. In addition, taking digital photographs of ground cover can be accomplished by one person, not two as is often used for fixed-frame monitoring. On steeper slopes, taking photographs to record quadrats is quicker and easier than reading attributes from a fixed frame.

Sampling by camera entails the same statistical design and plot layout as the fixed-frame method. The photograph is taken within a quadrat frame with 11-inch by 14-inch dimensions, covering approximately 1 square foot. In the office, digital photographs are catalogued electronically and a 20-point overlay is placed on the image that identifies the data points to be described (Figure 6-6). The data points are described in the same manner as for the fixed-frame method.



**Figure 6-4 | Fixed frame for measuring soil cover**

A fixed frame for measuring soil cover is placed at predetermined distances on a transect.

*Photo credit: David Steinfeld*

**For the Designer**  
Find this and many other fillable spreadsheets in the [Native Revegetation Resource Library](#) by typing “xls” into the search field.

|   | A2             |         |                  |        |       | fx       |          |              |      |             | Transect |          |          |  |  |  |  |
|---|----------------|---------|------------------|--------|-------|----------|----------|--------------|------|-------------|----------|----------|----------|--|--|--|--|
| 1 | A              | B       | C                | D      | E     | F        | G        | H            | I    | J           | K        | L        | M        |  |  |  |  |
| 2 | Sampling Area: |         |                  |        |       |          |          |              |      |             |          |          |          |  |  |  |  |
| 3 | Transect       | Quadrat | Number of Points |        |       |          |          |              |      | % of Points |          |          |          |  |  |  |  |
| 4 | 1              | 1       | Soil             | Gravel | Mulch | Live Veg | Dead Veg | Total Points | Soil | Gravel      | Mulch    | Live Veg | Dead Veg |  |  |  |  |
| 5 | 1              | 2       | 4                | 7      | 8     | 1        | 0        | 20           | 20   | 35          | 40       | 5        | 0        |  |  |  |  |
| 6 | 1              | 3       | 0                | 1      | 12    | 3        | 4        | 20           | 0    | 5           | 60       | 15       | 20       |  |  |  |  |
| 7 | 1              | 4       | 1                | 2      | 8     | 3        | 6        | 20           | 5    | 10          | 40       | 15       | 30       |  |  |  |  |
| 8 | 2              | 1       | 3                | 5      | 12    | 0        | 0        | 20           | 15   | 25          | 60       | 0        | 0        |  |  |  |  |
| 9 | 2              | 2       | 0                | 1      | 10    | 6        | 3        | 20           | 0    | 5           | 50       | 30       | 15       |  |  |  |  |

The FHWA and the U.S. Forest Service have developed a computer program to evaluate digital photographs called Cover Monitoring Assistant (CMA). This program configures each digital photograph taken at a quadrat so it is easy to evaluate in the office. It randomly places 20 points on the photograph and navigates the user to each point where the soil cover attribute is recorded. The data for each photograph is summarized in spreadsheets for statistical analysis. The CMA program has reduced field time and increased data quality significantly. The [CMA program information](#) and the [User's Guide](#) are available in the Native Revegetation Resource Library.

**Figure 6-5 | Data collection forms and statistical packages are available on the Native Revegetation Resource Library**

A data collection sheet and summary form spreadsheets, such as this one for soil cover, can be downloaded from the [Native Revegetation Resource Library](#) by typing in “xls” into the search field and selecting the appropriate workbook. Each section in this chapter has corresponding spreadsheets with instructions, field forms, and statistical packages.

### 6.3.2 SPECIES COVER

The species cover procedure is used to assess the above-ground abundance of plant species. This method is a point intercept method of estimating cover in a defined area (quadrat). This procedure can be used when DFC targets state that a certain percentage of vegetative cover be composed of native species. For example, a DFC target from the revegetation plan states that over 50 percent of the vegetative cover existing on the site two years after construction will be composed of native forb species for pollinator habitat and that this cover will consist of species that were in the seed mix. Monitoring vegetative cover with this procedure will determine whether this goal was achieved.

Another advantage of evaluating species cover is the elucidation of patterns of co-occurrences. Weedy species can occur in isolated patches, but often co-occur with other weeds and even among native species. Understanding these distributional patterns and the density of the species assemblages, as they change and develop with the project site, can help the designer select the most effective treatment methods, reduce redundant treatment costs, and prevent damage to the existing native plant community.

Because grass and forb cover changes during the growing season, the timing of species cover monitoring is important. It is ideal to monitor when most plants are flowering, typically mid-May through July for the western United States and July through August for the upper mid-western and eastern states. Bloom periods for forb species also coincide with the optimum period for monitoring pollinator species, so monitoring for species cover and pollinators can be done during the same field visit ([Section 6.4](#)).

This sampling procedure typically benefits from a botanists' knowledge to identify species and another person to help lay out the plots and record data. Sampling can be done either with a fixed frame or digital photographs using the CMA program.

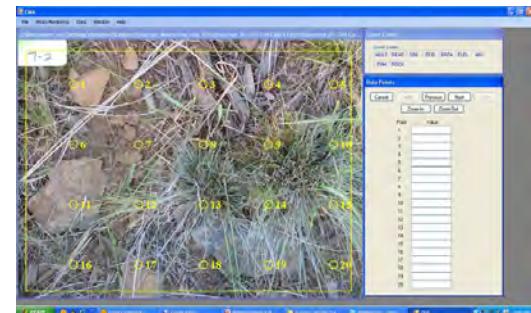
#### Sampling Species Cover Using a Fixed Frame

Using the fixed frame discussed in [Section 6.3.1](#), 20 points are identified in the quadrat frame. At each point, the species is identified and recorded in a ruggedized computer or on a field recording sheet. Species cover is quantified at each quadrat based on readings from a 20-point fixed frame. At each point, the species is recorded on a spreadsheet which is available on the [Native Revegetation Resource Library](#).

#### Sampling Species Cover & Floral Density using Digital Photographs and CMA

The CMA program, discussed in [Section 6.3.1](#), can also be used for determining species cover when plant species are easy to identify from digital photographs. These include species that are in bloom or with seed heads. Species of interest, such as those species used in a seed mix, may be the only species to identify for the analysis, requiring the recorder to know the appearance of just a few species. It may also be helpful to take a range of photographs of each species of interest for later reference when photographs are being analyzed. As with using CMA for determining soil cover, a 20-point grid is placed on each digital photograph and species cover points are evaluated (Figure 6-6).

Floral density can be quantified using the CMA program for describing the quality of pollinator foraging habitat while photographs are evaluated for species cover. In this analysis, the number of flowers of each species of interest are counted within the quadrat frame of each photograph (Figure 6-7). A spreadsheet for this procedure is available in the [Native Revegetation Resource Library](#).



**Figure 6-6 / The CMA program reduces field time**

Using the CMA program for determining soil cover and/or species cover, each quadrat photo is overlain with a 20-point grid that is randomly placed. Beginning with the first point, the user determines that point lands on a rock and that cover code is entered. The program moves the recorder to the next point until all 20 points are entered. The cover codes are decided by the recorder. In this example, plant cover was detected along with soil, mulch, and rock cover codes. In this case, three plant species were identified (shown as the abbreviations BRCA, EEL, and FEID) as discussed in [Section 6.3.2](#).

*Photo credit: David Steinfeld*

### 6.3.3 SPECIES PRESENCE

The Species Presence procedure is an alternative to the Species Cover method of determining whether species are present on the site. While this method still necessitates a person knowledgeable in plant identification in the field, it takes far less time per quadrat because only the presence of a species is recorded. In this method, a fixed frame is placed on the surface of the soil at each quadrat location and the presence or absence a “species of interest” is recorded. Depending on the specific project’s success criteria, only species of interest are recorded. Species of interest may include those species that have been seeded, important pollinator supporting species, and undesirable non-native plants. If ecological restoration is a project objective, then all species may be identified.

The size of the fixed frame is based on several factors, such as the growth pattern of the species of interest and the frequency that the species is present on the site. Large frames are not necessary for most grass and forb plant communities. The 1-square-foot frame used for the Soil Cover and Species Cover procedures may suffice (the tall grass prairie systems may be an exception, requiring larger frames). Larger native woody species and undesired weedy species are often measured in quadrats with an area of 1 square meter. This size of quadrat can accommodate the patchy distribution that weedy species frequently display and, when extrapolated, can create a fair representation of the plant species’ frequency across the project site. An Excel workbook with field forms and a data analysis package is available in the [Native Revegetation Resource Library](#) for this procedure.



**Figure 6-7 /** **Floral density can be determined using CMA**

Floral density can be determined by counting the number of flowers for each species of interest in a quadrat, then evaluated. In this photograph, Oregon sunshine (*Eriophyllum lanatum*) is the main species that is counted.

*Photo credit: David Steinfeld*

### 6.3.4 PLANT DENSITY

Trees, shrubs, and wetland plants are typically established from containerized plants grown in nurseries and planted at a specified density depending on the project objectives.

Monitoring the density of live plants after they are planted is important to determine how many plants have survived and whether the site will need to be replanted.

The quadrat in this procedure is a circular plot with a specified radius. For trees and shrubs, it is recommended a 1/100 acre plot be used, which has an 11.7-foot radius and covers 436 ft<sup>2</sup>. A staff is placed at the plot center and a tape or rope is stretched 11.7 feet. While one person holds the staff, the other walks the circumference of the plot and counts the number of plants of each species within the quadrat. This information is recorded on a spreadsheet, such as that shown in the form available in the [Native Revegetation Resource Library](#). This spreadsheet summarizes the total plants per acre and the number of plants per acre by species. Note: for linear sampling areas, only one quadrat is placed randomly within the transect.

Plant density monitoring that is used to determine survival rates are called survival surveys and these are usually conducted six months to a year after planting. If the site is planted in fall or spring and monitoring occurs the following fall, the results are referred to as the “first year survival.” Plant density monitoring thereafter is referred to as the years after planting (e.g., third year sampling is “third year survival”).

### 6.3.5 PLANT ATTRIBUTES

This procedure can be used to assess plant growth responses to determine how well plants are growing or whether DFC targets pertaining to growth targets were met. Typically, only sensitive areas, such as visual corridors or wetlands, have stated growth targets. This procedure might have limited applicability to most revegetation projects.

Any part of a plant can be measured for growth, and the selection of an attribute is

dependent on the species being sampled. Common attributes include the following:

- **Total height**—trees and most shrubs
- **Last season's leader length**—most trees
- **Stem diameter**—shrubs and trees
- **Crown cover**—shrubs, forbs, and grasses

Total height is typically measured from the ground surface to the top of the bud. If the plant has several leaders (or tops), the most dominant leader is used for measurement. Last season's leader growth can be observed in most tree species. For conifers, leader growth is measured from the last whorl of branches to the base of the bud. Comparing leader growth from year to year can indicate whether plants are healthy and actively growing.

Stem diameter is also used to measure trees and shrubs. In this method, the basal portion of the plant is measured with calipers at the ground surface. Another attribute is crown cover, which is more conducive to spreading plants, such as shrubs. A large frame with grids or points is placed over the plant to obtain an aerial coverage.

Attributes for shrub and tree species uses the sampling layout and design described for plant density ([Section 6.3.4](#)) and focuses only on those species of interest that have been identified in the revegetation plan. If there are many plants of a species of interest in a quadrat, only five plants are measured. An unbiased way to select which plants of each species to monitor is to sample those plants closest to the plot center. If there are not enough plants in the quadrat to sample, then the nearest plants outside of the quadrat are sampled. The measurement for each plant is recorded on a field data sheet, available in the [Native Revegetation Resource Library](#). Note: for linear sampling areas, only one quadrat is placed randomly within the transect.

### 6.3.6 SAMPLING UNIT DESIGN

The shape of the sampling unit determines how quadrats are laid out. Procedures have been developed for linear sampling areas, such as long roadsides; rectilinear sampling areas for rectangular-shaped areas such as restored borrow areas; and dispersed sampling areas, which are revegetation areas that are in patches or clumps. These three sampling designs are described below, as well as how to calculate the minimum number of quadrats to obtain an accurate representation of the sampling unit.

#### Linear Areas

Linear areas, such as cut slopes, fill slopes, and abandoned roads, are sampled from equally spaced transects placed along the entire length of the sampling unit. Each transect has varying numbers of quadrats, depending on the length of the transect. When the sampling unit is uniform in width, each transect will have an approximately equal number of quadrats; when the width of the revegetation unit is variable, there will be an unequal number of quadrats per transect. In either case, each transect is treated as the primary experimental unit instead of the quadrat (the primary experimental unit in this report is either the transect, quadrat, or dispersed area and is the unit for which statistical analysis is conducted).

Figure 6-8 shows a typical sampling design for a linear area. In this example, the sampling unit included only those portions of the cut slope that were seeded; it did not include road shoulders or the ditch line. For statistical analysis, the number of transects (n) to collect was estimated to be 20 based on pre-monitoring data. Spacing the 20 transects equally along the sampling unit was calculated by dividing the total length of the sampling unit by

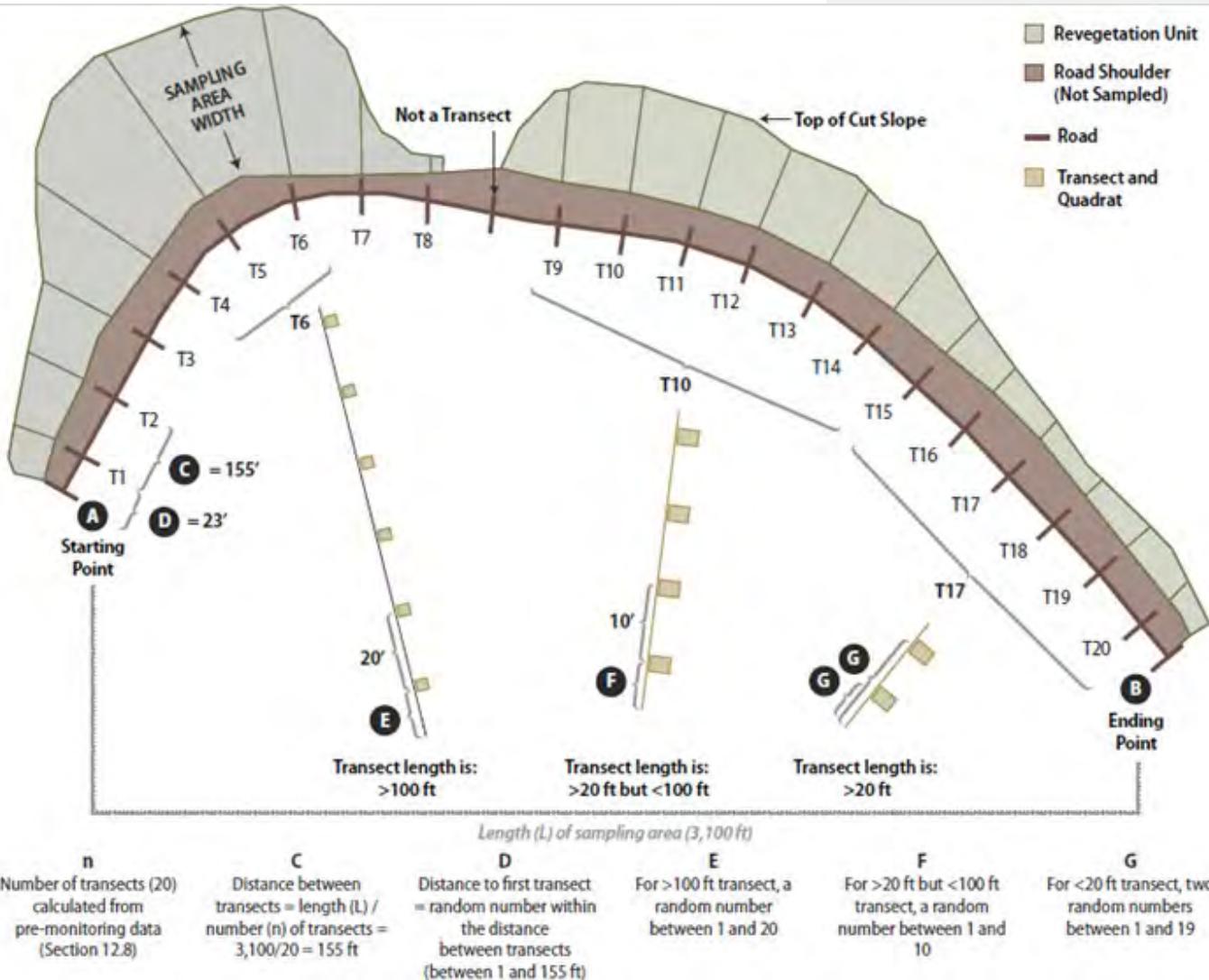
the number of transects to obtain the distance between transects ( $3,100/20 = 155$  feet). Locating the first transect is done in an unbiased way by generating a random number between 1 and 155. A random numbers table is available in the [Native Revegetation Resource Library](#).

Transects are laid out by establishing a line, typically a measuring tape, perpendicular to the edge of the road. The transect begins at the edge of the road beyond the ditch line and where seeding or other treatments were conducted. Each transect contains a series of quadrats spaced as follows:

- **<20 foot transect length**—For transects where lengths are less than 20 feet, two quadrats are placed. Two random numbers are generated between the numbers 1 and 19. These numbers indicate the location of each transect.
- **>20 foot and <100 foot transect lengths**—For transects where lengths are between 20 and 100 feet, quadrats are placed at 10-foot intervals. The distance to the first quadrat is based on a random number between 1 and 10 feet.
- **>100 foot transect lengths**—Long transects have 20 feet between quadrats. The distance to the first quadrat is based on a random number between 1 and 20 feet.

**Figure 6-8 / Linear sampling areas**

Linear sampling areas are long units with varying widths. Road cuts, fills, and abandoned roads typically fit this sampling design. The design of linear sampling areas uses a series of equally spaced transects with uniformly spaced quadrats within each transect. The distance between quadrats varies by the length of the transect. Transects longer than 100 feet have 20 feet of spacing between quadrats (T<sub>4</sub> through T<sub>6</sub>), transects between 20 and 100 feet have 10 feet between quadrats (T<sub>9</sub> through T<sub>14</sub>), and transects less than 20 feet long have two randomly placed quadrats (T<sub>15</sub> through T<sub>20</sub>).



## Rectilinear Areas

When sampling units are more elliptical or rectangular in shape, or composed of several large irregular polygons, the sampling design is based on a rectangular grid of quadrats

systematically located with a random starting point. Figure 6-9 illustrates an example of such a sampling design. Notice that this design is different from the linear sampling design in that there are no transects. The quadrat in this sampling design is the primary experimental unit, not the transect.

To determine the grid spacing (E) for the quadrat locations, the area of the sampling unit is calculated from maps, and the number of quadrats to be sampled (n) is determined from pre-monitoring data. The following equation gives the length of each side of a square grid (E):

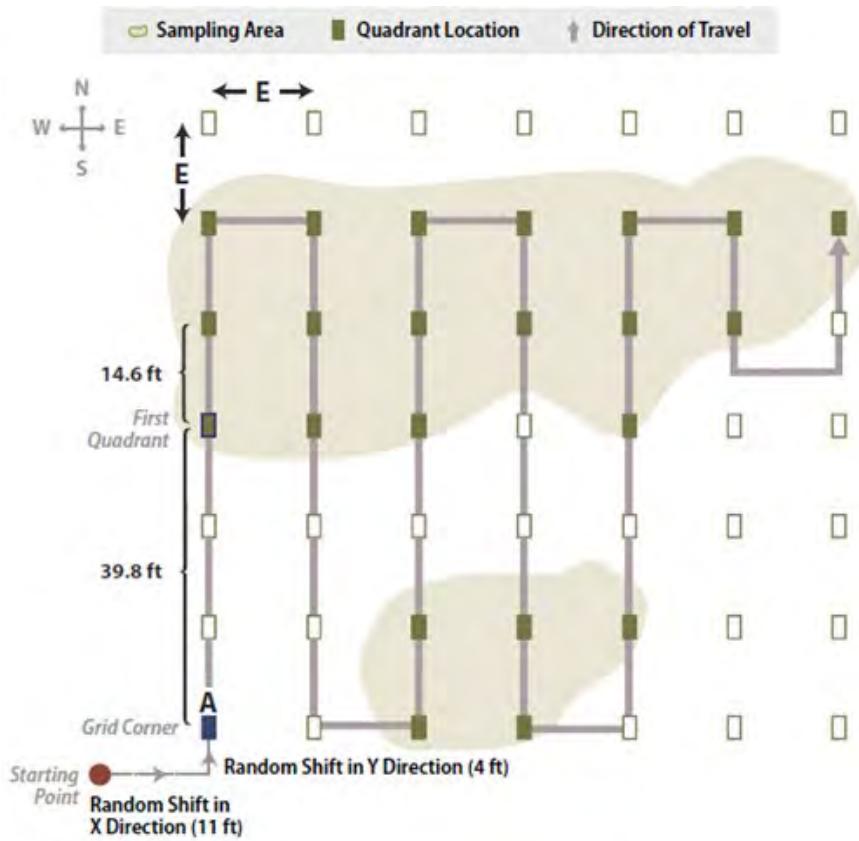
$$E = \sqrt{\frac{Area}{n}}$$

For example, the sampling unit in Figure 6-9 covers 4,262 ft<sup>2</sup> and it has been determined from pre-monitoring data that approximately 20 quadrats are necessary to attain the statistical sampling requirements for this sampling unit. The equation indicates that each side (E) of the grid is 14.6 feet:

$$E = \sqrt{\frac{4,262}{20}} = 14.6 \text{ ft}$$

A starting point of the grid is arbitrarily located in any corner of the study area. To avoid biasing the placement of the quadrats, the corner of the grid is shifted by assigning random numbers to the x and the y coordinates, as shown in Figure 6-9. This is called a systematic design with a random start. It provides equal likelihood that any point in the study area is included in the sample unless there are obvious systematic patterns in the site, such as planting rows. In this example, the random number shifts were 11 feet in the horizontal direction and 4 feet in the vertical direction.

The monitoring team locates the random starting point in the field and measures 39.8 feet ( $10.6 + 14.6 + 14.6 = 39.8$ ) north to the first quadrat. From that quadrat, they measure 14.6 feet north to the second quadrat. After they collect data from the last quadrat in that line, they travel east 14.6 feet to locate the next plot. This system of sampling continues in this fashion until the entire area has been sampled.

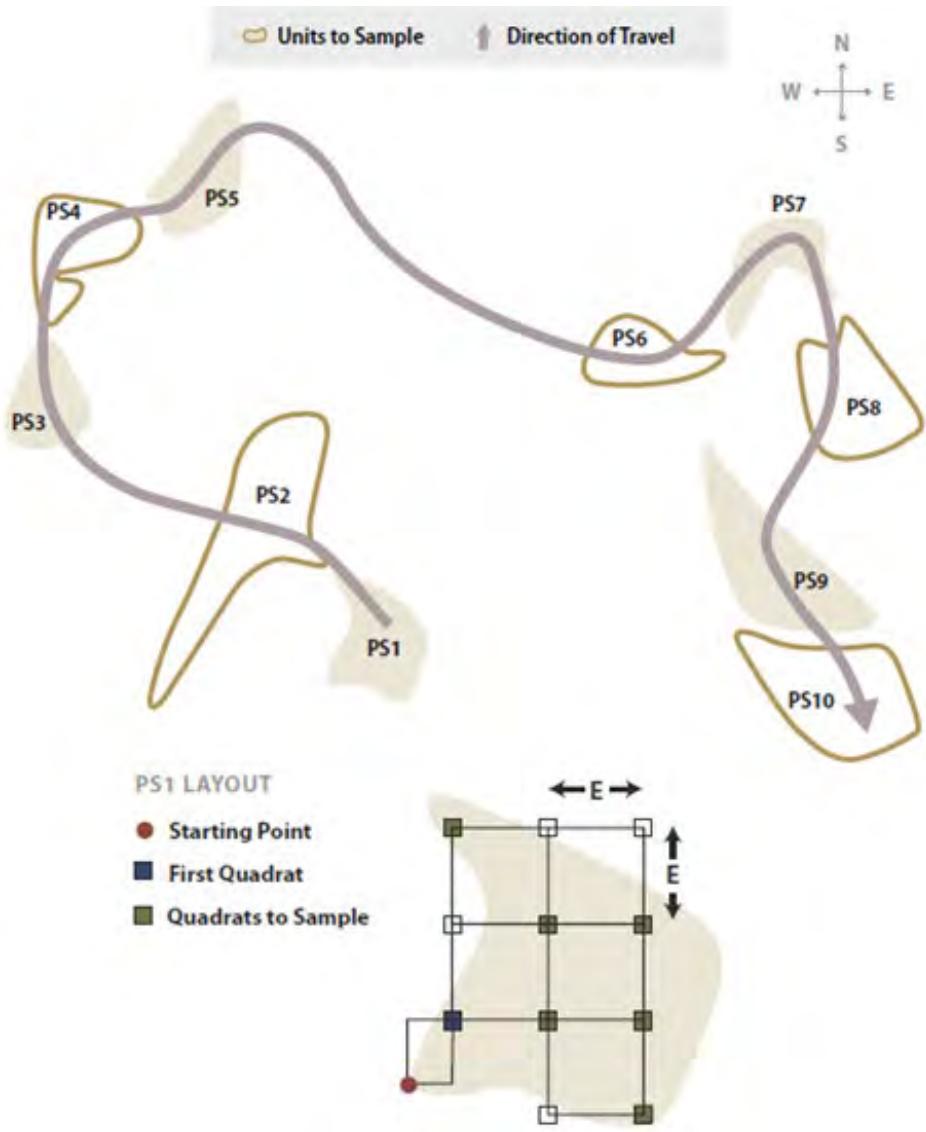


**Figure 6-9 | Rectilinear sampling areas**

For rectangular or elliptical sampling areas, a grid composed of quadrats is used. The length between grid cells ( $E$ ) becomes the standard distance between quadrats for this sampling unit. In this example,  $E = 14.6$  feet. To avoid bias, the x and y axes of the grid were randomly offset from the starting point by 11 and 4 feet to a new, random starting point (A). Then 39.8 feet was measured to the first quadrat. The monitoring team follows compass or GPS bearings to locate all quadrats.

## Dispersed Areas

When sampling units occur as small, distinct areas, a two-stage sampling design is recommended. The first stage is to determine which dispersed areas to sample, and the second stage is to determine how to sample within each selected area. For the first sampling stage, it is suggested that a minimum of 20 dispersed areas be monitored within each revegetation unit. If there are 20 dispersed areas or fewer, then all dispersed areas are sampled. If there are more than 20 areas, then sampling is conducted with one of two sampling designs. For dispersed areas that are mapped, the systematic sampling design is used. If the dispersed areas are small or are not mapped, then a grid sampling design is used. Both sampling designs are described in more detail below. These areas include planting islands and planting pockets. In most cases, the grid sampling method is recommended.



**Figure 6-10 | Systematic sampling of dispersed areas**

A systematic sample of dispersed areas, shown in this example, is based on 50 percent sampling (the pink areas). Alternating dispersed areas were sampled. Quadrats were located in each sampling unit by first locating a starting point and then measuring off random x and y offset coordinates to locate the first quadrat of the sampling grid.

## Systematic Sampling of Dispersed Areas

### First Stage

The systematic sampling method of dispersed areas assumes that the dispersed areas have been mapped. The dispersed area in this method is the experimental unit. In this approach, the dispersed areas are numbered sequentially by progressing from one dispersed area to the next closest dispersed area. Determination of the number ( $n$ ) of dispersed areas to sample is presented in [Section 6.3.6](#) (see [Sample Size Determination](#)). Odd or even-numbered dispersed areas are selected based on a random number using the RANDBETWEEN function. For example, if there were 40 dispersed areas ( $N$ ) but only 20 dispersed areas needed to be sampled ( $n$ ), then 50 percent of the dispersed areas would be sampled ( $n/N$ ). A random number using RANDBETWEEN (1, 2) is used. If the function returns a "1," the odd-numbered dispersed areas are selected; if 2, then the even numbered areas are selected. Figure 6-10 provides a schematic depiction of such a design.

### Second Stage

Once the dispersed areas have been selected, the layout of quadrats within each dispersed area is determined. A grid-based sampling design is used. To determine the grid spacing ( $E$ ) for the quadrat locations, determine whether the size of the dispersed area is  $>1,600 \text{ ft}^2$  or  $<1,600 \text{ ft}^2$  ( $1,600 \text{ ft}^2$  is a 40-by-40-foot area). Depending on the sample

size, the number of quadrats will be as follows:

- **Less than 1,600 feet**—four quadrats
- **More than 1,600 feet**—eight quadrats

Using the following equation, the grid sides (E) are determined where  $n = 4$  or  $8$  (depending on the size of the dispersed area) and the area is the estimated size of the dispersed area.

$$E = \sqrt{\frac{\text{Area}}{a n}}$$

This calculation is made for each dispersed area. To avoid biasing the placement of quadrats, a predetermined starting point is made (e.g., the northwest corner of each dispersed area). Random x and y coordinates are generated for each dispersed area that is within the predetermined length of the square grid (E). The corner of the grid is placed at this point and oriented north. The location of the first quadrat is x and y feet from the starting point, as shown in Figure 6-10.

For the plant density and plant attribute procedures, only one quadrat is randomly located within each dispersed sampling area. The location is determined by assigning the RANDBETWEEN random number function to the number of quadrats determined for the dispersed area. The random number that is generated is the quadrat selected for sampling.

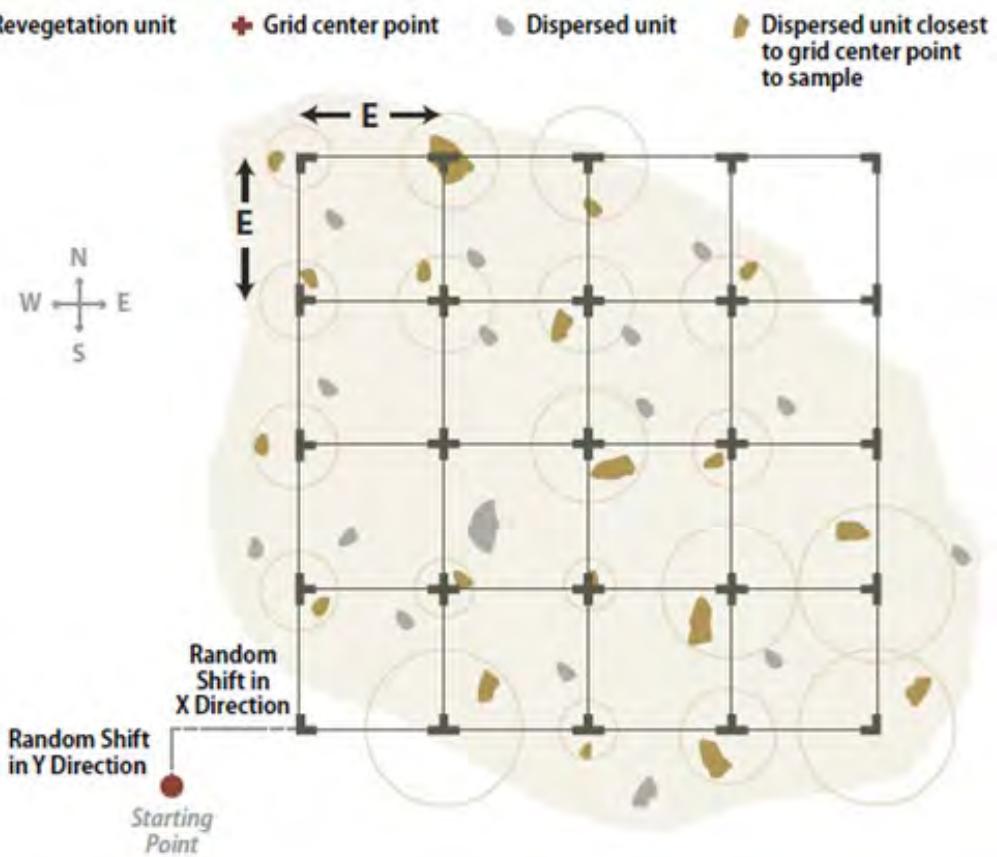
### **Grid Sampling of Dispersed Areas**

#### ***First Stage***

A grid sampling design is used on projects where dispersed areas have not been mapped and there are more than 30 areas. This sampling design is used on projects where dispersed areas are numerous and the sizes of the areas are small. In this sampling design, a grid is placed over the revegetation unit, as shown in Figure 6-11, and the dispersed area nearest to a grid crossing is selected for monitoring. One quadrat is randomly placed in each dispersed area and this becomes the experimental unit.

Determining the grid cell dimensions (E) is accomplished by using the following equation:

$$E = \sqrt{\frac{\text{Area}}{a n}}$$



The grid is laid out in an unbiased manner by locating a starting point just outside the revegetation unit and assigning random numbers as offsets for the x and y coordinates. The corner of the grid is placed at the x and y offset point and oriented north. At each grid crossing, the closest dispersed area is selected for monitoring. If there are no dispersed areas found within half the distance between grid centers ( $E/2$ ), then the monitoring team moves on to the next grid center.

#### **Second Stage**

Since the dispersed areas are small (e.g., planting pockets, planting islands, benches), all measurements for seedling density and seedling attributes may be sampled within each dispersed area. For other procedures, such as soil cover, species cover, or species presence procedures, one quadrat is located randomly in the dispersed area.

#### **Sample Size Determination**

This section covers how to take pilot data to determine the number of experimental units (transects or quadrats) needed to statistically monitor a sampling unit. It is important to note that an alternative to taking pilot data is to simply to sample a minimum of 20 primary experimental units per sampling unit. For example, for a linear sampling area, 20 transects are laid out; for rectilinear areas—20 quadrats; and for a dispersed area sampling design—20 dispersed areas. Twenty primary experimental units will provide adequate data to accurately estimate means and confidence limits and to understand the uncertainty (i.e., width of confidence intervals). The risk is that, in some cases, the intervals may be too wide to provide meaningful estimates, which means that more samples would be needed, requiring revisiting of the sampling unit. Generally, if data are moderately variable and symmetrically distributed, 20 experimental units will often be adequate (personal communication: John Kern, Kern Statistical Services Inc., February 10, 2017). The following section is intended for projects where a more exact sample size, using pilot data, is desired to achieve the precision requirements of the monitoring plan.

**Figure 6-11 | Sampling dispersed areas with an offset grid**

A systematic sample of dispersed areas, shown in this example, is based on 50 percent sampling (the pink areas). Alternating dispersed areas were sampled. Quadrats were located in each sampling unit by first locating a starting point and then measuring off random x and y offset coordinates to locate the first quadrat of the sampling grid.

Sample size determination methods are tailored to the monitoring objectives and sampling area design. This section outlines four sampling methods: (1) comparing means with DFCtarget for linear area sampling design, (2) comparing means with DFC targets for rectilinear sampling area design, (3) comparing means with DFC target for dispersed areas, and (4) determining sample size for comparing treatment areas.

### **Comparing Means with DFC Targets—Linear Sample Size Determination**

To calculate the minimum sample size for linear sampling areas, the expected sample mean and the range in values are approximated. A visual estimate of the mean and the range of values may be adequate. For more precise estimates, four transects are sampled in the field to establish estimates of the mean and range of values.

The following is a quick method to determine the number of transects to sample based on pre-monitoring data collection:

- Driving the entire revegetation unit and noting extremes in vegetation
- Finding four areas that represent the extremes and laying out a transect in each area
- Randomly placing two quadrats within each transect to collect data on the attribute of interest (e.g., percent ground cover)
- For each transect, averaging the quadrat values (four averages)
- Calculating the average of the four transect averages and the range (difference between highest and lowest values of the four averages)
- Applying the range and the average to the following equation to obtain the minimum number of transects needed to monitor the revegetation unit (equation based on a sample size of 20 percent of the true population value with 90 percent confidence):

$$n = (0.838 * \text{Range})^2 / (0.2 * \bar{x})^2$$

The number of transects ( $n$ ) obtained from this quick assessment is used to determine the layout of the monitoring design, as described in [Section 6.3.5](#).

**Example:** Suppose that monitoring is to be conducted along a road cut to determine the percentage of soil cover. The monitoring objective is to estimate mean percent cover to within 20 percent of the true population value with 90 percent confidence. Four transects that represent a range in conditions are laid out within the revegetation unit. These transects do not need to be located randomly but rather can be sited to capture the range of values observed in the sampling unit. It is better to over-estimate the range of values, as this will tend to result in a conservative estimate of the sample size. The data set and equation are shown in Figure 6-12.

Using the equation presented above, an estimated 13 transects were calculated for a minimum sample size

### Comparing Means with DFC Targets—Rectilinear Sample Size Determination

When a grid of quadrats is to be implemented (Section 6.3.6), the sample size calculations use quadrat measurements as opposed to the transect averages that were used in linear sampling areas. The steps involved in calculating the number ( $n$ ) of quadrats are as follows:

- Visiting the entire revegetation unit and noting extremes in revegetation
- Finding four areas that represent the extremes and laying out a transect in each area
- Randomly placing two to three quadrats within each transect to collect data on the variable of interest (e.g., percent ground cover)
- Averaging the quadrat values (eight quadrat values to average)
- Calculating the range of all eight samples (difference between highest and lowest values)
- Applying the range and the average to the following equation to obtain the minimum number ( $n$ ) of transects needed to monitor the revegetation unit:

$$n = (0.838 * \text{Range})^2 / (0.2 * \bar{x})^2$$

**Example:** Using the data from the previous example (Figure 6-12), the eight quadrats would be averaged (instead of four transect averages being averaged). While the mean in this example remains 17, the range has spread to 26 (maximum 32 minus the minimum 6). The number of quadrats to sample would be 41:

$$n = (0.838 * 26)^2 / (0.2 * 17.0)^2 = 41.126$$

### Comparing Means with DFC Targets—Dispersed Area Sample Size Determination

**Systematic Sampling Method:** The systematic sampling method is used when the dispersed areas are mapped (see Systematic Sampling of Dispersed Areas). The primary experimental unit in this design is the dispersed area. The number of quadrats to sample is estimated by following these steps:

- Finding four dispersed areas that represent the extremes of the attribute of interest
- Randomly placing two quadrats within each dispersed area to collect data on the attribute of interest (e.g., percent ground cover)
- For each dispersed area, calculating the average of the four dispersed areas' averages and the range
- Applying the range and the average to the following equation to obtain the minimum number of dispersed areas needed to monitor in a revegetation unit (equation based on a sample size of 20 percent of the true population value with 90 percent confidence):

$$n = (0.838 * \text{Range})^2 / (0.2 * \bar{x})^2$$

An example of the data set and equation are shown in Figure 6-12. Note: substitute “dispersed area” for “transect” in the first column heading.

**Grid Sampling Method:** The grid sampling method is used for revegetation units that have small dispersed areas that have not been mapped (see Grid Sampling of Dispersed Areas). Since only one quadrat is in a dispersed area, the quadrat becomes the primary experimental unit. The number of quadrats to sample is estimated by following these steps:

| QUADRAT  |    |    |          |
|----------|----|----|----------|
| Transect | Q1 | Q2 | Averages |
| T1       | 10 | 15 | 12.5     |
| T2       | 12 | 22 | 17       |
| T3       | 18 | 6  | 12       |
| T4       | 32 | 21 | 26.5     |

$$\text{Estimated Mean} = 17$$

$$\text{Range} (26.5 - 12.5) = 14.5$$

$$n = (0.838 * 14.5)^2 / (0.2 * 17.0) = 12.8 \text{ transects}$$

**Figure 6-12 | Determining the number of transects using pilot data**

Example of how to calculate the number of transects needed, based on the pre-monitoring data set.

- Visiting a range of dispersed areas and selecting eight dispersed areas that represent extremes of the attribute of interest
- Randomly placing one quadrat in each dispersed area and collecting data on the attribute of interest (e.g., percent ground cover)
- Calculating the average and range in values
- Applying the range and the average to the following equation to obtain the minimum number (n) of dispersed areas to sample:

$$n = (0.838 * \text{Range})^2 / (0.2 * \bar{x})^2$$

**Example:** Using the data from the previous example (Figure 6-12), the eight quadrats would be averaged (instead of four transect averages being averaged). While the mean in this example remains 17, the range has spread to 26 (maximum 32 minus the minimum 6). The number of quadrats to sample would be 41:

$$n = (0.838 * \text{Range})^2 / (0.2 * \bar{x})^2$$

### Comparing Means among Treatment Groups—Sample Size Determination

When determining the sample size for comparing treatment differences (Section 6.3.7), it is not necessary to differentiate between linear, rectilinear, or dispersed sampling area designs. Sample size determinations follow these steps:

- Reviewing each treatment area to be compared. These can be different revegetation units or different types of revegetation treatments.
- From each treatment area, collecting data from two transects with each transect consisting of two quadrats (total of four transects) that represent a range of extremes.
- Determining the range between the low and high quadrat values.
- Determining delta (also represented by  $\Delta$ ). Delta defines the level of significance that is needed for monitoring, or the meaningful difference in measurement output. For example, calculating bare soil at 1 percent difference in means would be unimportant, that is, the difference between 8 and 9 percent bare soil is too fine a distinction to make. A 5 percent difference might be important if the amount of data that is needed to be collected was not great. More than likely, a 10 percent difference (e.g., the difference between 10 and 20 percent bare soil) would be an acceptable delta value for bare soil cover. It is important to note that the smaller the delta value, the more samples that need to be collected.

The number of transects (n) can be calculated using a simplified equation:

$$n = 15.68 / (\Delta * 2.059 / \text{Range})^2$$

The number of transects determined from these calculations are applied equally to the two (or more) areas being compared. This equation assumes that tests will be conducted at the delta level of significance and that there will be a difference detected at or greater than an 80 percent probability.

**Example:** A test was set up to determine whether a commercial product could increase plant cover by at least 10 percent one year after application, as advertised. In one area, the product was applied and in another similar area, it was not. To determine the number of transects to install in each area, a year after application, two transects were set up in the treated area and two transects in the untreated area. From the quadrat readings, the

range in percent vegetative ground cover values was found to be 22. Assuming a delta level of significance of 10 percent (important to be able to detect a difference in 10 percent cover with at least 80 percent probability), the following number of transects required for each treatment area were determined to be 18:

$$n = 15.68 / (10 * 2.059 / 22)^2 = 17.9 \text{ transects}$$

In general, using the simple equation provided above, and possibly adding 10 to 20 percent additional samples as a level of conservatism, is a reasonable approach.

### 6.3.7 ANALYZE DATA

This section provides statistical methods for analyzing data collected for all monitoring procedures described in the previous section. There are three types of analysis based on the objective of the area being monitored:

- **Compliance**—Determine whether DFC targets were met
- **Treatment differences**—Determine whether there were differences between treatments or changes between years
- **Trends**—Determine the degree of vegetation or soil cover change over time

Only one of these procedures is selected for an analysis depending on the monitoring objective used. These procedures use confidence intervals to determine the statistical significance of the monitoring data set.

#### Analysis of Compliance

The objective behind most monitoring is to answer these three questions:

- Was the project successful?
- Were DFC targets met?
- Were the commitments made to the community and those described in planning and compliance documents and reports met in terms of protecting soil, reestablishing native vegetation, and maintaining or improving pollinator habitat?

To answer these questions, the means of the attribute data of interest (e.g., bare soil, species presence) are compared with the DFC targets. For example, a project has a DFC target of at least 70 percent soil cover on a road cut near a live stream one year after road construction.

Using the soil cover procedure, data are collected on 20 transects and a mean of 81 percent soil cover is determined. At this point, the designer might conclude that the targets were met. From a statistician's point of view, however, the data displayed in this context is inconclusive because the variability of the soil cover in the sampling unit is not known or cannot be accounted for. In other words, how good is the number? Is it really depicting what is happening on the site? If another person were to use the soil cover procedure in the same sampling unit but at a different spot, would the soil cover be exactly 81 percent? This is highly unlikely because of the high variability of soil cover.

Confidence intervals provide a means of predicting, at a chosen level of certainty, whether the soil cover value collected anywhere in the sampling unit will be within a stated range. Confidence intervals are an alternative to saying, “we think the soil cover at any point in the sampling unit will be around 81 percent.” Using confidence intervals, it can be said instead that, “we are 90 percent confident that if data collection was repeated at this site 10 times, 9 out of 10 times the average soil cover estimate would be within our confidence limits.” If a higher confidence level is desired (most scientists working in health fields want to be more certain), confidence intervals can be based on 95 percent or even 99 percent certainty. In this case, the confidence interval would be much wider.

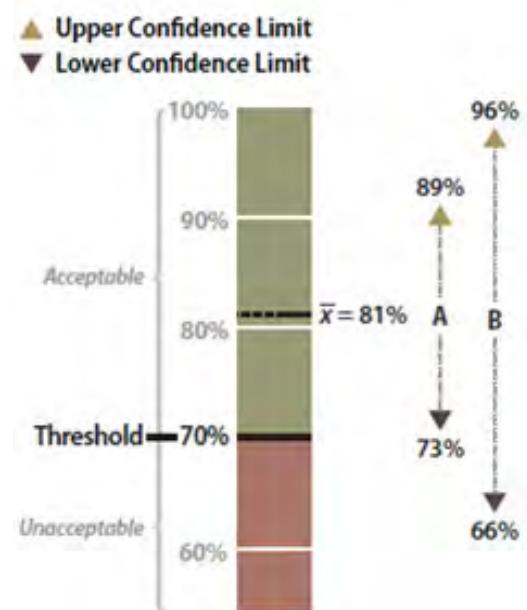
The data sets from very different revegetation units are shown in Figure 6-13 to convey the concept of confidence intervals. While both data sets have the same mean of 81 percent soil cover, the confidence intervals are very different. Data set A was taken at a site with very uniform soil cover, while data set B had much more variability. For both data sets, a 90 percent confidence interval is desired. Notice that the confidence interval for data set B is much wider than data set A since it has greater variability.

With confidence intervals, it can be said with greater certainty whether the DFC target or threshold of 70 percent soil cover was met. It can be stated with 90 percent confidence that data set A met the target because the lower confidence limit (73 percent) is above the target of 70 percent. Alternatively, data set B poses some problems. It cannot be said with 90 percent confidence that the average is above the threshold of 70 percent because the lower confidence limit of data set B is 66 percent, or 4 percent below the threshold. The designer might argue that, because the mean is above the target, the target was met. To statisticians, however, the fact that the lower confidence interval is below the stated target indicates a fair amount of uncertainty. They would have to obtain more data or change how confident the prediction is before they could be certain the DFC target was met.

### Calculating Confidence Intervals

Workbooks are available in the Native Revegetation Resource Library website to calculate confidence intervals depending on the sampling unit design (Section 6.3.6):

- **Linear sampling design**—Select [this Excel workbook](#) for linear sampling designs.
- **Rectilinear sampling design**—Select [this Excel workbook](#) for rectilinear sampling designs.
- **Dispersed sampling design**—For this sampling design, there are two workbooks to choose from. If a grid sampling design was used, [this Excel workbook](#) is used, and if a systematic sampling design was used, [this workbook](#) is used.



**Figure 6-13 / Example data analysis results**

Data set A has an upper confidence limit of 89 percent and a lower confidence limit of 73 percent. Since the lower confidence limit is above the target, or threshold, of 70 percent, it can be stated with 90 percent confidence that the target was met. Data set B has a wider confidence interval and a lower confidence limit of 66 percent, which is below the target. In this case, there is uncertainty at the 90 percent confidence limit that the target was met.

## Interpreting Confidence Intervals for Compliance

Confidence intervals, as stated previously, are used to evaluate the success of a revegetation project relative to a specified DFC target. For example, an objective of a revegetation project was to establish at least 65 percent total cover over the soil to protect against surface erosion. Or stated another way, the objective was that no more than 35 percent bare soil would be exposed one year after revegetation treatments were applied. During monitoring, it was found that the lower confidence level was 23.8 and the upper confidence level was 30.8, both below the target of 35 percent. It could be said with a 90 percent level of confidence that the objectives were met. But what if the confidence interval was either above the target or straddled the target?

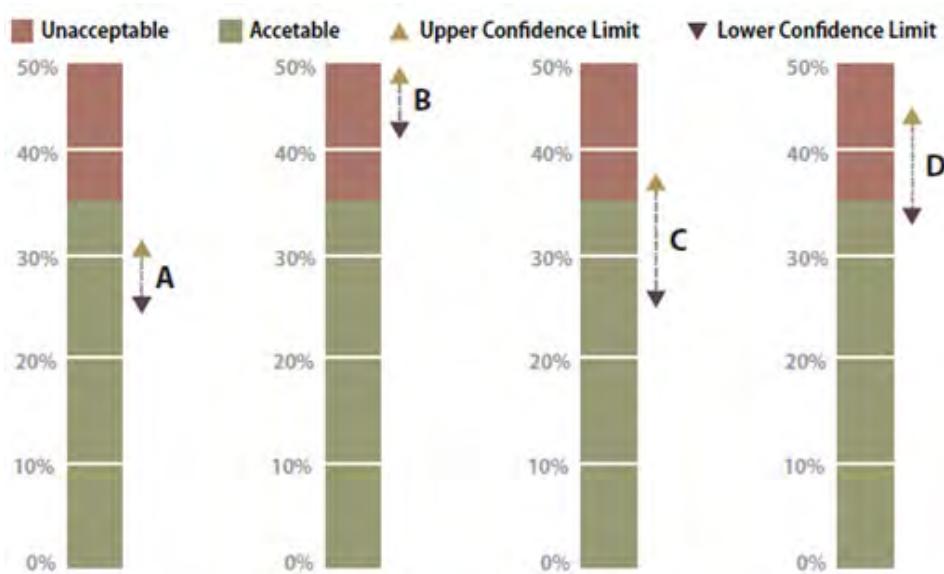


Figure 6-14 shows four possible scenarios for confidence intervals that may be encountered. Scenario A is a case in which the data supports the conclusion at 90 percent confidence that the project met the target. Scenario B did not meet the target because the lower confidence limit was above 35 percent bare soil.

Scenario C is a case where the mean meets the target, but the upper confidence limit is above the target. It cannot be said with 90 percent confidence that the targets were met. At this point it might be asked how important it is to know whether the target was met. If the site directly influences a live stream, it might be very important. However, if there are no streams nearby, it might suffice to report that there was some uncertainty whether the target was met. Additional data collection can help address a lack of certainty.

Scenario D is a case where the mean does not meet the target, but the lower confidence limit is below the target of 35 percent bare soil. One might state that this project did not meet the target, but this still could not be stated with 90 percent confidence. In this scenario, more transects could be taken to narrow the confidence interval and hope that the results do not straddle the target.

Another option for Scenarios C and D might be to implement measures to decrease the amount of bare soil. This could include more seeding or application of mulch. The site would be resampled after an interval of time to determine whether the DFC target had been met.

**Figure 6-14 | Possible scenarios when comparing targets to confidence intervals**

Four possible scenarios that can be encountered when comparing targets to confidence intervals. Confidence interval A is clearly acceptable, whereas confidence interval B is clearly unacceptable. Uncertainty arises when the lower confidence limit is acceptable but the upper confidence limit is unacceptable. This is shown in cases where the mean is in the acceptable range (C) and in the unacceptable range (D).

## Analysis of Treatment Differences

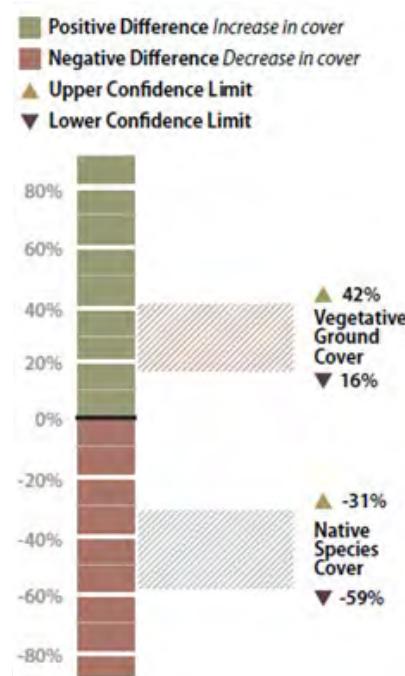
There may be opportunities to compare the effects of different revegetation products or methods on plant establishment and growth using monitoring data. Some of these opportunities will be planned (e.g., trying a new product), and some will be mistakes (e.g., inadvertently doubling the rate of mulch application). Planned or unplanned, when different revegetation activities have occurred within a revegetation unit, monitoring can be designed to assess whether there is a different vegetative response between those activities or treatments. The monitoring design outlined in this section will not replace a well-designed study or experiment; it is suggested that if more conclusive results of treatment differences are desired, a study would be designed with statistical oversight. An Excel workbook is available in the [Native Revegetation Resource Library](#) for this analysis.

The confidence interval concept is applied in this subsection to determine differences between new revegetation treatments (new treatments) and routine revegetation methods (standard treatments). Three possible outcomes are possible when new treatments are compared to standard methods: (1) the new treatment results in a favorable increase in the measured parameters over the standard treatment (positive difference), (2) the new treatment results in a decrease (negative difference), or (3) there is no positive or negative difference (no difference). Using confidence intervals, it is possible to determine which of these outcomes is statistically supported for any monitoring data set. In this method, the means and variance of means are calculated for both the new treatment and standard treatment and a confidence interval is calculated for the certainty of the treatment differences.

The following example demonstrates how a confidence interval is determined and how it can be used to interpret two data sets. During a hydroseeding operation, it is discovered that fertilizer was mistakenly applied at twice the normal rate in one area. This area was staked in the field when the realization was made and visited by the project team a year later. Some on the team believed that there was more vegetative ground cover where fertilizer was doubled; others felt that there was less. One or two on the team did not believe they could make either call. Since monitoring was going to occur a few weeks later, they decided to design a monitoring procedure to answer the question, "Was there a positive, negative, or no response of vegetative ground cover to the application of additional fertilizer?"

Within the framework of monitoring that was already scheduled for this revegetation unit, a monitoring strategy was developed. The species cover procedure was used ([Section 6.3.2](#)) along with the linear sampling design since this was a road cut ([Section 6.3.6](#)). Each treatment area was considered a separate sampling unit, and monitoring of each treatment area took place independently of the others. The data from each treatment area was recorded in the Species Cover spreadsheet (see [Species Cover Monitoring Procedure workbook](#)) to obtain means and variance of means. These values were then entered into the Comparing Treatment Differences Monitoring Procedure spreadsheet to obtain confidence intervals.

The results of this analysis showed that the standard rates of fertilizer had an average vegetative ground cover of 33 percent as compared to 62 percent for double the fertilizer rate. While this looks like an obvious difference, how certain could the team be? In this example, the confidence interval (at 90 percent confidence) showed that additional fertilizer significantly increased vegetative ground cover. This can be shown graphically in Figure 6-15. The doubled fertilizer treatment increased ground cover a minimum of 16 percent (lower confidence interval) over the standard treatment to as much as 42 percent ground cover (upper confidence limit).



**Figure 6-15 | Interpreting results with confidence intervals**

In the example presented in the text, confidence intervals are used to answer: (1) how vegetative ground cover responds to  $2\times$  the fertilizer rate, and (2) how native species cover responds to  $2\times$  the fertilizer rates. The confidence interval collected for the first question was found to be positive, indicating that vegetative ground cover responds positively to twice the fertilizer. The confidence interval for the data set collected for the second question was negative, indicating that native species cover responded negatively to more fertilizer.

The team accepted these results and commented that fertilizer rates be increased for future projects. One member posed the question, "We might have achieved better vegetative cover on this site during the first year, but how did additional fertilizer affect the native species cover?" Since the monitoring team was still on the project site, they resampled the two areas using the species cover procedure (Section 6.3.2). In this procedure, native and non-native annuals and perennials were recorded at each quadrat. Confidence intervals were determined for each treatment for native perennial cover and they learned, in this case, that additional fertilizer had a negative effect on the establishment of native perennial cover (Figure 6-15). But what if the upper confidence interval in this example had been positive and the lower confidence interval had been negative? In this case, it would have to be concluded that there was no difference between treatments at 90 percent confidence.

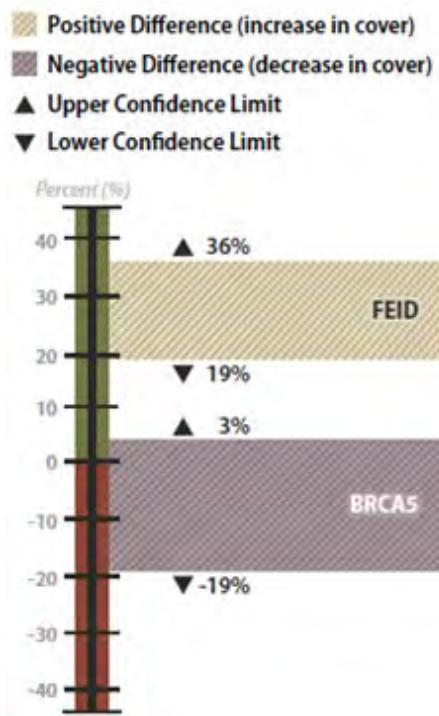
## Analysis of Trends

The last of the three objectives for roadside monitoring is assessing trends, or the degree that attributes, such as vegetative or soil cover, change over time. One of the main reasons to perform this type of monitoring is to understand how plant growth or successional patterns change. Many monitoring procedures employ permanent monitoring plots or transects that can be repeatedly and accurately revisited for sampling. This approach does not work as well for roadside monitoring however, because of the hazards to road maintenance personnel and to the public of placing permanent stakes in road corridors. In addition, permanent markers are often hard to relocate years later or can move due to the instability of steep cut and fill slopes. In this section, a statistical analysis that does not entail locating and resurveying of exact quadrats is offered. An Excel workbook is available in the [Native Revegetation Resource Library](#) for this procedure

The confidence interval approach is applied in this section to determine if there are differences in attributes from one sampling date to another. Three outcomes are possible when comparing data from one sampling date to the next: (1) attributes have increased since the last sampling period (positive difference), (2) attributes have decreased since the last sampling period (negative difference), or (3) either there was no change in attributes or the number of samples was inadequate to detect the amount of change that occurred. Using confidence intervals, it is possible to make statistically valid statements regarding the observed outcomes.

This question necessitated the use of the Species Cover procedure (Section 6.3.2) since dominance was being expressed as percent crown cover for each species. Linear Sampling Design was used for both monitoring dates because the sampling unit was a long cut slope. The data from each sampling date for California brome and Idaho fescue was entered into the spreadsheet shown in the Species Coverspreadsheet (see [Species CoverMonitoring Procedure workbook](#)) to obtain means and variance of means. These values were then entered into the spreadsheet shown in the [Analyzing Trends Monitoring Procedure workbook](#) to produce confidence intervals for comparison. Data entry was also conducted in the same manner for Idaho fescue.

The results of this analysis showed that California brome had an average crown cover of 35 percent in 2001 but decreased to 27 percent in 2006, five years later. Was this a statistically significantly difference? Using confidence intervals, it was determined that the means were not statistically different at 90 percent confidence. This can be shown graphically in Figure 6-16. Because the upper confidence limit was positive and the lower was negative, the team could be 90 percent confident that crown cover did not increase from 2001 to 2006. Alternatively, Idaho fescue did show an increase in mean crown cover from 2001 to 2006. This increase was found to be significant because both the upper and



**Figure 6-16 / Example results with confidence intervals**

In the example, confidence intervals were used to determine whether California brome (BRCA5) and Idaho fescue (FEID) increased, decreased, or stayed the same from 2001 to 2006. The percent crown cover of California brome was found not to have changed during this period because the lower confidence limit was negative and the upper confidence limit was positive. The confidence interval for the Idaho fescue showed an increase between 2001 and 2006. Since the upper and lower confidence limits were positive, the differences in the means between sampling dates were significant at 90 percent confidence.

lower confidence limits were positive. The team could be 90 percent confident that there was a true increase in crown cover.

## 6.4 POLLINATOR MONITORING PROCEDURES

The monitoring procedures presented in this section provide instructions for monitoring bees, butterflies, and monarch butterflies to assess the quality of revegetated roadsides as pollinator habitat. The data collected from these procedures will allow the project designer to determine if the revegetation project successfully addressed pollinator-specific DFCs. These procedures can be used to assess roadside habitat in a number of ways, such as determining (1) whether revegetation efforts have increased pollinator richness and abundance (by including nearby non-revegetated roadsides in samples as reference sites), (2) if pollinator richness and abundance change over time, (3) the effects of different roadside management strategies or seed mixes on pollinators, and (4) habitat quality and survival rates for monarch butterflies.

Three monitoring procedures are presented in this section.

- **Bee abundance procedure**—This is a standardized, streamlined monitoring procedure that provides estimates of bee abundance and involves minimal time and training. It involves establishing transects in the project and conducting timed assessment to observe and count the abundance of bees on flowers. This procedure can be used to statistically compare the quality of sites or seed mixes for pollinators and assess changes in pollinator populations over time. This procedure can be conducted on the same transects as those established for soil and plant monitoring ([Section 6.3](#)).
- **Bee and butterfly diversity procedure**—This procedure provides methods for measuring morphogroups of bee and butterfly species along transects in the project. While it involves some training and practice, this procedure generates robust data that is useful for detecting community-level changes in bee or butterfly abundance and richness.
- **Monarch butterfly reproduction and habitat procedure**—This procedure outlines methods to measure the abundance of host plants for monarch butterflies and the density of monarch eggs and larvae. Obtaining this data entails a moderate amount of time and expertise, but they can be used to quantify host plant availability and survival rates of monarchs.

In all of these procedures, standardizing the sampling effort and weather conditions are effective practices. Weather conditions strongly influences pollinator behavior. Pollinators are uncommon during cold, windy, or overcast weather, so it is best to monitor under optimal and consistent conditions when monitoring adult bees or butterflies. Optimal conditions for sampling pollinators include sampling between 10 a.m. and 4 p.m. on days with air temperatures over 60° F, wind speeds less than 10 miles per hour, and skies mostly clear. If sampling over time, monitoring is conducted using the same procedure over the same area at roughly the same phenology stage or the same weeks and months every year and under similar weather conditions.

It is important to note that counting individual European honey bees alone cannot provide a measure of the value of habitat for native bees or other pollinators because the number of individual honey bees visiting habitat is primarily determined by the number of managed hives in the vicinity. Although the presence of honey bees does indicate that the vegetation supports bees, it does not demonstrate how well the vegetation increases abundance and diversity of unmanaged bees and other pollinators.

Pollinator populations vary over the course of the growing season and from year to year. If a species of interest is to be targeted using monitoring (e.g. monarch butterflies), it is useful to schedule monitoring according to their flight period. Pollinator populations also vary annually, increasing as plants become established and mature, which may take several years after seeding or planting. For this reason, monitoring procedures are conducted for multiple years after a revegetation project has been completed.

## 6.4.1 BEE ABUNDANCE MONITORING PROCEDURE

### Overview of procedure

- **What the procedure will measure**—Wild bee and honey bee abundance
- **Sampling design**—Transects within the project area
- **Sampling frequency**—Two visits per growing season (at minimum)
- **Sampling timing**—Warm, sunny, and calm days, between 10 a.m. and 4 p.m.
- **Level of identification needed**—Distinguish bees from all flower visitors; distinguish native bees from European honey bees
- **Equipment needed**—Stopwatch or clock, data sheets, clipboard and pencils, long measuring tape (100 meters), flags/stakes to mark transects, GPS device, this procedure and pollinator identification guide
- **Personnel needed**—Can be conducted by a single person, but easier with two
- **Workbooks**—Bee Abundance Monitoring Procedure workbook

This streamlined monitoring procedure provides a quantitative measurement of bee abundance and can be conducted with minimal time and training, especially when the surveyor is able to recognize bees from other insects visiting flowers, and distinguish honey bees from native bees (see [Native Revegetation Resource Library guide](#) for more information about this level of identification). Two separate visits are made in a growing season to conduct a timed assessment to observe and count bees on flowers. During each visit, observations are made along transects. Surveys are conducted in the middle of the growing season, at least one month apart. In California, for example, May through July is the ideal survey window, while in the Upper Midwest or Northeast, July through August is preferred. Monitoring is conducted only when weather conditions are warm, sunny, and calm, with air temperatures over 60° F, wind speeds less than 10 miles per hour, and skies mostly clear. Pollinators are most active between 10 a.m. and 4 p.m.

This procedure can be implemented with one person, but it is easier with a two-person team (Figure 6-17). When using two people, one person observes and counts bees while the other records the data and tracks the time and weather conditions. A field data and analysis form in the Bee Abundance Monitoring procedure workbook can be downloaded [here](#).

Bee counts are made along multiple transects perpendicular to the road across the entire length of a roadside sampling area, as described in [Section 6.3.6](#). The bee counts can be conducted on the same transects and at the same time that Soil Cover, Species Cover, and Species Presence procedures ([Section 6.3.1](#), [Section 6.3.2](#) and [Section 6.3.3](#)) are conducted. However, the bee abundance monitoring is conducted prior to the Soil Cover, Species Cover, or Species Presence procedures to reduce disturbance that could disrupt bee activity.

For each transect, a measuring tape is laid out from the edge of the road shoulder to the outer roadside edge (Figure 6-18). The length of each transect determines the time spent

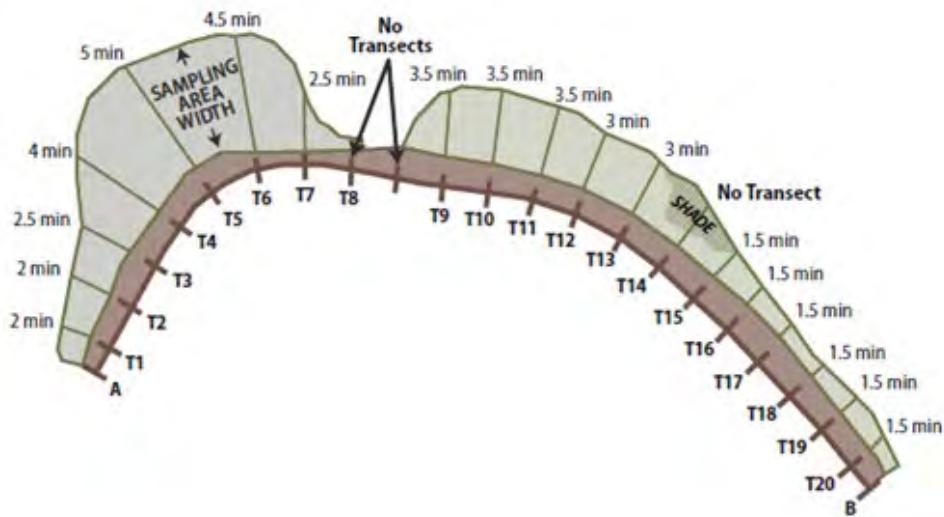


**Figure 6-17 | Two-person monitoring teams**

With a two-person monitoring team, one person counts pollinators and the other records the data and tracks time.

*Photo credit: Michael Kent, Cape Atlantic Conservation District*

counting bees on each transect. Plan on spending approximately one minute counting bees for every 10 feet of transect. For example, if the transect length is 35 feet, then time spent counting bees for that transect would be 3.5 minutes. To reduce the disturbance to bees prior to sampling, only one person lays out the measuring tape and then stays clear of the tape while walking back to the beginning of the transect.



**Figure 6-18 / Transect layout on roadsides for Bee Abundance monitoring procedures**

Multiple transects for pollinator surveys are laid out similarly to those described in Section 6.3.6 for each roadside sampling area. In this example, survey times are based on 1 minute per 10 feet of transect. Transects that are shaded from the sun are not sampled, nor are transects that are less than 10 feet.

Transects typically start from the road and end at the outer edge of the roadside. In situations where the shadow of the surveyor is directly in front of the direction of travel, the transect recording begins at the outer edge of the roadside and travels to the road. If the entire transect is shaded, then this is noted on the data sheet. Bee activity declines in shaded areas.

At the start of each transect, the data recorder starts the stopwatch and the observer begins to count bees, covering about 10 feet of the transect per minute. The observer chooses the side of the transect with the least shadow and observes the flowers within 3 feet of the transect line. While walking transects, each bee that lands on the reproductive structures of the flower is recorded. Bees that are perching on leaves or petals are not counted. If performing the monitoring with just one person, a timer is used and the timer is paused as needed to record a bee. The timer is restarted when observations resume.

While moving along the transect, a tally of the number of native bees and honey bees is recorded. The same individual bee is not counted twice, even if it visits multiple flowers, because the overall goal is to count the number of individuals using the site rather than the rate of floral visitation.

At the end of each transect, the recorder tallies the number of pollinators, the travel time, time of day, the length of transect, transect number, and weather conditions. If other monitoring procedures (e.g., Soil Cover, Species Cover, and Species Presence) are to be conducted, the team returns to the beginning of the transect and conducts these procedures.

Records of roadside sampling areas also include the site name, sampling area designation, date, name of the sampler, weather conditions, time of visit to the site, and any pertinent notes about the site. It is also valuable to record the plants that are blooming along the transect at the time of the survey, as well as to collect data on the floral associations of the bees at the site. The data sheet is available at the website mentioned previously. This procedure was adapted from a procedure developed by researchers with the University of California, Davis; Rutgers University; Michigan State University; and The

Xerces Society (Ward et al 2014).

## Data Analysis

To determine overall abundance per roadside sampling area, the number of individual bees observed during each monitoring event is tallied. Then, the monitoring events per year are averaged. To determine the number of honey bees and the number of native bees per transect-foot, the total number of bee counts of all transects in a field visit is summarized and divided by the total transect distance. This number can be used to compare with other field monitoring visits.

A second analysis method is based on the statistical methods outlined in [Section 6.3.7](#). Using this method, field data is entered into the [Bee Abundance Monitoring procedure workbook](#), which calculates mean and confidence intervals for honey bees and native bees for each sampling area. Confidence intervals can be used to determine if there are significant changes in populations between sampling areas or dates. This analysis can also be used to correlate findings from Soil Cover, Species Cover, and Species Presence procedures. For example, using data from each transect, a regression analysis of bee populations to floral density ([Section 6.3.2](#)) could be analyzed. Other regressions could include percent bare soil, specific plant species, and species richness.

The number of native bees counted through this monitoring procedure correlates positively with the overall diversity of native bees at a site; therefore, if a large abundance of individuals is present, that indicates higher diversity (Ward et al 2014). If multiple roadside sampling areas are surveyed, the differences observed in native bee abundance reflect differences in habitat quality among sites. Native bee counts can be used to rank the quality of sites or the quality of seed mixes, or can be used over time to assess changes at a site. Note that honeybee abundance cannot provide a measure of the value of habitat for native bees or other pollinators because the number of individual honey bees visiting habitat is determined by the number of managed hives in the vicinity.

### 6.4.2 BEE AND BUTTERFLY DIVERSITY PROCEDURE

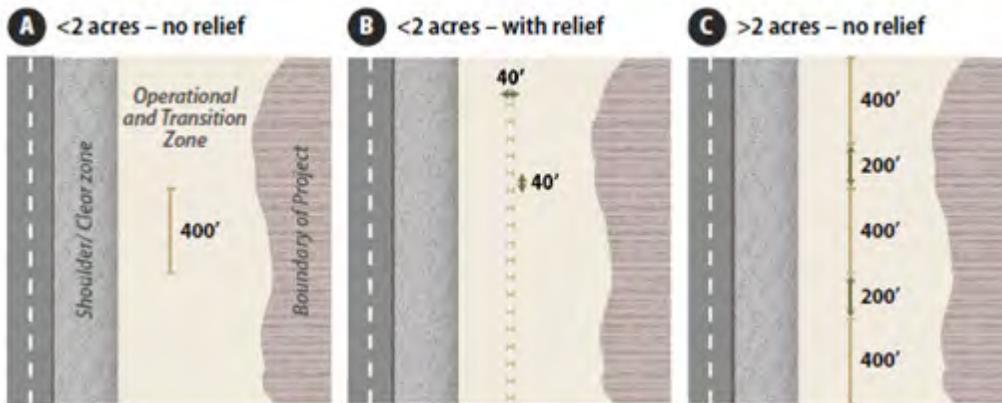
#### Overview of procedure

- **What the procedure will measure**—Abundance and richness of bees and butterflies
- **Sampling design**—Transects within the project area
- **Sampling frequency**—Three visits per growing season, at a minimum—late spring, mid-summer, late summer
- **Sampling timing**—Warm, sunny, and calm days, between 10 a.m. and 4 p.m.
- **Level of identification needed**—Identify morphological groups (morphogroups) of bees and groups of butterflies
- **Personnel needed**—Two people: one person to identify pollinators, one to keep time and record data
- **Equipment needed**—Stopwatch or clock, data sheets, clipboard and pencils, long measuring tape (100 meters), flags/stakes to mark transects, GPS device, this procedure and pollinator identification guide, and plant lists and plant identification guide ([see Native Revegetation Resource Library](#))
- **Workbooks**—Bee and Butterfly Diversity Monitoring Procedure workbook ([see Native Revegetation Resource Library](#))

## Procedure Details

This procedure provides measurements of bee and butterfly abundance, as well as the number of bee species (richness). It can be challenging to identify pollinators. For example, most native bee species can only be identified to species by examining pinned specimens under the microscope and obtaining confirmation from experts. In fact, standard pollinator monitoring techniques typically employ destructive sampling, with specimens collected with nests or traps, pinned, and then identified by a taxonomist. While these collection techniques provide the most robust data, they can be labor intensive, time consuming, and expensive. Collecting observational data on pollinators is an economical and effective alternative to monitoring pollinators, as long as the observers have had some training and practice in order to recognize and monitor pollinators. This procedure can be used to collect consistent observational data on bee and butterfly communities. The procedure is based on a standardized method designed by researchers and practitioners (Kremen et al 2011; Minnerath et al 2016).

The goal of this procedure is to identify specific associations between pollinators and their habitat. Monitoring takes place along transects, which are located within sampling units that have relatively uniform vegetation and site characteristics (Figure 6-19). If the project is large or composed of several different plant communities, then several sampling units may be delineated and sampled separately. The number of transects placed in a sampling unit depends on the size of the unit and topographic relief:



**Figure 6-19 | Transect layout on roadsides for Bee and Butterfly Diversity monitoring procedures**

- **<2 acres – no relief**—establish one 400 feet transect through the middle of the area outside of the clear zone, running parallel to the road (Figure 6-19A)
- **<2 acres – with relief**—orient multiple transects of equal length that add up to a total of 400 ft. running perpendicular to road (for ease of walking the transect) and spaced apart evenly. Figure 6-19B shows a sampling area where 10 transects averaging 40 feet in length were laid out for a total of 400 feet.
- **>2 acres – no relief**—establish three 400 feet transect through the middle of the area outside of the clear zone, running parallel to the road separated evenly (Figure 6-19C)
- **>2 acres – with relief**—orient multiple transects of equal length that add up to a total of 1,200 ft. running perpendicular to road (for ease of walking the transect) and spaced apart evenly.

If making comparisons between sites, it is best to keep the individual transect length and the distances between transects constant. For sites that are greater than 2 acres in size, three 400-foot-long transects are used that are spaced evenly throughout the site. Transects are located in full sun where possible (or notations are made on data sheets when in shade) because pollinator activity declines in shade. GPS coordinates are taken to allow others to monitor the area in the future.

Monitoring is conducted at least three times a growing season, ideally monthly. This is because many bee and butterfly species fly for a limited number of weeks each growing season, and the communities may differ greatly between samplings. It is important to be consistent from year to year in the frequency of monitoring so that comparisons between years can be made.

Weather conditions strongly influence bee and butterfly behavior. Bees avoid activity on cold, windy, or overcast days, so monitoring is optimal when sampling on days with air temperatures over 60° F, wind speeds less than 10 miles per hour, and skies mostly clear. Standardizing the time of day that sampling occurs is also important. Pollinators are most active between 10 a.m. and 4 p.m.

These procedures can be implemented by observers working on their own or in pairs, with one person to act as an observer and the other to record observations and data. It is easiest to conduct observations of bees and butterflies on separate walks of the transects to help focus identification.

When making observations of floral visitors in a transect best practices include:

- Observing Insects carefully and only identified to a level at which the surveyor is confident.
- Leaving insects visiting flowers undisturbed before an observation is made. Walking slowly and avoiding sudden movements can help. Since insects will respond to shadows passing overhead, surveyors may try to walk so as not to cast a shadow where observations are to be made.

### Bee Monitoring

The transect is walked at a steady pace, such as 10 feet per minute; expect each 400-foot-transect to take 40 to 60 minutes. The surveyor walks slowly but consistently, not spending more than a couple of observational minutes at any flower or group of flowers. Consistency in time spent walking each transect allows for better comparisons between samples.

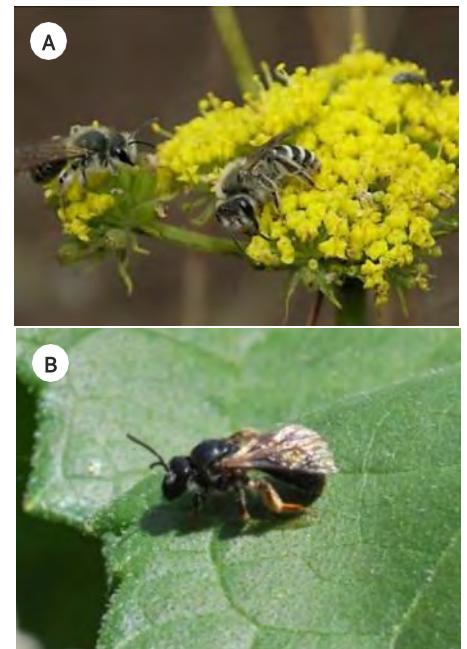
Only bees that are on the reproductive parts of the flower are identified and recorded (Figure 6-20). Bees sitting on petals, leaves, or in flight are not recorded. When a bee is visiting a flower, the bee is observed and identified using the identification guides presented in the [Native Revegetation Resource Library](#). It can also be helpful to note other floral visitors beyond bees, and the species of the flower on which the bees are observed. If more than one floral visitor is observed on a single flower, the number of visitors is noted first and then they are identified. After the bee data is collected, each additional flower species that is in bloom but did not have floral visitors during the survey is noted.

### Butterfly Monitoring

This monitoring procedure is based on the standardized Pollard Walk (Pollard et al 1975) and can be used to collect observational data on the abundance and richness of butterflies. The objective of this procedure is not to count all butterflies present at the entire site or within the habitat being monitored, but to count those individuals that occur in, or move through, the transect's sampling area while the surveyor is walking a steady pace of 10 feet per minute. The surveyor identifies and counts butterflies on flowers or in flight if they occur within approximately 15 feet on each side of the transect or overhead of the surveyor. Butterflies that fly in from behind the surveyor are not counted to avoid counting the same individual twice. Butterflies are identified to the level most comfortable to the surveyor (refer to butterfly identification guide) and recorded as whether the butterfly was observed in flight or nectaring on a flower. If the butterfly is nectaring, it can be helpful to record the species of the plant on which the butterfly was feeding.

Considerations include:

- It is important to be consistent, using the same level of effort with each time



**Figure 6-20 | Recording bee visits**

When recording bee visits, only bees present on reproductive parts of flowering plants (A) but not on leaves, petals, or in flight (B) are recorded.

*Photo credit: Mace Vaughan, Xerces Society (A), Sara Morris, Xerces Society (B)*

transect.

- The time spent walking the transect is recorded each time.
- It is recommended that records of observations include the site name, date, name of the sampler, weather conditions, time of visit to the site, and any pertinent notes about the site.
- It is also valuable to record the plants that are blooming along the transect at the time of the survey, as well as to collect data on the floral associations of the pollinators at the site.

## Data Analysis

To draw meaningful conclusions about the effects of roadside habitat on bees and butterflies, data is collected consistently over time in the same areas. To determine changes in abundance over time, the number of individuals observed during each monitoring event is tallied (transects can be combined if multiple transects are sampled). These numbers can be averaged for each year (e.g., the results from late spring, mid-summer, and late summer can be averaged) and plotted on a graph or table to show changes between years. The numbers of individuals can also be totaled for each monitoring event and evaluated separately. Honey bee counts may be excluded in some analysis because the placement of hives influences their abundance.

To calculate species richness for each sampling period or each year, the total number of bee morphogroups and butterfly groups that were observed are tallied. Richness can then be plotted to record changes over time, whether comparing the average or total number of groups observed across all sample periods for each year, or comparing the total number of groups observed during a certain sampling period from year to year.

As noted before, pollinator populations vary over the course of the growing season and from year to year, so it is important to schedule monitoring accordingly. Pollinator populations also vary annually, increasing as plants become established and mature, which may take several years after seeding or planting. For this reason, monitoring is conducted for several years or more after the project has been completed, ideally 3 to 5 years. The longer sites are surveyed, the more meaningful the results.

### 6.4.3 MONARCH BUTTERFLY REPRODUCTION AND HABITAT PROCEDURE

#### Overview of Procedure

- **What the procedure will measure**—abundance of milkweed plants, abundance of nectar plants in bloom, abundance of monarch eggs and larvae
- **Sampling design**—Transects within the project area, with uniformly spaced quadrats along each transect
- **Sampling frequency**—Once a growing season during the breeding season of monarchs, at a minimum
- **Sampling timing**—Warm, sunny, and calm days between 10 a.m. and 4 p.m.
- **Level of identification needed**—Recognize monarch butterfly eggs and larvae, identify milkweed species, identify target nectar plants
- **Personnel needed**—Two people: one person to identify plants, one to search for monarch eggs and larvae
- **Equipment needed**—Stopwatch or clock, data sheets, clipboard and pencils, long measuring tape (100 meters), flags/stakes to mark transects, GPS device, 1m x 1m quadrat, this procedure and **monarch egg and larval identification guides**, and

plant lists and plant identification guide (see Native Revegetation Resource Library)

- **Workbooks**—Monarch Butterfly Monitoring Procedure (see Native Revegetation Resource Library)

## Procedure Details

Monarch butterfly populations have recently declined to dangerously low levels and are being considered for listing under the Endangered Species Act. If the project area is within the distribution of the monarch butterfly, it may be important to conduct monarch monitoring. Monarch butterflies have a range that extends across most the United States (exceptions include western Washington and parts of western Oregon where their host plants do not occur) and into southern Canada and northern Mexico.

This procedure outlines how to measure (1) abundance of milkweed, (2) abundance of nectar plants, and (3) density of monarch eggs and larvae. Measurements are made along multiple transects perpendicular to the road across the entire length of a roadside, as described in [Section 6.3.6 \(see Linear Areas\)](#) and as shown in Figure 6-18. This procedure can be conducted on the same transects and at the same time that Soil Cover ([Section 6.3.1](#)), Species Cover ([Section 6.3.2](#)), and Species Presence procedures ([Section 6.3.3](#)) are conducted. However, the monarch egg and larval monitoring is conducted prior to the Soil Cover, Species Cover, or Species Presence procedures to reduce disturbance that could dislodge caterpillars from milkweed plants.

For each transect, a measuring tape is laid out from the edge of the road shoulder to the outer roadside edge. Vegetation is sampled at quadrats spaced every 20 feet along the transects. Milkweeds, nectar sources, and monarch eggs and larvae are sampled within each quadrat. A 1m x 1m quadrat frame is constructed with PVC pipe and corner connectors.

A workbook that contains data forms for milkweed abundance, nectar plant abundance, and monarch eggs and larvae density is available in the [Native Revegetation Resource Library](#).

## Milkweed Abundance

The number of plants and stems of milkweed species that are rooted within the quadrat are recorded. A milkweed plant is defined as all above-ground stems originating from a single identifiable common point in the ground. One milkweed plant may be composed of one or multiple stems, depending on the species. For example, green antelopehorn milkweed plants (*Asclepias viridis*), butterfly milkweed (*A. tuberosa*), or swamp milkweed (*A. incarnata*) may grow multiple stems per plant. When these plants are found, each cluster of stems originating from a single point is counted as a single plant (Figure 6-21). Multiple stalks from a single point are recorded as a single plant with multiple stems. Some species, such as common milkweed (*A. syriaca*) or whorled milkweed (*A. verticillata*), grow multiple stalks, and it is impossible to discern if the stalks are from the same plant without excavating the roots. Each individual stalk that does not originate from a central point and is separated by soil is therefore counted as a separate plant. Data on the number of plants, as well as the number of stems per plant, is recorded to determine milkweed density per area.

## Nectar Plant Abundance

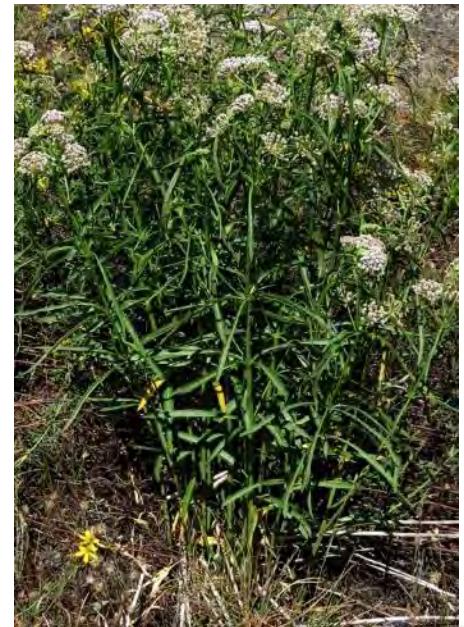
Within each quadrat, the species with blooming flowers and the number of plants is recorded. After monitoring is completed, at the end of the growing season, gaps in bloom or times when species diversity was low can be identified.

## Density of Monarch Eggs and Larvae

To determine the monarch density per plant, monarch butterflies are recorded within each quadrat and within a 10-foot swath between quadrats. Each milkweed that is encountered within a quadrat is examined for monarch eggs and larvae. The numbers of eggs and larvae are recorded for each quadrat. For plants within the 10-foot swath between quadrats, the number of milkweed plants and stems by species and the number of eggs and larvae observed on these stems are recorded separately (e.g., quadrat 3—quadrat 4). The collected data is used to determine the monarch density at the site, calculated as a proportion of milkweed plants with monarchs.

All parts of the milkweed plant are carefully examined, including the bottoms of the leaves, the area within the small leaves at the top of the plant, and buds and flowers if they are present. Eggs are very tiny, about 1 mm wide, and are cream colored (Figure 6-22). Early instars of caterpillars are also very small, 2–6 mm in length. Monarch caterpillars leave clues such as chew marks on the leaves that indicate their presence (Figure 6-23). It is important to handle the plants carefully to avoid dislodging any larvae. Photographs of the egg and caterpillar are helpful to distinguish monarchs from other insects. Monarch eggs and larvae can be hard to find.

These procedures were adapted and modified from the Monarch Conservation Science Partnership Monarch Monitoring Trial Protocol, the Monarch Larva Monitoring Protocol, and the Prairie Reconstruction Initiative Retrospective Research protocol (personal communications: Laura Lukens, University of Minnesota, January 3, 2017; Diane Larson, United States Geological Survey, January 6, 2017).



**Figure 6-21 | Milkweeds may have multiple stems from one plant**

Milkweeds with multiple stems are recorded as a one plant.

## Figure 6-22 | Monarch butterfly eggs and caterpillars

Monarch eggs are often deposited singly on the underside of milkweed leaves (A), and early instar caterpillars can be found on milkweed vegetation or on flowers (B).

*Photo credits: Stephanie McKnight/Xerces Society (A); Anne Stine/Xerces Society (B)*



#### 6.4.4 POLLINATOR PLANT MONITORING

Monitoring plant species found on a project site is a method of assessing potential pollinator habitat. This involves collecting and evaluating data from any or all plant monitoring procedures outlined in [Section 6.3](#). To do this, plant species recorded during these surveys are categorized as to whether they are pollen or nectar producers, flowering times, plant structure, or any other characteristics important for pollinator habitat. Some of these plant characteristics are available on the ERA.

DFC targets developed during planning can be referred to for grouping plant species. For example, a DFC target may be “50 percent of the plant cover will be composed of flowering species.” Using the Species Cover monitoring procedure ([Section 6.3.2](#)), plant species are grouped as “flowering” and “non-flowering,” and the percent cover for each group is summarized. Using the analysis procedures described in [Section 6.3.7](#) it could be determined statistically whether the DFC target was achieved.

DFC targets can also include overlapping bloom time throughout the growing season so that flowering species are available sequentially for pollinators.

### 6.5 PHOTO POINT MONITORING PROCEDURES

Photo point monitoring is a method of recording landscape changes in vegetation over time and showing the success or failure of a revegetation project. Photo monitoring can be established for long-term monitoring where a permanent point has not been established and subsequent photographs are taken ([Section 6.5.1](#)) or where permanent locations have not been located from historic photographs or photo monitoring can be reconstructed from historic photographs ([Section 6.5.2](#)).



**Figure 6-23 / Chew marks on milkweed leaves indicate the presence of monarch butterflies**

An indication of the presence of monarch butterflies is the chew marks on milkweed leaves.

Landscape images from photo point monitoring, in conjunction with data collected from other monitoring methods, can be a powerful way to describe the results of a revegetation project. *Hall's Photo Point Monitoring Handbook* (2002a, 2002b) provides a thorough coverage of this subject.

## 6.5.1 ESTABLISHING PHOTO POINTS FOR LONG-TERM MONITORING

### Locating Photo Points

The first step in establishing a long-term photo-monitoring point is to determine where the photo will be taken. The locations of the photo point are often based on the objectives of the revegetation project. For example, if the objectives for revegetation are visual enhancement, photo points are located where motorists view the vegetation of interest from the road. With time, small shrub and tree seedlings planted along roadsides will fill in the entire picture frame, obscuring any long-distance views. If it is important to photograph long-distance views, then the location and direction of the camera are placed where the foreground is free of potentially tall growing vegetation.

Photo point locations are recorded with GPS coordinates and described in detail so they can easily be found years later by other personnel. Permanent structures that can be easily described and located by others make good reference points for photo locations. These include large boulders, culvert inlet and outlet, concrete headwalls, drain inlet frames or concrete aprons, topographical features, guardrail posts, permanent survey markers, mile post markers, road signs, road intersections, railroad lines, telephone poles, and other types of utility structures. Since these features are not always permanent, it is important to locate more than one structure in the field.

The photograph is either taken from a permanent structure or from a measured distance and direction from the structure. For example, a photographer might decide to take a picture on the fog line directly above a culvert outlet. This spot would be easy to describe and locate in the field by another person several years later. A photographer might also locate a photo point using a sign post as a permanent reference. In this case, an azimuth reading and measured distance from the sign post would be recorded in the notes.

A common method for locating a photo point is to drive a stake (rebar) at the spot. This method works in some cases but has drawbacks: the stake may interfere with maintenance activities, such as mowing or traffic safety if it is not set back far enough from the roadway; the stake is driven several feet into the soil, which is difficult on most cut slopes and some fill slopes; and the stake is identified with a tag that can be readable over time. One problem associated with poorly placed stakes is that they are often displaced through active soil movement, animal traffic, vandalism, or road maintenance activities. Using a more permanent feature of the roadside, such as a concrete structure, guardrail, or signpost, is generally a better alternative.

### The Camera

When using a digital camera for photo point monitoring, the highest resolution is used. Using cameras with good lens quality is also essential for optimum photographs. The camera is situated so that the zoom lens does not have to be used. The zoom lens can pose problems in reproducing the original photograph (Hall 2002b) and also result in lower picture resolution.

### Taking the First Picture

The camera is set on a tripod over the identified photo point location and the image is previewed on the LCD screen or through the lens. With a compass aligned to the side of the camera, an azimuth reading is taken. Using a clinometer on the top of the camera, the degrees from horizontal are recorded. A log or electronic document is kept that includes the following information about each photograph:

- Project name

Roadside Revegetation: An Integrated Approach to Establishing Native Plants and Pollinator Habitat

**Figure 6-24 | Establishing photo point locations from historic photographs**

To relocate the photo point from the original photograph (A), a large print was made and taken to the general location in the field. Since the original photograph was taken from the paved road surface on the inside of the guardrail, there was little doubt that the elevation of the second photograph (B) would be approximately the same.

Locating the original photo point along the guardrail was accomplished by establishing a relatively vertical centerline connecting two identifiable reference points on both the original photograph (A) and the photograph to be taken (B). In this example, the centerline is drawn from the left side of the culvert (1) to the 12th post of the guardrail (2) from the reflector stake (3). Picture B is a close approximation to the photo point location of photo A, but notice that the centerline is not quite in the same direction as in photo A. To find the more exact location, the photographer would move to the right along the inside of the guardrail until the centerline lines up in the same direction as in photograph A.



Photo credits: David Steinfeld

- Photograph ID
- Date
- Time
- Photographer
- GPS location
- Location on plan map
- Location description
- Azimuth of camera
- Angle of camera
- Manufacturer and model of camera
- Large imported jpeg image of photograph

## Taking Subsequent Photographs

At later dates, the photo point is relocated using the map, GPS point, and/or location description. The camera azimuth and vertical angle are set, and the image is viewed through the LCD screen or lens and compared to the print of the original photograph. If the original spot cannot be located, then relocating the photo point can be attempted using the procedures outlined in the following section.

### 6.5.2 ESTABLISHING PHOTO POINTS FROM HISTORIC PHOTOGRAPHS

If no records exist for the original photo locations, then a close approximation can be determined (after Hall 2002a). Prior to monitoring, all historic photographs of interest are printed out in large format. The general locations along the road corridor where these photographs may have been taken are located. Based on the photograph, permanent features, such as guardrails, culverts, trees, shrubs, road cuts, and fills, are found. The general area is walked while comparing the large photograph with the surrounding area. Finding several permanent reference points in the distance, such as mature trees or large rocks, and moving until all features are positioned similar to the photograph is a quick means of establishing a general location for the original photograph. Figure 6-24 shows how a photo point was located using guardrails and a culvert as reference points.

## 6.6 DEVELOPING A MONITORING REPORT

The purpose of a monitoring report is to document how well a revegetation plan was implemented. This report is written and shared so that corrective measures can be made and that the lessons learned from implementing the revegetation project can be applied to future projects. Most projects fail to deliver a monitoring report for the following reasons:

- The monitoring plan was too complicated (massive amounts of extraneous data were generated)
- The monitoring was not designed in a meaningful or statistical manner
- The monitoring objectives were poorly stated
- Insufficient data were collected to draw any meaningful conclusions
- Data were lost

## 6—MONITORING

More than likely, the main reasons were that “there wasn’t enough time” or “there were more important things to get done.” In other words, writing a monitoring report often is not completed because it does not seem important at the time. It stands to reason, however, if time was taken to collect field data, time also be given to analyze and present the findings.

The value of a monitoring report is that it is often the only record of what was done on a revegetation project and how well it was executed. It is a statement to management as to whether revegetation objectives were met. It can guide revegetation specialists and road managers to improve revegetation methods and reduce costs on future projects.

The monitoring report does not have to be long. In fact, a two- to three-page report summarizing the important findings is often sufficient for most projects. That is also the appropriate length of a report that most people have time to read. The details of the data collection and analysis can be included in an appendix. Examples of several monitoring reports can be found [here](#) by entering “Plans and Reports” in the Report Type dropdown menu and “Monitoring” in the Topic Type dropdown menu.

Every monitoring report will be different, but most reports address the following questions in some form:

- Who did the monitoring?
- When did monitoring occur?
- Where in the project was monitoring conducted?
- What was monitored?
- How was it monitored?
- How was it analyzed?

The above questions were addressed in the monitoring plan, facilitating their documentation. The report then answers these questions:

- Were the objectives met?
- Are corrective actions needed?
- What lessons were learned?
- Is there further monitoring that needs to occur?

Many monitoring reports are followed by appendices that contain some or all of the following:

- Maps
- Data analysis
- Photo point monitoring
- Project diaries—A detailed account of all activities that took place during the project
- A summary of the revegetation treatments or activities that occurred on the project

Revegetating highly disturbed sites with native plants to maintain or increase pollinator habitat is a relatively new field of study. Well-designed and executed monitoring projects can provide useful information to a wider audience of practitioners, designers, scientists, and managers working in this field. Conferences, societies, newsletters, journals, and trade publications are some venues to share this knowledge. The small-scale trials that were tested during implementation of a revegetation plan (e.g., different rates of fertilizer, tackifier, seeds, and hydromulch) are likely of great interest to other designers. By taking

## 6—MONITORING

the time to share monitoring results, the science and practice of revegetating highly disturbed sites can be advanced and future revegetation projects can be improved.

# 7—Operations & Maintenance

- 7.1   Introduction**
- 7.2   Decision Process for Treating Unwanted Vegetation**
- 7.3   Vegetation Treatment Options**
- 7.4   Prevention**
- 7.5   Protection**

## 7.1 INTRODUCTION

Roadside maintenance is the final step in the revegetation timeline. It occurs when the revegetation efforts and responsibilities are transferred to the road maintenance staff for the long-term management of roadside vegetation. If the transfer is successful, revegetation objectives are carried out for many years with good results. The intended audience for this chapter is maintenance staff. In handing off a roadside revegetation project, it is helpful if the objectives and reasoning behind the revegetation plan are understood by maintenance staff. This may have occurred if maintenance staff were included during the planning phases of the revegetation plan. In such instances, maintenance staff can provide insights into the development of a vegetation management strategy (Section 3.2) and bring to the discussion a thorough understanding of how vegetation responds to various management activities specific to the project environment.

Many state DOTs have a statewide Integrated Vegetation Management (IVM) plan or an Integrated Roadside Vegetation Management (IRVM) plan. Some states have more detailed IVM plans for regions within the state. IVM plans are typically updated annually based on monitoring of the previous year's work.

The IRVM is “an approach to right-of-way maintenance that combines an array of management techniques with sound ecological principles to establish and maintain safe, healthy and functional roadsides” (Brant et al 2015). It applies many of the Integrated Pest Management concepts developed for agriculture, horticulture, and forestry to roadside vegetation management. IVM plans typically include sections on preventing the introduction of unwanted vegetation, protecting rare plants, preserving natural areas, and developing a decision-making process for treating vegetation.

## 7.2 DECISION PROCESS FOR TREATING UNWANTED VEGETATION

Most states have a decision-making process within the IVM for treating unwanted vegetation. The process includes some or all the steps outlined in Figure 7-1. An IVM decision-making process can be used to evaluate the control of an individual target weed species (e.g., noxious weed control) or control the vegetation on an entire section of roadside. In this process, roadsides are inventoried and objectives are set for unwanted vegetation. All methods for treating unwanted vegetation are considered, including no action, mowing, herbicide application, manual removal, prescribed burns, grazing, and biological control. A treatment plan is developed based on a thorough understanding of a species or plant community. Treatment plans often outline the “action threshold” or “tolerance level” that, when exceeded, constitutes the implementation of a vegetation treatment. The effectiveness of the treatment is determined based on monitoring. If the treatment is ineffective, then the treatment options are reviewed and updated, if appropriate. If the treatments are effective, then no action is taken and monitoring continues.

**For the Designer**  
To ensure that revegetation objectives are carried through during the maintenance phase, maintenance staff can be informed of these objectives and how they relate to maintenance activities.



**Figure 7-1 /**  
**Components of a**  
**decision-making**  
**process for treating**  
**unwanted vegetation**

## 7.2.1 INVENTORY OF ROADSIDES

Roadsides are inventoried for weeds or unwanted vegetation. For some states, invasive and noxious weeds are reported using GPS equipment and GIS mapping technology. The associated maps often prioritize sections of road that require some form of vegetation control. Roadside inventories can also show areas of desirable vegetation, rare plants, threatened species or populations, natural areas, pollinator habitat, and vegetative types. Vegetation assessments conducted during the planning stages of a revegetation plan may be useful in inventorying roadside vegetation by identifying target weed species and prioritizing roadsides for treatment (Section 3.6.1).

## 7.2.2 DEFINING ROADSIDE OBJECTIVES

The backbone of a treatment plan is composed of clearly stated road maintenance objectives. These include objectives for road safety as well as roadside resources. Objectives are set at a statewide level in the IVM plan or at the local or regional level within a state.

Many states assign maintenance objectives to roadside zones similar to those shown in Figure 3-8. Oregon and Washington, for example, identify three roadside zones (WSDOT 2016, Oregon DOT 2013):

- **Zone 1**—This zone borders the roadway pavement with vegetative objectives of preventing pavement breakup, preventing noxious weeds, facilitating a “soft return” to the travel lane should a driver accidentally veer, and maintaining maximum visibility.
- **Zone 2**—This zone is spatially located between zone 1 and zone 3, and the vegetative objectives are to maintain low growing vegetation for maximum sight distance, enhance visual qualities, and maintain hydraulically functioning ditches. Pollinator habitat can also be an objective.
- **Zone 3**—This zone is farthest from the roadway and has fewer restrictions for maintaining low-growing vegetation for sight distance. These areas may contain shrubs and trees, and can serve as pollinator habitat.

Each of these zones is treated differently and may have separate action thresholds, treatment plans, monitoring activities, and maintenance schedules.

Roadside objectives are also developed for the important resources associated with roadsides. These resources include pollinator habitat, water, natural areas, and visual resources. These objectives are brought forth from the prevention and protection portions of the plan (Section 7.3 and Section 7.4) of the IVM. It is important that resources, safety, and road maintenance objectives are clearly stated so that treatment options can be developed accordingly.

It is important to note that state departments of transportation manage areas that are not roadway or right-of-way. These areas include rest areas, “back 40” property, stockpile lots, maintenance yards, office grounds, bike paths, scenic viewpoints, and points of interest pull-outs. Management objectives for these areas may integrate well with creating or improving pollinator habitats because alternative uses are unlikely to conflict with maintenance activities or expose people (maintenance staff or volunteers) to the dangers of the roadside.

## 7.2.3 EVALUATING TREATMENT OPTIONS

In this step of the decision-making process, all possible treatments are evaluated for how each best meets the road objectives. The details of each vegetation treatment are described in Section 7.3. In evaluating each treatment, it is important to consider how well each treatment meets the roadside safety, maintenance, and resource objectives. If there are conflicts, how can the treatment be

modified?

To evaluate treatments, it is helpful to have a working knowledge of the target weed species or plant community of concern. It is also helpful to understand how a weed species or an entire plant community changes over time through succession, how they respond to various disturbances, how they can be controlled or maintained, and how treatments change the direction and rate of plant succession (Figure 3-9). Some of this information can be obtained from the vegetation management strategy outlined in the revegetation plan or from the designers of the revegetation plan ([Section 3.2](#)). Adjacent maintenance departments, land owners, and local agricultural extension specialists are good contacts to discuss the best controls for the target weed species present on the project site. Plant Guides located on the profile page of the [USDA PLANTS](#) database provide control treatments for many weed species. Other resources include a publication by Harper-Lore et al (2013) that describes how to control 40 common weed species in the United States.

In evaluating treatments, it is also important to understand how each treatment affects roadside resources. Pollinators are affected by most vegetation treatment methods, and a knowledge of the important pollinators on the roadsides, their life history, and their habitat requirements can help in developing the appropriate treatments. Water quality can be affected by vegetation maintenance, especially where bare soil is created near drainages and stream courses. Reviewing the Storm Water Pollution Prevention Plan for the project area or road corridor, if it is available, can help the designer identify appropriate treatments.

#### 7.2.4 ESTABLISHING A VEGETATION TREATMENT PLAN

The vegetation treatment plan prioritizes the treatments that best meet the road objectives. It addresses the action threshold for treatment implementation to control a target weed species or a plant community on a section of roadside. The action thresholds may be based on the phenological condition of the vegetation (e.g., treat when average grass height is 2 feet), presences of a noxious weed (e.g., control all noxious weeds no later than flowering stage), and safety threat (e.g., trim or remove trees when they are considered hazardous to road or neighboring property). When the action threshold is exceeded, the plan details the treatment that will be used (e.g., timing, rates, equipment, personnel).

The treatment plan prioritizes the areas to be treated and sets site limitations (e.g., distance from streams or bodies of water, slope steepness, saturated soils). It also addresses the effects of the treatment on other roadside resources and options for mitigating negative effects. For example, the effects of mowing on pollinators and pollinator habitat can be minimized if mowing is done outside the flowering period of most forb species.

#### 7.2.5 MONITORING TREATMENTS

The treatment plan outlines how roadsides will be monitored. Setting up monitoring for operations and maintenance is similar to the steps outlined for vegetation monitoring ([Section 6.2](#)). Monitoring in an IVM plan addresses the following:

- **Purpose**—Determines if the action threshold was exceeded, treatments were applied as prescribed, treatments were effective, or other resources were negatively affected
- **Who**—Identifies personnel or expertise needed
- **What**—Determines what is being monitored (e.g., noxious weed presence, vegetation height)
- **When**—Defines frequency and time of year
- **Where**—Delineates sections of road to be monitored
- **How**—Determines intensity and methodology

- **Logistics**—Timeline, budget, and equipment

Monitoring is the feedback loop for effectively controlling unwanted vegetation over time. As outlined in Figure 7-1, monitoring is conducted to determine if an “action threshold” is exceeded. If it is not, then monitoring continues as scheduled and no treatments are made.

If an action threshold is exceeded, then a treatment is applied according to the treatment plan.

Implementation monitoring during treatment application records whether the treatment was applied as prescribed or if changes were made based on equipment or site factors. Post-treatment monitoring evaluates the effectiveness of the treatment and whether the treatment affected other resources (e.g., pollinator, water quality, visual resources). If the treatment was not effective in controlling unwanted vegetation or other resources were adversely affected, then a reevaluation of the treatment is usually made and, if necessary, the treatment plan can be changed.

## 7.3 VEGETATION TREATMENT OPTIONS

When developing a vegetation treatment plan, it is important that the target vegetation and the objectives outlined in the IRVM plan are understood. It is also important to understand the resources of value that may be affected with a vegetation treatment. These include water quality, wildlife habitat, visual resources, and pollinators. This section describes common vegetation treatments and how they affect these resources, with an emphasis on enhancing pollinators and pollinator habitat.

Because pollinator species can be strongly affected by weed control treatments, it is helpful to know the important pollinator species that inhabit the area during maintenance activities and develop weed treatments that minimize the negative effects on their populations and habitat. This section outlines vegetation management practices that can control target vegetation while supporting pollinators. Additional details are provided in two recent publications from the U.S. Department of Transportation (Hopwood et al 2015, Hopwood et al 2016), which review the best management practices (BMPs) for controlling roadside vegetation while optimizing pollinator habitat and reducing pollinator mortality. In addition, a review of [Section 3.11.6](#), which covers control of unwanted vegetation before and during construction, may be useful.

Given the complexity of developing treatments that control target vegetation while maintaining or improving pollinator habitat, some general pollinator strategies may aid in designing vegetation treatments specific to a project environment. These are outlined in Figure 7-2.

### 7.3.1 NO ACTION

While not taking an action to control vegetation is not a treatment, it is included in this section as a reminder that there are times when vegetation does not need to be treated. Reviewing vegetation management objectives in the IRVM plan, especially how treatments may affect resource objectives such as water quality and pollinator health, may provide justification for the “no action” treatment. On roadsides where rare plant species or at-risk pollinator species are present, taking no-action may be the most prudent approach to protecting species.

### 7.3.2 MOWING

Mowing is frequently used to maintain roadside vegetation by reducing invasive weeds and woody plants, improving driver sight lines, allowing vehicle pull off, and reducing the risk of wildfires. Typically, vegetation in the clear zone, a band of vegetation directly adjacent to the pavement or shoulder, is mowed regularly to keep the vegetation short for drivers who need to regain control of their vehicle and to lower the fuel level for spread of fires. Periodic mowing within the clear zone creates a habitat that is not typically used by pollinators. However, roadside vegetation beyond the

clear zone can support pollinators.

Mowing at certain times can directly kill pollinator eggs or larvae present on the vegetation. It can also indirectly affect adult pollinators by temporarily removing host and flowering plants (e.g., food sources). For these reasons, it is important to carefully time or limit mowing during the growing season. This is especially true if the habitat supports endangered or rare and sensitive pollinator species. Higher mowing frequencies reduce native plant growth, plant diversity (Parr and Way 1988), and the ability of forbs to compete with grasses (Williams *et al.* 2007), and decrease the amount of nectar and pollen present on the roadside. More frequent mowing can also increase the roadkill of pollinators, especially butterflies (Skorka *et al.* 2013). Therefore, minimizing the number of times a roadside is mowed benefits many pollinator species.

Reducing routine mowing of the entire right-of-way (fence to fence) benefits pollinators by allowing wildflowers to bloom and thereby supply nectar and pollen as food. It is best to restrict routine mowing to the clear zone as much as possible, and mow beyond the clear zone only when there are well-defined objectives, such as reducing brush or maintaining lines of sight. Reducing mowing beyond the clear zone to two or fewer times a growing season is best for wildflowers and pollinators. In some regions, mowing can be reduced to once a year per site, every other year, or even every two to three years, depending on the regional intervals of mowing needed to control woody plant encroachment or to reinvigorate populations of wildflowers.

When timed appropriately, mowing can reduce the effects on pollinators and promote plant diversity. It is optimal to delay mowing until autumn or after the first frost if regional constraints allow. When mowing is delayed, butterflies and other pollinators with larvae that reside on vegetation are able to complete their full life cycles, and flowering plants are able to bloom and provide pollen and nectar to pollinators uninterrupted throughout the growing season. If mowing must occur during the growing season, consider selecting a time to mow that balances vegetation management needs (e.g., noxious weed control) with the resource needs of pollinators (e.g., presence of flowers and host plants). For example, mowing at a time that promotes the growth of wildflowers benefits pollinators in the long term. Timing will vary with region. Wildflower growth is promoted by a mid-summer mow in some regions, while in others mowing after spring bloom might be optimal. It is also worth considering varying the season when mowing occurs every few years to increase plant diversity. Mowing consistently at the same time every year will select some plants over others. Plant diversity can be maintained by occasionally varying the timing of mowing, which will favor different plants and prevent certain plants from dominating.

It is important to time mowing to avoid vulnerable stages of the life cycle of any rare or declining species that are present. For example, in Texas, to reduce harm to monarch butterflies, it is best to avoid mowing before March and between May and August, time periods during which monarchs are breeding in the region. Finally, it is helpful to adjust the height of the mower. By mowing vegetation at a height of 12 to 16 inches, vegetation recovers more quickly and plant stress is reduced, particularly during dry periods or drought. Mowing also leaves a greater depth of vegetation for pollinators to use.

There may be a public perception that by reducing mowing, road departments are not taking care of roadsides, which can be an obstacle to implementing mowing strategies that benefit pollinators. Public education may change this perception with time as the public becomes more aware of the benefits of roadsides to pollinator populations and health.

**Figure 7-2 | Considerations for developing a vegetation treatment sensitive to pollinators**

|                             |  |
|-----------------------------|--|
| <b>Timing and Frequency</b> | Treatments are timed to avoid or minimize effects on flowering plants or treatments are timed to maximize plant diversity<br><br>The number of treatments are kept to a minimum to reduce the impacts on pollinators<br><br>Treatments are timed to avoid detrimental effects on breeding or nesting of at-risk pollinators<br><br>Treatments are avoided during periods when floral resources are scarce<br><br>Prescribed fire and grazing are timed carefully to avoid negatively affecting life cycles of imperiled or sensitive pollinators<br><br>Avoiding mowing, non-targeted herbicide applications, burning, and grazing during adult flight periods or when butterfly or moth larvae are feeding on host plants<br><br>Treating perennial weeds with herbicides in late summer and fall when it is most effective<br><br>Applying herbicides when they are most effective — early plant stages<br><br>Applying herbicides when pollinators are least active — before sunrise, after sunset, cool temperatures<br><br>Rotational burns, conducted 3 to 5 years apart, will allow time for pollinator populations to recover<br><br>Varying the timing of broadcast burns from year to year<br><br>Treatments maintain some undisturbed vegetation<br><br>Using of herbicides beyond the safety strip is targeted to noxious and non-native plants<br><br>Grazing is conducted to have minimal trampling impacts that may affect nesting habitat<br><br>Haying is done at the end of the growing season<br><br>Selecting herbicides that are selective to target weeds minimizes the impacts to non-target plants species<br><br>Applying non-selective herbicides when desirable vegetation is dormant<br><br>Using herbicides with low toxicity to pollinators<br><br>Using appropriate equipment and weather conditions to avoid herbicide drift<br><br>Selecting grazers that can be controlled<br><br>Where appropriate, leaving snags and trees with cavities, and down wood for nesting habitat<br><br>Setting mower blades at 12" to 16" will reduce the impact to vegetation structure<br><br>Distance from edge is less than 50 feet - pollinators have places to go for habitat and food during treatments<br><br>Leaving untreated adjacent areas within 100 feet of center of treated area |
| <b>Effect</b>               | Treatments are patchy - <50 percent of the area is treated<br><br>Brush removal is used to soften forest edges and to maintain stems or other sites for tunnel nesting bees<br><br>Leaving sections of the road corridor unburned  |
| <b>Scale</b>                |  |

### 7.3.3 MANUAL REMOVAL

Hand-pulling weeds can be the most complete method of controlling specific weeds because it is highly targeted. Hand-weeding is well suited for removal of weeds that occur in low numbers or that are scattered throughout a site. Hand-weeding is often the least intrusive method of removing weedy species as it has the least effect on pollinators.

Tools that help with hand-weeding include hoes, picks or pulaski axes, trowels, and shovels. Tree and weed puller tools are designed to provide grip and leverage for removing deep-rooted species such as scotch broom. It is important to target weeds during their active growth stages before the weeds have flowered and set seed. For perennial and rhizomatous species, it is best to remove as much of the root material as possible because many plants can re-sprout from root fragments.

One strategy for hand-pulling is the Bradley Method (Brock 2016) which prioritizes the areas to be weeded, beginning in undisturbed areas first, and then working out toward more heavily infested areas. When pulling weeds by hand or with tools, it is useful to minimize soil disturbance so that

Roadside Revegetation: An Integrated Approach to Establishing Native Plants and Pollinator Habitat

#### *For the Designer*

The timing and frequency of mowing can have large effects on pollinators and the vegetation they rely upon and be carefully considered by maintenance staff.

weeds do not become established. It is also important to dispose of weeds in designated areas. If weed removal results in large patches of bare soil, re-colonization by unwanted species can be reduced by seeding with desirable species.

### 7.3.4 HERBICIDES

Herbicides are used to control woody vegetation as well as target weed species on roadsides. The use of herbicides can benefit pollinators by suppressing undesirable plants and encouraging the valuable native plants that provide them with food or shelter. However, used indiscriminately, herbicides can reduce the quality of roadside habitat by removing floral resources and host plants, and may be directly toxic to some pollinators (Mader et al 2010; Russell and Schultz 2010; LaBar and Schultz 2012). Overuse of herbicides can also weaken stands of vegetation, making them more vulnerable to weed invasions, which also indirectly affect pollinators (Inset 7-1). By using herbicides as efficiently as possible, maintenance staff can reduce both the amount applied and the effect on plants that benefit pollinators. Using products selectively, timing applications carefully, and following label directions can increase the effectiveness of herbicide use and decrease impacts to pollinators and other resources.

#### **Inset 7-1 | Weeds and Pollinators**

Some weeds can provide resources for pollinators (e.g., Harmon-Threatt and Kremen 2015). However, nonnative plants typically only support a subset of the overall pollinator community (e.g., Tallamy and Shropshire 2009). Additionally, noxious weeds reduce overall plant diversity, which also reduces pollinator diversity (Memmott and Wasser 2002; Zuefle et al 2008). When noxious and invasive weeds are removed and plant diversity recovers, pollinator abundance and diversity rebounds as well (e.g., Hanula and Horn 2011; Fiedler et al 2012).

It is important to use the appropriate products and application rates that are effective in controlling the target weed species yet have minimal effects on non-target plant species. Whenever possible, the use of selective herbicides—those formulated to control specific weeds or groups of weeds—can reduce damage to nontarget plants. Nonselective herbicides—those that are broad-spectrum and kill or damage all plants—can also be used selectively to reduce effects on nontarget plants. For example, nonselective herbicides can be used selectively by applying them on weeds when desirable native plants are dormant and by using directed or targeted applications (e.g., spot spraying). In addition, understanding how quickly the herbicide being applied degrades on the site ensures that seed germination and restoration planting are not negatively affected. Herbicide labels include information about selectivity and persistence.

Applications can be timed to be most effective based on the herbicide's mode of action and the application technique. For example, when using a systemic herbicide (absorbed by the plant and transported throughout the plant by the vascular system), perennial weeds can be treated in late summer and fall. During this period, perennials begin to move sugars down to their roots, and the herbicide is translocated to vegetative reproductive structures where it is most effective at controlling the plant. Applications of herbicides at the stage of growth when the weed is most vulnerable can make applications most successful. For many weeds, this is the seedling or rosette stage.

Reducing off-site movement of herbicides and the use of nonselective broadcast applications can help avoid damage to non-target plants that provide pollinators with food or shelter. Referring to road inventories of unwanted vegetation (Section 7.2.1), conducting training in weed identification, and using plant identification reference materials to recognize noxious and invasive weeds will help

distinguish these species from similar non-target species. In order to avoid weakening non-target species, weeds can be targeted using spot treatment applications made with a backpack sprayer, weed wiper, or similarly appropriate technology. Using highly targeted applications on cut stems, stumps, or under bark can also reduce unnecessary effects to desirable plants. Broadcast treatments or pellet dispersal are recommended only for dense infestations of weeds or for clear zone or guardrail treatments.

The off-site movement of herbicides can be reduced by selecting appropriate spray equipment, periodically calibrating equipment, and adhering to the pesticide label. Nozzles that produce larger droplets are less likely to cause herbicides to drift off target. Equipment that is calibrated regularly limits over and under applications. It is best to avoid applications when wind speeds are greater than 15 mph and during a temperature inversion (when warmer air above traps cooler air near the ground) when herbicides and other pesticides can linger in the air and move long distances offsite with air movement.

To reduce direct contact exposure to pollinators, herbicides can be applied during a time of day when pollinators are less active. Many pollinators (but not all) are less active before the sun rises or after the sun sets, and are also less active at cooler temperatures (below 50 degrees Fahrenheit). Additionally, avoiding broadcast applications of systemic herbicides and herbicides with long residuals reduces exposure to butterfly and moth caterpillars that can be poisoned by consuming contaminated vegetation.

### 7.3.5 GRAZING

Grazing is used to limit tree and shrub invasion, provide structural diversity, and encourage the growth of nectar-rich plants. However, livestock grazing is only beneficial to plant diversity, and in turn, pollinators, at low to moderate levels during short periods of time separated by long recovery periods (Hopwood et al 2016). Grazing can negatively affect insect communities by changing the plant community structure and diversity (Kruess and Tscharntke 2002). Insufficient forage from grazing can decrease bumble bee populations (Carell 2002; Hatfield and LeBuhn 2007) and destroy potential nest sites through trampling (Sugden 1985). Intensive grazing can also affect butterfly populations through trampling (Warren 1993) and altering plant community composition (Stoner and Joern 2004).

Development of a grazing plan that includes careful consideration of the type of grazer, its food preference, and how well the grazer can be managed is important for managing invasive species and compatibility with pollinator health and other resources. For example, goats and sheep can be controlled through herding when they are brought onto a site and when they are removed, bracketing periods of time when pollinators and pollinator habitat are least affected. Goats and sheep preferentially eat broadleaf plants and are therefore the preferred grazers for sites where broadleaf weeds are an issue. At specific densities and duration, goats and sheep can control large infestations of invasive weeds. In addition, they can be controlled in areas near water, thereby reducing effects on water quality, and can be effective in inaccessible spots, such as steep slopes.

It is best to introduce grazers at a time when they preferentially feed on the target weed species. Grazing is most effective, for example, when target weeds are palatable; however, this period may be detrimental to pollinators. If rare or imperiled pollinators are present, timing grazing so as to avoid breeding and foraging periods is best. Avoiding grazing during the adult flight period or when imperiled butterfly or moth larvae are feeding on host plants reduces the effect on pollinators. It is also important to avoid grazing during periods when floral resources are already scarce, as grazing during such times can eliminate pollinators from sites over time. Lastly, the stocking density of grazers can help to determine the duration of grazing. If stocking density is high, it is best to keep the duration relatively short so that desirable vegetation is not affected.

### 7.3.6 FIRE

Prescribed fire is used to manage roadside vegetation and rejuvenate plant diversity in some regions of the United States that have a history of natural fires. Prescribed fire can benefit pollinators through restoration or maintenance of suitable habitat (e.g., Huntzinger 2003), but it can also be harmful when not applied appropriately and have long-term effects on the populations of some species (e.g., Ne’eman et al 2000). For example, burns during the growing season destroy eggs, caterpillars, and above-ground nests and remove vegetation at a time when pollinators need floral resources, host plants, and nesting materials, while winter burns destroy species that overwinter in leaf litter or stems. The scale of the prescribed fire can also affect pollinators. For example, an extremely large and expansive fire may kill pollinators overwintering in above-ground biomass, and such fires during the growing season may eliminate all floral resources in a given area. Smaller, dispersed fires, on the other hand, conserve floral resources and support pollinators in the area by providing refuges.

Burns that are timed so as to have the least effect on pollinators and limiting the scale and frequency of each burn are important pollinator-friendly practices. It is best to use prescribed burning on sections of the roadside corridor rather than the entire corridor. By leaving unburned roadside habitat, enough pollinators remain to recolonize the burned areas. Rotational burning, such as burns conducted three to five or more years apart, allow time for pollinator populations to recover. Rotational burning can provide the benefits of prescribed fire without irreparably damaging the local pollinator community (Black et al 2011). Varying the timing of prescribed burns can also reduce harm to pollinators by avoiding continuously affecting certain pollinators and components of the roadside plant community. Burns affect pollinators no matter when the burn occurs, so altering the timing of burns can reduce negative effects to a particular group or suite of pollinators.

### 7.3.7 BIOLOGICAL CONTROL

Biological control is the process of introducing natural enemies of the target weed that occur in the geographic region of the weed. Although biological weed control is not currently widely implemented by state departments of transportation, several have released natural enemies to such weeds as purple loosestrife (*Lythrum salicaria*), leafy spurge (*Euphorbia esula*) (Johnson 2000), yellow star thistle (*Centaurea solstitialis*), and Russian thistle (*Salsola kali*) (Harper-Lore et al 2013).

Biological control can be an effective and focused approach to weed control. However, there are ecological and economic risks associated with introducing a species outside of its natural range, including unpredictable and irreversible consequences (Simberloff and Stiling 1996). The Eurasian weevil (*Rhinocyllus conicus*), for example, was introduced to control musk thistle (*Carduus nutans*), but expanded its host plants to include native thistles after introduction, including several rare thistle species (Louda et al. 1997). The loss of native thistles or other native species affects the wildlife that depend upon the plants, including pollinators that visit the plants for pollen and nectar or use the plants as hosts for their caterpillars.

It is best to avoid using natural enemies that have expanded their hosts to include native plants. Coordinating with state agencies, keeping records about locations of releases, and monitoring the target weed populations and potential non-target native species can be used to evaluate the potential impacts of biocontrol agents.

### 7.3.8 MECHANICAL REMOVAL

Weed populations can be opportunistically removed during road construction. Removal of populations is accomplished by excavating soils within the populations at least a foot deep and transporting the weed-contaminated material offsite to an approved storage area.

Brush may also be removed from roadsides. Brush removal can benefit pollinator health when  
Roadside Revegetation: An Integrated Approach to Establishing Native Plants and Pollinator Habitat

brush and trees that pose no risk to motorists are left on site and by opening up the canopy along forest edges. Mechanical trimming to remove problematic shrubs or trees and selective trimming to partially remove woody vegetation can benefit pollinators by creating opportunities for wildflowers to grow. However, complete removal of trees and shrubs is not always beneficial because many butterflies and moths use woody native plants as hosts and to roost during the flight period, and some tunnel-nesting bees use the stems of some shrubs as nesting sites.

Removal of any brush or trees that could be hazardous to motorists is important. This includes plants that could impede sight distance, become dangerous fixed objects, fall onto the highway, or shade the road in winter creating patches of ice. When possible, consider leaving snags or trees with cavities in areas where they are set back from the road and pose no safety risk. Snags can provide nesting habitat for some bees, as well as habitat for birds and bats.

Transitional areas between forest and grass can be created by using brush removal to feather or soften forest edges adjacent to clear zones. Edge feathering involves thinning portions of the forest canopy along the edge next to grassy areas and removing undesirable or unhealthy trees. Periodic cutting to maintain healthy growth and an open canopy benefits remnant patches of savanna, forest, or other habitat dominated by woody vegetation, improves the quality of the habitat for pollinators and many birds, and is aesthetically pleasing.

### 7.3.9 HAYING

In some states, landowners are permitted to cut and remove roadside vegetation for animal fodder. States might grant emergency hay permits under drought conditions, for example, or allow annual haying by adjacent landowners on certain roads throughout the growing season. Haying is not a tool typically used by roadside maintenance staff, although it does affect roadside vegetation and thus pollinators by the abrupt removal of flowers and host plants from a site. In general, haying once in the middle of the growing season can favor wildflowers and cool-season grasses that are often suppressed by dominant warm-season grasses. However, too-frequent haying in a growing season can reduce roadside revegetation over time (Jacobsen et al. 1990), thereby reducing floral resources for pollinators. A poorly timed haying may have severe consequences for rare or endemic pollinator species. When possible, maintenance staff who communicate with landowners can suggest they hay a portion of the entire site at a time, leaving a refuge for pollinators. Additionally, setting the mower blades at 12 to 16 inches reduces the effect on vegetation structure that provides nesting and overwintering habitat, and allows vegetation to recover and bloom more quickly.

## 7.4 PREVENTION

Ideally, when maintenance staff takes over a roadside revegetation project, the site is weed-resistant (Section 3.11.4). Maintaining a healthy native plant community thereafter greatly reduces the possibility for future weed invasion. The role of maintenance then, is to prevent or minimize unwanted vegetation from becoming established in a weed-free revegetation project. Prevention is the first line of defense in vegetation management and it is accomplished by maintaining a weed-resistant roadside environment, quickly treating disturbances, and protecting natural areas.

### 7.4.1 MAINTAINING A WEED-RESISTANT ROADSIDE ENVIRONMENT

Roadside vegetation changes over time through successional processes and land uses. If the revegetation project is implemented successfully, the roadside vegetation should continue to be weed-resistant. That can change, however, if the vegetation is disturbed, creating bare soil where weeds can become established. Some practices to maintain a weed-resistant roadside environment are described below.

## Minimizing Ground Disturbance

Maintaining a roadside free of ground disturbance is not always possible, but minimizing the amount of disturbance can reduce the area affected. Ground disturbances, and potential solutions, include the following:

- Rutting from vehicle run-offs (repairing roadsides soon after crashes)
- Mowing slopes that exceed  $3(H):1(V)$  often cause rutting and erosion from equipment
- Mowing when soils are too wet, compacting vegetation and soil, and causing erosion
- Landslides (maintaining stable cut and fill slopes)
- Gullies and rills from road runoff (improving road drainage, soil structure, and groundcover)
- Ditch cleaning (limiting ditch cleaning or maintaining a groundcover with occasional mowing)
- Unauthorized trails and off-road vehicle disturbances (controlling access)
- Grazing (controlling animals)
- Vegetation maintenance (performing maintenance activities to prevent soil exposure)

## Disposal of Soil in Designated Areas

Road maintenance often necessitates the disposal of soil that comes from maintaining the road. Material from landslides, ditch cleaning, and winter graveling operations is sometimes pushed over the roadsides or deposited in areas along the roadside, potentially covering roadside vegetation with exposed soils. It is important that excavated soils be removed and deposited in designated areas that have been reviewed for their offsite effects on water quality and other resources.

## Controlling Noxious Weeds

If there are no sources of weed seeds, then exposed soils revegetate from the seeds that are sown by the maintenance staff. It is important to reduce or eliminate unwanted seed sources by controlling noxious weeds prior to disturbances ([Section 7.3](#)).

## Retaining Shade

Many weeds require full or partial sunlight to thrive (Penny and Neal 2003); therefore, retaining shade from existing native vegetation is one strategy to control some weed species. Cutting trees and shrubs or mowing vegetation can increase light and space for invasive weeds (Schooler et al 2010).

## Mulching of Woody Material On Site

Maintenance activities that produce material from processed slash or excess vegetation from right-of-way clearing can be shredded and spread over the roadsides as mulch, especially on areas that have bare soils. Strategic placement of this material can reduce the potential for weed establishment.

### 7.4.2 TREATING DISTURBANCES FOR QUICK RECOVERY

When ground disturbances do occur, a quick response by maintenance staff to treat these areas can ensure that unwanted vegetation does not become established, as described below.

## Limiting or Controlling the Activity Causing the Disturbance

The first response is to assess the activity that caused the disturbance and to fix it before proceeding to revegetate the site. For example, if runoff from a road surface causes erosion on a fill slope, then it is important to fix the drainage before restoring the vegetation. If a large vehicle runoff creates rutting of the roadside, it is recommended that soil filling and regrading of the area occur prior to reseeding. If off-road vehicle use creates bare soil, then access to the area would be limited prior to revegetating the site.

## Having an Appropriate Seed Mix Readily Available

Having an appropriate seed mix available prior to a disturbance is important because locating the appropriate seed mixes at the quantities needed can be difficult on short notice. Seed mixes may be available commercially or from the designer of the revegetation project. At a minimum, a list of appropriate plant species in a seed mix can be obtained from the revegetation plan or using the ERA tool. If a seed mix is maintained for these disturbances, keeping the mix in favorable long-term storage environments maintains its viability ([Section 5.3.4](#), see [Seed Storage](#)). The disturbance may need to be stabilized until it is favorable to seed ([Section 3.8.6](#)).

## Improving Soils

Sites that have been disturbed often have poor soils because the topsoil has been removed or mixed with the subsoil (e.g., landslide scars, gullies). Compost blankets are a quick way of covering bare soils and increasing the soil productivity at the same time ([Section 5.2.3](#), see [Seed Covering](#)).

## Oversowing Seeds

On disturbed sites, environmental conditions for seed germination are often poor for many native grass and forb species. Applying seeds at excessive rates increases the probability of greater plant establishment. It is important to understand the site's environmental conditions that facilitate seed germination and establishment, as all sites have a maximum seed load they are able to support. If seeding is conducted above this rate, a net gain may not be realized.

## Applying a Weed-free Mulch

After seeds are sown on a disturbed area, applying a mulch over the seeds ensures that soil erosion is kept to a minimum and seeds have a favorable germination environment. Most seeds can germinate through mulches applied  $\frac{1}{2}$  to 1 inch thick on top of the seed. A variety of mulches are available that include wood fiber, hydromulch, straw, hay, erosion mats, and manufactured wood strands ([Section 5.2.3](#)). It is important to obtain weed-free mulches so the introduction of unwanted vegetation is kept to a minimum. Having weed-free sources available prior to disturbances expedites a quick recovery.

### *For the Designer*

**Seeding at excessive rates can benefit the project to a point. Understanding conditions that contribute to, or limit, seed establishment will help determine maximum seeding rates.**

## Using Clean Equipment

Inspecting and cleaning vehicles and equipment that will apply seeds and mulch ensures that unwanted vegetation is not brought onto the site. The equipment is typically pre-washed by the contractor at an approved off-site facility. Washing with high-powered, high-temperature water (steam cleaning) is effective. After cleaning, equipment can be inspected at the wash site or another agreed upon location prior to arriving at the project site. Overlooked areas to inspect are the hoppers and hoses of hydroseeding equipment.

## Avoiding Nitrogen Fertilizer the First Year

Consider applying a slow-release fertilizer (instead of fast-release fertilizers) after native plants have

established to reduce the nitrogen levels available for annual weed establishment ([Section 3.11.4](#)).

## 7.5 PROTECTION

Roadsides may contain unique plant communities and rare plants that may need protection. These areas are identified in the planning process but are managed during the operations and maintenance phase of the project. Many states have programs to protect listed species, species of interest, or remnant plant communities (AASHTO 2013a) on roadsides. They are called by a variety of names, including Special Management Areas (SMAs), natural heritage remnants, wildflower research areas, and Biological Management Areas (BMAs). These areas have some or all of the following features in common (adapted from AASHTO 2013a):

- **Collaboration**—Most states with established management areas in right-of-way have accomplished this through collaboration with maintenance staff; conservation groups; and federal, state, and local agencies.
- **GIS**—Many states use GIS to identify rare plant populations and remnant habitats. Known locations of rare species are obtained from state departments of natural resources, the U.S. Fish and Wildlife Service, the U.S. Forest Service, the Bureau of Land Management, counties, local environmental groups, and individuals.
- **Management plans and BMPs**—Most special management areas have plans or BMPs for how to preserve remnant sites or species of interest. These practices include how vegetation control treatments, such as mowing, herbicides, prescribed burns, and grazing, will be used to preserve and enhance remnant areas or species of interest. Maintenance treatments that support species of interest, such as minimizing shading from trees or competition from surrounding vegetation, can be a part of the BMPs. In areas where species of interest require full sunlight for survival, trees and shrubs around these plants may be removed. Where competing vegetation or noxious plant species threaten species of interest, herbicide may be applied or manual removal of vegetation around these species may occur. In areas where potential soil erosion and landslides threaten species of interest or remnant areas, soil stabilization measures may be taken. Public access may be curtailed if it affects plant survival.
- **Signage**—Some states place special signs along the boundaries of the management areas and have instructions on the signs indicating what maintenance activities are allowed.
- **Training**—Some states have developed training programs for maintenance staff specific to managing rare species. This training covers the identification of species of interest, habitat, and preferred management options.

# 8—Case Studies

- 8.1 I-35 Corridor (aka “The Monarch Highway”) Case Studies**
- 8.2 Florida resolves to protect wildflowers on roadsides**
- 8.3 Mapping and planning benefit Washington State pollinators**
- 8.4 Establishing native plants in Arizona**



Chapter 8 provides five case studies to highlight successful pollinator-friendly practices and strategies in various roadside projects from multiple parts of the country. Several of these case studies are a subset of those found in Hopwood et al 2016a.

## 8.1 I-35 CORRIDOR (AKA “THE MONARCH HIGHWAY”) CASE STUDIES

The 2015 “National Strategy to Promote the Health of Honey Bees and Other Pollinators” is a strategy calling for, among other things, a multi-state partnership called the “Monarch Highway” along the I-35 corridor. Running through Minnesota, Iowa, Missouri, Kansas, Oklahoma, and Texas, the I-35 highway spans the central flyway monarch butterflies take during their annual migration. The “Monarch Highway” effort hopes to serve as a national model of cooperation to enhance pollinator habitat along transportation rights-of-way. The six states involved in the strategy have agreed to coordinate efforts to establish best practices and promote monarch butterfly and pollinator conservation. A list of I-35 plant species and their pollinator attributes is available in the [Native Revegetation Resource Library](#).

The two following case studies highlight successful projects from Iowa, one of the six states that comprise the I-35 corridor where agencies are focused on restoring habitat to support the imperiled Monarch butterfly and other pollinators.

### 8.1.1 BRINGING PRAIRIE BACK TO IOWA: IOWA’S INTEGRATED ROADSIDE VEGETATION MANAGEMENT PROGRAM AND LIVING ROADWAY TRUST FUND

Prairie once dominated Iowa’s landscape, covering more than 85 percent of the state. With less than 0.1 percent of virgin prairie remaining, and more than 95 percent of Iowa’s original wetlands destroyed, Iowa had the nation’s most altered landscape as of the 1980s. Prior to the mid-1980s, roadside weed control in Iowa relied heavily on blanket spraying, putting large amounts of herbicide into the environment with undesirable consequences. Recognizing Iowa’s lost heritage and the need to protect groundwater and surface waters, Iowa roadside managers began making some changes. For example, they began using native prairie grasses and wildflowers for erosion control and reintroduced “a little wildness,” according to Kirk Henderson, retired State IRVM Specialist from the Native Roadside Vegetation Center at the University of Northern Iowa.

In 1989, the Iowa legislature passed IRVM legislation to promote an integrated approach to roadside management while maintaining a safe travel environment (Code of Iowa, Chapter 314). Section 314.21 created the Living Roadway Trust Fund to develop and implement IRVM plans. Section 314.22, titled “Integrated roadside vegetation management” established goals and responsibilities for the State DOT that other roadside managing agencies could



Butterfly milkweed and prairie cone flower flourish along an Iowa roadside.

*Photo credit: Iowa Living Roadway Trust Fund*

adopt. Iowa is widely seen as a leader in IRVM, in large part because of this legislation. The Living Roadway Trust Fund includes an annual competitive grant program that the Iowa DOT administers to provide funding for school, city, county, and state projects, as well as research projects involving IRVM.

Iowa's road use tax, along with several other sources, funds the Living Roadway Trust Fund. Roadside managers can submit applications to obtain resources to help implement IRVM, including vegetation inventories, purchasing native seed, equipment for burns or plant establishment, GPS units, signage, workshops, and more. Roadsides are seeded with mixes of species that are appropriate for a particular site, including many native wildflowers that are attractive to pollinators. Seed mixes also contain species that bloom at different times throughout the growing season, which helps support pollinators all season long. The targeted vegetation management practiced by Iowa's roadside managers also benefits pollinators (Ries et al 2001).

Research projects have also been supported by the Living Roadway Trust Fund, including studies of restoration techniques, as well as studies of the impact of roadside habitat on butterflies (Ries et al 2001) and bees (Hopwood et al 2010). Since the 1989 law was passed, more than 100,000 acres of Iowa's nearly 600,000 acres of state and county roadsides have been planted to native vegetation (Brandt et al 2015). In the process, Iowa has fostered the development of experienced roadside managers who are equipped to collaborate with other land managers around the state and bring habitat and wildlife, such as pollinators, back to Iowa's landscape.

### 8.1.2 IOWA'S NATURAL SELECTIONS PROGRAM INCREASES IOWA NATIVE SEED

The Natural Selections program was formed to build the native seed industry in Iowa to meet the demands for high quality, regionally adapted, and genetically diverse sources of native seed for prairie restorations, including roadside restorations. For native regional seed to be priced to compete with cultivars, it has to be produced in commercial quantities. The Natural Selections program is a collaboration between state and federal agencies, as well as private corporations. Past or present partners include the Iowa Crop Improvement Association, the Living Roadway Trust Fund administered by Iowa DOT, USDA Natural Resources Conservation Service Elsberry Plant Materials Center, the University of Northern Iowa, and independent seed producers. Seed is collected by hand from remnant populations by the project manager and by volunteers throughout the state. Seed is collected from remnant roadsides, natural areas, and private land in three regional zones within Iowa. Iowa's Natural Selections program uses provenance zones large enough to support a market but narrow enough to retain regional distinctiveness. Collectors do not collect seed to intentionally select for certain traits. Foundation seed plots at the University of Northern Iowa amplify the seed, which is then further increased. Once seed has been increased, it is released to qualified native seed growers with production certified by the Iowa Crop Improvement Association. About 6 to 8 years after the initial collection, enough seed is available to sell to the public.

Successes of the project include increasing seed of 70 species, with nearly 120 ecotypes of 60 species released for commercial production and 180,000 to 200,000 pounds of ecotype source-identified seed produced annually. More information can be found at the Tallgrass Prairie [website](#).



*Photo credit: Carl Kurtz*



*Photo credits: Kirk Henderson*

## 8.2 FLORIDA RESOLVES TO PROTECT WILDFLOWERS ON ROADSIDES

The following case study from Florida highlights a unique citizen-based effort to protect pollinator habitat through county policy resolutions that were subsequently supported by pollinator-friendly management plans.

Florida, home to a great diversity of plants and animals, was once dubbed the “land of flowers” by a Spanish explorer in 1513. Many of these wildflowers can be found on Florida’s 200,000 acres of roadsides. However, wildflower proliferation along roadsides can be limited by the frequency of mowing. Roadside mowing can be very intensive in some parts of Florida, particularly in urban areas. When showy stands of wildflowers were mowed during bloom when pollinators were present, concerned citizens contacted Florida DOT. Jeff Caster, State Transportation Landscape Architect with Florida DOT, describes the situation: “There would be butterflies on the side of the road feasting on the native vegetation and we would come in and mow it all down and we would get people naturally upset with us. Environmentally conscious citizens called us to complain that we were mowing down wildflowers and butterfly habitat.”

Florida DOT is not able to alter management plans based on direct requests from a garden club or an individual that wants less roadside mowing. But citizens in Wakulla County found another way. They worked with their county commissioners to draft a resolution that made it county policy to preserve existing stands of roadside wildflowers. Then, county staff worked with Florida DOT to develop a roadside management plan to accommodate the resolution.

Recognizing the cultural, historical, and environmental significance of native wildflowers, 27 out of Florida’s 67 counties have moved to adopt similar wildflower resolutions. These counties “make a commitment to saying they want to enjoy the visibility of wildflowers for whatever reason, some do it to attract nature-based tourism, some doing it to help their farms,” Caster says. A model resolution can be found on the [Florida Wildflower Foundation’s website](#).

Florida’s unique grassroots approach to protecting wildflowers at the county level has great potential for pollinator conservation.



*Photo credits: Maria Urice*

Native wildflowers along a Florida turnpike.

*Photo credit: Jeff Norcini*





This interchange, in the heart of Washington's grape and hops growing region, is undergoing integrated management to reduce the dominant invasive species kochia and cereal rye. Native milkweed occurs naturally at this site, serving as potential breeding habitat for monarch butterflies.

*Photo credit: Washington State DOT*

## 8.3 MAPPING AND PLANNING BENEFIT WASHINGTON STATE POLLINATORS

Washington State DOT (WSDOT) maintains about 100,000 acres of roadside. WSDOT has been implementing integrated vegetation management for many years with the overarching goal of reducing undesirable vegetation while encouraging desirable vegetation. Many of their practices and policies to manage vegetation also promote pollinator habitat.

Whenever possible, WSDOT preserves existing native habitat that can provide food, host plants, shelter, and nesting for pollinators. WSDOT also identifies roadsides for "managed succession" (WSDOT 2003). These are areas that have desirable vegetation that could flourish under a strategy that allows natural plant succession to proceed to a stable plant community. Outside of the clear zone or lines of sight where repeated mowing still occurs, roadsides managed for natural succession have multi-year treatment strategies that employ a variety of vegetation management tools, including reduced mowing and targeted herbicide use. Managed succession allows native vegetation to emerge and flourish, with sagebrush (*Artemesia tridentata*), greasewood (*Sarcobatus vermiculatus*), and native grasses taking hold in prairie regions, and understory shrubs that are attractive to pollinators like snowberry (*Symphoricarpos albus*), Oregon grape (*Mahonia nervosa*), and spirea (*Spiraea* sp) in other regions.

When roadside revegetation is needed, WSDOT prioritizes the use of native plants, including a diversity of native wildflowers and flowering shrubs and trees. Landscape designers focus on native plants that can establish with minimal input and compete with weeds. Key considerations for pollinators during the planning process include sequential bloom periods of flowering plants and high plant diversity. Maintenance staff are included in the planning process to provide input on long-lived plants that can be managed minimally.

Mapping and planning are key elements of Washington DOT's approach to roadside revegetation and maintenance. Vegetation inventories help to identify areas with weed infestations as well as areas that are conducive to managed succession. All maintenance staff have tablets, used to record data and view maps and aerial images.

WSDOT uses the best available science to inform its actions and also undertake its own research to determine the right methods for their management needs. For example, experimental plots are used for testing seed mixes and planting methods. WSDOT also monitors the effectiveness of maintenance techniques on vegetation and pollinators. More information about the actions WSDOT is taking to protect pollinators and promote pollinator health can be found at its [website](#).



A site identified through GIS analysis as a priority location for managed succession to replace nonnative species with desirable native species. Careful planning and management for a number of years has increased native plants such as camas, snowberry, serviceberry, and Oregon grape, but invasive scotch broom still remains and requires selective treatment with herbicides.

*Photo: to Washington State Department of Transportation*

## 8.4 ESTABLISHING NATIVE PLANTS IN ARIZONA

In 1992, Arizona DOT began to use native species in all roadside revegetation efforts. This was implemented by drafting a plant list from State maps of biotic communities grouped by eco-regions. From this list, they identified species that were available commercially. Over the years, available plant mixes included a diversity of 15 to 25 species, composed of annual, biennial, perennial wildflowers, shrubs, and 3 to 5 species of native grasses. After observing that grasses out-competed flowers and even shrubs over time in plantings composed of 50 percent grass and 50 percent wildflowers/ shrubs, Arizona DOT scaled back the proportion of grasses to 25 percent. The availability of plant materials, particularly seeds of native species, has increased greatly since 1992.

Arizona includes noxious and invasive species control in construction specifications. In practice, this specification means that noxious and invasive weeds are controlled before, during, and after road construction. By proactively controlling problematic weeds that can outcompete beneficial plants, this practice helps the desired species establish quickly and maintain the long-term integrity of the planting.



Native wildflowers along Arizona's Interstate 10.

*Photocredits: Arizona Department of Transportation*

## References

- Ahmad M. 2013. Biochar as a sorbent for contaminant management in soil and water: A review. Chemosphere. In Press. URL: <http://dx.doi.org/10.1016/j.chemosphere.2013.10.071> (accessed 5 October 2018).
- Alaux C, Ducloz, F, Crauser D, Le Conte, Y. 2010. Diet effects on honeybee immunocompetence. Biology Letters: rsbl20090986.
- Alexander R. 2003a. A composting update: the way we were, and where we're going. Municipal Stormwater Management 13(1). 7 p.
- Alexander R. 2003b. Developing and implementing national erosion/sediment control specifications for composted products. Durham (NH): Recycled Materials Resource Center. URL: [https://www.researchgate.net/publication/238798244\\_Developing\\_and\\_Implementing\\_National\\_ErosionSediment\\_Control\\_Specifications\\_for\\_Composted\\_Products](https://www.researchgate.net/publication/238798244_Developing_and_Implementing_National_ErosionSediment_Control_Specifications_for_Composted_Products) (accessed 26 December 2018).
- Allen E, Espejel C, Sigüenza C. 1997. Role of mycorrhizae in restoration of marginal and derelict land and ecosystem sustainability. In: Palm M, Capela I, editors. Mycology in sustainable development: expanding concepts, vanishing borders. Boone (NC): Parkway Publishers Inc. p. 147-159.
- Amaranthus M. 2002. Declaration of interdependence: mycorrhizal fungi are the 'tiny little secrets' for planting success. Oregon Association of Nurserymen Digger. 46(4):48-53.
- Amaranthus M, Perry D. 1987. The effect of soil transfer on ectomycorrhiza formation and the survival and growth of conifer seedlings on old, nonforested clearcuts. Canadian Journal of Forest Research 17:944–950.
- Amaranthus M, Steinfeld D. 1997. Soil compaction after yarding of small-diameter Douglas-fir with a small tractor in southwest Oregon. Portland (OR): USDA Forest Service, Pacific Northwest Research Station. Research Paper PNW-RP-504. 7 p.
- Amaranthus M, Steinfeld D. 2005. Arbuscular mycorrhizal inoculation following biocide treatment improves *Calocedrus decurrens* survival and growth in nursery and outplanting sites. In: Riley LE, Dumroese RK, Landis TD, technical coordinators. National proceedings: forest and conservation nursery associations—2004. Fort Collins (CO): USDA Forest Service, Rocky Mountain Research Station. RMRS-P-35. p. 103-108.
- Ament R, Begley J, Powell S, Stoy P. 2014. Roadside Vegetation and Soils on Federal Lands – Evaluation of the Potential for Increasing Carbon Capture and Storage and Decreasing Carbon Emissions. FHA Report. 47 p. Report for the Federal Highway Administration, # DTFH68-07-E-00045. URL: [http://www.montana.edu/spowell/FLMA%20Roadside%20Carbon%20Report%20Final\\_January2014.pdf](http://www.montana.edu/spowell/FLMA%20Roadside%20Carbon%20Report%20Final_January2014.pdf) (accessed 26 December 2018).
- Anderson H, Baumgartner D, editors. Forest soils of the Douglas-fir region. Pullman (WA): Washington State University Cooperative Extension Service. p. 113-120.

Anderson N, Hart J, Sullivan D, Christensen N, Hornectk D, Pirelli G. 2013. Applying lime to raise soil pH for crop production (western Oregon). Oregon State University Extension Service. EM 9057. URL: <https://catalog.extension.oregonstate.edu/em9057> (accessed 5 October 2018).

Andrus C, Froehlich H. 1983. An evaluation of four implements used to till compacted forest soils in the Pacific Northwest. Corvallis (OR): Forest Research Laboratory. Research Bulletin 45. 12 p.

Anonymous. [nd]. Mycorrhizae and revegetation of disturbed lands. Bozeman (MT): Montana State University. URL: <https://studylib.net/doc/7253715/mycorrhizae-and-revegetation-of-disturbed-lands> (accessed 27 December 2018).

Araratyan L, Zakharyan S. 1988. On the contamination of snow along main highways. Biologicheskii Zhurnal Armenii 41: 511-519.

Archuleta J, Baxter E. 2008. Subsoiling promotes native plant establishment on compacted forest sites. Native Plants Journal. 9:2 17-122.

Applegate G, Bragg A. 1989. Improved growth rates of red cedar (*Toona australis* (F. Muell.) Harms) seedlings in grow tubes in north Queensland. Australian Forestry Journal 52(4):293-297.

Ashby W, Vogel W. 1994. Tree planting on mined lands in the Midwest: a handbook. Carbondale (IL): Southern Illinois University, Coal Research Center. 115 p. URL: [https://books.google.com/books/about/Tree\\_planting\\_on\\_mined\\_lands\\_in\\_the\\_Midw.html?id=iJEdAQAAMAAJ](https://books.google.com/books/about/Tree_planting_on_mined_lands_in_the_Midw.html?id=iJEdAQAAMAAJ) (accessed 26 December 2018).

American Association of State Highway and Transportation Officials [AASHTO]. 2013. Chapter 8 Winter Operations and Salt, Sand, and Chemical Management. URL: [https://sicop-dev.transportation.org/wp-content/uploads/sites/36/2017/07/NCHRP\\_20-07318\\_Final-Report-2013.pdf](https://sicop-dev.transportation.org/wp-content/uploads/sites/36/2017/07/NCHRP_20-07318_Final-Report-2013.pdf) (accessed 9 October 2018).

American Association of State Highway and Transportation Officials (AASHTO). 2013a. Chapter 9.1 Inventory of and management for rare species and sensitive resources in the ROW. URL: [http://environment.transportation.org/environmental\\_issues/construct\\_maint\\_prac/compendium/manual/9\\_1.aspx](http://environment.transportation.org/environmental_issues/construct_maint_prac/compendium/manual/9_1.aspx) (accessed 19 January 2016).

Association of Official Seed Analysts [AOA]. 2002. Rules for testing seeds. Stillwater (OK): Association of Official Seed Analysts. 166 p.

Aubry C, Shoal R, Erickson V. 2005. Grass cultivars: their origins, development, and use on national forests and grasslands in the Pacific Northwest. Portland (OR): USDA Forest Service. 41 p.

Bainbridge D, Tisler J, MacAller R, Allen M. 2001. Irrigation and mulch effects on desert shrub transplant establishment. Native Plants Journal 2(1):25-29.

Bainbridge D. 1994. Tree shelters improve establishment on dry sites. Tree Planters' Notes 45(1):13-16.

- Bainbridge D. 2006a. Beyond drip irrigation—hyper-efficient irrigation. In: American Society of Agricultural and Biological Engineers Annual International Meeting; 2006 July 9-12; Portland, OR. St Joseph (MI): American Society of Agricultural and Biological Engineers. Paper 062071. 10 p.
- Bainbridge D. 2006b. Deep pipe irrigation. Holualoa (HI): Agroforestry Net Inc. 8p. URL: [https://www.agroforestry.org/books/6-downloads?layout\\_](https://www.agroforestry.org/books/6-downloads?layout_) (accessed 15 May 2007).
- Bainbridge D. 2006c. Irrigation for remote sites. San Diego (CA): San Diego State University, Soil Ecology and Restoration Group. Restoration Bulletin 6. 20 p.
- Ballard T. 1981. Physical properties and physical behavior of forest soils. In: Heilman P, Anderson H, Baumgartner D, editors. Forest soils of the Douglas-fir region. Pullman (WA): Washington State University Cooperative Extension Service. P. 113-120.
- Barnar J. 2007. Personal communication. Bend (OR): USDA Forest Service, Bend Seed Extractory Manager.
- Barnum S. 2003. Identifying the best locations to provide safe highway crossing opportunities for wildlife. In 2003 Proceedings of the International Conference on Ecology and Transportation, edited by Irwin C, Garrett P, McDermott K. Raleigh, NC: Center for Transportation and the Environment, North Carolina State University, 2003. 246-252.
- Barnum S, Rinehart K, Elbroch M. 2007. Habitat, highway features, and animal-vehicle collision locations as indicators of wildlife crossing. In Proceedings of the 2007 International Conference on Ecology and Transportation, edited by Irwin C, Nelson D, McDermott K. Raleigh, NC: Center for Transportation and the Environment, North Carolina State University, 2007. 511-518.
- Barvenik F. 1994. Polyacrylamide characteristics related to soil applications. Soil Science 158(4): 235-243.
- Basey A, Fant J, Kramer A. 2015. Producing native plant materials for restoration: 10 rules to collect and maintain genetic diversity. Native Plants Journal 16(1):37-52.
- Baskin J, Baskin C. 1989. Seasonal changes in the germination responses of buried seeds of *Barbarea vulgaris*. Canadian Journal of Botany 67:2131-2134.
- Baxter E, Stephan E. 2011. Using biosolids to establish native grasses on timber haul roads. Learning Summary. URL: [http://resources.nativerevegetation.org/files/baxter2008\\_learnsum.pdf](http://resources.nativerevegetation.org/files/baxter2008_learnsum.pdf) (accessed 5 October 2018).
- Bean T, Smith S, Karpiscak M. 2004. Intensive revegetation in Arizona's hot desert—the advantages of container stock. Native Plants Journal 5(2):173-180.
- Bedrossian T. 1983. Watershed mapping in northern California. Californian Geology 36:140-147.
- Beesley L, Moreno-Jimenez E, Gomex-Eyles, Harris E, Robinson B, Sizmur T. 2011. A review of biochars' potential role in the remediation, revegetation and restoration of contaminated soils. Environmental Pollution. 159 (2011) 3269-3282.

- Bellot J, Ortiz De Urbina J, Bonet A, Sanchez J. 2002. The effects of tree shelters on the growth of *Quercus coccifera* L. seedlings in a semiarid environment. *Forestry* 75(1):89-106.
- Bentrup G, Hoag J. 1998. The practical streambank bioengineering guide. Aberdeen (ID): USDA Natural Resources Conservation Service, Plant Materials Center. 150 p. URL: [https://efotg.sc.egov.usda.gov/references/public/NM/BIO-48\\_The\\_Practical\\_Streambank\\_Bioengineering\\_Guide.pdf](https://efotg.sc.egov.usda.gov/references/public/NM/BIO-48_The_Practical_Streambank_Bioengineering_Guide.pdf) (accessed 27 December 2018).
- Berger R. 2005. Integrated roadside vegetation management. Washington (DC): Transportation Research Board. 80 p.
- Bergez J, Cupraz C. 1997. Transpiration rate of *Prunus avium* L. seedlings inside an unventilated treeshelter. *Forest Ecology and Management* 97:255-264.
- Black S, Shepherd M, Vaughan M. 2011. Rangeland management for pollinators. *Rangelands* 33(3):9-13.
- Black S, Fallon C, Hatfield R, Mazzacano, C. 2014. Controlled Burning and Mardon Skipper: Summary of Mardon Skipper Coon Mountain Burn Site Occupancy Study Data from 2009 to 2013. Report to the U.S. Forest Service, Oregon Zoo, and U.S. Fish and Wildlife Service: 29.
- Bloomfield H, Handley J, Bradshaw A. 1982. Nutrient deficiencies and the aftercare of reclaimed derelict land. *Journal of Applied Ecology* 19:151-158.
- Bogemams J, Nicrinck L, Stassart, J. 1989. Effect of deicing chloride salts on ion accumulation in spruce (*Picea abies* (L.) sp.). *Plant and Soil* 113:3-11.
- Bollen W. 1974. Soil microbes. In: Cramer O, editor. Environmental effects of forest residues management in the Pacific Northwest. Portland (OR): USDA Forest Service, Pacific Northwest Forest and Range Experiment Station. General Technical Report PNW-24. P. B1-B41.
- Blomqvist G, Johansson E, 1999. "Airborne spreading and deposition of deicing salt case study", *Science of the Total Environment*, 235, 161-168.
- Bonar R. 1995. The northern pocket gopher—most of what you thought you might want to know, but hesitated to look up. Missoula (MT): USDA Forest Service, Technology and Development Program. Technical Services, Reforestation TE02E11. 61 p.
- Borland J. 1990. Mulch—examining the facts and fallacies behind the uses and benefits of mulch. *American Nurseryman* 172(4):132-141.
- Bouchard N, Osmond D, Winston R, Hunt W. 2013. The capacity of roadside vegetated filter strips and swales to sequester carbon. *Ecological Engineering*. Vol 54:227-232.
- Bower A, St. Clair J, Erickson V. 2014. Generalized provisional seed zones for native plants. *Ecological Applications*, 24(5), pp:913-919.

Bowman D, Cramer G, Devitt D. 2006. Effect of Salinity and Nitrogen Status on Nitrogen Uptake by Tall Fescue Turf. *Journal of Plant Nutrition* 29:1481-1490.

Bradshaw A, Marrs R, Roberts R, Skeffington R. 1982. The creation of nitrogen cycles in derelict land. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences* 296:557-561.

Bradshaw A. 1992. The reclamation of derelict land and the ecology of ecosystems. In: Jordan W, Gilpin M, Aber J, editors. *Restoration ecology: a synthetic approach to ecological restoration*. Cambridge (England): Cambridge University Press. p. 53-74.

Brandle J and Nickerson H. 1996. Windbreaks for snow management. 4 p. URL: <https://digitalcommons.unl.edu/cgi/viewcontent.cgi?article=1126&context=natrespapers> (accessed 27 December 2018).

Brant J, Henderson K, Utley J. 2015. Integrated roadside vegetation management - technical manual. USDA Natural Resource Conservation Service and Iowa DOT's Living Roadway Trust Fund. 105 p. URL: [http://www.tallgrassprairiecenter.org/sites/default/files/irvm-technical-manual-2015-2\\_0.pdf](http://www.tallgrassprairiecenter.org/sites/default/files/irvm-technical-manual-2015-2_0.pdf) (accessed 10 July 2016).

Breed M, Stead M, Ottewell K, Gardner M, and Lowe A. 2013. Which provenance and where? Seed sourcing strategies for revegetation in a changing environment. *Conservation Genetics*. Vol 14:1-10.

Brejda J, Yocom D, Moser L, Waller S. 1993. Dependence of 3 Nebraska Sandhills warm-season grasses on vesicular-arbuscular mycorrhizae. *Journal of Range Management* 46(1):14-20.

Brock T. 2016. The Bradley method for control of invasive plants. URL: [http://courses.washington.edu/ehuf462/462\\_mats/bradley\\_method.pdf](http://courses.washington.edu/ehuf462/462_mats/bradley_method.pdf) (accessed 11 July 2016).

Brooks C, Blaser R. 1964. Effect of fertilizer slurries used with hydroseeding on seed viability. Washington (DC): Highway Research Board. *Highway Research Record* 53:30-34.

Brown J. 2016. A field study of biochar amended soils: water retention and nutrient removal from stormwater runoff. Presented at 2016 US Biochar Initiative Conference. Corvallis OR. 29p.

Brown J, Maddox J, Splittstoesser W. 1983. The effect of fertilizer and strip mine spoil on germination and growth of legumes. In: Graves D, editor. *Proceedings 1983 symposium on surface mining, hydrology, sedimentology and reclamation*. Lexington (KY): University of Kentucky. P. 419-422.

Brown M, Wolf D, Morse R, Neal J. 1982. Survival of rhizobium inoculum in hydroseeding slurries. In: Graves D, editor. *Proceedings 1982 symposium on surface mining, hydrology, sedimentology and reclamation*. Lexington (KY): University of Kentucky. p. 681-685.

Brown R, Amacher M. 1999. Selecting plant species for ecological restoration: a perspective for land managers. In: Holzworth L, Brown R, compilers. *Revegetation with native species: Proceedings, 1997 Society for Ecological Restoration annual meeting; 1997 November 12- 15; Ft Lauderdale, FL*. Ogden (UT): USDA Forest Service, Rocky Mountain Research Station. *Proceedings RMRS-P-8*. p. 1-16.

- Brown R, Gores J. 2011. The use of soil amendments to improve survival of roadside grasses. *HortScience* 46(10): 1404-1410.
- Brunsen D, Prior D. 1984. Slope instability. Chichester (UK): John Wiley and Sons Ltd. 620 p.
- Brunzel S, Elligsen H, Frankl R. 2004. Distribution of the Cinnabar moth *Tyria jacobaeae* L. at landscape scale: use of linear landscape structures in egg laying on larval hostplant exposures. *Landscape Ecology* 19(1), 21-27.
- Bryson G, Barker A. 2002. Sodium accumulation in soils and plants along Massachusetts roadsides. *Communications in Soil Science and Plant Analysis* 33(1-2), 67-78.
- Brzozowski C. 2003. Carpets of grass [online]. Santa Barbara (CA): Forester Communications Inc., Erosion Control. URL: [http://www.forester.net/ecm\\_0301\\_carpets.html](http://www.forester.net/ecm_0301_carpets.html) (accessed 18 August 2004).
- Brzozowski C. 2004. Hydroseeding: a versatile alternative [online]. Santa Barbara (CA): Forester Communications Inc, Erosion Control. URL: [http://www.forester.net/ecm\\_0401\\_hydroseeding.html](http://www.forester.net/ecm_0401_hydroseeding.html) (accessed 18 August 2004).
- Bureau of Land Management [BLM]. 1994. Record of decision for the final supplemental environmental impact statement on management of habitat for late-successional and old growth forest related species within the range of the northern spotted owl. Portland (OR): USDA Forest Service. 475 p.
- Bureau of Land Management [BLM]. 2002. A collaborative approach for reducing wildland fire risks to communities and the environment: 10-year comprehensive strategy. URL: <https://www.loc.gov/item/2002418530/> (accessed 27 December 2018).
- Burghardt K, Tallamy D, Shriver, W. 2009. Impact of native plants on bird and butterfly biodiversity in suburban landscapes. *Conservation Biology* 23:219–224.
- Burroughs Jr E, King J. 1989. Reduction of soil erosion on forest roads. Ogden (UT): USDA Forest Service, Intermountain Research Station. General Technical Report INT-264. 22 p.
- Busse M. 2000. Ecological significance of nitrogen fixation by actinorrhizal shrubs in interior forests of California and Oregon. In: Powers R, Hauxwell D, Nakamura G, technical coordinators. Proceedings of the California Forest Soils Council conference on forest soils biology and forest management. Albany (CA): USDA Forest Service, Pacific Southwest Research Station. General Technical Report GTR-PSW-178. p. 23-41.
- Calderone N. 2012. Insect pollinated crops, insect pollinators and US agriculture: trend analysis of aggregate data for the period 1992-2009. *PLoS ONE* 7(5) e37235.
- California Department of Transportation [Caltrans]. 2003. Caltrans erosion control new technology report. Sacramento (CA): California Department of Transportation. CTSW-RT-03-049. 463 p.
- California Stormwater Quality Association [CASQA]. 2003a. EC-3 Hydraulic mulch. In: Construction Handbook [online]. Menlo Park (CA): California Stormwater Quality Association. 3 p. URL: <http://www.cabmphandbooks.com> (accessed 20 January 2006).

California Stormwater Quality Association [CASQA]. 2003b. EC-5 Soil binders. In: Construction Handbook [online]. Menlo Park (CA): California Stormwater Quality Association. 7 p. URL: <http://www.cabmphandbooks.com> (accessed 20 January 2006).

Cane J. 1991. Soils of ground-nesting bees (Hymenoptera: Apoidea): texture, moisture, cell depth and climate. *Journal of the Kansas Entomological Society*: 64(4): 406-413.

Campbell K, Helgested E, Shaw E, Hawkes G. 1980. Western fertilizer handbook. Sacramento (CA): California Fertilizer Association. 269 p.

Campbell R. 1986. Mapped genetic variation of Douglas fir to guide seed transfer in southwest Oregon. *Silvae Genetica* 35:85-96.

Casement B, Timmermans J. 2007. Field shelterbelts for soil conservation. Agdex 277/20-3. 8p. URL: [http://www1.agric.gov.ab.ca/\\$department/deptdocs.nsf/all/agdex2073/\\$file/277\\_20-3.pdf?Open](http://www1.agric.gov.ab.ca/$department/deptdocs.nsf/all/agdex2073/$file/277_20-3.pdf?Open) (accessed 5 October 2018).

Carr W, Ballard T. 1979. Effects of fertilizer salt concentration on viability of seed and Rhizobium used for hydroseeding. *Canadian Journal of Botany* 57(7):701-704.

Carr W, Ballard T. 1980. Hydroseeding forest roadsides in British Columbia for erosion control. *Journal of Soil and Water Conservation* 35(1):33-35.

Carson M, Kirkby M. 1972. Hillslope form and process. London (UK): Cambridge University Press. 475 p.

Carvell C. 2002. Habitat use and conservation of bumble bees (*Bombus* spp.) under different grassland management regimes. *Biological Conservation* 103:33–49.

Chaplin-Kramer R, Tuxen-Bettman K, Kremen C. 2011. Value of wildland habitat for supplying pollination services to Californian agriculture. *Rangelands* 33(3):33-41.

Charvat I, Tallaksen J, White J, Agwa H, Raley M, Slack S, Hebberger J, Gould E. 2000. Mycorrhizal/ plant factors involved in roadside reclamation. St Paul (MN): Minnesota Department of Transportation. Final Report - MN/RC-2000-30. 131 p.

Childs S, Flint L. 1987. Effect of shadecards, shelterwoods, and clearcuts on temperature and moisture environments. *Forest Ecology and Management* 18:205-217.

Claassen V, Carey J. 2004. Regeneration of nitrogen fertility in disturbed soils using composts. *Compost Science and Utilization* 12(2):145-152.

Claassen V, Hogan M. 1998. Generation of water-stable soil aggregates for improved erosion control and revegetation success. Sacramento (CA): California Department of Transportation. Research Technical Agreement 53X461. 111 p.

Claassen V, Hogan M. 2002. Soil nitrogen pools associated with revegetation of disturbed sites in the Lake Tahoe area. *Restoration Ecology* 10(2):195-203.

- Claassen V, Marler M. 1998. Annual and perennial grass growth on nitrogen-depleted decomposed granite. *Restoration Ecology* 6(2):175-180.
- Claassen V, Zasoski R, Southard R. 1995. Soil conditions and mycorrhizal infection associated with revegetation of decomposed granite slopes. Sacramento (CA): California Department of Transportation. Technical Report FHWA-CA-TL 96/1. 151 p.
- Claassen V, Zasoski R. 1994. The effects of topsoil reapplication on vegetation reestablishment. Sacramento (CA): California Department of Transportation. Technical Report FHWA-CA-TL- 94/18. 52 p.
- Claassen V, Zasoski R. 1998. A comparison of plant available nutrients on decomposed granite cut slopes and adjacent natural soils. *Land Degradation and Development* 9:35-46.
- Claassen V. 2006. Soil resource evaluation. Sacramento (CA): California Department of Transportation. URL: <http://citeseerx.ist.psu.edu/viewdoc/download?doi=10.1.1.641.6065&rep=rep1&type=pdf> (accessed 27 December 2018).
- Clark T, Stevenson M, Ziegelmayer K, Rutherford M, editors. 2001. Species and ecosystem conservation: an interdisciplinary approach. New Haven (CT): Yale University Press.
- Cleary B, Greaves R, Hermann R, editors. 1988. Regenerating Oregon forests, a guide for the regeneration forester. 5th printing. Corvallis (OR): Oregon State University Extension Service. 287 p.
- Clevenger, A, Huijser M. *Wildlife Crossing Structure Handbook Design and Evaluation in North America*. Lakewood Co.: U.S. Federal Highway Administration Central Federal Lands Highway Division, 2011. Print.
- Clevenger A, Walther N. 2000. Factors influencing the effectiveness of wildlife underpasses in Banff National Park, Alberta, Canada. *Conservation Biology*, 14(1), 47-56.
- Clevenger A, Walther N. 2005. Performance indices to identify attributes of highway crossing structures facilitating movement of large mammals. *Biological Conservation* 121:453-464.
- Clewel A, Rieger J, Munro J. 2005. Guidelines for developing and managing ecological restoration projects. 2nd ed. Tuscon (AZ): Society for Ecological Restoration International. URL: <http://www.ser.org> (accessed 20 Aug 2006).
- Cole D, Johnson D. 1981. The cycling of elements within forests. In: Heilman P, Anderson H, Baumgartner D, editors. *Forest soils of the Douglas-fir region*. Pullman (WA): Washington State University Cooperative Extension Service. p. 185-198.
- Cook D, Nelson S. 1986. Effect of polyacrylamide on seedling emergence in crust-forming soil. *Soil Science* 141:328-333.

Corkiki L, Allen E, Merhaut D, Allen M, Downer J, Bohn J, Evans M. 2005. Effectiveness of commercial mycorrhizal inoculants on the growth of *Liquidambar styraciflua* in plant nursery conditions. Journal of Environmental Horticulture 23(2):72-96.

Cox T, Glover J, Van Tassel D, Cox C, DeHaan L. 2006. Prospects for Developing Perennial Grain Crops. BioScience. Vol. 56 No.8. August 2006. URL: [https://landinstitute.org/wp-content/uploads/2007/07/Bioscience\\_PerennialGrains.pdf](https://landinstitute.org/wp-content/uploads/2007/07/Bioscience_PerennialGrains.pdf) (accessed 12/8/2016).

Crémieux L, Bischoff A, Müller-Schärer H, Steinger T. 2010. Gene flow from foreign provenances into local plant populations: fitness consequences and implications for biodiversity restoration. American Journal of Botany. 97(1): 94–100.

Crowder W, Darris D. 1999. Producing Pacific Northwest native trees and shrubs in hardwood cutting blocks or stooling beds. Portland (OR): USDA Natural Resources Conservation Service. Technical Notes—Plant Materials No 24. 13 p.

Croxton P, Hann J, Greatorex-Davis J, Sparks T. 2005. Linear hotspots? The floral and butterfly diversity of green lanes. Biological Conservation 121:579–584.

Curtis M, Claassen V. 2005. Compost incorporation increases plant available water in a drastically disturbed serpentine soil. Soil Science 170(12):13.

Dancer W, Handley J, Bradshaw A. 1977. Nitrogen accumulation in kaolin mining wastes in Cornwall. I. Natural communities. Plant and Soil 48:153-167.

Dancer W. 1975. Leaching losses of ammonium and nitrate in the reclamation of sand spoils in Cornwall. Journal of Environmental Quality 4(4):499-504.

Darboux F, Huang C. 2005. Does soil roughness increase or decrease water and particle transfer? Soil Science Society of America Journal 69:748-756.

Darris D, Williams D. 2001. Native shrubs as a supplement to the use of willows as live stakes and facines in western Oregon and western Washington. In: Haase D, Rose R, editors. Proceedings of the conference: native plant propagation and restoration strategies; 2001 Dec 12-13; Eugene, OR. Corvallis (OR): Oregon State University, Nursery Technology Cooperative. p. 112-120.

Darris D. 2002. Native shrubs as a supplement to the use of willows as live stakes and facines in western Oregon and western Washington. Portland (OR): USDA Natural Resources Conservation Service. Technical Notes—Plant Materials No 31. 10 p.

Daugovish O, Downer J, Faber B, McGiffen M. 2006. Weed survival in yard waste mulch. Weed Technology 21(1):59-65.

Davies R. 1988a. Sheet mulching as an aid to broadleaved tree establishment. I. The effectiveness of various synthetic sheets compared. Forestry 61(2):89-105.

Davies R. 1988b. Sheet mulching as an aid to broadleaved tree establishment II. Comparison of various sizes of black polythene mulches and herbicide treated spot. Forestry 61(2):107-124.

- DeByle N. 1969. Black polyethylene mulch increases survival and growth of a Jeffrey pine plantation. *Tree Planters' Notes* 19(4):7-11.
- Deere D, Deere D. 1988. The rock quality designation (RQD) index in practice. In: Kirkaldie L, editor. Rock classification systems for engineering purposes. Philadelphia (PA): American Society of Testing Materials. ASTM Special Publication 984. p. 91-101.
- Denning C, Prellwitz R, Reneria R, Rose E. 1994. Parameters for stability analysis. In: Hall D, Long M, Remboldt M, editors. Slope stability reference guide for national forests in the United States. Volume II. Washington (DC): USDA Forest Service, Engineering Staff. EM 7170-13. p. 327-721.
- DePuit E, Coenenberg J. 1979. Responses of revegetated coal strip mine spoils to variable fertilization rates, longevity of fertilization program, and season of seeding. Bozeman (MT): Montana State University, Montana Agricultural Experiment Station. Research Report 150. 81 p.
- Di Pasquale G, Salignon M, Le Conte Y, Belzunces L, Decourtye A, Kretzschmar A, Suchail S, Brunet J, Alaux C. 2013. Influence of pollen nutrition on honey bee health: do pollen quality and diversity matter. *PLOS One* 8(8): e72016.
- Dirig R, Cryan J. 1991. The status of silvery blue subspecies (*Glauopsyche lygdamus lygdamus* and *G. l. couperi*: Lycaenidae) in New York. *Journal of the Lepidopterists' Society* 45(4):272-290.
- Dixon R, Carr A. 2001a. Land imprinting specifications for ecological restoration and sustainable agriculture. In: Ascough II J, Flanagan D, editors. Soil erosion research for the 21st Century, proceedings of an international symposium; 2001 Jan 3-5; Honolulu, HI. St Joseph (MI): American Society of Agricultural Engineers. ASAE.701P0007. p. 195-197.
- Dixon R, Carr A. 2001b. Preferential flow control: key to sustainable land management. In: Bosch D, King K, editors. Preferential flow water: movement and chemical transport in the environment, proceedings of the 2nd international symposium; 2001 Jan 3-5; Honolulu, HI. St Joseph (MI): American Society of Agricultural Engineers. ASAE.701P0006. p. 287-289.
- Dixon R, Simanton J. 1977. A land imprinter for revegetation of barren land areas through infiltration control. IN Hydrology and water resources in Arizona and the southwester. Vol 7, p 79-88.
- Dorner J. 2002. An introduction to using native plants in restoration projects. Seattle (WA): Plant Conservation Alliance and Environmental Protection Agency. 66 p.
- Dreesen D, Harrington J. 1997. Propagation of native plants for restoration projects in the southwestern US—preliminary investigations. In: Landis T, Thompson J, technical coordinators. National proceedings, forest and conservation nursery associations. Portland (OR): USDA Forest Service, Pacific Northwest Research Station. General Technical Report PNW- GTR-419. p. 77-88.

Dumroese R, Jacobs D, Landis T. 2005. Successful stock production for forest regeneration: what foresters should ask nursery managers about their crops (and vice versa). In: Colombo S, editor. The thin green line: a symposium on the state-of-the-art in reforestation, proceedings; 2005 July 26-28; Thunder Bay, Ontario. Sault Ste Marie (Ontario): Ontario Ministry of Natural Resources, Forest Research Information Paper 160:14-20.

Dumroese R, Stumph T, Wenny D. 1998. Revegetating Idaho's Henry's Fork: a case study. In: Rose R, Haase DL, editors. Native plants: propagation and planting; 1998 Dec 9-10; Corvallis, OR. Corvallis (OR): Oregon State University, Nursery Technology Cooperative. p. 108-112.

Dumroese R, Landis T, Barnett J, Burch F. 2005. Forest Service Nurseries: 100 years of ecosystem restoration. *Journal of Forestry*: 241-247. URL: <https://www.treesearch.fs.fed.us/pubs/25823> (accessed 5 October 2018).

Dumroese R. 2006. Personal communication. Moscow (ID): USDA Forest Service, Southern Research Station. Research Plant Physiologist.

Dupraz C, Bergez J. 1997. Carbon dioxide limitation of the photosynthesis of *Prunus avium* L. seedlings inside an unventilated treeshelter. *Forest Ecology and Management* 119(1):89-97.

Dyer K, Curtis L, Willie R, Crews J. 1984. Response of vegetation to various mulches used in surface mine reclamation in Alabama and Kentucky—7-year case history. Broomhall (PA): USDA Forest Service, Northeastern Forest Experiment Station. General Technical Report NE-93. 11 p.

Eilers E, Kremen C, Greenleaf S, Garber A, Klein AM. 2011. Contribution of pollinator-mediated crops to nutrients in the human food supply. *PLoS one* 6(6)e21363.

Elzinga, C, Salzer D, Willoughby J. 1998. Measuring and monitoring plant populations. Denver (CO): USDI Bureau of Land Management. BLM Technical Reference 1730-1. 492 p.

Environmental Protection Agency [EPA]. 1994. A plain English guide to the EPA Part 503 Biosolids Rule. EPA/832/H-93/003. EPA Office of Waste Management. 183p.

Environmental Protection Agency [EPA]. 1995. Process design manual - land application of sewage sludge and domestic septage. EPA/625/R-95/001. 301 p.

Environmental Protection Agency [EPA]. 2006. Environmental Protection Agency (EPA) National Pollutant Discharge Elimination System (NPDES) Appendix A - Definitions and Acronyms: "Final Stabilization." Washington (DC): US EPA, Office of Wastewater Management. URL: [https://www3.epa.gov/npdes/pubs/cgp\\_appendixa.pdf](https://www3.epa.gov/npdes/pubs/cgp_appendixa.pdf) (accessed 27 December 2018).

Epstein E. 1997. The science of composting. Boca Raton (FL): CRC Press LLC. 487 p.

Epstein E. 2002. Land application of sewage sludge and biosolids. Boca Raton (FL): CRC Press LLC. 201 p.

Erickson V, Mandel N, Sorensen F. 2004. Landscape pattern of phenotypic variation and population structuring in a selfing grass, *Elymus glaucus* (blue wildrye). *Canadian Journal of Botany* 82:1776-1789.

Erickson V, Wood J, Riley S. 2003. Guidelines for revegetation of invasive weed sites and other disturbed areas on national forests and grasslands in the Pacific Northwest. Portland (OR): USDA Forest Service, Pacific Northwest Region. 80 p.

Erickson V. 2008. Developing native plant germplasm for national forests and grasslands in the Pacific Northwest. Native Plants Journal 9:255–266.

Evans R. 2006. Review of the application of polyacrylamides for soil erosion reduction through sprinklers. Sidney (MT): USDA Agricultural Research Service. 4 p. URL:  
<https://www.ars.usda.gov/ARSUserFiles/21563/Review%20of%20the%20App%20of%20Polyacrylamides%20for%20Soil%20Erosion.pdf> (accessed 27 December 2018).

Fava E. 2015. Biosolids as a roadside soil amendment. Dissertations and Master's Theses (Campus Access) at the University of Rhode Island DigitalCommons@URI. URL:  
<http://digitalcommons.uri.edu/dissertations/AAI10005034> (accessed 5 October 2018).

Feber R, Smith H, Macdonald D. 1996. The effects on butterfly abundance of the management of uncropped edges of arable fields. Journal of Applied Ecology 33:1191–1205.

Federal Highway Administration [FHWA]. 1992. Drainable pavement systems - participant notebook. (Demonstration Project 87). Publication No. FHWA-SA-92-008. Federal Highway Administration, Washington, DC.

Federal Highway Administration [FHWA]. 2005. Project development and design manual (PDDM). Publication Number FHWA-DF-88-003.

Federal Highway Administration [FHWA]. 2005. Project development process flow chart. Vancouver (WA): Western Federal Lands Highway Division. 206 p. URL:  
<https://flh.fhwa.dot.gov/resources/design/tools/wfl/process/PD-Process.pdf> (accessed 5 October 2018).

Federal Highway Administration [FHWA]. 2008. 2008 Status of the Nation's highways, bridges, and transit: conditions and performance. FHWA, Washington, D.C. URL:  
<http://www.fhwa.dot.gov/policy/2008cpr/pdfs/cp2008.pdf> (accessed 5 October 2018).

Federal Highway Administration [FHWA]. 2008(B). Vegetation Control for Safety – A guide for Local Highway and Street Maintenance Personnel. FHWA, Washington, D.C. URL:  
[http://www.safety.fhwa.dot.gov/local\\_rural/training/fhwasa07018/](http://www.safety.fhwa.dot.gov/local_rural/training/fhwasa07018/) (accessed 6 April 2017).

Federal Highway Administration [FHWA]. 2010. Carbon Sequestration Pilot Program, estimated land available for carbon sequestration in the National Highway System. 24 p.

Federal Highway Administration [FHWA]. 2012. Road Weather Management Program, Snow and Ice. Website updated November 2012. URL:  
[http://ops.fhwa.dot.gov/weather/weather\\_events/snow\\_ice.htm](http://ops.fhwa.dot.gov/weather/weather_events/snow_ice.htm) (accessed 5 October 2018).

Federal Highway Administration [FHWA]. 2017. Clear Zones. URL: [https://safety.fhwa.dot.gov/roadway\\_dept/countermeasures/safe\\_recovery/clear\\_zones/](https://safety.fhwa.dot.gov/roadway_dept/countermeasures/safe_recovery/clear_zones/) (accessed 5 October 2018).

Federal Highway Administration [FHWA]. 2010. <https://rosap.ntl.bts.gov/view/dot/8692> (accessed 5 October 2018).

Federal Register. Presidential Memorandum – Creating a Federal Strategy to Promote the Health of Honey Bees and Other Pollinators, The White House, Washington, D.C., June 20, 2014. 79 FR 35903.

Ferrick M, Gatto L. 2004. Quantifying the effect of a freeze-thaw cycle on soil erosion: laboratory experiments. Hanover (NH): US Army Corps of Engineers, Engineer Research and Development Center, Cold Regions Research and Engineering Laboratory. Letter Report A670134. 47 p.

Fidelibus M, Bainbridge D. 2006. Microcatchment water harvesting for desert revegetation [online]. San Diego (CA): San Diego State University, Soil Ecology and Restoration Group. Restoration Bulletin 5. 12 p. URL: <http://www.sci.sdsu.edu/SERG/techniques/microcatch.pdf> (accessed 5 December 2006).

Fiedler A, Landis D, Arduser M. 2012. Rapid shift in pollinator communities following invasive species removal. *Restoration Ecology* 20(5):593–602.

Fisher P, Argo W. 2005. Electrical conductivity of growing media: why is it important? *Greenhouse Management and Production* 25(5):54-58.

Flint A. 1983. Soil physical properties and available water capacity of southwest Oregon forest soils [MS thesis]. Corvallis (OR): Oregon State University. 61 p.

Flint L, Childs SW. 1984. Seedling responses to heat and moisture environments in clearcuts and shelterwoods. *Forestry Intensified Research Report* 6(2):3-4.

Foltz R, Dooley J. 2003. Comparison of erosion reduction between wood strands and agricultural straw. *Transactions of the American Society of Agricultural Engineers* 46(5):1389-1396.

Foltz. Wagenbrenner, N.S. 2010. An evaluation of three wood shred blends for post-fire erosion control using indoor rain events on small plots. *Catena*. 80: 86-94. doi:10.1016/j.catena.2009.09.00.

Forman R, Alexander L. 1998. Roads and their major ecological effects. *Annual Review of Ecology and Systematics* 29: 207-231.

Forman R, editor. 2003. *Road ecology: science and solutions*. Washington (DC): Island Press. 481 p.

Frankie G, Thorp GW, Schindler M, Ertter B, Przybylski M. 2002. Bees in Berkeley? *Fremontia* 30(3-4):50–58.

Frearson K, Weiss N. 1987. Improved growth rates within tree shelters. *Quarterly Journal of Forestry* 81:184-187.

- Fredricksen RL, Harr RD. 1981. Soil, vegetation, and watershed management. In: Heilman P, Anderson H, Baumgartner D, editors. Forest soils of the Douglas-fir region. Pullman (WA): Washington State University Cooperative Extension Service. p. 231-260.
- Fifield J. 2004. Designing for effective sediment and erosion control on construction sites - 2nd edition. Forester Press.
- Fritzsche C. 1992. Calcium magnesium acetate deicer: An effective alternative for salt-sensitive areas. Water Environ Tech WAEIJ. 4(1), 44-51.
- Froehlich H, McNabb D. 1984. Minimizing soil compaction in the Pacific Northwest forests. In: Stone EL, editor. Forest soils and treatment impacts. Proceedings of the 6th North American Forest Soil Conference; 1983 June 19-23; Knoxville, TN. Knoxville (TN): University of Tennessee, Department of Forestry. p. 159-192.
- Froehlich H, Miles D. 1984. Winged subsoiler tills compacted forest soil. Forest Industries 111(2):42-43.
- Froehlich H, Robbins R, Miles D, Lyons JK. 1983. Monitoring recovery of compacted skid trails in central Idaho. Contract 43-0256-2-543 [Special report for Payette National Forest]. Located at: Payette National Forest, McCall, ID.
- Froese J, Cruse R, Ghaffarzadeh M. 1999. Erosion mechanics of soils with an impermeable subsurface layer. Soil Science Society of America Journal 63:1836-1841.
- Fryrear D and Skidmore E. 1985. Chapter 24. Methods for controlling wind erosion. In Soil Erosion and Crop Productivity. ASA-CSSA-SSSA. P 443-457.
- Fulbright T. 1988. Effects of temperature, water potential, and sodium chloride on Indiangrass germination. J. Range Manage. 41:207-210.
- Gade K. 2013. Freeways as Corridors for Plant Dispersal: A Case Study from Central Arizona. Proceedings of the 2013 International Conference on Ecology and Transportation. Available online at [http://www.icoet.net/ICOET\\_2013/documents/papers/ICOET2013\\_Paper403B\\_Gade.pdf](http://www.icoet.net/ICOET_2013/documents/papers/ICOET2013_Paper403B_Gade.pdf) (accessed 27 December 2018).
- Gallai N, Salles J, Settele J, Vaissière B. 2009. Economic valuation of the vulnerability of world agriculture confronted with pollinator decline. Ecological Economics 68:810-821.
- Gassman S. 2001. Hydraulic seeding: it's all in the slurry. Overland Park (KS): Grounds Maintenance. URL: [http://www.grounds-mag.com/mag/grounds\\_maintenance\\_hydraulic\\_seeding\\_slurry/](http://www.grounds-mag.com/mag/grounds_maintenance_hydraulic_seeding_slurry/) (accessed 4 Jan 2005).
- Gatto L, Ferrick M, White K. 2004. Inclusion of freeze-thaw-induced soil and bank erosion in CoE planning, engineering, O&M, and model development. Hanover (NH): US Army Corps of Engineers, Engineer Research and Development Center, Cold Regions Research and Engineering Laboratory. Technical Note 04-2. 5 p.

- Gedney D, Weber W. 1978. Design and construction of soil slopes. In: Schuster R, Krizek R, editors. Landslides, analysis and control. Washington (DC): National Research Council, Transportation Research Board Commission on Sociotechnical Systems. Special Report 176. p. 172-191.
- Gelbard J, Belnap J. 2003. Roads as conduits for exotic plant invasions in a semiarid landscape. *Conservation Biology* 17(2):420–432.
- Ghazoul J. 2006. Floral diversity and the facilitation of pollination. *Journal of Ecology* 94:295–304.
- Gibson D, Carrington M. 2007. Road salt germination inhibition of native plants used in roadside plantings. *Transactions of the Illinois State Academy of Science*. Vol. 101, #1&2, p13-22.
- Gifford G. 1975. Beneficial and detrimental effects on range improvement practices on runoff and erosion. In: Watershed management, proceedings of a symposium conducted by the irrigation and drainage division of the American Society of Civil Engineers; 11-13 Aug 1975; Logan, UT. Reston (VA): American Society of Civil Engineers. p. 216-248.
- Gjessing E, Lygren E, Berglind L, Gulbrandsen T, Skanne R. 1984. Effect of highway runoff on lake water quality. *Science of the Total Environment* 33:247-257.
- Gobar C. 2006. Personal communication. Albuquerque (NM): USDA Forest Service. Forest Wildlife Biologist.
- Goheen D. 2005. Personal communication. Medford (OR): USDA Forest Service. Zone Pathologist.
- Goodrich M, Dulak L, Freidman M, Lech J. 1991. Acute and long-term toxicity of water- soluble cationic polymers to rainbow trout (*Oncorhynchus mykus*) and the modification of toxicity by humic acid. *Environmental Toxicology and Chemistry* 10:509-551.
- Gray D, Leiser A. 1982. Biotechnical slope protection and erosion control. Malabar (FL): Robert E Krieger Publishing Co. 271 p.
- Greaves R, Black H, Hooven E, Hansen E. 1978. Plantation maintenance. In: Cleary B, Greaves R, Hermann R, editors. Regenerating Oregon forests, a guide for the regeneration forester. 5th printing. Corvallis (OR): Oregon State University Extension Service. p. 187-203.
- Greaves R, Hermann R. 1978. Planting and seeding. In: Cleary B, Greaves R, Hermann R, editors. Regenerating Oregon forests, a guide for the regeneration forester. 5th printing. Corvallis (OR): Oregon State University Extension Service. p. 131-161.
- Grismer M, Hogan M. 2004. Simulated rainfall evaluation of revegetation/mulch erosion control in the Lake Tahoe Basin - 1: method assessment. *Land Degradation Development* 15:1-16.
- Grismer M, Hogan M. 2007. Simulated rainfall evaluation at Sunriver and Mt Bachelor Highways, Oregon. Report to WFHLD. 8 p.
- Grismer M, Hogan M, Steinfeld DE. 2011. Highway 35 treatment slopes and bio-retention steps monitoring plan: 1 laboratory column analyses. Integrated Environmental Restoration Services. 18 p.

Grubb B. 2007. Personal communication. Fort Collins (CO): Colorado State Forest Service Nursery. Nursery Grower.

Grubb P. 1977. The maintenance of species richness in plant communities and the importance of the regeneration niche. *Biological Reviews* 52:107-145.

Guinon M. 1993. Promoting gene conservation through seed and plant procurement. In: Landis T, editor. *Proceedings, Western Forest Nursery Association; 1992 Sept 14-18; Fallen Leaf Lake, CA*. Fort Collins (CO): USDA Forest Service, Rocky Mountain Forest and Range Experiment Station. General Technical Report RM-221. p. 38-46.

Gustafson D, Gibson D, Nickrent D. 2005. Using local seeds in prairie restoration – data support the paradigm. *Native Plant Journal* 6(1):25-28.

Habte M, Osorio N. 2001. Arbuscular mycorrhizas: producing and applying arbuscular mycorrhizal inoculum. Honolulu (HI): University of Hawaii, College of Tropical Agriculture and Human Resources. 48 p.

Haddad N. 1999. Corridor and distance effects on interpatch movements: a landscape experiment with butterflies. *Ecological Applications* 9:612–622.

Haddad N, Baum KA. 1999. An experimental test of corridor effects on butterfly densities. *Ecological Applications* 9:623–633.

Haferkamp M, Miller R, Sneva F. 1985. Seeding rangelands with a land imprinter and rangeland drill in the Palouse vegetative prairie and sagebrush-bunchgrass zone. In: 39th annual report, vegetative rehabilitation and equipment workshop; 1985 February 10-11; Salt Lake City, UT. Missoula (MT): USDA Forest Service, Missoula Equipment Development Center. p. 19-22.

Halbritter D, Daniels, J, Whitaker D, Huang L. 2015. Reducing mowing frequency increases floral resource and butterfly (Lepidoptera: Hesperioidae and Papilionoidea) abundance in managed roadside margins. *Florida Entomologist* 98(4): 1081-1092.

Hall D, Long M, Remboldt M, editors. 1994. Slope stability reference guide for National Forests in the United States, Volume II. Moscow (ID): USDA Forest Service, Rocky Mountain Research Station. EM-7170-13.

Hall F. 2002a. Photo point monitoring handbook: part A - field procedures. Portland (OR): USDA Forest Service, Pacific Northwest Research Station. General Technical Report PNW-GTR- 526. p. 1-48.

Hall F. 2002b. Photo point monitoring handbook: part B - concepts and analysis. Portland (OR): USDA Forest Service, Pacific Northwest Research Station. General Technical Report PNW-GTR- 526. p. 49-134.

Hamm P, Campbell S, Hansen E, editors. 1990. Growing healthy seedlings: identification and management of pests in northwest forest nurseries. Corvallis (OR): Oregon State University, Forest Research Laboratory. Special Publication 19. 110 p.

- Hansen M, Clevenger A. 2005. The influence of disturbance and habitat on the presence of non-native plant species along transport corridors. *Biological Conservation* 125(2): 249–259.
- Hanula J, Horn S. 2011. Removing an invasive shrub (Chinese privet) increases native bee diversity and abundance in riparian forests of the southeastern United States. *Insect Conservation and Diversity* 4(4):275–283.
- Harmon-Threatt A, Kremen C. 2015. Bumble bees selectively use native and exotic species to maintain nutritional intake across highly variable and invaded local floral resource pools. *Ecological Entomology*: 4: 471-478.
- Harper-Lore B, Johnson M, and Ostrum W. 2013. Vegetation Management: An Ecoregional Approach. U.S. Department of Transportation Federal Highway Administration. URL: [https://www.environment.fhwa.dot.gov/ecosystems/veg\\_mgmt\\_rpt/vegmgmt\\_ecoregional\\_approach.asp](https://www.environment.fhwa.dot.gov/ecosystems/veg_mgmt_rpt/vegmgmt_ecoregional_approach.asp) (accessed 12 July 2016).
- Harrington J, Meikle T. 1992. Road salt effects on the germination of eight select prairie species. Proceedings of the thirteenth North American prairie conference. Department of Parks and Recreation, Windsor, Ontario, Canada, 183-192.
- Harper-Lore B, Wilson M, editors. 2000. Roadside use of native plants. Washington (DC): Island Press. 665 p.
- Hatchett B, Hogan M, Grismer M. 2006. Mechanical mastication thins Lake Tahoe forest with few adverse impacts. *California Agriculture* 60(2):77-82.
- Hatfield R, Jepsen S, Mader E, Black S, Shepherd M. 2012. Conserving Bumble Bees, Guidelines for Creating and Managing Habitat for America's Declining Pollinators. 32 pp. Portland, OR: The Xerces Society for Invertebrate Conservation.
- Hatfield R and LeBuhn G. 2007. Patch and landscape factors shape community assemblage of bumble bees, *Bombus* spp. (*Hymenoptera: Apidae*), in montane meadows. *Biological Conservation* 139:150–158.
- Hatfield R, Colla S, Jepsen S, Richardson L, Thorp R. 2015. International Union for the Conservation of Nature (IUCN) Assessments for North American *Bombus* spp. for the North American IUCN Bumble Bee Specialist Group. The Xerces Society for Invertebrate Conservation, Portland, OR.
- Havens K, Vitt P, Still S, Kramer A, Fant J, Schatz K. 2015. Seed sourcing for restoration in an era of climate change. *Nature Areas Journal* 35:122-133.
- Havlin J, Beaton J, Tisdale S, Nelson W. 1999. Soil fertility and fertilizers, an introduction to nutrient management. 6th edition. Upper Saddle River (NJ): Prentice-Hall, Inc. 499 p.
- Herrera C. 1989. Pollinator abundance, morphology, and flower visitation rate: analysis of the “quantity” component in a plant-pollinator system. *Oecologia*, 80: 241-248.

Heilman P. 1981. Minerals, chemical properties, and fertility of forest soils. In: Heilman P, Anderson H, Baumgartner D, editors. Forest soils of the Douglas-fir region. Pullman (WA): Washington State University Cooperative Extension Service. p. 121-136.

Helgerson O, Newton M, deCalesta D, Schowalter T, Hansen E. 1992. Protecting young regeneration. In: Hobbs S, Tesch S, Owston P, Stewart R, Tappener II J, Wells G, editors. Reforestation practices in southwestern Oregon and northern California. Corvallis (OR): Oregon State University, Forest Research Laboratory. p. 384-420.

Helgerson O. 1990. Heat damage in tree seedlings and its prevention. *New Forests* 3:17-42.

Henderson K. 2000. Integrating all the management tools: integrated roadside vegetation management. In: Harper-Lore B, Wilson M, editors. Roadside use of native plants. Washington (DC): Island Press. p. 30-31.

Herrick J, Van Zee J, Havstad K, Burkett L, Whitford W. 2005a. Monitoring manual for grassland, shrubland and savanna ecosystems, vol I: quick start. Las Cruces (NM): USDA Agricultural Research Service, Jornada Experimental Range. 36 p. URL: [https://www.researchgate.net/publication/285321006\\_Monitoring\\_manual\\_for\\_grassland\\_shrubland\\_and\\_savanna\\_ecosystems](https://www.researchgate.net/publication/285321006_Monitoring_manual_for_grassland_shrubland_and_savanna_ecosystems) (accessed 27 December 2018).

Herrick J, Van Zee J, Havstad K, Burkett L, Whitford W. 2005b. Monitoring manual for grassland, shrubland and savanna ecosystems, vol II: design, supplementary methods and interpretation. Las Cruces (NM): USDA Agricultural Research Service, Jornada Experimental Range. 200 p. URL: [https://www.researchgate.net/publication/285321006\\_Monitoring\\_manual\\_for\\_grassland\\_shrubland\\_and\\_savanna\\_ecosystems](https://www.researchgate.net/publication/285321006_Monitoring_manual_for_grassland_shrubland_and_savanna_ecosystems) (accessed 27 December 2018).

Hoag J, Simonson B, Cornforth B, St John L. 2001. Waterjet Stinger: a tool for planting dormant nonrooted cuttings. *Native Plants Journal* 2:84-89.

Hoag J. 2003. Willow clump plantings. Boise (ID): USDA Natural Resources Conservation Service. Plant Materials Technical Note No. 42. 8 p.

Hoag J. 2006a. Personal communication. Aberdeen (ID): USDA Natural Resources Conservation Service, Aberdeen Plant Materials Center. Plant Ecologist and Wetland Specialist.

Hoag J. 2006b. The pot planter: a new attachment for the Waterjet Stinger. *Native Plants Journal* 7(1): 100-101.

Hoag J. 2007. Personal communication. Aberdeen (ID): USDA National Resource Conservation Service, Aberdeen Plant Materials Center. Plant Ecologist and Wetland Specialist.

Hobbs S. 1982. Performance of artificially shaded container-grown Douglas-fir seedlings on skeletal soils. Corvallis (OR): Oregon State University, Forest Research Laboratory. Research Note 71. 5 p.

Hogan M. 2007. Personal communication. Tahoe City (CA): Integrated Environmental Restoration Services. Consultant in revegetation.

- Hopwood J, Black S, Fleury S. 2016a. Pollinators and Roadsides: Best Management Practices for Managers and Decision Makers. 96 pp. Washington, D.C.: Federal Highway Administration. URL: [https://www.environment.fhwa.dot.gov/env\\_topics/ecosystems/Pollinators\\_Roadsides/BMPs\\_pollenators\\_roadsides.aspx](https://www.environment.fhwa.dot.gov/env_topics/ecosystems/Pollinators_Roadsides/BMPs_pollenators_roadsides.aspx) (accessed 27 December 2018).
- Hopwood J, Black S, Fleury S. 2016b. Roadside Best Management Practices that Benefit Pollinators: Handbook for Supporting Pollinators through Roadside Maintenance and Landscape Design. 88 pp. Washington, D.C.: Federal Highway Administration. URL: [https://www.environment.fhwa.dot.gov/env\\_topics/ecosystems/Pollinators\\_Roadsides/BMPs\\_pollinators\\_landscapes.aspx](https://www.environment.fhwa.dot.gov/env_topics/ecosystems/Pollinators_Roadsides/BMPs_pollinators_landscapes.aspx) (accessed 27 December 2018).
- Hopwood J, Black S, Lee-Mäder E, Charlap A, Preston R, Mozumder K, Fleury S. 2015. Literature Review: Pollinator Habitat Enhancement and Best Management Practices in Highway Rights-of-Way. 68 pp. Washington, D.C.: Federal Highway Administration. URL: [https://www.environment.fhwa.dot.gov/env\\_topics/ecosystems/pollinator\\_reports/pollinators\\_BMPs\\_in\\_highway\\_ROW.aspx](https://www.environment.fhwa.dot.gov/env_topics/ecosystems/pollinator_reports/pollinators_BMPs_in_highway_ROW.aspx) (accessed 27 December 2018).
- Hopwood J, Winkler L, Deal B, Chivvis M. 2010. Use of roadside prairie plantings by native bees. Iowa Living Roadway Trust Fund Final Report 90-00-LRTF-011 URL: <http://ilr.winningit.com/ResearchProjects/90-00-LRTF-011.aspx> (Accessed 28 July 2016)
- Hopwood J. 2008. The contribution of roadside grassland restorations to native bee conservation. Biological Conservation 141:2632-2640.
- Horneck D, Hart J, Topper K, Koepsell B. 1989. Methods of soil analysis used in the soil testing laboratory at Oregon State University. Corvallis (OR): Oregon State University, Agricultural Experiment Station. SM 89:4. 21 p.
- Horning M, McGovern T, Darris D, Mandel N, Johnson R. 2010. Genecology of *Holodiscus discolor* (Rosaceae) in the Pacific Northwest, U.S.A. Restoration Ecology 18(2): 235-243.
- Houseal G, Smith D. 2000. Source-identified seed: The Iowa roadside experience. Ecological Restoration 18(3):173-183.
- Hufford K, Mazer S. 2003. Plant ecotypes: genetic differentiation n the age of ecological restoration. Trends in Ecology and Evolution 18:147-155.
- Huijser M. Wildlife-vehicle Collision Reduction Study: Report to Congress. McLean, VA: U.S. Dept. of Transportation, Federal Highway Administration, 2008. Print.
- Huijser M, Kociolek A, Allen T, McGowen P, Cramer P, and Venner M. 2015. Construction Guidelines for Wildlife Fencing and Associated Escape and Lateral Access Control Measures. NCHRP 25-25 Task 84. URL: [http://onlinepubs.trb.org/onlinepubs/nchrp/docs/NCHRP25-25%2884%29\\_FR.pdf](http://onlinepubs.trb.org/onlinepubs/nchrp/docs/NCHRP25-25%2884%29_FR.pdf) (accessed 5 October 2018).
- Humbert J, Ghazoul J, Sauter G, Walter T. 2010. Impact of different meadow mowing techniques on field invertebrates. Journal of Applied Entomology 134(7):592–599.

Huntzinger M. 2003. Effects of fire management practices on butterfly diversity in the forested western United States. *Biological Conservation* 113(1):1–12.

Indiana Department of Transportation, 2008. The Indiana Procedural Manual for Preparing Environmental Documents, Section II.B.3.f Context Sensitive Solutions, Indiana Department of Transportation (Bloomington, IN: Indiana Department of Transportation, 2008).

International Seed Testing Association [ISTA]. 1976. International rules for seed testing. *Seed Science and Technology* 4:4-180.

Jablonski B, Koltowsk Z, Marcinkowski J, Rybak-Chmielewska H, Szczesna T, Warakomska Z. 1995. Zawartosc metali ciezkich [Pb, Cd, i Cu] w nektarze, miodzie i pylku pochodzacym z roslin rosnacych przy szlakach komunikacyjnych [The content of heavy metals (Pb, Cd and Cu) in the nectar, honey and pollen collected from roadside plants]. *Pszczelnicze Zeszyty Naukowe* 39:129–144.

Jackson L, Strauss R, Bartolome J. 1988. Plant and soil dynamics in California annual grassland. *Plant and Soil* 110:9-17.

Jacobs D, Rose R, Haase D, Alzugaray P. 2004. Fertilization at planting impairs root system development and drought avoidance of Douglas-fir seedlings. *Annals of Forest Science* 61:643-651.

Jacobs D, Steinbeck K. 2001. Tree shelters improve the survival and growth of planted Engelmann spruce seedlings in southwestern Colorado. *Western Journal of Applied Forestry* 16(3):114-120.

Jacobsen R, Albrecht N, Bolin K. 1990. Wildflower Routes: Benefits of a Management Program for Minnesota Right-of-Way Prairies. *Proceedings of the Twelfth North American Prairie Conference* 153-158.

Jasper D, Roson A, Aboot L. 1979. Phosphorus and the formation of vesicular arbuscular mycorrhizas. *Soil Biology and Biochemistry* 11:501-505.

Jenny H. 1980. The soil resource origin and behavior. *Ecological Studies* 37. 1st edition. New York (NY): Springer-Verlag Inc. 377 p.

Jepsen S, Schweitzer D, Young B, Sears N, Ormes M, Black S. 2015. Conservation status and ecology of Monarchs in the United States. 36 pp. NatureServe, Arlington, Virginia, and the Xerces Society for Invertebrate Conservation, Portland, Oregon.

Johnson G, Stritch L, Olwell P, Lambert S, Horning M, Cronn R. 2010. What are the best seed sources for ecosystem restoration on BLM and USFS lands? *Native Plants Journal* 11:117–131.

Johnson R, Hellier B, Vance-Borland K. 2013. Genecology and seed zones for tapertip onion in the US Great Basin. *Botany* 91(10):686-694.

Johnson, A. 2000. Best Practices Handbook on Roadside Vegetation Management. Minnesota Department of Transportation, Report # 2000-19, p. 78, University of Minnesota 132. URL: <https://www.lrrb.org/media/reports/200820.pdf> (accessed 12 July 2016).

- Johst, K., Drechsler M, Thomas J, Settele J. 2006. Influence of mowing on the persistence of two endangered large blue butterfly species. *Journal of Applied Ecology* 43(2):333–342.
- Jones P, Jeffrey B, Watler P, Hutchon H. 1992. Environmental Impact of Road Salting. In F. M. D'Itri (Ed.), *Chemical deicers and the environment* (pp. 1-116). Lewis Publishers, Boca Raton, FL. P 1-116.
- Kay B, Evans R, Young J. 1977. Soaking procedures and hydroseeder damage to common bermudagrass seeds. *Agronomy Journal* 69(4):555-557.
- Kay B. 1972a. Hydro-seeding limitations and alternatives. Davis (CA): University of California Davis, Agronomy and Range Science. *Agronomy Progress Report #43*. 3 p.
- Kay B. 1972b. Revegetation of ski slopes and other disturbed mountain sites in California. Davis (CA): University of California Davis, Agronomy and Range Science. *Agronomy Progress Report #42*.
- Kay B. 1974. Erosion control treatments on coarse decomposed granite. Davis (CA): University of California Davis, Agronomy and Range Science. *Agronomy Progress Report #60*. 7 p.
- Kay B. 1978. Mulch and chemical stabilizers for land reclamation in dry regions. In: Schaller F, editor. *Reclamation of drastically disturbed lands*. Madison (WI): American Society of Agronomy, Crop Science Society of America, Soil Science Society of America. p. 467-483.
- Kay B. 1983. Straw as an erosion control mulch. Davis (CA): University of California Davis, Agronomy and Range Science. *Agronomy Progress Report #140*. 11 p.
- Kearns C, Inouye D, Waser N. 1998. Endangered mutualisms: the conservation of plant–pollinator interactions. *Annual Review of Ecology & Systematics* 29:83–113.
- Keller G, Sherar J. 2003. Low-volume roads engineering: best management practices field guide. US AID and USDA Forest Service. 158 p.
- Kerr J, Pindar A, Galpern P, Packer L, Potts S, Roberts, S, Rasmont P, Schweiger P, Colla S, Richardson L, Wagner D, Gall L, Sikes D, Pantoja A. 2015. Climate change impacts on bumblebees converge across continents. *Science*. 349(6244): 177-180.
- Ketcheson G, Megahan W, King J. 1999. "R1-R4" and "BIOSED" sediment prediction model tests using forest roads in granitics. *Journal of the American Water Resources Association* 35(1):83-98.
- Ketcheson G, Megahan W. 1996. Sediment production and downslope sediment transport from forest roads in granitic watersheds. Ogden (UT): USDA Forest Service, Intermountain Research Station. Research Paper INT-RP-486. 12 p.
- Kill D, Foote L. 1971. Comparison of long and short-fibered mulches. *Transactions of the American Society of Agricultural Engineers* 14:942.
- Kjelgren R, Montague D, Rupp L. 1997. Establishment in treeshelters. II. Effect of shelter color on gas exchange and hardiness. *HortScience* 32(7):1284-1287.

- Klein A, Vaissiere B, Cane J, Steffan-Dewenter I, Cunningham S, Kremen C, Tscharntke T. 2007. Importance of pollinators in changing landscapes for world crops. *Proceedings of the Royal Society of London B: Biological Sciences*, 274(1608):303-313.
- Klock G. 1981. Some engineering aspects of forest soils. In: Heilman P, Anderson H, Baumgartner D, editors. *Forest soils of the Douglas-fir region*. Pullman (WA): Washington State University Cooperative Extension Service. p. 269-277.
- Kloetzel S. 2004. Revegetation and restoration planting tools: an in-the-field perspective. *Native Plants Journal* 5(1):34-42.
- Knapp E, Rice K. 1994. Starting from seed: genetic issues in using native grasses for restoration. *Restoration and Management Notes* 12:40-45.
- Kramer A, Havens K. 2014. Plant conservation genetics in a changing world. *Trends in Plant Science* 14(11):599-607.
- Kramer A, Larkin D, Fant J. 2015. Assessing potential seed transfer zones for five forb species from the Great Basin floristic region, USA. *Natural Areas Journal*. 35: 174–188.
- Kruess A, Tscharntke T. 2002. Contrasting responses of plant and insect diversity to variation in grazing intensity. *Biological Conservation* 106(3):293–302.
- Kruess A, Tscharntke T. 2002. Grazing intensity and the diversity of grasshoppers, butterflies, and trap-nesting bees and wasps. *Conservation Biology* 16:1570–80.
- Landis T, 2009. The target plant concept. In: Dumroese RK, Luna T, Landis TD, editors. *Nursery manual for native plants: A guide for tribal nurseries – Volume 1: Nursery Management*. Washington (DC): USDA Forest Service. P. 15-31.
- LaBar C, Schultz C. 2012. Investigating the role of herbicides in controlling invasive grasses in prairie habitats: effects on non-target butterflies. *Natural Areas Journal* 32(2):177–189.
- Landis T, Dumroese R. 2006. Monitoring Electrical Conductivity in Soils and Growing Media. *Forest Nursery Notes*, Summer 2006. R6-CP-TP-04-2006. USDA-Forest Service, Pacific Northwest Region: 6-10. URL: <https://rngr.net/publications/fnn/2006-summer/articles/monitoring-electrical-conductivity-in-soils-and-growing-media?searchterm=Electrical+Conductivity> (accessed 5 October 2018).
- Landis T. 2011. Using organic fertilizers in forest and native plant nurseries. *Forest Nursery Notes* 31(2): 1-17.
- Landis T, Tinus R, McDonald S, Barnett J. 1989. The biological component: nursery pests and mycorrhizae. *The container tree nursery manual*. Vol 5. Washington (DC): USDA Forest Service. Agricultural Handbook 674. 171 p.

Landis T, Tinus RW, McDonald S, Barnett J. 1994. Nursery planning, development, and management. The container tree nursery manual. Vol 1. Washington (DC): USDA Forest Service. Agricultural Handbook 674. 188 p.

Landis T, Dumroese R, Haase D. 2010. Seedling processing, storage and outplanting, volume 7, the container tree nursery manual In: Luna T, Dumroese R, editors. Nursery manual for native plants: a guide for tribal nurseries. Washington (DC): USDA Agricultural Handbook. 199 p.

Landis T, Steinfeld D, Watson R. 2002. The long tube stocktype and expandable stinger. Forest Nursery Notes R6-CP-TP-06-01. p. 15-17.

Landis T, Steinfeld D. 1990. Salt injury. In: Hamm P, Campbell S, Hansen E, editors. Growing healthy seedlings: identification and management of pests in northwest forest nurseries. Corvallis (OR): Oregon State University, Forest Research Laboratory. p. 83-86.

Landis T. 1981. Irrigation water quality in tree nurseries in the inland west. In: Huber R, editor. Proceedings of the 1981 Intermountain Nurserymen's Association meeting; 1981 Aug 11-13; Edmonton, Alberta. Edmonton (Alberta): Canadian Forestry Service, Northern Forest Research Centre. Information Report NOR-X-241. p. 60-67.

Landis T, Dreesen D, Dumroese R. 2003. Sex and the single Salix: considerations for riparian restoration. Native Plants Journal 4(2):111-117.

Lavelle M. 2014. What's that Growing Along the Roadside? Carbon Storage. The Daily Climate - July 28, 2014. URL: <https://www.mnn.com/earth-matters/wilderness-resources/stories/whats-that-growing-along-the-roadside-carbon-storage> (accessed 27 December 2018).

Laycock W, Richardson B. 1975. Long-term effects of pocket gopher control on vegetation and soils of a subalpine grassland. Journal of Range Management 28(6):458-462.

Leger E. 2008. The adaptive value of remnant native plants in invaded communities: an example from the Great Basin. *Ecological Applications* 18: 1226-1235.

Leharne S, Charesworth D, Chowdhry B. 1992. A survey of metal levels in street dusts in an inner London neighbourhood. Environment International 18:263-270.

Lesica P, Allendorf F. 1999. Ecological genetics and the restoration of plant communities: mix or match? Restoration Ecology 7:42–50.

Lewis L. 2000. Soil bioengineering, an alternative for roadside management—a practical guide. San Dimas (CA): USDA Forest Service, San Dimas Technology and Development Center. Technical Report 0077 1801 - SDTDC. 44 p.

Ley T, Stevens R, Topielec R, Neibling W. 1994. Soil water monitoring and measurement. Pullman (WA): Washington State University. Pacific Northwest Publication PNW0475.

Linhart Y. 1995. Restoration, revegetation, and the importance of genetic and evolutionary perspectives. In: Roundy B, McArthur E, Haley J, Mann D, editors. Proceedings, Wildland shrub and arid land restoration symposium, 1993 October 19-21; Las Vegas, NV. USDA Forest Service, General Technical Report. INT-GTR-315. p. 271-287.

Linsley E. 1958. The ecology of solitary bees. *Hilgardia* 27:543-599.

Louda S, Kendall C, Connor J, Simberloff D. 1997. Ecological effects of an insect introduced for the biological control of weeds. *Science* 277:1088–1090.

Louis Berger Group. 2011. Implementing Measures to Reduce Highway Impacts on Habitat Fragmentation. NCHRP 25-25 Task 68. URL: [http://onlinepubs.trb.org/onlinepubs/nchrp/docs/NCHRP25-25\(68\)\\_FR.pdf](http://onlinepubs.trb.org/onlinepubs/nchrp/docs/NCHRP25-25(68)_FR.pdf) (accessed 5 October 2018).

Lövei G, Macleod A, Hickman J. 1998. Dispersal and effects of barriers on the movement of the New Zealand hover fly *Melanostoma fasciatum* (Dipt., Syrphidae) on cultivated land. *Journal of Applied Entomology* 122(1-5), 115-120.

Lowenstein H, Ptikin F. 1970. Ponderosa pine transplants aided by black plastic mulch in Idaho plantation. *Tree Planters' Notes* 21(4):23-24.

Luce C. 1997. Effectiveness of road ripping in restoring infiltration capacity of forest roads. *Restoration Ecology* 5(3):265-270.

Lyles L, Krauss R. 1971. Threshold velocities and initial particle motion as influenced by air turbulence. *Tans. ASAE* 14:563-566.

Mackay D. 1972. The measurement of viability. In: Roberts EH, editor. *Viability of seeds*. Syracuse (NY): Syracuse University Press. p. 172-208.

Mader E, Spivak M, Evans E. 2010. *Managing Alternative Pollinators: A Handbook for Beekeepers, Growers, and Conservationists*. Sustainable Agriculture Research and Education, Handbook 11. College Park, MD: University of Maryland, Sustainable Agriculture Research and Extension and Ithaca, NY: Cornell University, Natural Resource, Agriculture, and Engineering Service.

Mallams K. 2006. Personal communication. Central Point (OR): USDA Forest Service, J Herbert Stone Nursery. Zone pathologist.

Margulis L, Sagan D, Thomas L. 1997. *Microcosmos: four billion years of evolution from our microbial ancestors*. Berkeley (CA): University of California Press. 304 p.

Martin L, Pezeshki S, Shields Jr F. 2005. Soaking treatment increases survival of black willow posts in a large-scale field study. *Ecological Restoration* 23(2):95-98.

Massu L. 2005. Common control measures for invasive plants of the Pacific Northwest. In: USDA Forest Service. Pacific Northwest Region invasive plant program: preventing and managing invasive plants. Final Environmental Impact Statement. Portland (OR): USDA Forest Service, Pacific Northwest Region. R6-NR-FHP-PR-02-05.

Mast B. 2007. Personal communication. Richland (WA): Wildlands Inc. President.

Maxwell W, Ward F. 1976a. Photo series for quantifying forest residues in the coastal Douglas-fir-hemlock type and coastal Douglas-fir-hardwood type. Portland (OR): USDA Forest Service, Pacific Northwest Research Station. General Technical Report PNW-51. 103 p.

Maxwell W, Ward F. 1976b. Photo series for quantifying forest residues in the ponderosa pine type, ponderosa pine and associated species type, lodgepole pine type. Portland (OR): USDA Forest Service, Pacific Northwest Research Station. General Technical Report PNW-52. 73 p.

Maxwell W, Ward F. 1979. Photo series for quantifying forest residues in the Sierra mixed conifer type, Sierra true fir type. Portland (OR): USDA Forest Service, Pacific Northwest Research Station. General Technical Report PNW-95. 79 p.

McCreary D, Tecklin J. 1997. Effects of seedling protectors and weed control on blue oak growth and survival. In: Pillsbury N, Verner J, Tietje W, technical coordinators. Proceedings of a symposium on oak woodlands: ecology, management, and urban interface issues; 1996 March 19-22; San Luis Obispo, CA. Albany (CA): USDA Forest Service, Pacific Southwest Research Station. General Technical Report PSW-GTR-160.

McDonald P, Helgerson OT. 1991. Mulches aid in regenerating California and Oregon forests: past present and future. Berkeley (CA): USDA Forest Service, Pacific Southwest Forest and Range Experiment Station. General Technical Report PSW-123. 19 p.

McDonald S. 1984. Irrigation in forest-tree nurseries: monitoring and effects on seedling growth. In: Duryea ML, Landis TD, editors. Forest nursery manual: production of bareroot seedlings. Boston (MA): Martinus Nijhoff/Dr E Junk Publishers. p. 107-121.

McElroy G, Dawson W. 1986. Biomass from short-rotation coppic willow on marginal land. Biomass 10:255-240.

McFrederick Q, Kathilankal J, Fuentes J. 2008. Air pollution modifies floral scent trails. Atmospheric Environment 42(10):2336-2348.

McGinnies W, Crofts K. 1986. Effects of N and P fertilizer placement on establishment of seeded species on redistributed mine topsoil. Journal of Range Management 39:118-121.

McGregor S. 1976. Insect pollination of cultivated crop plants. USDA Agriculture Handbook No. 496. URL: <https://www.ars.usda.gov/ARSUserFiles/20220500/OnlinePollinationHandbook.pdf> (accessed 27 December 2018).

McKay J, Christian C, Harrison S, Rice K. 2005. "How local is local?" – A review of practical and conceptual issues in the genetics of restoration. Restoration Ecology. 13:432–440.

- McLean E. 1973. Testing soils for pH and lime requirement. In: Walsh LM, Beaton JD, editors. *Soil testing and plant analysis*. Madison (WI): Soil Science Society of America. p. 77-95.
- McKenna D, McKenna K, Malcom S, Berenbaum M. 2001. Mortality of Lepidoptera along roadways in central Illinois. *Journal of the Lepidopterists Society* 55(2):63–68.
- McLendon T, Redente E. 1992. Effects of nitrogen limitation on species replacement dynamics during early succession on a semiarid sagebrush site. *Oecologia* 91:312-317.
- Megahan W. 1974. Deep-rooted plants for erosion control on granitic road fills in the Idaho Batholith. Ogden (UT): USDA Forest Service, Intermountain Research Station. Research Paper INT-161. 18 p.
- Memmott J, Waser N. 2002. Integration of alien plants into a native flower–pollinator visitation web. *Proceedings of the Royal Society of London Series B: Biological Sciences* 269:2395–2399.
- Menz M, Phillips R, Winfree R, Kremen C, Aizen M, Johnson S, Dixon K. 2011. Reconnecting plants and pollinators: challenges in the restoration of pollination mutualisms. *Trends in Plant Science* 16(1):4-12.
- Meyer L, Wischmeier WH, Daniel WH. 1971. Erosion, runoff, and revegetation of denuded construction sites. *Transactions of the American Society of Agricultural Engineers* 14:138-141.
- Mijnsbrugge K, Bischoff A, Smith B. 2010. A question of origin: Where and how to collect seed for ecological restoration. *Basic and Applied Ecology*. 11(4): 300-311.
- Michener C. 2007. *The Bees of the World*, 2nd Ed. 992 pp. Baltimore: John Hopkins University Press.
- Millar C, Libby W. 1989. Disneyland or native ecosystem: genetics and the restorationist. *Restoration and Management Notes* 7(1):18-24.
- Miller R, Jastrow J. 1992. The application of VA mycorrhizae to ecosystem restoration and reclamation. In: Allen MF, editor. *Mycorrhizal functioning: an integrative plant-fungal process*. New York (NY): Chapman and Hall. p. 438-467.
- Miller R, May S. 1979. Below-ground ecosystems project. In: Land Reclamation Program: annual report 1978. Argonne (IL): Argonne National Laboratory. ANL/LRP 5. p. 64-70.
- Minnerath A, Vaughan M, Lee-Mader E. 2016. Maritime Northwest citizen science monitoring guide native bees & butterflies. Xerces Society. 60p.
- Minore D. 1971. Shade benefits Douglas-fir in southwestern Oregon cutover area. *Tree Planters' Notes* 22(1):22-23.
- Mok J, Landphair H, Naderi J. 2006. Landscape Improvement Impacts on Roadside Safety in Texas, *Landscape and Urban Planning*, Vol. 78, 2006, pp. 263-274.

- Monsen S, Stevens R. 2004. Seedbed preparation and seeding practices. In: Monsen S, Stevens R, Shaw N, compilers. Restoring western ranges and wildlands. Fort Collins (CO): USDA Forest Service, Rocky Mountain Research Station. General Technical Report RMRS-GTR-136 - volume 1. p. 121-154.
- Montalvo A, Williams S, Rice K, Buchmann S, Cory C, Handel S, Nabhan G, Primack R, Robichaux RH. 1997. Restoration biology: a population biology perspective. *Restoration Ecology* 5: 277–290.
- Morgan J. 1994. Soil impoverishment: a little-known technique holds potential for establishing prairie. *Restoration and Management Notes* 12:55-56.
- Morandin L, Kremen C. 2013a. Bee preference for native versus exotic plants in restored agricultural hedgerows. *Restoration Ecology* 21(1):26–32.
- Morandin L, Kremen C. 2013b. Hedgerow restoration promotes pollinator populations and exports native bees to adjacent fields. *Ecological Applications* 23(4):829-839.
- Morgenson G. 2007. Personal communication. Bismark (ND): Lincoln-Oakes Nurseries. Nursery Manager.
- Moroń D, Szentgyorgyi H, Grzes' I, Wantuch M, Rozej E, Laskowski R, Woyciechowski M. 2010. The effect of heavy metal pollution on development of wild bees. *Atlas of Biodiversity Risks – From Europe to the Globe, From Stories to Maps* (eds Settele J, Penev LD, Georgiev TA, Grabaum R, Grobelnik V, Hammen V et al.), pp. 224–225. Penssoft, Sofia & Moscow.
- Mulder C, Aldenberg T, de Zwart D, van Wijnen H, Breure A. 2005. Evaluating the impact of pollution on plant-Lepidoptera relations. *Environmetrics* 16:357–373.
- Munguira M, Thomas J. 1992. Use of road verges by butterfly and burnet populations, and the effect of roads on adult dispersal and mortality. *Journal of Applied Ecology* 29:316–329.
- Munshower F. 1994. Practical handbook of disturbed land revegetation. Boca Raton (FL): CRC Press. 265 p.
- National Research Council. 2005. Assessing and managing the ecological impacts of paved roads. Washington (DC): National Academies Press. 294 p.
- National Research Council. 2007. Status of pollinators in North America. Washington, D.C.: National Academies Press. 312 p.
- National Roadside Vegetation Management Association [NRVMA]. 1997. How to develop and implement an integrated roadside vegetation management program - a guide for township, city, county, parish, state, turnpike and other roadside authorities. 48 p.
- Ne'eman G, Dafni A, Potts S. 2000. The effect of fire on flower visitation rate and fruit set in four core-species in east Mediterranean scrubland. *Plant Ecology* 146:97–104.
- Neufeld J, Davison J. 2000. Practical considerations when selecting a soil testing laboratory for an educational program [online]. *Journal of Extension* 38(4). 9 p. URL: <http://www.joe.org/joe/2000august/rb2.html> (accessed 3 Apr 2007).

Nieminen M, Nuorteva P, Tulisalo E. 2001. The effect of metals on the mortality of *Parnassius apollo* larvae (Lepidoptera: Papilionidae). *Journal of Insect Conservation* 5(1):1–7.

Nitrogen-Fixing Tree Association Staff. 1986. Actinorrhizal trees useful in cool to cold regions. In: NFT Highlights: a quick guide to useful nitrogen-fixing trees from around the world. Morrilton (AR): Winrock International.

Norcini J, Thetford M, Klock-Moore K, Bell M, Harbaugh B, and Aldrich J. 2001. Growth, Flowering, and Survival of Black-eyed Susan from Different Regional Seed Sources. *HortTechnology* 11(1): 26-30.

Norland M. 2000. Use of mulches and soil stabilizers for land reclamation. In: Barnhisel RI, Darmody RG, Daniels WL, editors. *Reclamation of drastically disturbed lands*. Madison (WI): American Society of Agronomy. Agronomy Monograph 41. p. 645-666.

Nursery Technical Cooperative. 2004. Nursery Technical Cooperative 2003-2004 Annual Report. Corvallis (OR): Oregon State University, Department of Forest Science.

Oberts G. 1986. Pollutants associated with sand and silt applied to roads in Minnesota. *Water Resources Bulletin* 22:479-483.

Odum E. 1971. *Fundamentals of ecology*. Philadelphia (PA): WB Saunders Co. 574 p.

Ollerton J, Winfree R, Tarrant S. 2011. How many flowering plants are pollinated by animals? *Oikos* 120:321-326

Onstad C, Wolfe M, Larson C, Slack D. 1984. Tilled soil subsidence during repeated wetting. *Transactions of the American Society of Agricultural Engineers* 27:733-736.

Oregon Department of Transportation. 2013. Oregon DOT Integrated Vegetation Management Statewide Plan. URL:  
[https://www.oregon.gov/ODOT/Maintenance/Documents/statewide\\_IVM\\_plan.pdf](https://www.oregon.gov/ODOT/Maintenance/Documents/statewide_IVM_plan.pdf) (accessed 5 October 2018). 12p.

Ottewell K, Donnellan S, Lowe A, Paton D. 2009. Predicting reproductive success of insect-versus bird-pollinated scattered trees in agricultural landscapes. *Biological Conservation* 142(4):888-898.

Ouin A, Aviron S, Dover J, Burel F. 2004. Complementation/supplementation of resources for butterflies in agricultural landscapes. *Agriculture, Ecosystems & Environment* 103:473-479.

Ozores-Hampton M, Obreza T, Stoffella P, Fitzpatrick G. 2002. Immature compost suppresses weed growth under greenhouse conditions. *Compost Science & Utilization* 10(2):105-113.

Ozores-Hampton M. 1998. Compost as an alternative weed control method. *HortScience* 33(6):938-940.

Page H, Bork E. 2005. Effect of planting season, bunchgrass species, and neighbor control on the success of transplants for grassland restoration. *Restoration Ecology* 13(4):651–658.

- Page I. 1977. Improving the efficiency of ripping. Rotorua (New Zealand): Forest Industries Review. Forest Research Institute. p. 7- 13.
- Palmer J. 1990. Nutrient cycling: the key to reclamation success. In: Chambers J, Wade G, editors. Evaluating reclamation success: the ecological consideration—proceedings of a symposium; 1990 Apr 23-26; Charleston, WV. Radnor (PA): USDA Forest Service, Northeastern Forest Experiment Station. General Technical Report NE-164. p. 27-36.
- Palumbo A, McCarthy J, Amonette J, Fisher L, Wullschleger S, Daniels W. 2004. Prospects for enhancing carbon sequestration and reclamation of degraded lands with fossil-fuel combustion by-products. *Advances in Environmental Research*. 8:425-438.
- Parent V, Koenig R. 2003. Solutions to soil problems. I. High salinity (soluble salts). Logan (UT): Utah State University Extension. AG/Soils/2003-01. 2 p.
- Parmesan C. 2006. Ecological and evolutionary responses to recent climate change. *Annual Review of Ecology, Evolution, and Systematics*. Vol 37:637-669.
- Parmesan C. 2007. Influences of species, latitudes and methodologies on estimates of phenological response to global warming. *Global Change Biology*. 13:1860-1872.
- Parmesan C, Yohe G. 2003. A globally coherent fingerprint of climate change impacts across natural systems. *Nature*. Vol 421:37-42.
- Parr T, Way J. 1988. Management of roadside vegetation: the long-term effects of cutting. *Journal of Applied Ecology* 25:1073-1087.
- Parsons Brinckerhoff, 2014. Appendix U Illiana Corridor Sustainability and Context Sensitive Solutions Technical Report, Illinois Department of Transportation and Indiana Department of Transportation, June 2014, pp.44. URL: [http://nepa.illianacorridor.org/pdfs/tier2\\_feis/appendix\\_u.pdf](http://nepa.illianacorridor.org/pdfs/tier2_feis/appendix_u.pdf) (accessed 14 Mar 2016).
- Pellett N, Heleba D. 1995. Chopped newspaper for weed control in nursery crops. *Environmental Horticulture* 13(2):77-81.
- Penny G, Neal J. 2003. Light, temperature, seed burial, and mulch effects on mulberry weed (*Fatoua villosa*) seed germination. *Weed Technology* 17(2):213-218.
- Perugini M, Manera M, Grotta L, Cesarina Abete M, Tarasco R, Amorena M. 2011. Heavy metal (Hg, Cr, Cd, and Pb) contamination in urban areas and wildlife reserves: honeybees as bioindicators. *Biological Trace Element Research* 140(2):170–176.
- Peterson, D. 2002. Hydrophilic polymers - effects and uses in the landscape. *Soviet Soil Science* 13(4): 111-115.
- Pezeshki S, Brown C, Elcan J, Shields Jr F. 2005. Responses of non-dormant black willow (*Salix nigra*) cuttings to pre-planting soaking and soil moisture. *Restoration Ecology* 13:1-7.

Pezeshki S, Shields F. 2006. Black willow cutting survival in streambank plantings, southeastern United States. *Journal of the American Water Resources Association* 42(1):191-200.

Pill W, Frett J, Williams I. 1997. Matric priming of Kentucky bluegrass and tall fescue seeds benefits seedling emergence. *HortSci* 32:1061-1063.

Pill W, Nesnow D. 1999. Germination of hydro-seeded Kentucky bluegrass (*Poa pratensis L.*) and perennial ryegrass (*Lolium perenne L.*) in response to seed agitation in the tank. *Journal of Turfgrass Management* 3(2):59-67.

Poff R. 1996. Effects of silvicultural practices and wildfire on productivity of forest soils. In: *Sierra Nevada Ecosystem Project: Final report to Congress. Vol II. Assessments and scientific basis for management options*. Davis (CA): University of California, Centers for Water and Wildland Resources. p. 477-493.

Polster D. 1997. Restoration of landslides and unstable slopes: considerations for bioengineering in interior locations. In: *Proceedings of the 21st Annual British Columbia Mine reclamation symposium; 1997 Sept 22-25; Cranbrook, BC. Victoria (BC): British Columbia Technical Research Committee on Reclamation*. p. 153-166.

Ponisio L, M'Gonigle L, Kremen C. 2015. On-farm habitat restoration counters biotic homogenization in intensively managed agriculture. *Global Change Biology* 22(2): 704-715.

Potts S, Biesmeijer J, Kremen C, Neumann P, Schweiger O, Kunin W. 2010. Global pollinator declines: trends, impacts and drivers. *Trends in Ecology and Evolution*. 25(6): 345-353.

Potts S, Vulliamy B, Dafni A, Ne'eman G, Willmer P. 2003. Linking bees and flowers: how do floral communities structure pollinator communities? *Ecology* 84:2628–2642.

Potts S, Vulliamy B, Roberts S, O'Toole C, Dafni A, Ne'eman G, Willmer P. 2005. Role of nesting resources in organizing diverse bee communities in a Mediterranean landscape. *Ecological Entomology* 30(1):78–85.

Powell D, Urban C, Johnson C. 1998. Personal communication. Baker City (OR): USDA Forest Service. Forest Silviculturist, Forest Botanist, Area Ecologist.

Preadjati A, Smits W, Tolkamp G. 2001. Vegetative propagation to assure a continuous supply of plant material for forest rehabilitation. In: Hillegers PJM, de Jongh HH. *The balance between biodiversity conservation and sustainable use of tropical rain forests: workshop proceedings; 1999 Dec 6-8; Balikpapan, Kalimantan, Indonesia*. p. 19-30.

Proudfoot J, Ponder D, Tindall D, Wysocki D. 2015. Guidebook for Designing and Managing Rights-of-Way for Carbon Sequestration and Biomass Generation. National Cooperative Highway Research Program. Report 804. 55p.

Quarles III H, Hanawalt R, Odum W. 1974. Lead in small mammals, plants, and soil at varying distances from a highway. *Journal of Applied Ecology*: 937-949.

- Racey G, Raitanen E. 1983. Seedling development as affected by fibre and straw mulching at midhurst Nursery. Ottawa (Ontario): Ontario Ministry of Natural Resources, Forest Resources Branch. Nursery Notes 94. 5 p.
- Rauch N. 2006. Personal communication. Bend (OR): USDA Forest Service Bend Seed Extractory, Seed Extractory Assistant Manager.
- Rauzi F, Tresler R. 1978. A preliminary report on herbage yields, stand evaluation, soils, and chemical content of selected grasses and a legume grown on topsoil, White River and Wind River geologic materials. Laramie (WY): University of Wyoming, Wyoming Agricultural Experiment Station. Research Journal 124.
- Reeder J, Sabey B. 1987. Nitrogen. In: Williams R, Schuman G, editors. Reclaiming mine soils and overburden in the Western United States. Ankeny (IA): Soil and Water Conservation Society. p. 155-184.
- Rehfeldt G. 1994. Evolutionary genetics, the biological species, and the ecology of the Interior cedar-hemlock forests. Pages 93–100 In: Proceedings of the interior cedar-hemlock-white pine forests: ecology and management. 1993 March 2–4; Spokane, WA. Department of Natural Resource Sciences, Washington State University, Pullman, Washington, USA.
- Resource Efficient Agricultural Production Canada [REAP Canada]. 1991. Effects of carbon addition on nitrogen fixation and P nutrition of hairy vetch and following crops. In: Abboud A, Patriquin D. Processes involved in mobilization of P in different farming systems in Southwestern Ontario: nutrient levels in plant tissues and soils. Harrow (ON): Agriculture Canada Ecological Services for Planning. Report to Canada-Ontario SWEEP (Soil and Water Enhancement Program), SWEEP Report #23 from REAP-Canada. p. 22-24.
- Re-Sourcing Associates, Inc, CPM Consultants Inc. 1997. Wood waste recovery: size reduction technology study. EPA Report No. CDL-97-3. 66 p. URL: <http://www.p2pays.org/ref/13/12639.pdf> (accessed 21 June 2007).
- Reuter J, Jassby A, Marjanovic P, Heyvaert A, Goldman C. 1998. Preliminary phosphorus and nitrogen budgets for Lake Tahoe. In: Annual Progress Report 1998, Lake Clarity and Watershed Modelling, Presidential Deliverable for Lake Tahoe, USEPA Water and Watershed Grant; 1998 July 18; Tahoe Research Group, Department of Civil and Environmental Engineering. Davis (CA): University of California, John Muir Institute for the Environment.
- Robichaud P, Ashmun L, Foltz R, Showers C, Groenier J, Kesler J, DeLeo C, Moore M. 2013. Production and aerial application of wood shreds as a post-fire hillslope erosion mitigation treatment. Gen. Tech. Rep. RMRS-GTR-307. Fort Collins, CO: U.S. Department of Agriculture, Forest Service, Rocky Mountain Research Station. 31 p.
- Rice R. 1982. Sedimentation in the chaparral: How do you handle unusual events? In: Sediment Budgets and Routing in Forested Drainage Basins, edited by F. J. Swanson and others. Forest Service, U.S. Dep. Agric., Washington, D. C. pp 39-49.

- Ries L, Debinski D, Wieland M. 2001. Conservation value of roadside prairie restoration to butterfly communities. *Conservation Biology* 15:401–411.
- Riley L, Steinfeld D, Riley S. 2013. Effects of polyacrylamide crystals on seedling establishment and growth of native species when applied to soils of highly disturbed sites. Internal report to WFHLD. 24 p.
- Riley S. 1999. Proposal to cage shrubs [Internal memo Umatilla National Forest]. Located at: Umatilla National Forest, Pendleton, OR. 10 p.
- Ritchie G. 1985. Root growth potential: principles, procedures and predictive ability. In: Duryea M, editor. *Proceedings - evaluating seedling quality: principles, procedures and predictive ability of major tests; 1984 Oct 16-18; Corvallis, OR*. Corvallis (OR): Oregon State University, Forest Research Laboratory. p. 93-104.
- Robidoux P, Delisle C. 2001. Ecotoxicological evaluation of three deicers (NaCl, NaFo, CMA)-effect on terrestrial organisms. *Ecotoxicology and Environmental Safety*. 48(2), 128-139.
- Rose R, Haase D, Boyer D. 1995. Organic matter management in forest nurseries: theory and practice. Corvallis (OR): Oregon State University, Nursery Technology Cooperative. 65 p.
- Rose R. 2002. Slow release fertilizers 101. In: Dumroese R, Riley L, Landis T, technical coordinators. National proceedings: forest and conservation nursery associations—1999, 2000, and 2001. Ogden (UT): USDA Forest Service, Rocky Mountain Research Station. Proceedings RMRS-P-24. p. 304-308.
- Rose R. 2004. Comparison among soil and tissue testing laboratories. In: *Nursery Technology Cooperative, 2003-2004 Annual Report*. Corvallis (OR): Oregon State University, Department of Forest Science. p. 39-47.
- Ross D, Cairns A. 1981. Nitrogen availability and microbial biomass in stockpiled topsoils in Southland. *New Zealand Journal of Science* 24:137.
- Rossi P. 1999. Length of cuttings in establishment and production of short-rotation plantations of *Salix aquatica*. *New Forests* 18:161-177.
- Roulston T, Goodell K. 2011. The role of resources and risks in regulating wild bee populations. *Annual Review of Entomology* 56: 293-312.
- Russell C, Schultz C. 2010. Investigating the use of herbicides to control invasive grasses: effects on at-risk butterflies. *Journal of Insect Conservation* 14(1):53-63.
- Saarinen K, Valtonen A, Jantunen J, Saarnio S. 2005. Butterflies and diurnal moths along road verges: Does road type affect diversity and abundance? *Biological Conservation* 123:403-412.
- Salt Institute. 2007. The Snowfighters Handbook. The Salt Institute, Alexandria, Virginia. 28 p. URL: <http://saltinstitute.org/wp-content/uploads/2015/02/SASS-Handbook-2016-1.pdf> (accessed 5 October 2018).

- Samways M, Caldwell P, Osborn R. 1996. Ground-living invertebrate assemblages in native, planted and invasive vegetation in South Africa. *Agriculture, Ecosystems & Environment* 59:19–32.
- Sardinas H, Kremen C. 2014. Evaluating nesting microhabitat for ground-nesting bees using emergence traps. *Basic and Applied Ecology*: 15(2): 61-168.
- Savolainen O, Pyhäärvi T, Knürr T. 2007. Gene flow and local adaptation in trees. *Annual Review of Ecology, Evolution, and Systematics* 38:595-619.
- Schaff S, Pezeshki S, Shields F. 2002. Effects of pre-planting soaking on growth and survival of black willow cuttings. *Restoration Ecology* 10(2):267-274.
- Saunders D, Hobbs R, Margules C. 1991. Biological consequences of ecosystem fragmentation – a review. *Conservation Biology* 5:18–32.
- Schaffers A, Raemakers I, Sykora K. 2012. Successful overwintering of arthropods in roadside verges. *Journal of Insect Conservation* 16(4):511–522.
- Schmidt L. 2000. Guide to handling of tropical and subtropical forest seed. Humlebaek (Denmark): Danida Forest Seed Centre.
- Schooler S, Cook T, Prichard P, Yeates A. 2010. Disturbance-mediated Competition: The Interacting Roles of Inundation Regime and Mechanical and Herbicidal Control in Determining Native and Invasive Plant Abundance. *Biological Invasions*, Vol. 12. p 3289–3298.
- Schröder R, Schröder R., Prasse, R. 2013. From nursery into nature: a study on performance of cultivated varieties of native plants used in re-vegetation, their wild relatives and evolving wild × cultivar hybrids. *Ecological Engineering*. 60:428-437.
- Scoggins H, Nelson P, Bailey D. 2000. Development of the press extraction method for plug substrate analysis: effects of variable extraction force on pH, electrical conductivity, and nutrient analysis. *HortTechnology* 10(2):367-36.
- Society for Ecological Restoration [SER]. 2004. The SER international primer on ecological restoration. Tuscon (AZ): Society for Ecological Restoration, International Science and Policy Working Group. URL: <http://www.ser.org> (accessed 20 Aug 2006).
- Settele J, Bishop J, Potts S. 2016. Climate change impacts on pollination. *Nature Plants*. Vol 2:1-3.
- Shaw N. 2004. Production and use of planting stock. In: Monsen S, Stevens R, Shaw N. Restoring western ranges and wildlands. Fort Collins (CO): USDA Forest Service, Rocky Mountain Research Station. General Technical Report RMRS-GTR-136 - volumes 1-3. p. 745-768.
- Sheley R. 2005. Personal communication. Madras (OR): USDA Forest Service, Crooked River National Grassland.
- Simberloff D, Stiling P. 1996. How risky is biological control? *Ecology* 77(7):1965-1974.

Skórka P, Lenda M, Moroń D, Kalarus K, and Tryjanowski P. 2013. Factors affecting road mortality and the suitability of road verges for butterflies. *Biological Conservation* 159:148-157.

Slick B, Curtis W. 1985. A guide for the use of organic materials as mulches in reclamation of coal minesoils in the Eastern United States. Broomall (PA): USDA Forest Service, Northeastern Forest Experimental Station. General Technical Report NE-98. 144 p.

Sloan R. 2001. Modified brush layers and live pole drains for landslide reclamation [online]. Santa Barbara (CA): Erosion Control. URL: [http://www.forester.net/ec\\_0107\\_biotech.html](http://www.forester.net/ec_0107_biotech.html) (accessed 1 Jan 2007).

Smallidge P, Leopold D. 1997. Vegetation management for the maintenance and conservation of butterfly habitats in temperate human-dominated habitats. *Landscape and Urban Planning* 38:259–280.

Smith M, Charvat I, Jacobson R. 1998. Arbuscular mycorrhizae promote establishment of prairie species in a tallgrass prairie restoration. *Canadian Journal of Botany* 76(11):1947-1954.

Snell-Rood E, Espeset A, Boser C, White W, Smykalski R. 2014. Anthropogenic changes in sodium affect neural and muscle development in butterflies. *Proceedings of the National Academy of Sciences*, 111(28): 10221-10226.

Sobek A, Skousen J, Fisher Jr S. 2000. Chemical and physical properties of overburdens and minesoils. In: Barnhisel RI, Darmody RG, Daniels WL, editors. *Reclamation of drastically disturbed lands*. Madison (WI): American Society of Agronomy. Agronomy Monograph 41. p. 77-104.

Soderstrom B, Hedblom M. 2007. Comparing movement of four butterfly species in experimental grassland strips. *Journal of Insect Conservation* 11(4):333–342.

Sotir R, Gray D. 1992. Soil bioengineering for upland slope protection and erosion reduction. In: *Engineering Field Handbook*. Washington (DC): USDA Natural Resources Conservation Service. EFH-210. 61 p.

South D, Mexal J. 1984. Growing the “best” seedling for reforestation success. Auburn (AL): Auburn University. Forestry Department Series 12. 11 p.

South Dakota Department of Agriculture. 2006. Windbreaks and snow management. 2p. URL: <https://sdda.sd.gov/legacydocs/Forestry/publications/PDF/Windbreaks%20and%20Snow%20Management.pdf> (accessed 5 October 2018).

Spangler M, Handy R. 1973. *Soil engineering*. 3rd edition. New York (NY): Intext Educational Publishers. 748 p.

Spellerberg I. 1998. Ecological Effects of Roads and Traffic: A Literature Review. *Global Ecology and Biogeography Letters* 7(5):317-333.

Spira T. 2001. Plant-pollinator interactions: a threatened mutualism with implications for the ecology and management of rare plants. *Natural Areas Journal* 21(1):78–88.

- Spokas K, Cantrell K, Novak J, Archer D, Ippolito J, Collins H, Akwasi A, Boateng A, Lima I, Lamb M, McAloon A, Lentz R, Nichols K. 2012. Journal of Environmental Quality. P 973-989.
- St Clair B, Johnson R. 2003. Structure of genetic variation and implications for the management of seed and planting stock. In: Riley L, Dumroese R, Landis T, technical coordinators. National proceedings: Forest and Conservation Nursery Associations–2003; 2003 June 9–12; Coeur d’Alene, ID; and 2003 July 14–17; Springfield, IL. Fort Collins (CO): USDA Forest Service, Rocky Mountain Research Station. Proceedings RMRS-P-33. p. 64-71.
- St Clair J, Kilkenny F, Johsnon R, Shaw N, Weaver G. 2013. Genetic variation in adaptive traits and seed transfer zones for *Pseudoroegneria spicata* (bluebunch wheatgrass) in the northwestern United States. Evolutionary Applications 6:933-938.
- St Clair J, Mandel N, Vance-Borland K. 2005. Genecology of Douglas fir in western Oregon and Washington. Annals of Botany 96:1199-1214.
- St John L, Ogle D, Scianna J, Winslow S, Holzworth L. 2003. Plant materials collection guide. Boise (ID): USDA Natural Resources Conservation Service. Plant Materials Technical Note No 1. 12 p.
- St John T. 1995. Specially-modified land imprinter inoculates soil with mycorrhizal fungi (California). Restoration and Management Notes 14(1):84.
- St John T. 1999. Nitrate immobilization of the mycorrhizal network for control of exotic ruderals. CalEPPC News 7(1):4-11.
- Saint-Amour, M. 1998. Evaluation of a powered auger for planting large container seedlings. Pointe-Claire (Quebec): Forest Engineering Research Institute of Canada. Field Note No. Silviculture-107. 2 p.
- State of Illinois General Assembly-a, 2013. “Statute 605 ILCS 5/ Illinois Highway Code, Article 4 Division 2 State Highways, Section 4-219 Context Sensitivity,” URL: <http://www.ilga.gov/legislation/ilcs/ilcs.asp> (accessed 5 October 2018) (Springfield, IL: Legislative Information System, 2013).
- Steed J, DeWald L. 2003. Transplanting sedges (*Carex* spp.) in southwestern riparian meadows. Restoration Ecology 11(2):247-256.
- Steinbrenner E. 1981. Forest soil productivity relationships. In: Heilman PE, Anderson HW, Baumgartner DM, editors. Forest soils of the Douglas-fir region. Pullman (WA): Washington State University Cooperative Extension Service. p. 199-229.
- Steinfeld D, Amaranthus M, Cazares E. 2003. Survival of ponderosa pine (*Pinus ponderosa* Doug. ex Laws.) seedlings outplanted with Rhizopogon mycorrhizae inoculated with spores at the nursery [online]. Journal of Arboriculture 29(4). URL: <http://joa.isa-arbor.com/request.asp?JournalID=1&ArticleID=94&Type=1> (accessed 3 Apr 2007).
- Steinfeld D. 2002. Blaine Road project: findings - willow seedlings and cuttings trials [Internal memo]. Located at: Ashland Ranger District, Rogue River National Forest, Ashland, OR. 7 p.

Steinfeld D. 2004. Soil temperatures under mulch and no mulch on the Chiloquin Highway revegetation project [personal files]. Located at: USDA Forest Service, Ashland Ranger District, Rogue River National Forest, Ashland, OR.

Steinfeld D. 2005. Summary of tree shelter temperature recordings in and outside of tree shelters at the Canyonville planting pocket site [personal files]. Located at: USDA Forest Service, Ashland Ranger District, Rogue River National Forest, Ashland, OR.

Steinfeld D, Landis T. 1990. Soil compaction. In: Hamm P, Campbell S, Hansen E, editors. Growing healthy seedlings. Corvallis (OR): Oregon State University, Forest Research Laboratory. p. 87-91.

Stevens R, Monsen S. 2004. Mechanical plant control. In: Monsen S, Stevens R, Shaw N, compilers. Restoring western ranges and wildlands. Fort Collins (CO): USDA Forest Service, Rocky Mountain Research Station. General Technical Report RMRS-GTR-136 - vol 1. p. 65-87.

Stevens R, Van Epps G. 1984. Seeding techniques to improve establishment of forage kochia (*Kochia prostrata* [L.] Schrad.) and fourwing saltbush (*Atriplex canescens* [Pursh.] Nutt.). In: Tiedemann A, McArthur E, Stutz H, Stevens R, Johnson K, compilers. Proceedings- symposium on the biology of Atriplex and related chenopods; 1983 May 2-6; Provo, UT. Ogden (UT): USDA Forest Service, Intermountain Forest and Range Experiment Station. General Technical Report INT-172. p. 269-272.

Stoner K, Joern A. 2004. Landscape vs. local habitat scale influences to insect communities from tallgrass prairie remnants. *Ecological Applications* 14:1306–1320.

Strauss S, Lau J, Carroll S. 2006. Evolutionary responses of natives to introduced species: what do introductions tell us about natural communities? *Ecology Letters* 9: 357-374.

Stryker J. 2001. Irrigation tutorials. URL: <http://www.irrigationtutorials.com/> (accessed 11 Feb 2007).

Sucoff E. 1975. Effects of deicing salts on woody vegetation along Minnesota roads. Minn. Agr. Expt. Sta. Tech. Bul. 303.

Sugden E. 1985. Pollinators of *Astragalus monoensis* Barneby (Fabaceae): new host records; potential impact of sheep grazing. *Great Basin Naturalist* 45:299–312.

Sullivan D, Cogger C, Bary A. 2007. Fertilizing with Biosolids. PNW 508-E. 18p. URL: [https://catalog.extension.oregonstate.edu/sites/catalog/files/project/pdf/pnw508\\_0.pdf](https://catalog.extension.oregonstate.edu/sites/catalog/files/project/pdf/pnw508_0.pdf) (accessed 5 October 2018).

Svensson B, Lagerlöf J, Svensson B. 2000. Habitat preferences of nest-seeking bumble bees (Hymenoptera: Apidae) in an agricultural landscape. *Agriculture, Ecosystems & Environment* 77(3):247-255.

Svhra P, Burger D, Harris R. 1993. Treeshelters for nursery plants may increase growth and be cost effective. *California Agriculture* 47(4):13-16.

Swaileh K, Hussein R, Abu-Elhaj S. 2004. Assessment of heavy metal contamination in roadside surface soil and vegetation from the West Bank. *Archives of Environmental Contamination and Toxicology* 47(1):23–30.

Swistock B, Mecum K, Sharpe W. 1999. Summer temperatures inside ventilated and unventilated brown plastic treeshelters in Pennsylvania. *Northern Journal of Applied Forestry* 16(1):7-10.

Tallamy D, Shropshire K. 2009. Ranking lepidopteran use of native versus introduced plants. *Conservation Biology* 23(4):941–947.

Tanaka Y. 1984. Assuring seed quality for seedling production: cone collection and seed processing, testing, storage, and stratification. In: Duryea M, Landis T, editors. *Forest nursery manual: production of bareroot seedlings*. Boston (MA): Martinus Nijhoff/Dr W Junk Publishers. p. 27-40.

Teidemann A, Lopez C. 2004. Assessing soil factors in wildland improvement programs. In: Monsen S, Stevens R, Shaw N, compilers. *Restoring western ranges and wildlands*. Fort Collins (CO): USDA Forest Service, Rocky Mountain Research Station. General Technical Report RMRS-GTR-136 - vol 1. p. 39-56.

Tesch S, Helms J. 1992. Regeneration methods. In: Hobbs SD, Tesch S, Owston P, Stewart R, Tappener II J, Wells G, editors. *Reforestation practices in southwestern Oregon and northern California*. Corvallis (OR): Oregon State University, Forest Research Laboratory. P. 166-201.

Tewksbury J, Levey D, Haddad N, Sargent S, Orrock J, Weldon A, Danielson B, Brinkerhoff J, Damschen E, Townsend P. 2002. Corridors affect plants, animals, and their interactions in fragmented landscapes. *Proceedings of the National Academy of Sciences* 99(20): 12923-12926.

Therrell L, Cole D, Claassen V, Ryan C, Davies M. 2006. *Wilderness and backcountry site restoration guide*. Missoula (MT): USDA Forest Service, Missoula Technology and Development Center. Pub # 0623 2815. 394 p.

Thomas G. 1967. Problems encountered in soil testing methods. In: *Soil testing and plant analysis. Part 1*. Madison (WI): Soil Science Society of America. SSSA Special Publication Series No. 2. p. 37-54.

Thorup R. 1984. *Ortho agronomy handbook - a practical guide to soil fertility and fertilizer use*. San Francisco (CA): Chevron Chemical Company. 454 p.

Tisdale S, Nelson W. 1975. *Soil fertility and fertilizers*. New York (NY): Macmillan Publishing Co. 694 p.

Trahan N, Peterson C. 2008. Impacts of magnesium chloride-based deicers on roadside vegetation. In: In: Transportation Research Board (Ed.), Proc. 6th Intl. Symposium on Snow Removal and Ice Control Technology. *Transportation Research Circular E-C126. SNOW08-050*, 171-186.

Trappe J, Bollen W. 1981. Forest soil biology. In: Heilman P, Anderson H, Baumgartner D, editors. *Forest soils of the Douglas-fir region*. Pullman (WA): Washington State University Cooperative Extension Service. p. 145-155.

Trent A, Nolte D, Wagner K. 2001. Comparison of commercial deer repellents. Missoula (MT): USDA Forest Service, Missoula Technology and Development Center. Technology Tip 0124- 2331-MTDC. 6 p.

Trombulak S, Frissell C. 2000. Review of ecological effects of roads on terrestrial and aquatic communities. *Conservation Biology* 14(1):18-30.

Trotti J. 2000. Hydraulic seeding and stabilization 101 [online]. *Erosion control* 7(8).10 p. URL: <https://foresternetwork.com/weekly/erosion-control-weekly/soil/hydraulic-seeding-and-stabilization-101/> (accessed 27 December 2018).

Tuley G. 1985. The growth of young oak trees in shelters. *Forestry* 58:181-195.

Tuttle R, Wenberg R, Sotir R. 1996. Chapter 16. Streambank and shoreline protection. In: *Engineering Field Handbook*. Washington (DC): USDA Natural Resources Conservation Service. EFH- 210, Part 650. 88 p.

Tyser RW, Worley C. 1992. Alien flora in grasslands adjacent to road and trail corridors in Glacier National Park, Montana (USA). *Conservation Biology* 6(2):253–262.

Ungar I. 1992. The effect of salinity and temperature on seed germination and growth of *Hordeum jubatum*. *Can. J. Bot.* 52:1357-1362.

University of Minnesota Extension Service [UMES]. 2004. Soluble salts. St Paul (MN): University of Minnesota Extension Service. BU-01731. 3 p.

University of Nevada. 2009. Effects of de-icing salts on vegetation in the Lake Tahoe Basin. URL: <http://portal.nifa.usda.gov/web/crisprojectpages/0205723-effects-of-de-icing-salts-on-vegetation-in-the-lake-tahoe-basin.html> (accessed 5 October 2018).

USDA Agricultural Research Service [ARS]. 2012. Plant hardiness zone map. URL: <https://www.ars.usda.gov/news-events/news/research-news/2012/usda-unveils-new-plant-hardiness-zone-map/> (accessed 5 October 2018).

USDA Forest Service [USFS]. 1991. J Herbert Stone Nursery Annual Report. Central Point (OR): USDA Forest Service, J Herbert Stone Nursery.

USDA Forest Service [USFS]. 2003. BAER Manual-FSM 2523. In: USDA Forest Service. Forest Service manual FSM 2500—watershed and air management. Washington (DC): USDA Forest Service National Headquarters. 44 p.

USDA Forest Service [USFS]. 2011. National strategy and implementation plan for invasive species management. URL: <https://pubs.acs.org/doi/abs/10.1021/bk-2011-1073.ch008> (accessed 27 December 2018).

USDA Forest Service [USFS]. 2008. FSM 2070—Vegetation Ecology. URL [https://www.fs.fed.us/wildflowers/Native\\_Plant\\_Materials/documents/FSM\\_2070.pdf](https://www.fs.fed.us/wildflowers/Native_Plant_Materials/documents/FSM_2070.pdf) (accessed 27 December 2018).

- USDA Forest Service [USFS]. 2006. Interior Columbia Basin ecosystem management project. URL: <https://www.fs.fed.us/r6/icbemp/> <http://www.icbemp.gov/> (accessed 27 December 2018).
- USDA Forest Service [USFS]. 2007. Invasive plant prevention. In: Center for Invasive Plant Management. Invasive plant management: CIPM online textbook. Bozeman (MT): Montana State University. URL: [http://www.weedcenter.org/textbook/8\\_prevention.html](http://www.weedcenter.org/textbook/8_prevention.html) (accessed 3 Apr 2007).
- USDA Natural Resources Conservation Service [NRCS]. 1994. Planning and design manual for the control of erosion, sediment, and stormwater. 1st edition [online]. Mississippi State (MS): Mississippi State University, Agricultural and Biological Engineering. URL: <http://www.abe.msstate.edu/csd/pdm/index.php> (accessed 3 Apr 2007).
- USDA Natural Resources Conservation Service [NRCS]. 1997. Collecting plant materials. Brooksville (FL): USDA Natural Resources Conservation Service, Brooksville Plant Materials Center. Technical Note No. 35. 4 p.
- USDA Natural Resources Conservation Service [NRCS]. 2018. Soil Textural Triangle. URL: [https://www.nrcs.usda.gov/wps/portal/nrcs/detail/soils/edu/?cid=nrcs142p2\\_054311](https://www.nrcs.usda.gov/wps/portal/nrcs/detail/soils/edu/?cid=nrcs142p2_054311) (accessed 26 December 2018)
- Valtonen A, Saarinen K. 2005. A highway intersection as an alternative habitat for a meadow butterfly: effect of mowing, habitat geometry and roads on the ringlet (*Aphantopus hyperantus*). *Annales Zoologici Fennici* 42(5):545-556.
- Valtonen A, Saarinen K, Jantunen J. 2006. Effect of different mowing regimes on butterflies and diurnal moths on road verges. *Animal Biodiversity and Conservation* 29:133–148.
- Varnes D. 1978. Slope movement types and processes. In: Schuster R, Krize R, editors. Landslides-analysis and control. Washington (DC): National Academy of Sciences, Transportation Research Board. Special Report 176. p. 12-33.
- Visser S, Fujikawa J, Griffiths C, Parkinson D. 1984. Effect of topsoil storage on microbial activity, primary production and decomposition potential. *Plant and Soil* 82(1):41-50.
- Visser S, Griffiths C, Parkinson D. 1984. Topsoil storage effects on primary production and rates of vesicular-arbuscular mycorrhizal development in *Agropyron trachycaulum*. *Plant and Soil* 82(1):51-60.
- Von der Lippe M, Kowarik I. 2007. Long-Distance Dispersal of Plants by Vehicles as a Driver of Plant Invasions. *Conservation Biology* 21(4):986–996.
- Walker R. 2002. Responses of Jeffrey pine on a surface mine site to fertilizer and lime. *Restoration Ecology* 10(2):204-212.

Walker R. 2005. Growth and nutrition of Jeffrey pine seedlings on a Sierra Nevada surface mine in response to fertilization three years after planting. *Western Journal of Applied Forestry* 20(1):28-35.

Ward K, Cariveau D, May E, Roswell M, Vaughan M, Williams N, Winfree R, Isaacs R, Gill K. 2014. Streamlined bee monitoring protocol for assessing pollinator habitat. 16 pp. The Xerces Society for Invertebrate Conservation. URL: [http://www.xerces.org/wp-content/uploads/2014/09/StreamlinedBeeMonitoring\\_web.pdf](http://www.xerces.org/wp-content/uploads/2014/09/StreamlinedBeeMonitoring_web.pdf) (accessed 5 October 2018).

Ward K, Cariveau D, May E, Roswell M, Vaughan M, Williams N, Winfree R, Isaacs R, and Gill K. 2014. Streamlined Bee Monitoring Protocol for Assessing Pollinator Habitat. Portland, OR: The Xerces Society for Invertebrate Conservation. 16 pp.

Waring RH, Schlesinger W. 1985. Forest ecosystems: concepts and management. Orlando (FL): Academic Press. 340 p.

Warren M. 1993. A review of butterfly conservation in central southern Britain: II. Site management and habitat selection of key species. *Biological Conservation* 64(1):37–49.

Wearstler Jr K. 2004. Considerations for collecting and vegetatively propagating poplar woody plant materials. In: Riley L, Dumroese R, Landis T, technical coordinators. National proceedings: forest and conservation nursery associations—2003. Fort Collins (CO): USDA Forest Service, Rocky Mountain Research Station. Proceedings RMRS-P-33. p. 55-56.

Wedin D, Tilman D. 1996. Influence of nitrogen loading and species composition on the carbon balance of grasslands. *Science* 274:1720-1723.

Welker J, Gordon D, Rice K. 1991. Capture and allocation of nitrogen by *Quercus douglasii* seedlings in competition with annual and perennial grasses. *Oecologia* 87:459-466.

Wert S, Thoms B. 1981. Effects of skid roads on diameter, height, and volume growth in Douglas-fir. *Soil Science Society of America Journal* 45(3):629-632.

White A, Fant J, Kramer A. In Prep. Developing genetically appropriate seed mixes of vulnerable plant species for restoration. A component of AB White's thesis for a Master of Science in Plant Biology and Conservation, anticipated from Northwestern University and Chicago Botanic Garden 2016.

Whiting D, Card A, Wilson C. 2007. Estimating soil texture: sandy, loamy or clayey. Fort Collins (CO): Colorado State University Extension, Colorado Master Gardener. URL: <https://studylib.net/doc/8198142/estimating-soil-texture---sandy--loamy-or-clayey%3F> (accessed 27 December 2018).

Wilde S, Corey R, Iyer J, Voigt G. 1979. Soil and plant analysis for tree culture. 5th revised edition. New Dehli (India): Oxford & IBH Publishing Co. 224 p.

Williams D, Jackson L, Smith D. 2007. Effects of frequent mowing on survival and persistence of forbs seeded into a species-poor grassland. *Restoration Ecology* 15:24–33.

- Williams N, Cariveau D, Winfree R, Kremen C. 2011. Bees in disturbed habitats use, but do not prefer, alien plants. *Basic and Applied Ecology* 12(4):332–341.
- Williams N, Crone E, T'ai H, Minckley R, Packer L, Potts S. 2010. Ecological and life-history traits predict bee species responses to environmental disturbances. *Biological Conservation*, 143: 2280-2291.
- Wilson G, Hetrick B, Schwab A. 1991. Reclamation effects on mycorrhizae and productive capacity of flue-gas desulfurization sludge. *Journal of Environmental Quality* 20(4):777-783.
- Winfree R. 2010. The conservation and restoration of wild bees. *Annals of the New York Academy of Sciences*, 1195: 69-197.
- Winfree R, Aguilar R, Vázquez D, LeBuhn G, Aizen M. 2009. A meta-analysis of bees' responses to anthropogenic disturbance. *Ecology* 90:2068–2076.
- Withrow-Robinson B, Johnson R. 2006. Selecting native plant materials for restoration projects: insuring local adaptation and maintaining genetic diversity. Corvallis (OR): Oregon State University Extension Service. .
- Wolf D, Blaser R, Morse R, Neal J. 1984. Hydro-application of seed and wood-fiber slurries to bind straw mulch. *Reclamation and Revegetation Research* 3(2):101-107.
- Woodmansee R, Reeder J, Berg W. 1978. Nitrogen in drastically disturbed lands. In: Youngerg CT, editor. Forest and soil land use. Fort Collins (CO): Colorado State University, Department of Forest and Wood Sciences. p. 376-392.
- Wu Y, Wang C, Zhang X, Zhao B, Jiang L, Chen J, Li B. 2009. Effects of saltmarsh invasion by *Spartina alterniflora*. *Biological Invasions* 11:635–649.
- Yetka L, Galatowitsch S. 1999. Factors affecting revegetation of *Carex lacustris* and *Carex stricta* from rhizomes. *Restoration Ecology* 7:162-171.
- Youngberg C. 1981. Organic matter of forest soils. In: Heilman P, Anderson H, Baumgartner D, editors. Forest soils of the Douglas-fir region. Pullman (WA): Washington State University Cooperative Extension Service. p. 137-144.
- Zoldoske D. 1998. Selecting a drip irrigation system for vineyards. Fresno (CA): Center for Irrigation Technology..
- Zuefle M, Brown W, Tallamy D. 2008. Effects of non-native plants on native insect community of Delaware. *Biological Invasions* 10:1159–1169.



U.S. Department of Transportation  
**Federal Highway Administration**